

Supplementary Tables

Supple. Table. S1 The primer sequences used for qRT-PCR amplification.

Table S1 The primer sequences used for qRT-PCR amplification

Primer usage	Forward primer sequence (5'-3')	Reverse primer sequence (5'-3')
<i>CjBPC1</i> RT-PCR	CCCATACCGGTTTGCTCTTG	ACCACATCCCCATCGGTAAC
<i>CjBPC2</i> RT-PCR	TCCCACCGCCCTCAAAA	TCAGCCCCGGTGGACAT
<i>CjBPC3</i> RT-PCR	GAAGAACGCAGCTGTTGAAGA A	TCCTCTATGGCTGCATTTTTT TC
<i>CjBPC4</i> RT-PCR	TTACTGATGCACCCCCAATGT	TTTAGTCCGCCTAGGCTTTGC
<i>CjBPC5</i> RT-PCR	TGCAGCCGAGGGTCATG	TGGCCCAGTGATCCTTGAG
<i>CjBPC6</i> RT-PCR	ACCCGGTATTGCTGCTGTTC	TCTCTGCCGGCTCAGACATA
<i>GAPDH</i> internal reference	GGGAATCCTTGTTACACTGAG	ACCCCATTCGTTGTCATACC
<i>CjBPC1</i> full-length cloning	AGCACAGTACTGCAATGCCT	AACAGACCAGATGCGATGGG
<i>CjBPC2</i> full-length cloning	GCTTCAGTTTCGGCAGTCCA	AAACAGACCACGAGCACTGG
<i>CjBPC3</i> full-length cloning	ATGGATGACGGTGGGAAAC	CTTGATGGTAATGTAGCGAT TTG
<i>CjBPC4</i> full-length cloning	ATGGATGCCAGTGGGCATC	CACGATATCCCAGTAGTACG CA
<i>CjBPC5</i> full-length cloning	AGATGGATTTTTAGTGGTTTCC CT	CACGATATCCCAGTAGTACG CA
<i>CjBPC6</i> full-length cloning	ATGGGGACACACCACGATAG	TTACTTAATTGTAACAAAC
<i>CjBPC1</i> EXCLONE adaptor	tcagcagtgaagagcATGGCTCAATCT GTAGTTTC	ttagcgtgtgaagagcCCTGATTGTC ACAAACTTG
<i>Transgenic</i> validation PCR <i>Hygromycin</i>	TGACCTATTGCATCTCCCGC	ATTTGTGTACGCCCCGACAGT
<i>CjBPC1-F1/R1</i>	CCCATACCGGTTTGCTCTTG	ACCACATCCCCATCGGTAAC
<i>CjBPC1-F2/R2</i>	CGAGATTGTGCGGTTTCAGA	GGTTTACCCAACTATCCCTCA CAT
<i>ACTIN</i> reference in <i>Arabidopsis</i>	AAGGAGAAGAAGGAGAAGGA ACAG	CAAGGAGTGCAATGACTGTG AGATA

<i>AtSTK</i> RT-PCR	GTTCTGATAGCACCAACACTAG C	ACTCATGCTTCTTGGACCTGA TC
<i>AtABI3</i> RT-PCR	AAGCAAAGCGACGTGGGTAA	GTGTGTCTCAGCTTCTTTTTT TGG
<i>AtFUS3</i> RT-PCR	GCTCACTTGCCGGCACTT	AACCGTCCAAATCTTCCATTC TT
<i>AtLEC2</i> RT-PCR	TGGCAAGAGAGAGGTGGTTTTTC	TCCTGTTGATCCTTGCCATCT
<i>AtSTK</i> ChIP-PCR	CGTCTGCGAAAAACCGAGCT	GGACCAATACCTTCATTGTA CTTTGAA
<i>AtABI3</i> ChIP-PCR	GCCTCCTTACTCACATACAAAC CC	TCATCAGCGTCTCCACCGAG TATT
<i>AtFUS3</i> ChIP-PCR	CGACGTATGATACTCCCGAAGG TGAT	ATCCCTTCCTTGCATTCAAGT GCC
<i>AtLEC2</i> ChIP-PCR	CGCTCGCACTTCACAACAGTCC	TCATCACCGCCGCCATCTGC

Supple. Table. S2 Characteristics of the *BASIC PENTACYSTEINE* family in *Camellia japonica*.

TableS2 Characteristics of the BASIC PENTACYSTEINE family in *Camellia japonica*

Gene name	GeneBank ID	Length (aa)	Molecular weight (KD)	PI	Subcellular location
<i>CjBPC1</i>	OL631154	289	32.0	9.40	Nuclear
<i>CjBPC2</i>	OL631155	298	32.6	9.67	Nuclear
<i>CjBPC3</i>	OL888113	312	34.9	9.03	Nuclear
<i>CjBPC4</i>	OL888114	332	37.1	9.53	Nuclear
<i>CjBPC5</i>	OL888115	305	34.1	8.81	Nuclear
<i>CjBPC6</i>	OL888116	232	25.1	9.68	Nuclear

Supplementary Figures

Figure. S1: the details of conserved motifs in BPC proteins. The upper panel indicates the distribution of motifs that are presented by colored boxes. The lower panel lists the detailed conserved amino acids of each motif.

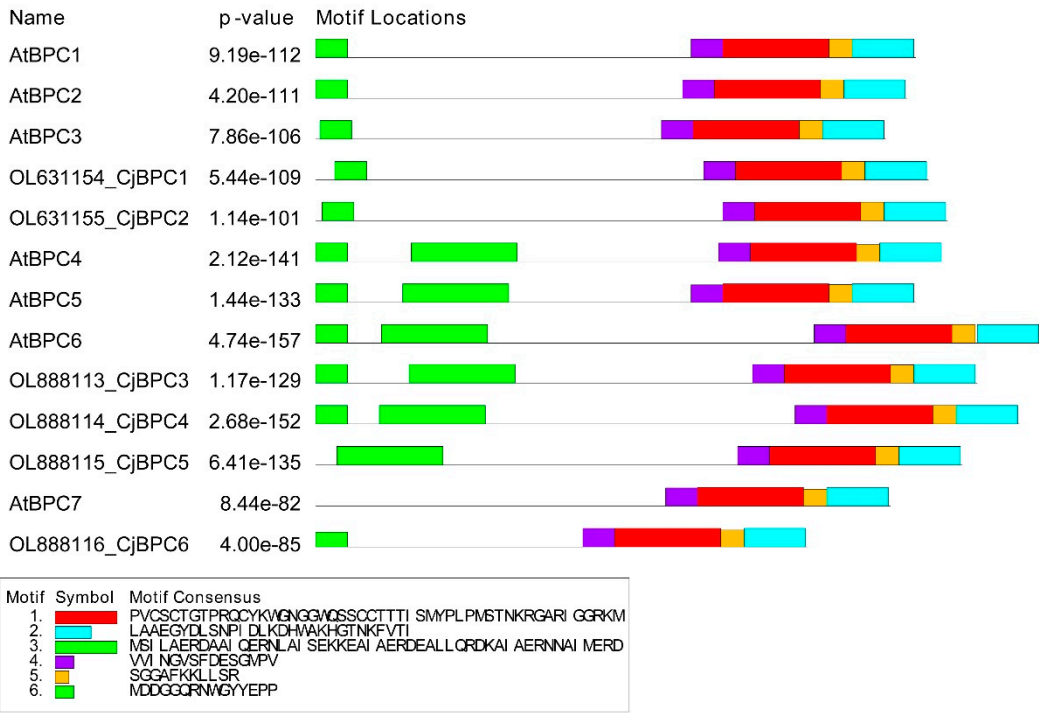


Figure. S2 Expression profiles of CjBPC genes in different tissues of *Camellia japonica*.

The expression *CjBPCs* in differe floral tissues of *Camellia japonica*. The expression is measured by the fragments per kilobase per million (FPKM). Three biological replicates are used for the expression analysis as described. CjYL, young leaves; CjFB, floral buds; CjPE, petal; CjEM, fruit endocarp; CjSD, Seed.

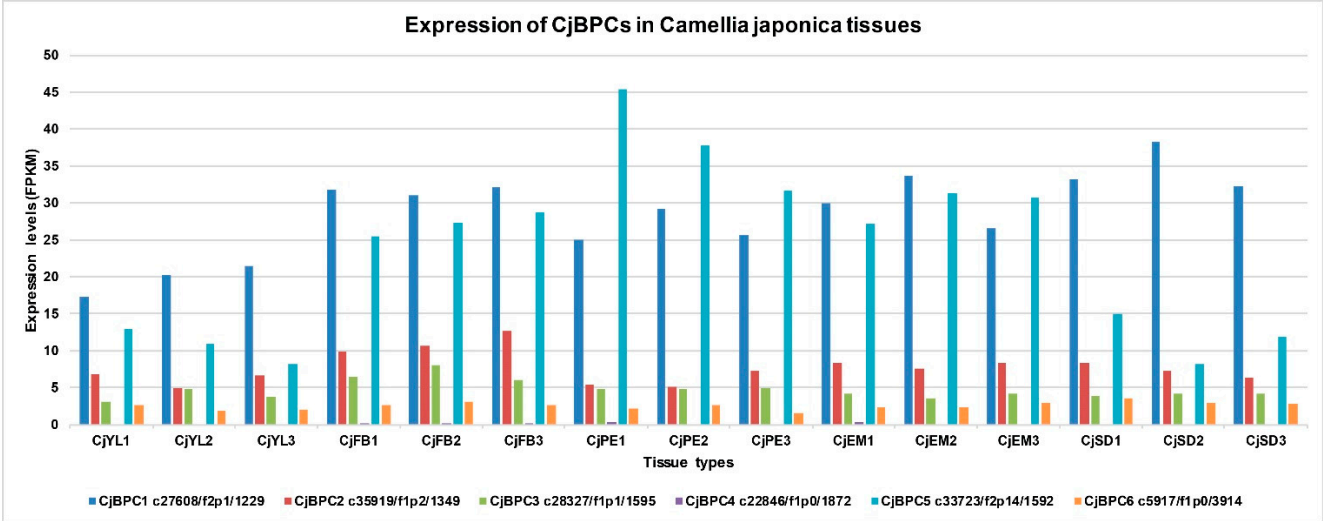


Figure. S3 Overexpression of *CjBPC1* in Arabidopsis and molecular validation of transgenic lines. A, the diagram of overexpression construct for Arabidopsis transformation. B, the PCR verification positive T1 transgenic lines. C, the relative expression of *CjBPC1* in different transgenic lines using gene-specific primers.

