



Review

Modulation of Neuron and Astrocyte Dopamine Receptors via Receptor–Receptor Interactions

Diego Guidolin ^{1,*}, Cinzia Tortorella ¹, Manuela Marcoli ², Chiara Cervetto ², Raffaele De Caro ¹, Guido Maura ² and Luigi F. Agnati ³

¹ Department of Neuroscience, University of Padova, 35122 Padova, Italy; cinzia.tortorella@unipd.it (C.T.); rdecaro@unipd.it (R.D.C.)

² Department of Pharmacy, University of Genova, 16126 Genova, Italy; manuela.marcoli@unige.it (M.M.); cervetto@difar.unige.it (C.C.); guido.maura@gmail.com (G.M.)

³ Department of Biomedical, Metabolic Sciences and Neuroscience, University of Modena and Reggio Emilia, 41121 Modena, Italy; luigi.agnati@gmail.com

* Correspondence: diego.guidolin@unipd.it

Abstract: Dopamine neurotransmission plays critical roles in regulating complex cognitive and behavioral processes including reward, motivation, reinforcement learning, and movement. Dopamine receptors are classified into five subtypes, widely distributed across the brain, including regions responsible for motor functions and specific areas related to cognitive and emotional functions. Dopamine also acts on astrocytes, which express dopamine receptors as well. The discovery of direct receptor–receptor interactions, leading to the formation of multimeric receptor complexes at the cell membrane and providing the cell decoding apparatus with flexible dynamics in terms of recognition and signal transduction, has expanded the knowledge of the G-protein-coupled receptor-mediated signaling processes. The purpose of this review article is to provide an overview of currently identified receptor complexes containing dopamine receptors and of their modulatory action on dopamine-mediated signaling between neurons and between neurons and astrocytes. Pharmacological possibilities offered by targeting receptor complexes in terms of addressing neuropsychiatric disorders associated with altered dopamine signaling will also be briefly discussed.

Keywords: receptor–receptor interactions; receptor complexes; GPCR; dopaminergic pathways; Parkinson's disease; Schizophrenia; drug addiction



Citation: Guidolin, D.; Tortorella, C.; Marcoli, M.; Cervetto, C.; De Caro, R.; Maura, G.; Agnati, L.F. Modulation of Neuron and Astrocyte Dopamine Receptors via Receptor–Receptor Interactions. *Pharmaceuticals* **2023**, *16*, 1427. <https://doi.org/10.3390/ph16101427>

Academic Editor: Szczepan Mogilski

Received: 11 September 2023

Revised: 29 September 2023

Accepted: 4 October 2023

Published: 8 October 2023



Copyright: © 2023 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (<https://creativecommons.org/licenses/by/4.0/>).

1. Introduction

Dopamine (DA) is a catecholamine, that is, an ethylamine with an attached catechol group (a phenyl group with two hydroxyl groups in meta- and para positions). DA-producing neurons were first identified and mapped in animals by Dahlström and Fuxe in 1964 [1,2], indicating the existence of neuronal circuits using DA as a neurotransmitter. In the years that followed, the characterization of the circuits which utilize DA, their organization, molecular signature, and cellular and functional features represented one of the most fertile fields of research in neuroscience (see [3]). Over the past decades, technological advances have also helped to expand the knowledge about the anatomical organization of the DA systems in the human brain. Dopamine neuronal populations have, indeed, been identified and characterized in the human brain from the level of gene transcription to the level of the distribution of related proteins by using post-mortem immunohistochemistry and in vitro autoradiography methods, as well as through in vivo neuroimaging techniques such as positron emission tomography and single photon emission tomography (see [4]). As summarized in Table 1, four major dopamine pathways and two additional ones can be described in the human brain [5,6]. They are involved in the regulation of both physiological and behavioral processes, including movement, endocrine control, cognition, reward, and motivation.

Table 1. Dopaminergic pathways [3,6].

Pathway	Description	Functional Features
Nigro-striatal	From the substantia nigra pars compacta to the dorsal striatum	Motor control
Mesolimbic	From the ventral tegmental area to the ventral striatum	Reward-/aversion-related cognition
Mesocortical	From the ventral tegmental area to the prefrontal cortex	Executive functions
Tubero-infundibular	From the hypothalamus to the pituitary gland	Regulation of prolactin secretion
Incerto-hypothalamic	From the zona incerta to the hypothalamus	Visceral and sensorimotor activities
Hypothalamo-spinal	From the hypothalamus to the spinal cord	Modulation of locomotor networks

The release of the catecholamine from nerve endings upon axonal stimulation certainly represents the main process of dopamine-mediated interneuronal communication. Released DA acts on postsynaptic and presynaptic receptors at the synapse and is mostly taken up back into nerve endings by the dopamine transporter protein, which belongs to the solute carrier transporter family [6]. In some regions of the central nervous system, however, dopamine signaling also occurs through processes of “volume transmission”, based on the diffusion of the molecule in the extra-cellular space to reach more distant targets (see [7–9]). Examples include globus pallidus [10], substantia nigra [11], ventral tegmental area [12], ventral subiculum [13], pedunculopontine nucleus [14], and retina [15]. In this respect, of significant interest is evidence indicating that DA, interacting with DA receptors expressed by astrocytes [16–18], may also act on these cells, leading to a modulation of neuron–astrocyte crosstalk (see [19]).

Dopamine receptors belong to the superfamily of G-protein-coupled receptors (GPCRs). A first indication of their existence was reported in 1972 [20]; they were identified in 1975 [21,22], and five different subtypes have been described so far. In view of the strong implication of DA signaling in a variety of neurological, psychiatric, and drug addiction disorders, with a relevant impact not only on afflicted individuals, but also on society, DA receptors have been the focus of intense research efforts and a variety of drugs have been designed to treat these illnesses by targeting DA receptors directly or indirectly (see [6]).

In recent decades, experimental evidence demonstrating that structural receptor–receptor interactions (RRIs) may occur between receptor proteins has been of interest [23–33]. The term RRI indicates a type of interaction needing a direct physical contact between the partner proteins, with the formation of oligomeric complexes at the cell membrane (see [34] for a recent review). Available studies indicated the formation of receptor complexes as a quite common process in the different receptor families, where the ion channel receptors are at one end of the spectrum (being assembled by multimerization) and GPCRs at the other. Thus, as pointed out by Changeux and Christopoulos in a detailed review [35], RRIs emerge as an efficient mechanism for modulating the functional properties of receptor proteins, including GPCRs that are able to signal as monomers. This mechanism, indeed, allows a sophisticated regulation of the intercellular communication already at the membrane level [9] and opens the possibility of new pharmacological strategies to modulate receptor signaling. In this context, several groups (including our group), have focused their attention on the detection of receptor complexes containing DA receptors in nervous tissues and on the role they can play in DA-mediated signaling in neurons and astrocytes. In the present review, published data concerning this modulatory process will be presented and discussed. Since the subject is quite broad, review articles focused on specific aspects of the topic will also be suggested for further information.

2. Dopamine Receptors

Five different subtypes of DA receptors (D_1 , D_2 , D_3 , D_4 , and D_5) have been identified in brain tissue (see [6] for a recent review), and based on their structure and pharmacological properties, they can be classified into two major groups [36]: D_1 -like receptors (including D_1 and D_5) and D_2 -like receptors (comprising D_2 , D_3 , and D_4). Binding studies have demonstrated some differences between the two groups in terms of affinity to DA, with D_2 -like receptors exhibiting a 10- to 100-fold greater affinity to DA than D_1 -like

receptors [37–39]. D₁- and D₂-like receptors also differ in their genetic structure. D₂-like receptor genes, indeed, have introns in their coding regions, while D₁-like receptor genes do not exhibit this feature [40]. This genetic organization, therefore, enables the generation of D₂-like receptor splice variants, and alternative splicing is particularly important for the D₂ receptor, leading to the generation of two distinct receptor isoforms: D₂-short and D₂-long [41,42], differing because of the insertion of 29 amino acids in the D₂-long intracellular domain, which may play a role in determining second messenger specificity [36,42].

Concerning signal transduction, it is commonly accepted that the receptors of the D₁-like group mainly mediate the stimulation of the second messenger adenylyl cyclase (cAMP) by coupling to the G_s protein, whereas receptors of the D₂-like group mainly exert inhibitory effects on this enzyme by coupling to G_{i/0} protein [6,43]. In addition to the just mentioned main pathway, D₁-like receptors may also couple to the G_q protein [44–46] and modulate phospholipase C [44,46,47], leading to an increase in intracellular calcium levels and activation of protein kinase C. In this respect, the regulation of intracellular calcium levels is a well-documented action of dopamine on astrocytes [48]. DA receptors are expressed by astrocytes [49], and D₂ receptor activation was reported to decrease intracellular Ca²⁺ levels in hippocampal [50] and ventral midbrain astrocytes [51], while D₁ receptor activation elevated astrocytic Ca²⁺ levels in the hippocampus [50], nucleus accumbens [52], and cerebellum [53].

DA signaling cascade, however, may also be modulated by the significant network of molecular interactions that DA receptors can establish in their environment [6,43], which interfere with the GPCRs activity. A first example [54,55] is provided by G-protein-coupled receptor kinases (GRKs). GRKs phosphorylate receptors in response to persistent stimulation [56]. Consequently, the receptor becomes a target for a scaffolding protein, named arrestin, blocking further activation of the GPCR [57] and allowing the GPCR–arrestin complex to engage a variety of G-protein-independent signaling pathways [58]. A second example [59,60] is represented by the regulators of G protein signaling (RGS). RGS are a family of more than 35 intracellular proteins (see [6]) that induce inhibitory effects on GPCRs. Concerning DA receptors, they mainly regulate the D₂-like class [61] and are important in order to stop signaling in the slow synaptic transmission elicited by D₂ receptors [38,62].

In this context, of particular interest is the possibility of direct RRI involving DA receptors with the formation of receptor complexes at the cell membrane [63,64]. In receptor complexes, indeed, the chain of events linking the recognition of a ligand by the single protomers to the signal transduction also depends on the neighboring receptors. This specific mechanism modulating DA signaling will be the focus of the next sections.

3. Structural Receptor–Receptor Interactions

Functional interactions between receptors, by mechanisms of transactivation or by sharing signaling pathways, are well-known processes that do not need a physical contact between the involved proteins [65]. In the 1980s, however, Agnati, Fuxe, and collaborators [23,66], through *in vitro* and *in vivo* experiments, provided indirect evidence that GPCR monomers can establish structural interactions (see [63] for historical details). These findings led to the hypothesis that neuron activity could be modulated by receptor complexes present at the cell membrane and formed by different types of GPCRs [64], a mechanism allowing (already at the membrane level) some integration of synaptic (wiring transmission) and extra-synaptic (volume transmission) signals [64]. The term RRI was subsequently proposed to emphasize the concept of an interaction between receptors requiring a direct physical contact between the molecules and leading to the formation of dimers or high-order molecular complexes at the cell membrane [67]. In the years that followed, several groups [23–33] provided direct evidence of the existence of this structural organization, and the amount of data supporting the existence of GPCR complexes further increased with the advent of biophysical techniques capable of detecting the spatial proximity of protein molecules [68–71]. The obtained results demonstrated that GPCRs

can signal not only as monomers, but also as part of receptor complexes [72] and indicated that receptor complexes represent a quite common molecular organization in the different families of receptors [34].

The basic molecular mechanism underlying the formation and the dynamics of these receptor assemblies are allosteric interactions (see [73]). Allostery (see [35,74–76] for extensive reviews) is a mode of communication between distant sites in a protein, in which the energy associated with dynamic or conformational changes at one site can be transferred (along specific pathways within the protein structure) to other sites, that, in turn, will change their conformational or dynamic features. Thus, when a quaternary structure is established via direct RRI between protomers, energy perturbations at some site of one protomer can propagate into the nearby protomers and change their conformational and functional properties, leading to a cooperative behavior of the whole complex [34,77]. In current research on receptor oligomerization, therefore, the identification of the residues forming the interface between protomers is of significant interest. They, indeed, influence the overall architecture that the receptor complex can assume. In this respect, to predict the interfaces available for RRI, several bioinformatics methods have been developed (see [78–80] for reviews on this topic). As a matter of fact, the number of ways GPCRs interact in the membrane to form complexes is probably limited. The vast majority of experimentally identified receptor complexes, indeed, are dimers. And some interfaces have been observed to be more exploited than others for RRI [81]. Nevertheless, oligomeric heteroreceptors have been detected (see [81–84]).

The signaling outcomes from a receptor complex, therefore, depend on several factors (including the composition and the topological organization of the complex and the effects of ligands on its stability and trafficking), which may strongly influence the cascade of events linking the recognition of a ligand by single protomers to the signal transduction (see [43,80,85]). Some of the possible modulations that allosteric RRI may induce on signaling when a receptor complex forms are summarized in Figure 1. They include changes in ligand recognition, G-protein activation, receptor desensitization [86], and switching to β -arrestin signaling [87]. In this context, a relevant aspect of receptor complex formation is also the possible appearance in the formed quaternary structure of novel specific allosteric sites allowing the binding of some modulator. Thus, ligands specific to the receptor complex as such may also exist (see [88]).

A final aspect deserving consideration (see [34] for a discussion) concerns the cell environment in which receptor complexes are located. In fact, the network of molecular interactions they can establish at the cell membrane with other biochemical components (the so-called “horizontal molecular networks” [89]) may influence their signaling. In this context, a specific aspect of interest is the lipid environment, since it was shown to influence receptor function [90]. In particular, changes in the membrane composition altering receptor signaling were associated with several health disorders during aging [90].

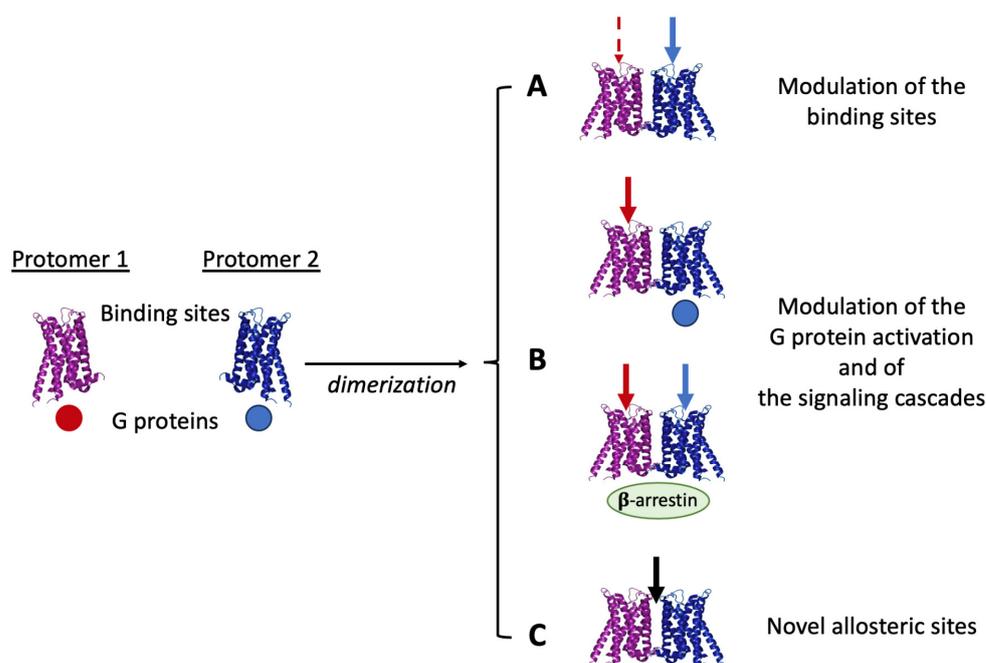


Figure 1. As a result of allosteric RRI, receptor complexes appear to be endowed with pharmacological features that cannot be fully derived from the characteristics of the single participating protomers (see text).

4. Receptor Complexes Involving Dopamine Receptors

DA receptors belong to class A GPCRs [91], well known for being able to signal as monomers [92]. In addition, however, the overall available evidence (obtained through multiple approaches with consistent results) strongly supports the presence of class A GPCR complexes in native systems [72]. In this respect, studies concerning the kinetics of complex formation and its dependence on the involved interaction energy [93] are of substantial interest. The observed half-lives of dimers indicate that they are often transient (lasting few hours) and may undergo recombination (“kiss-and-run” encounters [80]). These processes may lead to a dynamic equilibrium between monomers and receptor complexes for class A GPCRs, as suggested by studies on the corticotropin-releasing factor receptor type 1 in the endoplasmic reticulum [94], indicating that the ratio of monomers/receptor complexes was maintained at an almost constant level in the plasma membrane, even in spite of agonist activation of the receptors. Receptor complexes including DA receptors (see also [95]) are shown in Table 2.

Table 2. Receptor complexes involving dopamine receptors.

Receptor Complex	Cell Location	Reference
A _{2A} -D ₂	Neurons	[68]
A _{2A} -D ₂	Astrocytes	[96]
A _{2A} -D ₃	Neurons	[97]
A _{2A} -D ₄	Neurons	[98]
NMDA-D ₂	Neurons	[99]
NTS ₁ -D ₂	Neurons	[100,101]
CB ₁ -D ₂	Neurons	[102]
D ₂ -5HT ₁	Astrocytes	[103]
D ₂ -5HT _{2A}	Neurons	[101,104]
D ₂ -OTR	Neurons	[105]
D ₂ -OTR	Astrocytes	[106]
D ₂ -GHS _{1A}	Neurons	[107]

Table 2. Cont.

Receptor Complex	Cell Location	Reference
D ₂ -D ₄	Neurons	[108]
α _{2A} -D ₄	Neurons	[108]
β ₂ -D ₄	Neurons	[109]
D ₄ -MOR	Neurons	[110]
CCK ₂ -D ₂ (putative)	Neurons	[66,111]
α ₁ -D ₂ (putative)	Astrocytes	[112]
A _{2A} -D ₂ -sigma1	Neurons	[113]
A _{2A} -D ₂ -mGluR ₅	Neurons	[84]
A _{2A} -D ₂ -CB ₁	Neurons	[82]
D ₁ -D ₂	Neurons	[114]
D ₁ -D ₃	Neurons	[115]
A ₁ -D ₁	Neurons	[116,117]
NMDA-D ₁	Neurons	[118]
GABA _A -D ₅	Neurons	[119]
α ₁ -D ₁ (putative)	Astrocytes	[112]

NMDA—N-methyl-D-aspartate glutamate receptor; 5HT₁, 5HT_{2A}—type 1 and type 2A serotonin receptors; GHS_{1A}—type 1a ghrelin receptor; OTR—oxytocine receptor; CB₁—type 1 cannabinoid receptor; NTS₁—type 1 neurotensin receptor; α₁, β₂—type α1 and type β₂ adrenergic receptor; sigma1—sigma1 receptor; mGluR₅—type 5 metabotropic glutamate receptor; MOR—μ-opioid receptor; type A₁ and type A_{2A}—adenosine receptor.

4.1. Receptor Complexes Involving D₂-like Dopamine Receptors

A first aspect emerging from the available data is that D₂ appears to be a hub receptor which interacts with many other GPCRs.

Probably, the most studied interaction is between dopamine D₂ and adenosine A_{2A} receptors, leading to the formation of A_{2A}-D₂ heterodimers (see [95,120] for reviews). By using pull-down and mass spectrometry techniques, it has been demonstrated [121,122] that the heteromerization between A_{2A} and D₂ receptors significantly depends on charged residues located at the intracellular part of the transmembrane helix 5 (TM5) of the D₂ receptor. The role of TM helix interactions within the A_{2A}-D₂ heteroreceptor complex interface has also been explored by using synthetic TM α-helix peptides of the D₂ receptor [123], and the results allowed for the identification of a TM4/5 interface between the two monomers. The A_{2A}-D₂ heterodimer is also representative of many aspects concerning the signaling outcome from a receptor complex. Experimental evidence has shown that the receptor complex formation modifies the signaling from the single protomers. In particular, early in vitro experiments on membrane preparations showed a reduction in the affinity of the high-affinity D₂-agonist-binding site after incubation with the A_{2A} agonist CGS21680 [124,125], demonstrating that antagonistic interactions occur in the A_{2A}-D₂ heterodimer. By using receptor autoradiography, this finding was subsequently confirmed by studies on brain tissue from rats and humans [126]. They showed a strong reduction in D₂ receptor affinity for dopamine in the nucleus accumbens core and shell after the A_{2A} receptor agonist treatment. By using functional, biochemical, and biophysical techniques (such as co-immunoprecipitation and proximity ligation assay), antagonistic interactions between A_{2A} and D₂ receptors were also recently demonstrated in astrocytes [96,127]. In this context, observations indicating that agonist activation of the A_{2A} protomer in the A_{2A}-D₂ heteroreceptor complex inhibits D₂ G_{i/o}-mediated signaling but increases the D₂ β-arrestin₂-mediated signaling are of interest. This marks a difference compared with the action of D₂ receptor antagonists, which block all the D₂ signaling pathways. Thus, through the allosteric receptor–receptor interaction, an A_{2A} agonist becomes a biased inhibitory modulator of the G_{i/o}-mediated D₂ signaling [128]. The possible formation, as a consequence of the formation of a receptor complex, of new allosteric sites allowing the binding of some ligand is a further modulatory mechanism that the A_{2A}-D₂ heteromer illustrates. Homocysteine can, indeed, bind to the heterodimer without interfering with the RRI between A_{2A} and D₂ and acts as an allosteric antagonist of the D₂ receptor [129]. Thus, the inhibitory effect of A_{2A} agonists is amplified by homocysteine. These modulatory

actions were demonstrated in striatal neurons [129], as well as in astrocytes [130], where homocysteine reduces the D₂-mediated inhibition of glutamate release. An intriguing process involving A_{2A} and D₂ receptors was highlighted by studies on cell lines [19] that demonstrated intercellular transfer of these GPCRs by exosomes, resulting in the incorporation of functional receptors into acceptor cells. As shown by photo-bleaching fluorescence resonance energy transfer, the transferred receptors may also undergo A_{2A}-D₂ receptor heteromerization in the target cell. Thus, the release of extracellular micro vesicles (the so-called “roamer type” of volume transmission [19]) may represent a significant mechanism for the modulation of neuron-neuron and astrocyte–neuron intercellular signaling.

Evidence has been provided indicating that the adenosine A_{2A} receptor can establish antagonistic RRIs with the other D₂-like receptors as well, namely, D₃ [97] and D₄ [98], leading to a reduction in the affinity of their binding site for DA. Antagonistic RRIs also characterize other receptor complexes involving the D₂ receptor, as, for instance, the heterodimers it can form with the glutamate NMDA [99] and mGluR₅ [84] receptors, the neurotensin NTS₁ [100] receptor, and the cannabinoid CB₁ [102,131] receptor. Higher-order heteroreceptor complexes, involving both A_{2A} and D₂, have also been identified. Examples include the heterotrimers formed by A_{2A} and D₂ receptors with the metabotropic glutamate receptor 5 (A_{2A}-D₂-mGluR₅ [84]), the sigma₁ receptor (A_{2A}-D₂-sigma₁ [113]), and the cannabinoid CB₁ receptor (A_{2A}-D₂-CB₁ [82]). In these receptor complexes, the pattern of allosteric interactions on the D₂ protomer also inhibits the recognition and signaling of the DA receptor.

Synergistic RRIs involving the D₂ receptor, however, were also identified. A first example is provided by the receptor complex between the D₂ receptor and the serotonin 5-HT_{2A} receptor [104], where the activation of the 5-HT_{2A} protomer by 5-HT_{2A} agonists produced an enhancement of D₂ signaling. In astrocytes, receptor complexes between the dopamine D₂ receptor and the serotonin 5-HT_{1A} receptor have been observed [103]. However, the functional consequences of the signaling pathways mediated by D₂-5-HT₁ heteromers in these cells are still not known in detail [132]. A further example is represented by the D₂-OTR heterodimer, involving D₂ and the oxytocin receptor. In neurons [105], oxytocin, via the allosteric RRI established in the heterocomplex, markedly increased D₂ receptor recognition (increased affinity of the high-affinity state) and increased the coupling of G_{i/o} to the receptor. The D₂-OTR heterodimer was recently identified in astrocytes as well [106], and the activation of OTR was shown to have a facilitatory effect on the response of D₂ receptors, causing them to be activated by subthreshold D₂ agonist concentrations and leading to an inhibition of glutamate release by the cells.

Synergistic RRIs are also in operation in the heterodimer involving the dopamine D₄ receptor and μ -opioid receptor (MOR) [110], since D₄ activation causes a substantial increase in the affinity of the MOR agonist binding sites. Evidence was also obtained that the D₄ and β_2 -adrenergic receptor may form a D₄- β_2 receptor complex that integrates G_s- and G_i-mediated regulation of adenylyl cyclase [109]. In this context, of particular interest are also studies (see [108]) focused on the dopamine D₄ receptor polymorphic variants D_{4.4} (four repeats in exon 3) and D_{4.7} (seven repeats in exon 3), both able to heterodimerize with the norepinephrine α_{2A} receptor. However, only heteromerization with D_{4.7}, but not with D_{4.4}, increases the potency of norepinephrine in terms of activating the α_{2A} receptor, indicating the possible polymorphic variants of a D₂-like receptor as a factor conferring significantly different pharmacological properties onto the receptor complexes it may form.

4.2. Receptor Complexes Involving D₁-like Dopamine Receptors

The potentiation of immediate early gene expression and of arachidonic acid release have been described as functional interactions between activated dopamine D₁ and D₂ receptors (see [45]). However, it was also demonstrated that stably co-expressed D₁ and D₂ receptors may form heteromeric units [114]. It is of substantial interest that the two receptors, when coactivated in the same cell, produce a phospholipase C-mediated calcium signal that is not seen when the receptors are activated alone. The pharmacological analysis

of this receptor complex indicated a specific coupling to the $G_{q/11}$ pathway to produce such a response. Activation of $G_{q/11}$, however, could not be elicited through activation of either receptor when activated alone. Thus, the recruitment of G proteins other than those expected for the monomers has been observed after D_1 - D_2 dimerization, a further mechanism of signal transduction modulation associated with receptor complex formation.

Antagonistic interactions between D_1 and the adenosine A_1 receptor, associated with the formation of A_1 - D_1 heterodimers [116,117], were also characterized. A_1 agonists, indeed, were found to reduce the number of D_1 agonist binding sites in the high-affinity state, and with receptor autoradiography, A_1 agonists were found to antagonistically modulate D_1 binding sites, causing a reduction in their affinity (see [133] for details).

Receptor complexes between dopamine D_1 and D_3 have been demonstrated using several techniques, giving evidence for synergistic intramembrane D_1 - D_3 interactions at the level of D_1 recognition, since D_3 activation was able to increase the affinity of the D_1 agonist binding sites [115]. Synergistic RRI also exist in the D_1 -NMDA heterodimer [118], by which NMDA receptor activation can recruit D_1 receptors to the plasma membrane, thereby leading to an increase in D_1 signaling and cAMP accumulation.

Recent interesting findings on prefrontal cortex astrocytes indicated a significant functional interaction between α_1 -adrenergic and DA receptors, driving downstream Ca^{2+} signaling [112]. Also, in light of the abovementioned data showing that DA receptors may form receptor complexes with adrenergic receptors [109,134], and of neuroanatomical data showing that D_1 and α_1 -adrenergic receptors colocalize on prefrontal cortex dendrites and may undergo co-trafficking [119], the hypothesis has been put forward that in cortical astrocytes as well, heterodimers involving DA receptors and adrenergic receptors could be present [112]. A direct experimental demonstration, however, is still lacking.

GABA_A and dopamine D_5 heteromerization, demonstrated by Liu and collaborators [135], was the first identification of a receptor complex involving a GPCR and an ion-channel receptor. The results indicated that co-activation of the monomers was required for the formation of the complex, which allowed for a bidirectional crosstalk, leading to a reduction in GABA_A signaling and a reduced coupling between D_5 and G_s proteins.

4.3. Possible Differences in Receptor Complex Dynamics in Neurons and Astrocytes

As briefly illustrated before, a number of receptor complexes (such as, for instance, the A_{2A} - D_2 heterodimer) are expressed both in neurons and astrocytes. In this respect, it is reasonable to assume that the conformation of a receptor complex in the two cases may exhibit some difference because of differences in the membrane microenvironment. Differences in the energy landscape, indeed, modulate the pattern of allosteric interaction between monomers and may lead to changes in the signaling features of the complex that they can form [80].

Differences in membrane potential between the two cell types, for instance, have been documented [136]. Unlike neurons, astrocytes do not generate action potentials, but they are electrically dynamic cells. Indeed, in contrast to most non-excitable cells that have relatively depolarized membrane potentials, astrocytes have a hyperpolarized membrane (at a level that typically rests significantly below that of neurons) and a low membrane resistance. For the present discussion, membrane composition is another factor deserving consideration. This aspect was the focus of an extensive lipidome analysis by Fitzner and collaborators [137], showing that each cell type was characterized by a unique lipid composition: neurons, for instance, exhibited quite high levels of cholesterol, while astrocytes were enriched in phosphatidylinositol.

All these features of the membrane microenvironment, therefore, have the potential to modulate the pharmacological properties of a given receptor complex. To illustrate this concept, the results of a simulation based on molecular modeling methods and focused on the A_{2A} - D_2 heterodimer in two different membrane environments (neuron-like and astrocyte-like) are shown in Figure 2.

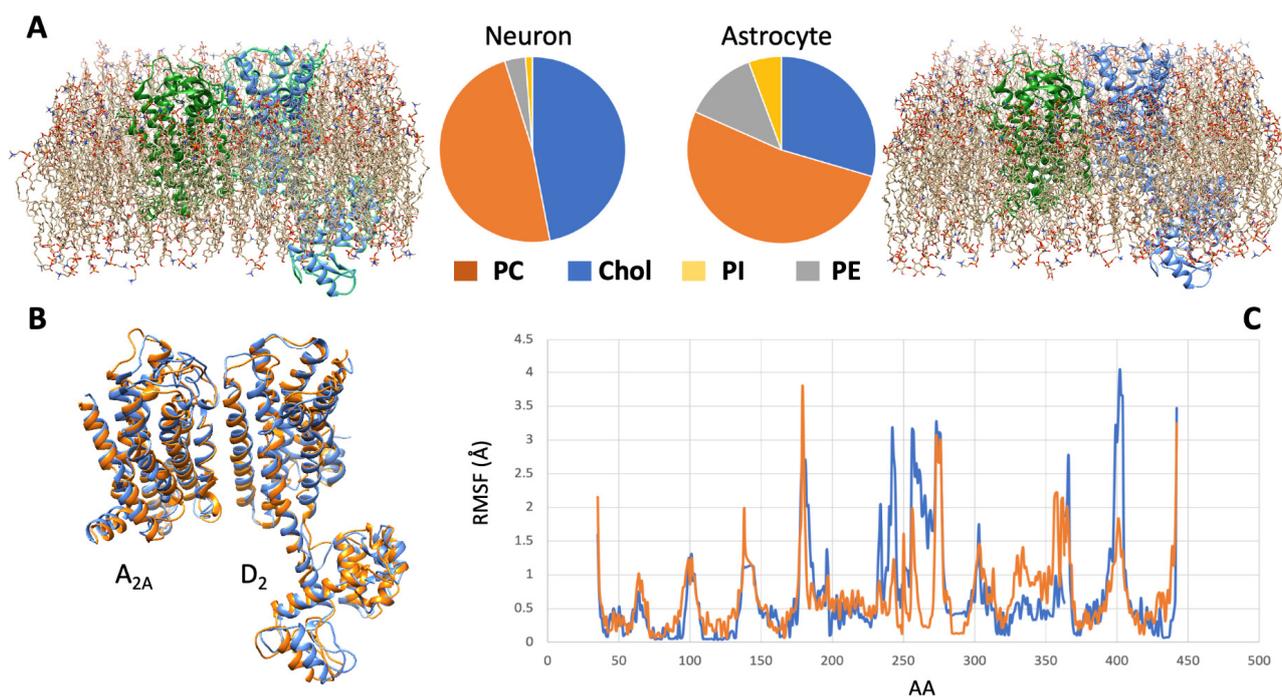


Figure 2. Molecular dynamics simulation of the A_{2A}-D₂ receptor complex in different cell membranes. (A) By using the CHARMM-GUI membrane builder web server (<http://www.charmm-gui.org/?doc=input> [138], accessed on 5 July 2023), four phospholipids, namely, phosphatidylcholine (PC), cholesterol (Chol), phosphatidylinositol (PI), and phosphatidylethanolamine (PE), were used to model two different membrane bilayers around the molecular model [120,123] of the heterodimer. The first (left panel) approximated the neuronal membrane composition, the second one (right panel) the astrocytic one (see [137]). A molecular dynamics procedure, based on the CABSflex method [139] and available as a web server (<https://biocomp.chem.uw.edu.pl/CABSflex2>, accessed on 6 July 2023), was then used to evaluate the conformations that the receptor complex may acquire in the two environments. (B) Configurations of minimal energy of the A_{2A}-D₂ heterodimer in neuronal (orange) and astrocytic (blue) membrane. (C) Root mean square fluctuations (RMSF) diagrams, per amino acid position, of the D₂ monomer chain when in neuronal (orange) and astrocytic (blue) membrane. The estimated differences in configuration and dynamical behavior of the heterodimer suggest that different membrane environments could represent a factor modulating the pharmacological properties of the receptor complex.

5. Complexes Involving Dopamine Receptors in the Main Dopaminergic Pathways: Impact on Neuropharmacology

The intermingling of findings from functional neuroanatomy (linking dopaminergic pathways to specific functions and diseases) with evidence emerging from chemical neuroanatomy (describing the distribution of receptor complexes involving DA receptors in brain cells and regions) may help to better appreciate the function that receptor complexes containing DA receptors can fulfill, and may contribute to the development of new pharmacological approaches with a potentially major impact on molecular medicine. In this respect, presently available information is limited to ascending dopaminergic pathways (nigrostriatal, mesolimbic, and mesocortical) and neuron–astrocyte crosstalk, being descending pathways (mentioned in Table 1) almost uninvestigated in terms of receptor complexes containing DA receptors. Thus, in the sections that follow, only the abovementioned signaling pathways will be considered (see also [108,133,140–142] for reviews). These are, however, of significant interest, being associated with an impact on neuropsychiatric diseases. Reported findings are summarized in Table 3.

Table 3. Complexes involving dopamine receptors in the ascending dopamine pathways and in neuron–astrocyte crosstalk [108,133,140–142].

DA Pathway	Receptor Complexes	Type of Interaction	Location	Major Pathologies
Nigro-striatal	A _{2A} -D ₂	Antagonistic	Dorsal striatum	PD
	CB ₁ -D ₂	Antagonistic		
	NMDA-D ₂	Antagonistic		
	A _{2A} -D ₂ -CB ₁	Antagonistic		
	A _{2A} -D ₂ -mGluR ₅	Antagonistic		
	A ₁ -D ₁	Antagonistic		
	D ₁ -D ₃	Synergistic		
Mesolimbic	A _{2A} -D ₂	Antagonistic	Ventral striatum	Addiction Schizophrenia
	A _{2A} -D ₃	Antagonistic		
	NMDA-D ₂	Antagonistic		
	NMDA-D ₁	Antagonistic		
	NTS ₁ -D ₂	Antagonistic		
	D ₂ -5HT _{2A}	Synergistic		
	D ₂ -OTR	Synergistic		
	D ₁ -D ₂	Signaling cascade change		
	D ₁ -D ₃	Synergistic		
	A _{2A} -D ₂ -sigma ₁	Antagonistic		
	A _{2A} -D ₂ -mGluR ₅	Antagonistic		
GABA _A -D ₅	Antagonistic			
D ₄ -MOR	Synergistic			
Mesocortical	α _{2A} -D ₄	Dependent on D ₄ polymorphism	Prefrontal cortex	ADHD
	β ₂ -D ₄	Signaling cascade change		
	D ₂ -D ₄	Dependent on D ₄ polymorphism		
	A _{2A} -D ₄	Antagonistic		
Neuron–Astrocyte crosstalk	A _{2A} -D ₂	Antagonistic	Astrocytes	PD Addiction Schizophrenia
	D ₂ -OTR	Synergistic		
	D ₂ -5HT ₁	Not detailed		

PD, Parkinson's disease; ADHD, attention-deficit hyperactivity disorder.

5.1. Nigro-Striatal Dopamine Pathway

The nigro-striatal pathway starts from dopamine-containing cells in the substantia nigra pars compacta (SNc) of the midbrain to establish multiple synaptic contacts with medium spiny neurons (MSNs) of the ipsilateral dorsal striatum [143]. MSNs also receive cortico-striatal glutamatergic afferents and are GABAergic projection neurons classified into three populations [3]. Island (patch) MSNs are localized in the so-called striatosomes [144] and send a feedback signal to neurons of the SNc, striato-nigral/entopeduncular MSNs project to the substantia nigra pars reticulata (SNr) and the entopeduncular nucleus (EPN) (nuclei from which the so-called direct pathway of motor control starts), and striato-pallidal MSNs project to the external globus pallidus (GPe) (nucleus from which the indirect pathway of motor control starts), which in turn modulates the subthalamic nucleus (STh). The direct pathway triggers a disinhibition of the target regions, whereas the indirect pathway triggers their inhibition, leading to activation and suppression of motor behavior, respectively. In terms of the dopaminergic modulation of these pathways, the direct pathway is dominated by D₁ receptors, expressed at a high level by striato-nigral/entopeduncular MSNs, while the indirect pathway is mainly regulated by D₂ receptors, well expressed by striato-pallidal MSNs [3].

As an endogenous neuroprotectant agent, adenosine is extensively distributed in the central nervous system, where it acts through specific receptors [145], and in the dorsal striatum, A₁ and A_{2A} adenosine receptors are widely expressed in both MSNs [133] and glutamatergic terminals [134]. It is not surprising, therefore, that receptor complexes involving adenosine and dopamine receptors were identified in the dorsal striatum. In

striato-nigral/entopeduncular MSNs, for instance, the presence of the A₁-D₁ heterodimer has been reported [116,146], while receptor complexes involving the adenosine A_{2A} and the D₂ receptors (namely, the A_{2A}-D₂ heterodimer and the heterotrimers A_{2A}-D₂-mGluR₅ and A_{2A}-D₂-CB₁) were found in striato-pallidal MSNs and their glutamate inputs [82,84,95]. STn is also innervated by collaterals of the nigro-striatal bundle [143], and co-localization of A_{2A} and D₂ receptors has been recently documented in this nucleus [147], opening the possibility of the presence (yet to be substantiated) of A_{2A}-D₂ heterodimers within the dorsal and medial aspects of the structure.

Parkinson's disease (PD) is a common disease, associated with neurodegeneration of the nigro-striatal pathway, leading to imbalance or loss of dopaminergic signaling to the dorsal striatum with the emergence of altered motor features, such as bradykinesias, tremor, and rigidity. The introduction of L-DOPA [148] revolutionized the management of this disease, leading to an effective symptomatic treatment. However, it soon became apparent that the drug offered only symptomatic relief and did not affect the underlying pathology. Moreover, chronic use of the drug was associated with a range of adverse effects, such as dyskinesias, toxicity, or loss of efficacy [149]. Current therapeutic protocols, therefore, seek to delay long-term complications of treatment for as long as possible. In this context, the antagonistic allosteric RRI described earlier, which characterize the receptor complexes involving adenosine and dopamine receptors, led to the hypothesis (schematically illustrated in Figure 3A) that by targeting these heteromers with antagonists of the adenosine receptors, antiparkinsonian effects could be obtained (see [150] for a specific review on this topic). This research effort mainly focused on A_{2A}-D₂ receptor complexes. Animal models of PD gave support to the hypothesis and clinical evidence was also obtained (see [120] for references). In this respect, it is of interest to mention the very recent approval in the United States of an A_{2A} antagonist (istradefylline) as an adjunctive treatment to L-DOPA [151] in PD. Following the same logic, D₁ signaling in the A₁-D₁ heterodimer could be modulated by targeting the adenosine A₁ receptor to obtain antiparkinsonian effects [133].

Other receptor complexes in the dorsal striatum, however, deserve a mention as possible pharmacological targets in PD. CB₁ antagonists targeting the CB₁-D₂ heterodimer, for instance, may represent possible antiparkinsonian drugs, since the antagonistic RRI, characterizing this receptor complex, can enhance D₂ signaling [152]. Behavioral correlates to the antagonistic receptor interactions in CB₁-D₂ heterodimers have also been obtained using the CB₁ receptor agonist HU-210, which has been found to reduce L-DOPA-induced rotations in 6-hydroxydopamine-lesioned rats [153]. In cortico-striatal glutamate terminals, the D₂-NMDA receptor complex (with antagonistic RRI) is constitutively present [99] and inspired the possibility that a dual approach in PD with low doses of selective D₂ agonists and NMDA antagonists could lead to antiparkinsonian actions with reduced development of dyskinesias [133].

5.2. Mesolimbic Dopamine Pathway

The mesolimbic pathway connects the ventral tegmental area (VTA), a dopaminergic nucleus of the midbrain, with the ventral striatum (occupying about 20% of the striatum), including the nucleus accumbens (NAc) and the olfactory tubercle, which are striatal regions receiving their major telencephalic input from the hippocampal formation and amygdala, and projecting to the ventral pallidum (VP) and SNr. From there, information is transferred to the anterior cingulate cortex and the orbitofrontal cortex [154]. Concerning the NAc, two main subterritories have been identified, namely, the shell and the core, the shell region being more closely associated with the limbic system than the other regions of the ventral striatum [3].

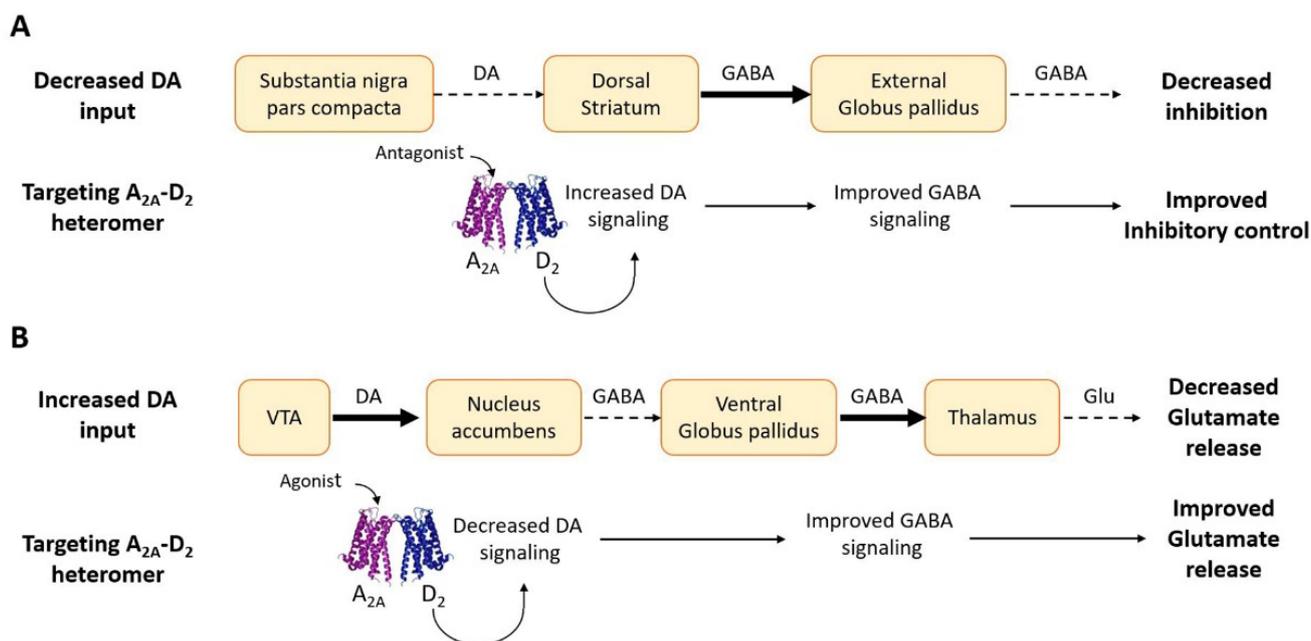


Figure 3. Schematic representation of pharmacological strategies to address imbalance of DA signaling by targeting the A_{2A} - D_2 receptor complex [120]. (A) Decreased DA signaling in the nigro-striatal pathway (as in Parkinson's disease) leads to a reduced D_2 activity and to a decreased inhibitory output from the external globus pallidus to the downstream structures, resulting in unbalanced motor control. Targeting A_{2A} - D_2 heteromers in the striatum with antagonists of the A_{2A} receptors may improve D_2 -mediated dopaminergic signaling and motor control. (B) Overactivity of the mesolimbic dopamine neurons increases the D_2 -mediated dopamine transmission to the ventral striatum, leading to a reduced glutamate drive from the mediodorsal thalamic nucleus. A_{2A} agonists targeting the antagonistic interactions between A_{2A} and D_2 receptors in the complex may improve this condition. Dashed arrows and thick arrows indicate decreased and increased signaling, respectively.

Ventral striatum neurons are MSNs, similar to those of the dorsal striatum, and their dopaminergic input are mainly regulated by D_2 receptors [155]. A_{2A} - D_2 heteroreceptor complexes with antagonistic RRIs were demonstrated in the ventral striatum [125], as were high-order receptor complexes including adenosine A_{2A} and dopamine D_2 receptors, such as, for instance, the A_{2A} - D_2 -mGluR₅ and A_{2A} - D_2 -sigma₁ heterotrimeric complexes [156]. Of interest is also the presence in ventral MSNs of cortico-accumbens terminals of receptor complexes involving dopamine D_2 , glutamate NMDA [99], neurotensin NTS₁ [100], serotonin 5-HT_{2A} [104], and oxytocin [105] receptors.

The mesolimbic pathway is a key element of the so-called reward circuit (see [154]), because the release of dopamine through this pathway regulates motivation and desire for rewarding stimuli (i.e., incentive salience), facilitates reinforcement- and reward-related motor function learning, and may also play a role in the subjective perception of pleasure. Thus, the dysregulation of the mesolimbic pathway and its downstream neurons plays a significant role in the development of significant neuropsychiatric diseases, including addiction and schizophrenia (see [156,157] for specific reviews).

A study [158], for instance, showed that chronic cocaine self-administration increased behavioral responses mediated by D_2 receptors, indicating the relevance of D_2 for cocaine use disorder. Furthermore, chronic cocaine self-administration persistently evoked more than 100% elevations of D_2 binding sites of the high-affinity type [159], and D_2 activation produced a strong relapse of cocaine seeking in animals [160]. In this respect, studies focused on the antagonistic RRIs in the A_{2A} - D_2 receptor complex as a possible pharmacological target indicated that A_{2A} agonists exhibited an inhibitory effect on cocaine reward [160], and A_{2A} activation, leading to D_2 -like receptor blockade, counteracted cocaine relapse.

It is also of interest that cocaine induces a selective increase in σ_1 receptors in the ventral striatum [161]. Thus, the A_{2A} - D_2 antagonistic interaction may become more present thanks to a higher presence of A_{2A} - D_2 - σ_1 receptor complexes. In this context, also results suggesting the existence of D_4 -MOR heterodimers [110] in the striatosomes and SNr, in which D_4 -MOR interactions are in operation, are also of interest. They may play a critical role in at least the early stages of the expression of the morphine effects. In view of the limbic-prefrontal–striatosome–nigral circuitry and its function (see [162]), this interaction may participate in reward-based motor learning and play a significant role in habit acquisition in drug addiction [133].

In schizophrenia, salience becomes exaggerated due, inter alia, to an increased D_2 recognition and signaling in the ventral striatum (mainly nucleus accumbens) [163]. Thus, the classic treatment [164] in schizophrenia is the use of DA receptor antagonists, typically haloperidol and chlorpromazine. Through the blockade of excessive D_2 -mediated DA transmission in the mesolimbic dopaminergic pathway, they allow an improvement of mental symptoms, but induce motor side effects due to the parallel block of the nigrostriatal pathway. Thus, based on the presence in the ventral GABAergic MSNs, in astrocytes, and in glutamatergic terminals of A_{2A} - D_2 containing heteroreceptor complexes with antagonistic A_{2A} - D_2 interactions, the use of A_{2A} receptor agonists (see Figure 3B) as a strategy for the treatment of schizophrenia has been proposed [133] and promising results in animal models have been found [165]. It is worth noting that A_{2A} agonist treatment, especially in combination with low doses of typical and/or atypical D_2 antagonists, could also represent a possible strategy for reducing the development of extrapyramidal side effects [133]. Facilitatory RRI in the 5-HT $_2A$ - D_2 receptor complex may represent a further target for treatments based on antagonists of the serotonin receptor (see [157]), and a reduction in the inhibitory D_2 signaling at the cortico-accumbens glutamatergic terminal level could be obtained by targeting NTS $_1$ - D_2 receptor complexes with agonists of the neurotensin receptor [101]. The D_2 -OTR heterodimer also deserves interest as a possible target in schizophrenia. Indeed, evidence was obtained that the molecular mechanism mediating the social salience was the formation of D_2 -OTR heteroreceptor complexes in the nucleus accumbens core [105]. In fact, being located to a special component of the ventral GABAergic MSNs involved in regulating a brain circuit reaching into the prefrontal cortex, the result of the activation of the D_2 -OTR heteroreceptor complex may produce social attachment and trust and the negative symptoms of schizophrenia may become markedly reduced [140]. Consistent with this hypothesis are data showing that oxytocin can induce antipsychotic actions [166], which appears to be true after being given to schizophrenic patients intranasally [167].

5.3. Mesocortical Dopamine Pathway

The mesocortical pathway connects the VTA to the prefrontal cortex, but dopaminergic axons branch within the cortex to reach multiple cortical areas [3]. By applying a modified Falck–Hillarp technique, Hököfelt and coworkers [168] identified a plexus of dopaminergic fibers in the limbic cortex with an uneven innervation of the entorhinal cortex. DA-containing varicosities preferentially establish synaptic contacts on pyramidal neurons [169].

This pathway is essential to the normal cognitive function of the dorsolateral prefrontal cortex (part of the frontal lobe) and is thought to be involved in cognitive control, motivation, and emotional response [170]. In this respect, it is closely associated with the mesolimbic pathway.

As recently discussed by Ferré and collaborators [108], an interesting aspect of this innervation pattern is the high expression of dopamine D_4 receptors in the cortex of mammals: most glutamatergic pyramidal neurons and about half of the GABAergic interneurons express D_4 . Considering the G_i -coupled D_4 as mostly inhibitory, the D_4 localized in neurons should be expected to exhibit an inhibitory effect on dopamine, while those localized in GABAergic interneurons should be expected to produce disinhibition. Several studies, however, indicate a more complex picture, associated with evidence indicating that D_4

receptors can form receptor complexes with adrenergic receptors [171]. As briefly discussed in Section 4.1, these receptor complexes may have significantly different pharmacological properties depending on the polymorphic variant of the D₄ receptor involved.

In this respect, available evidence associating D₄ polymorphisms with individual differences in impulse control-related neuropsychiatric disorders is of interest, with the most consistent associations found between the gene encoding D_{4.7} and attention-deficit hyperactivity disorder (ADHD) [172]. On this basis, it has been proposed that receptor complexes involving the D₄ receptor should be investigated as possible therapeutic targets for ADHD, as well as for restless legs syndrome [108].

5.4. Neuron–Astrocyte Crosstalk

Increasing evidence (see [141] for a specific review) indicates that astrocytes are directly involved in the regulation of neuronal excitability and action potential propagation. According to this view, a bidirectional relationship exists between astrocytes and neurons, where neural activity influences astrocytic activation, which in turn modulates the activity of neurons [173].

Astrocytes, indeed, monitor the extracellular environment through specific receptors, including many neurotransmitter receptors (such as those for DA). Single astrocytes integrate this information through the elevation of intracellular Ca²⁺ [141] and can propagate this information over large distances by communicating with each other through calcium waves [174]. Such calcium dynamics are considered a key step leading to the release of gliotransmitters (D-serine, ATP, and glutamate) that regulate ongoing neural activity [175]. As indicated by several experimental studies (see [173] for a review), this intercellular crosstalk significantly influences synaptic plasticity and, consequently, higher CNS functions such as, for instance, learning and memory.

In this context, extensive available data indicate that RRI may play a significant role. Relevant examples include the heterodimers A_{2A}-D₂ and D₂-OTR [96,106], formed by the association of the dopamine D₂ receptor with the adenosine A_{2A} or the oxytocin receptor, respectively. These receptor complexes are present in astrocytes and regulate the release of glutamate from these cells [106,127], a process relevant for the control of glutamatergic transmission in striatum and with potential roles in the dysregulation of glutamatergic transmission in various neuropsychiatric diseases (see [176] for a specific review on this topic).

The results of a study [177], showing that knocking down the striatal astrocytic glutamate transporter GLT-1 induces PD-like changes in rodents, illustrate the importance of the regulation of the striatal extracellular glutamate level by astrocytes in this pathology. Furthermore, dopamine-mediated glutamate release from striatal astrocyte processes can modulate the activation of NMDA and metabotropic glutamate receptors on striatal MSN [178], suggesting the abovementioned receptor complexes as potential targets to counteract striatal glutamatergic transmission disfunctions and circuit derangement in PD [176]. In this respect, an interesting possibility was suggested by findings showing that homocysteine (an allosteric modulator of the A_{2A}-D₂ heterodimer, see Section 4.1) was able to counteract the DA-mediated inhibition of glutamate release by astrocytes [130]. The relevance of this finding from a physio-pathological standpoint can be appreciated when considering that L-DOPA treatment can trigger synthesis of homocysteine in astrocytes and their release into the extracellular space [179].

Evidence indicating astrocyte involvement in schizophrenia has also been collected [180,181], where glial abnormalities were proposed to contribute to glutamatergic and dopaminergic neurotransmission dysfunctions [182]. In a mouse model of astrocytic A_{2A} receptor knockout, for instance, impaired glutamate homeostasis associated with enhanced behavioral sensitization to psychoactive drugs and reduced working memory (two behavioral symptoms of the pathology) was reported [180]. Thus, the astrocytic A_{2A}-D₂ heteromers may represent a possible target for A_{2A} agonist or other drugs (see [183,184]) in order to ameliorate the impaired glutamate homeostasis in schizophrenia.

Regulation of astrocytic RRI involving the D₂ receptor can also be of importance for the pathophysiology and treatment of drug addiction. Accumulating evidence, indeed, indicates that drugs of abuse can trigger glutamatergic dysregulation through astroglial mechanisms (see [185]). On this background, D₂-containing heteromers in astrocytes may provide new perspectives in the search for drug addiction therapies.

6. Concluding Remarks

Since the discovery of DA as a neurotransmitter, the relationship between the dopaminergic signaling network and essential physiological and pathological processes in the nervous systems has become clear. The dopaminergic system is a complex system, organized in parallel and segregated functional streams consisting of motor, reward (limbic), and associative (cognitive) control pathways [186]. However, evidence also exists that the system also exploits integrative mechanisms by which information is transferred between these functional circuits (see [3]). Furthermore, it extensively interacts with other critical signaling pathways [6]. Such a complex intercellular communication occurs through both synaptic and volume transmission (see [64]) and is mediated by a set of GPCRs.

In this respect, extensive evidence has been provided showing that DA receptors can also establish direct allosteric RRI with other receptor proteins, leading to the formation of receptor complexes and allowing a modulation of signal decoding already at the membrane level and characterized by specific pharmacological profiles; these are potentially of interest to devise new strategies to address relevant disorders. As briefly discussed here, in recent decades, an increasing number of receptor complexes involving DA receptors have been identified and studied. Several aspects, however, remain to be addressed to better understand their function and the possibilities that their targeting may offer.

As previously suggested [157], a first point (of a neuroanatomical nature) we would like to emphasize concerns the need for a more detailed mapping of the different DA-receptor-containing receptor complexes to better understand their distribution in the dopaminergic pathways and to better characterize their location at the cellular level. In this regard, of particular interest would be the study of the descending dopamine pathways, since almost no data concerning the distribution of receptor complexes containing DA receptors in these districts have been obtained so far. A second point (of a pharmacological nature) involves a more detailed assessment of how typical and atypical neuropsychiatric drugs may act on the different receptor complexes in order to optimize existing pharmacological treatments or to develop completely new pharmacological strategies. In this respect, however, the development of receptor-complex-specific ligands appears another very promising strategy. Indeed, the possibility to develop bivalent ligands [187] or to exploit allosteric modulators that are selective for structural domains in the heteroreceptor complexes [129,130] has been demonstrated.

Finally, it should be noted that the research effort to identify and characterize RRI and receptor complexes has been mainly focused on neurons, given that available data on RRI and on receptor complexes in astrocytes are more limited. However, a more intense effort in pharmacological research applied to receptor complexes in astrocytes may represent a topic of particular interest, not only to reach a better understanding of the role of neuron–astrocyte crosstalk in dopaminergic systems, but also from a therapeutical standpoint. Such a research effort, indeed, may open the possibility of exploring novel, glia-mediated strategies to address neurodegenerative and functional DA-related disorders (see [141]).

Author Contributions: Conceptualization, D.G. and L.F.A.; data collection, C.T., M.M. and C.C.; writing—original draft preparation, D.G.; writing—review and editing, G.M., R.D.C. and L.F.A. All authors have read and agreed to the published version of the manuscript.

Funding: This research received no external funding.

Institutional Review Board Statement: Not applicable.

Informed Consent Statement: Not applicable.

Data Availability Statement: Not applicable.

Conflicts of Interest: The authors declare no conflict of interest.

References

1. Dahlström, A.; Fuxe, K. A method for the demonstration of monoamine-containing nerve fibres in the central nervous system. *Acta Physiol.* **1964**, *60*, 293–294. [[CrossRef](#)] [[PubMed](#)]
2. Andén, N.-E.; Carlsson, A.; Dahlström, A.; Fuxe, K.; Hillarp, N.A.; Larsson, K. Demonstration and mapping out of nigro-striatal dopamine neurons. *Life Sci.* **1964**, *3*, 523–530. [[CrossRef](#)] [[PubMed](#)]
3. Bentivoglio, M.; Morelli, M. The organization and circuits of mesencephalic dopaminergic neurons and the distribution of dopamine receptors in the brain. In *Dopamine*; Dunnet, S.B., Bentivoglio, M., Björklund, A., Hökfelt, T., Eds.; Handbook of Chemical Neuroanatomy; Elsevier: Amsterdam, The Netherlands, 2005; Volume 21, pp. 1–107.
4. Hurd, Y.L.; Hall, H. Human forebrain dopamine systems: Characterization of the normal brain and in relation to psychiatric disorders. In *Dopamine*; Dunnet, S.B., Bentivoglio, M., Björklund, A., Hökfelt, T., Eds.; Handbook of Chemical Neuroanatomy; Elsevier: Amsterdam, The Netherlands, 2005; Volume 21, pp. 525–571.
5. Albanese, A.; Altavista, M.C.; Rossi, P. Organization of central nervous system dopaminergic pathways. *J. Neural Transm.* **1986**, *22*, 3–17.
6. Klein, M.O.; Battagello, D.S.; Cardoso, A.R.; Hauser, D.N.; Bittencourt, J.C.; Correa, R.G. Dopamine: Function, signaling, and association with neurological diseases. *Cell. Mol. Neurobiol.* **2019**, *39*, 31–59.
7. Fuxe, K.; Rivera, A.; Jacobsen, K.X.; Höistad, M.; Leo, G.; Horvath, T.L.; Staines, W.; De la Calle, A.; Agnati, L.F. Dynamics of volume transmission in the brain. Focus on catecholamine and opioid peptide communication and the role of uncoupling protein 2. *J. Neural Transm.* **2005**, *112*, 65–76. [[CrossRef](#)]
8. Agnati, L.F.; Guidolin, D.; Guescini, M.; Genedani, S.; Fuxe, K. Understanding wiring and volume transmission. *Brain Res. Rev.* **2010**, *64*, 137–159.
9. Guidolin, D.; Marcoli, M.; Maura, G.; Agnati, L.F. New dimensions of connectomics and network plasticity in the central nervous system. *Rev. Neurosci.* **2017**, *28*, 113–132.
10. Eid, L.; Parent, M. Chemical anatomy of pallidal afferents in primates. *Brain Struct. Funct.* **2016**, *221*, 4291–4317.
11. Rice, M.E.; Cragg, S.J. Dopamine spillover after quantal release: Rethinking dopamine transmission in the nigrostriatal pathway. *Brain Res. Rev.* **2008**, *58*, 303–313. [[CrossRef](#)]
12. Rice, M.E.; Patel, J.C. Somatodendritic dopamine release: Recent mechanistic insights. *Phil. Trans. R. Soc. Lond. B Biol. Sci.* **2015**, *370*, 20140185. [[CrossRef](#)]
13. Goto, Y.; Otani, S.; Grace, A.A. The yin and yang of dopamine release: A new perspective. *Neuropharmacology* **2007**, *53*, 583–587. [[CrossRef](#)] [[PubMed](#)]
14. Floresco, S.B. Dopaminergic regulation of limbic-striatal interplay. *J. Psychiatry Neurosci.* **2007**, *32*, 400–411. [[PubMed](#)]
15. Hirasawa, H.; Contini, M.; Raviola, E. Extrasynaptic release of GABA and dopamine by retinal dopaminergic neurons. *Phylos. Trans. R. Soc. Lond. B Biol. Sci.* **2015**, *370*, 20140186. [[CrossRef](#)] [[PubMed](#)]
16. Miyazaki, I.; Asanuma, M.; Diaz-Corrales, F.J.; Miyoshi, K.; Ogawa, N. Direct evidence for expression of dopamine receptors in astrocytes from basal ganglia. *Brain Res.* **2004**, *1029*, 120–123. [[CrossRef](#)] [[PubMed](#)]
17. Mladinov, M.; Mayer, D.; Brčić, L.; Wolstencroft, E.; thi Man, N.; Holt, I.; Hof, P.R.; Morris, G.E.; Šimic, G. Astrocyte expression of D2-like dopamine receptors in the prefrontal cortex. *Transl. Neurosci.* **2010**, *1*, 238–243. [[CrossRef](#)]
18. Montoya, A.; Elgueta, D.; Campos, J.; Chovar, O.; Falcón, P.; Matus, S.; Alfaro, I.; Bono, M.R.; Pacheco, R. Dopamine receptor D3 signalling in astrocytes promotes neuroinflammation. *J. Neuroinflamm.* **2019**, *16*, 258. [[CrossRef](#)]
19. Guidolin, D.; Tortorella, C.; Marcoli, M.; Cervetto, C.; Maura, G.; Agnati, L.F. Receptor-receptor interactions and microvesicle exchange as mechanisms modulating signaling between neurons and astrocytes. *Neuropharmacology* **2023**, *231*, 109509.
20. Keibarian, J.W.; Petzold, G.L.; Greengard, P. Dopamine-sensitive adenylate cyclase in caudate nucleus of rat brain, and its similarity to the “dopamine receptor”. *Proc. Natl. Acad. Sci. USA* **1972**, *69*, 2145–2149. [[CrossRef](#)]
21. Burt, D.R.; Enna, S.J.; Creese, I.; Snyder, S.H. Dopamine receptor binding in the corpus striatum of mammalian brain. *Proc. Natl. Acad. Sci. USA* **1975**, *72*, 4655–4659. [[CrossRef](#)]
22. Seeman, P.; Chau-Wong, M.; Tedesco, J.; Wong, K. Brain receptors for antipsychotic drugs and dopamine: Direct binding assays. *Proc. Natl. Acad. Sci. USA* **1975**, *72*, 4376–4380. [[CrossRef](#)]
23. Fuxe, K.; Agnati, L.F.; Benfenati, F.; Celani, M.; Zini, I.; Zoli, M.; Mutt, V. Evidence for the existence of receptor-receptor interactions in the central nervous system. Studies on the regulation of monoamine receptors by neuropeptides. In *Basic Aspects of Receptor Biochemistry*; Springer: Vienna, Austria, 1983; Volume 18, pp. 165–179.
24. Bockaert, J.; Pin, J.P. Molecular tinkering of G protein coupled receptors: An evolutionary success. *EMBO J.* **1999**, *18*, 1723–1729. [[CrossRef](#)] [[PubMed](#)]
25. Marshall, F.H.; White, J.; Main, M.; Green, A.; Wise, A. GABA(B) receptors function as heterodimers. *Biochem. Soc. Trans.* **1999**, *27*, 530–535. [[CrossRef](#)] [[PubMed](#)]

26. Xie, Z.; Lee, S.P.; O'Dowd, B.F.; George, S.R. Serotonin 5-HT_{1B} and 5-HT_{1D} receptors form homodimers when expressed alone and heterodimers when co-expressed. *FEBS Lett.* **1999**, *456*, 63–67. [[CrossRef](#)] [[PubMed](#)]
27. Lee, S.P.; Xie, Z.; Varghese, G.; Nguyen, T.; O'Dowd, B.F.; George, S.R. Oligomerization of dopamine and serotonin receptors. *Neuropsychopharmacology* **2000**, *23*, S32–S40. [[CrossRef](#)]
28. Overton, M.C.; Blumer, K.J. G protein-coupled receptors function as oligomers in vivo. *Curr. Biol.* **2000**, *10*, 341–344. [[CrossRef](#)]
29. Zeng, F.; Wess, J. Molecular aspects of muscarinic receptor dimerization. *Neuropsychopharmacology* **2000**, *23*, S19–S31. [[CrossRef](#)]
30. Angers, S.; Salahpour, A.; Bouvier, M. Biochemical and biophysical demonstration of GPCR oligomerization in mammalian cells. *Life Sci.* **2001**, *68*, 2243–2250. [[CrossRef](#)]
31. Dean, M.K.; Higgs, C.; Smith, R.E.; Bywater, R.P.; Snell, C.R.; Scott, P.D.; Upton, G.J.; Howe, T.J.; Reynolds, C.A. Dimerization of G protein-coupled receptors. *J. Med. Chem.* **2001**, *44*, 4595–4614.
32. Kenakin, T. Drug efficacy at G protein-coupled receptors. *Annu. Rev. Pharmacol. Toxicol.* **2002**, *42*, 349–379. [[CrossRef](#)]
33. Waldhoer, M.; Fong, J.; Jones, R.M.; Lunzer, M.M.; Sharma, S.K.; Kostenis, E.; Portoghese, P.S.; Whistler, J.L. A heterodimer-selective agonist shows in vivo relevance of G protein-coupled receptor dimers. *Proc. Natl. Acad. Sci. USA* **2005**, *102*, 9050–9055. [[CrossRef](#)]
34. Guidolin, D.; Marcoli, M.; Tortorella, C.; Maura, G.; Agnati, L.F. Receptor-receptor interactions as a widespread phenomenon: Novel targets for drug development? *Front. Endocrinol.* **2019**, *10*, 53.
35. Changeux, J.P.; Christopoulos, A. Allosteric modulation as a unifying mechanism for receptor function and regulation. *Diabetes Obes. Metabol.* **2017**, *19*, 4–21. [[CrossRef](#)] [[PubMed](#)]
36. Baik, J.-H. Dopamine Signaling in reward-related behaviors. *Front. Neural Circuits* **2013**, *7*, 152. [[CrossRef](#)] [[PubMed](#)]
37. Missale, C.; Nash, S.R.; Robinson, S.W.; Jaber, M.; Caron, M.G.; Wishart, D.S.; Anselmi, L.; Toti, L.; Bove, C.; Prakash, Y.S.; et al. Dopamine receptors: From structure to function. *Physiol. Rev.* **1998**, *78*, 189–225.
38. Beaulieu, J.-M.; Gainetdinov, R.R. The physiology, signaling, and pharmacology of dopamine receptors. *Pharmacol. Rev.* **2011**, *63*, 182–217.
39. Tritsch, N.X.; Sabatini, B.L. Dopaminergic modulation of synaptic transmission in cortex and striatum. *Neuron* **2012**, *76*, 33–50.
40. Gingrich, J.A.; Caron, M.G. Recent advances in the molecular biology of dopamine receptors. *Annu. Rev. Neurosci.* **1993**, *16*, 299–321. [[CrossRef](#)]
41. Dal Toso, R.; Sommer, B.; Ewert, M.; Herb, A.; Pritchett, D.B.; Bach, A.; Shivers, B.D.; Seeburg, P.H. The dopamine D2 receptor: Two molecular forms generated by alternative splicing. *EMBO J.* **1989**, *8*, 4025–4034. [[CrossRef](#)]
42. Giros, B.; Sokoloff, P.; Martres, M.P.; Riou, J.-F.; Emorine, L.J.; Schwartz, J.-C. Alternative splicing directs the expression of two D2 dopamine receptor isoforms. *Nature* **1989**, *342*, 923–926. [[CrossRef](#)]
43. Kim, K.-M. Unveiling the differences in signaling and regulatory mechanisms between dopamine D₂ and D₃ receptors and their impact on behavioral sensitization. *Int. J. Mol. Sci.* **2023**, *24*, 6742.
44. Jose, P.A.; Yu, P.-Y.; Yamapchi, I.; Eisner, G.M.; Mouradian, M.M.; Felder, C.C.; Felder, R.A. Dopamine D1 receptor regulation of phospholipase C. *Hypertens. Res.* **1995**, *18* (Suppl. 1), S39–S42. [[CrossRef](#)]
45. Rashid, A.; O'Dowd, B.F.; Verma, V.; George, S.R. Neuromal Gq/11-coupled dopamine receptors: An uncharted role for dopamine. *Trends Pharmacol. Sci.* **2007**, *28*, 551–555. [[CrossRef](#)]
46. Sahu, A.; Tyeryar, K.R.; Vongtau, H.O.; Sibley, D.R.; Undieh, A.S. D5 dopamine receptors are required for dopaminergic activation of phospholipase C. *Mol. Pharmacol.* **2009**, *75*, 447–453. [[CrossRef](#)] [[PubMed](#)]
47. Felder, C.C.; Jose, P.A.; Axelrod, J. The dopamine-1 agonist, SKF 82526, stimulates phospholipase-C activity independent of adenylate cyclase. *J. Pharmacol. Exp. Ther.* **1989**, *248*, 171–175. [[PubMed](#)]
48. Kofuji, P.; Araque, A. G-protein-coupled receptors in astrocyte-neuron communication. *Neuroscience* **2021**, *456*, 71–84. [[PubMed](#)]
49. Oda, S.; Funato, H. D1- and D2-type dopamine receptors are immunolocalized in pial and layer I astrocytes in the rat cerebral cortex. *Front. Neuroanat.* **2023**, *17*, 1111008. [[CrossRef](#)] [[PubMed](#)]
50. Jennings, A.; Tyurikova, O.; Bard, L.; Zheng, K.; Semyanov, A.; Henneberger, C.; Rusakov, D.A. Dopamine elevates and lowers astroglial Ca²⁺ through distinct pathways depending on local synaptic circuitry. *Glia* **2017**, *65*, 447–459. [[CrossRef](#)]
51. Xin, W.; Schuebel, K.E.; Jair, K.W.; Cimbrow, R.; De Biase, L.M.; Goldman, D.; Bonci, A. Ventral midbrain astrocytes display unique physiological features and sensitivity to dopamine D2 receptor signaling. *Neuropsychopharmacology* **2019**, *44*, 344–355.
52. Corkrum, M.; Covelo, A.; Lines, J.; Bellocchio, L.; Pisansky, M.; Loke, K.; Quintana, R.; Rothwell, P.E.; Lujan, R.; Marsicano, G.V.; et al. Dopamine-evoked synaptic regulation in the nucleus accumbens requires astrocyte activity. *Neuron* **2020**, *105*, 1036–1047.
53. Li, C.; Saliba, N.B.; Martin, H.; Losurdo, N.A.; Kolahdouzan, K.; Siddiqui, R.; Medeiros, D.; Li, W. Purkinje cell dopaminergic inputs to astrocytes regulate cerebellar-dependent behavior. *Nat. Commun.* **2023**, *14*, 1613.
54. Gurevich, V.V.; Gurevich, E.V. The structural basis of arrestin-mediated regulation of G-protein-coupled receptors. *Pharmacol. Ther.* **2006**, *110*, 465–502. [[PubMed](#)]
55. Komolov, K.E.; Benovic, J.L. G protein-coupled receptor kinases: Past, present and future. *Cell. Signal.* **2018**, *41*, 17–24. [[PubMed](#)]
56. Pitcher, J.A.; Freedman, N.J.; Lefkowitz, R.J. G protein-coupled receptor kinases. *Annu. Rev. Biochem.* **1998**, *67*, 653–692.
57. Lohse, M.J.; Benovic, J.L.; Codina, J.; Caron, M.G.; Lefkowitz, R.J. beta-Arrestin: A protein that regulates beta-adrenergic receptor function. *Science* **1990**, *248*, 1547–1550. [[CrossRef](#)] [[PubMed](#)]
58. Luttrell, L.M.; Lefkowitz, R.J. The role of beta-arrestins in the termination and transduction of G-protein-coupled receptor signals. *J. Cell Sci.* **2002**, *115*, 455–465. [[CrossRef](#)]

59. Hollinger, S.; Hepler, J.R. Cellular regulation of RGS proteins: Modulators and integrators of G protein signaling. *Pharmacol. Rev.* **2002**, *34*, 527–559. [[CrossRef](#)]
60. Woodard, G.E.; Jardin, I.; Berna-Erro, A.; Salido, G.M.; Rosado, J.A. Regulators of G-protein-signaling proteins: Negative modulators of G-protein-coupled receptor signaling. *Int. Rev. Cell Mol. Biol.* **2015**, *317*, 97–183.
61. Kovoov, A.; Seyffarth, P.; Ebert, J.; Barghshoon, S.; Chen, C.-K.; Schwarz, S.; Axelrod, J.D.; Cheyette, B.N.R.; Simon, M.I.; Lester, H.A.; et al. D2 dopamine receptors colocalize regulator of G-protein signaling 9-2 (RGS9-2) via the RGS9 DEP domain, and RGS9 knock-out mice develop dyskinesias associated with dopamine pathways. *J. Neurosci.* **2005**, *25*, 2157–2165. [[CrossRef](#)]
62. Cerver, J.; Sharma, M.; Kovoov, A. RGS9-2 mediates specific inhibition of agonist-induced internalization of D2-dopamine receptors. *J. Neurochem.* **2010**, *114*, 739–749. [[CrossRef](#)]
63. Fuxe, K.; Canals, M.; Torvinen, M.; Marcellino, D.; Terasmaa, A.; Genedani, S.; Leo, G.; Guidolin, D.; Diaz-Cabiale, Z.; Rivera, A.; et al. Intramembrane receptor-receptor interactions: A novel principle in molecular medicine. *J. Neural Transm.* **2007**, *114*, 49–75.
64. Guidolin, D.; Tortorella, C.; Marcoli, M.; Maura, G.; Agnati, L.F. Intercellular communication in the central nervous system as deduced by chemical neuroanatomy and quantitative analysis of images: Impact on neuropharmacology. *Int. J. Mol. Sci.* **2022**, *23*, 5805.
65. Prezeau, L.; Rives, M.L.; Comps-Agrar, L.; Maurel, D.; Knlazeff, J.; Pin, J.P. Functional crosstalk between GPCRs: With or without oligomerization. *Curr. Opin. Pharm.* **2010**, *10*, 6–13. [[CrossRef](#)] [[PubMed](#)]
66. Agnati, L.F.; Fuxe, K.; Giardino, L.; Calzà, L.; Zoli, M.; Battistini, N.; Benfenati, F.; Vanderhaeghen, J.J.; Guidolin, D.; Ruggeri, M. Evidence for cholecystokinin-dopamine receptor interactions in the central nervous system of the adult and old rat. Studies on their functional meaning. *Ann. N. Y. Acad. Sci.* **1985**, *448*, 315–333. [[CrossRef](#)] [[PubMed](#)]
67. Kenakin, T.; Agnati, L.F.; Caron, M.; Fredholm, B.; Guidolin, D.; Kobilka, B.; Lefkowitz, R.W.; Lohse, M.; Woods, A.; Fuxe, K. International workshop at the Nobel Forum, Karolinska Institutet, on G protein-coupled receptors: Finding the words to describe monomers, oligomers, and their molecular mechanisms and defining their meaning. Can a consensus be reached? *J. Recept. Signal Transduct. Res.* **2010**, *30*, 284–286. [[CrossRef](#)] [[PubMed](#)]
68. Trifilieff, P.; Rives, M.L.; Urizar, E.; Piskowski, R.A.; Vishwasrao, H.D.; Castrillon, J.; Schmauss, C.; Stätman, M.; Gullberg, M.; Javitch, J.A. Detection of antigen interactions ex vivo by proximity ligation assay: Endogenous dopamine D2-adenosine A2A receptor complexes in the striatum. *Biotechniques* **2011**, *51*, 111–118. [[CrossRef](#)]
69. Fernández-Dueñas, V.; Gómez-Soler, M.; Valle-León, M.; Watanabe, M.; Ferrer, I.; Ciruela, F. Revealing adenosine A2A-dopamine D2 receptor heteromers in Parkinson's disease post-mortem brain through a new AlphaScreen-based approach. *Int. J. Mol. Sci.* **2019**, *20*, 3600. [[CrossRef](#)]
70. Petazzi, R.A.; Aji, A.K.; Chiantia, S. Fluorescence microscopy methods for the study of protein oligomerization. In *Progress in Molecular Biology and Translational Science*; Giraldo, J., Ciruela, F., Eds.; Academic Press: Cambridge, MA, USA, 2020; Volume 169, pp. 1–42.
71. De Oliveira, P.; Moreno, E.; Casajuana-Martin, N.; Casadó-Anguera, V.; Cai, N.-S.; Camacho-Hernandez, G.A.; Zhu, H.; Bonifazi, A.; Hall, M.D.; Weinshenker, D.; et al. Preferential Gs protein coupling of the galanin Gal₁ receptor in the μ -opioid-Gal₁ receptor heterotetramer. *Pharmacol. Res.* **2022**, *182*, 106322. [[CrossRef](#)]
72. Franco, R.; Martínez-Pinilla, E.; Lanciego, J.L.; Navarro, G. Basic pharmacological and structural evidence for class A G-protein-coupled receptor heteromerization. *Front. Pharmacol.* **2016**, *7*, 76.
73. Changeux, J.P. The origins of allostery: From personal memories to material for the future. *J. Mol. Biol.* **2013**, *425*, 1396–1406.
74. Kenakin, T.; Miller, I.J. Seven transmembrane receptors as shape shifting proteins: The impact of allosteric modulation and functional selectivity on new drug discovery. *Pharm. Rev.* **2010**, *62*, 265–304.
75. Smith, N.J.; Milligan, G. Allostery of G protein-coupled receptors homo- and heteromers: Uncharted pharmacological landscapes. *Pharm. Rev.* **2010**, *62*, 701–725. [[PubMed](#)]
76. Liu, J.; Nussinov, R. Allostery: An overview of its history, concepts, methods and applications. *PLoS Comput. Biol.* **2016**, *12*, e1004966. [[CrossRef](#)] [[PubMed](#)]
77. Ferré, S.; Ciruela, F.; Woods, A.S.; Lluís, C.; Franco, R. Functional relevance of neurotransmitter receptor heteromers in the central nervous system. *Trends Neurosci.* **2007**, *30*, 440–446. [[CrossRef](#)] [[PubMed](#)]
78. Filizola, M.; Weinstein, H. The study of G-protein coupled receptor oligomerization with computational modeling and bioinformatics. *FEBS J.* **2005**, *272*, 2926–2938. [[PubMed](#)]
79. Simpson, L.M.; Taddese, B.; Wall, I.D.; Reynolds, C.A. Bioinformatics and molecular modelling approaches to GPCR oligomerization. *Curr. Opin. Pharm.* **2010**, *10*, 30–37. [[CrossRef](#)] [[PubMed](#)]
80. Guidolin, D.; Ciruela, F.; Genedani, S.; Guescini, M.; Tortorella, C.; Albertin, G.; Fuxe, K.; Agnati, L.F. Bioinformatics and mathematical modeling in the study of receptor-receptor interactions and receptor oligomerization. Focus on adenosine receptors. *Biochim. Biophys. Acta* **2011**, *1808*, 1267–1283. [[CrossRef](#)]
81. Borroto-Escuela, D.O.; Tarakanov, A.O.; Brito, I.; Fuxe, K. Glutamate heteroreceptor complexes in the brain. *Pharmacol. Rep.* **2018**, *70*, 936–950.
82. Pinna, A.; Bonaventura, J.; Farré, D.; Sánchez, M.; Simola, N.; Mallol, J.; Lluís, C.; Costa, G.; Baqi, Y.; Müller, C.E.; et al. L-DOPA disrupts adenosine A(2A)-cannabinoid CB(1)-dopamine D(2) receptor heteromer cross-talk in the striatum of hemiparkinsonian rats: Biochemical and behavioral studies. *Exp. Neurol.* **2014**, *253*, 180–191. [[CrossRef](#)]

83. Cabello, N.; Gandia, J.; Bertarelli, D.C.; Watanabe, M.; Lluís, C.; Franco, R.; Ferré, S.; Luján, R.; Ciruela, F. Metabotropic glutamate type 5, dopamine D2 and adenosine A2a receptors form higher-order oligomers in living cells. *J. Neurochem.* **2009**, *109*, 1497–1507. [[CrossRef](#)]
84. Beggiano, S.; Tomasini, M.C.; Borelli, A.C.; Borroto-Escuela, D.O.; Fuxe, K.; Antonelli, T.; Tanganelli, S.; Ferraro, L. Functional role of striatal A2A, D2, and mGlu5 receptor interactions in regulating striatopallidal GABA neuronal transmission. *J. Neurochem.* **2016**, *138*, 254–264. [[CrossRef](#)]
85. Farran, B. An update on the physiological and therapeutic relevance of GPCR oligomers. *Pharmacol. Res.* **2017**, *117*, 303–327. [[PubMed](#)]
86. Gainetdinov, R.R.; Premont, R.T.; Bohn, L.M.; Lefkowitz, R.J.; Caron, M.G. Desensitization of G protein-coupled receptors and neural functions. *Annu. Rev. Neurosci.* **2004**, *27*, 107–144. [[CrossRef](#)] [[PubMed](#)]
87. Smith, J.S.; Rajagopal, S. The β -arrestin: Multifunctional regulators of G protein-coupled receptors. *J. Biol. Chem.* **2016**, *291*, 8969–8977. [[PubMed](#)]
88. Agnati, L.F.; Leo, G.; Genedani, S.; Andreoli, N.; Marcellino, D.; Woods, A.; Piron, L.; Guidolin, D.; Fuxe, K. Structural plasticity in G-protein coupled receptors as demonstrated by the allosteric actions of homocysteine and computer-assisted analysis of disordered domains. *Brain Res. Rev.* **2008**, *58*, 459–474. [[PubMed](#)]
89. Agnati, L.F.; Guidolin, D.; Vilardaga, J.P.; Ciruela, F.; Fuxe, K. On the expanding terminology in the GPCR field: The meaning of receptor mosaics and receptor heteromers. *J. Recept. Signal Transduct. Res.* **2010**, *30*, 287–303. [[PubMed](#)]
90. Alemany, R.; Perona, J.S.; Sánchez-Dominguez, J.M.; Montero, E.; Cañizares, J.; Bressani, R.; Escribà, P.V.; Ruiz-Gutierrez, V.G. protein-coupled receptor systems and their lipid environment in health disorders during aging. *Biochim. Biophys. Acta* **2007**, *1768*, 964–975.
91. Foord, S.M.; Jupe, S.; Holbrook, J. Bioinformatics and type II G-protein-coupled receptors. *Biochem. Soc. Trans.* **2002**, *30*, 473–479. [[CrossRef](#)]
92. Whorton, M.R.; Bokoch, M.P.; Rasmussen, S.G.F.; Huang, B.; Zare, R.N.; Kobilka, B.; Sunahara, R.K. A monomeric G protein-coupled receptor isolated in a high-density lipoprotein particle efficiently activates its G protein. *Proc. Natl. Acad. Sci. USA* **2007**, *104*, 7682–7687.
93. Gurevich, V.V.; Gurevich, E.V. How and why do GPCRs dimerize? *Trends Pharmacol. Sci.* **2008**, *29*, 234–240.
94. Teichmann, A.; Gibert, A.; Lampe, A.; Grzesik, P.; Rutz, C.; Furkert, J.; Schmoranzner, J.; Krause, G.; Wiesner, B.; Schüle, R. The specific monomer/dimer equilibrium of the corticotropin-releasing factor receptor type 1 is established in the endoplasmic reticulum. *J. Biol. Chem.* **2014**, *289*, 24250–24262. [[CrossRef](#)]
95. Fuxe, K.; Marcellino, D.; Guidolin, D.; Woods, A.; Agnati, L.F. Dopamine Receptor Oligomerization. In *The Dopamine Receptors*; Neve, K.A., Ed.; Humana Press: Totowa, NJ, USA; Springer: Berlin/Heidelberg, Germany, 2010; pp. 255–280.
96. Pelassa, S.; Guidolin, D.; Venturini, A.; Averna, M.; Frumento, G.; Campanini, L.; Bernardi, R.; Cortelli, P.; Buonauro, G.C.; Maura, G.; et al. A2A-D2 heteromers on striatal astrocytes: Biochemical and biophysical evidence. *Int. J. Mol. Sci.* **2019**, *20*, 2457. [[CrossRef](#)] [[PubMed](#)]
97. Torvinen, M.; Marcellino, D.; Canals, M.; Agnati, L.F.; Lluís, C.; Franco, R.; Fuxe, K. Adenosine a2a receptor and dopamine d3 receptor interactions: Evidence of functional a2a/d3 heteromeric complexes. *Mol. Pharmacol.* **2005**, *67*, 400–407. [[PubMed](#)]
98. Fuxe, K.; Borroto-Escuela, D.O. *Receptor-Receptor Interactions in the Central Nervous System*; Humana Press: New York, NY, USA, 2018; Volume 140, p. 346.
99. Liu, X.Y.; Chu, X.P.; Mao, L.M.; Wang, M.; Lan, H.X.; Li, M.H.; Zhang, G.C.; Parelkar, N.K.; Fibuch, E.E.; Haines, M.; et al. Modulation of D2R–NR2B interactions in response to cocaine. *Neuron* **2006**, *52*, 897–909. [[PubMed](#)]
100. Koschätzky, S.; Tschammer, N.; Gmeiner, P. Cross-receptor interactions between dopamine D2L and neurotensin NTS1 receptors modulate binding affinities of dopaminergics. *ACS Chem. Neurosci.* **2011**, *2*, 308–316.
101. Plach, M.; Schäfer, T.; Borroto-Escuela, D.O.; Weickert, D.; Gmeiner, P.; Fuxe, K.; Friedland, K. Differential allosteric modulation within dopamine D₂R–neurotensin NTS1R and D₂R–serotonin 5-HT_{2A}R receptor complexes gives bias to intracellular calcium signaling. *Sci. Rep.* **2019**, *9*, 16312.
102. Przybyla, J.A.; Watts, V.J. Ligand-induced regulation and localization of cannabinoid CB1 and dopamine D2L receptor heterodimers. *J. Pharmacol. Exp. Ther.* **2010**, *332*, 710–719. [[CrossRef](#)]
103. Kolasa, M.; Solich, J.; Faron-Górecka, A.; Żurawek, D.; Pabian, P.; Łukasiewicz, S.; Kuśmider, M.; Szafran-Pilch, K.; Szlachta, M.; Dziejzicka-Wasylewska, M. Paroxetine and Low-dose Risperidone Induce Serotonin 5-HT_{1A} and Dopamine D2 Receptor Heteromerization in the Mouse Prefrontal Cortex. *Neuroscience* **2018**, *377*, 184–196.
104. Borroto-Escuela, D.O.; Romero-Fernandez, W.; Tarakanov, A.O.; Marcellino, D.; Ciruela, F.; Agnati, L.F.; Fuxe, K. Dopamine D2 and 5-hydroxytryptamine 5-HT_{2A} receptors assemble into functionally interacting heteromers. *Biochem. Biophys. Res. Commun.* **2010**, *401*, 605–610. [[CrossRef](#)]
105. Romero-Fernandez, W.; Borroto-Escuela, D.O.; Agnati, L.F.; Fuxe, K. Evidence for the existence of dopamine D2–oxytocin receptor heteromers in the ventral and dorsal striatum with facilitatory receptor–receptor interactions. *Mol. Psychiatry* **2013**, *18*, 849–850. [[CrossRef](#)]
106. Amato, S.; Averna, M.; Guidolin, D.; Ceccoli, C.; Gatta, E.; Candiani, S.; Pedrazzi, M.; Capraro, M.; Maura, G.; Agnati, L.F.; et al. Heteromerization of Dopamine D2 and Oxytocin Receptor in Adult Striatal Astrocytes. *Int. J. Mol. Sci.* **2023**, *24*, 4677.

107. Kern, A.; Albarran-Zeckler, R.; Walsh, H.E.; Smith, R.G. Apo-grelin receptor forms heteromers with DRD2 in hypothalamic neurons and is essential for anorexigenic effects of DRD2 agonism. *Neuron* **2012**, *73*, 317–332. [[CrossRef](#)] [[PubMed](#)]
108. Ferré, S.; Becher, A.M.; Bonaventura, J.; Quiroz, C.; Sanchez-Soto, M.; Casadó-Anguera, V.; Cai, N.-S.; Moreno, E.; Boateng, C.A.; Keck, T.M.; et al. Functional and pharmacological role of the dopamine D4 receptor and its polymorphic variants. *Front. Endocrinol.* **2022**, *13*, 1014678.
109. Rebois, R.V.; Maki, K.; Meeks, J.A.; Fishman, P.H.; Hébert, T.E.; Northup, J.K. D2-like dopamine and β -adrenergic receptors form a signaling complex that integrates Gs- and Gi-mediated regulation of adenylyl cyclase. *Cell. Signal.* **2012**, *24*, 2051–2060.
110. Gago, B.; Fuxe, K.; Agnati, L.; Penafiel, A.; De La Calle, A.; Rivera, A. Dopamine D(4) receptor activation decreases the expression of mu-opioid receptors in the rat striatum. *J. Comp. Neurol.* **2007**, *502*, 358–366. [[PubMed](#)]
111. Petkova-Kirova, P.; Giovannini, M.G.; Kalfin, R.; Rakovska, A. Modulation of acetylcholine release by cholecystokinin in striatum: Receptor specificity; role of dopaminergic neuronal activity. *Brain Res. Bull.* **2012**, *89*, 177–184.
112. Pittolo, S.; Yokoyama, S.; Willoughby, D.D.; Taylor, C.R.; Reitman, M.E.; Tse, V.; Wu, Z.; Etchenique, R.; Li, Y.; Poskanzer, K.E. Dopamine activates astrocytes in prefrontal cortex via α 1-adrenergic receptors. *Cell Rep.* **2022**, *40*, 111426.
113. Pinton, L.; Borroto-Escuela, D.O.; Narváez, M.; Oflijan, J.; Agnati, L.F.; Fuxe, K. Evidence for the existence of dopamine D2R and Sigma 1 allosteric receptor–receptor interaction in the rat brain: Role in brain plasticity and cocaine action. *Springerplus* **2015**, *4*, P37. [[CrossRef](#)]
114. Rashid, A.J.; So, C.H.; Kong, M.M.C.; Furtak, T.; El-Ghundi, M.; Cheng, R.; O’Dowd, B.F.; George, S.R. D1-D2 dopamine receptor heterooligomers with unique pharmacology are coupled to rapid activation of Gq/11 in the striatum. *Proc. Natl. Acad. Sci. USA* **2007**, *104*, 654–659. [[CrossRef](#)]
115. Marcellino, D.; Ferré, S.; Casado, V.; Cortés, A.; Le Foll, B.; Mazzola, C.; Drago, F.; Saur, O.; Stark, H.; Soriano, A.; et al. Identification of dopamine D1–D3 receptor heteromers: Indications for a role of synergistic D1–D3 receptor interactions in the striatum. *J. Biol. Chem.* **2008**, *283*, 26016–26025.
116. Ginés, S.; Hillion, J.; Torvinen, M.; Le Crom, S.; Casadó, V.; Canela, E.I.; Rondin, S.; Lew, J.Y.; Watson, S.; Zoli, M.; et al. Dopamine D1 and adenosine A1 receptors form functionally interacting heteromeric complexes. *Proc. Natl. Acad. Sci. USA* **2000**, *97*, 8606–8611. [[CrossRef](#)]
117. Franco, R.; Lluis, C.; Canela, E.I.; Mallol, J.; Agnati, L.; Casadó, V.; Ciruela, F.; Ferré, S.; Fuxe, K. Receptor–receptor interactions involving adenosine A1 or dopamine D1 receptors and accessory proteins. *J. Neural Transm.* **2007**, *114*, 93–104. [[PubMed](#)]
118. Lee, F.J.; Xue, S.; Pei, L.; Vukusic, B.; Chéry, N.; Wang, Y.; Wang, Y.T.; Niznik, H.B.; Yu, X.-M.; Liu, F. Dual regulation of NMDA receptor functions by direct protein–protein interactions with the dopamine D1 receptor. *Cell* **2002**, *111*, 219–230. [[PubMed](#)]
119. Mitrano, D.A.; Pare, J.F.; Smith, Y.; Weinschenker, D. D1-dopamine and α 1-adrenergic receptors co-localize in dendrites of the rat prefrontal cortex. *Neuroscience* **2014**, *258*, 90–100.
120. Guidolin, D.; Marcoli, M.; Tortorella, C.; Maura, G.; Agnati, L.F. Adenosine A2A-Dopamine D2 Receptor-Receptor Interaction in Neurons and Astrocytes: Evidence and Perspectives. *Prog. Mol. Biol. Transl. Sci.* **2020**, *169*, 247–277. [[PubMed](#)]
121. Ciruela, F.; Burgueno, J.; Casado, V.; Canals, M.; Marcellino, D.; Goldberg, S.R.; Bader, M.; Fuxe, K.; Agnati, L.F.; Lluis, C.; et al. Combining mass spectrometry and pull-down techniques for the study of receptor heteromerization. Direct epitope–epitope electro- static interactions between adenosine A2A and dopamine D2 receptors. *Anal. Chem.* **2004**, *76*, 5354–5363. [[CrossRef](#)] [[PubMed](#)]
122. Woods, A.S.; Ciruela, F.; Fuxe, K.; Agnati, L.F.; Lluis, C.; Franco, R.; Ferré, S. Role of electrostatic interaction in receptor–receptor heteromerization. *J. Mol. Neurosci.* **2005**, *26*, 125–132. [[CrossRef](#)] [[PubMed](#)]
123. Borroto-Escuela, D.O.; Rodriguez, D.; Romero-Fernandez, W.; Kapla, J.; Jaiteh, M.; Ranganathan, A.; Lazarova, T.; Fuxe, K.; Carlsson, J. Mapping the interface of a GPCR dimer: A structural model of the A2A adenosine and D2 dopamine receptor heteromer. *Front. Pharmacol.* **2018**, *9*, 829.
124. Ferré, S.; von Euler, G.; Johansson, B.; Fredholm, B.B.; Fuxe, K. Stimulation of high-affinity adenosine A2 receptors decreases the affinity of dopamine D2 receptors in rat striatal membranes. *Proc. Natl. Acad. Sci. USA* **1991**, *88*, 7238–7241. [[CrossRef](#)]
125. Fuxe, K.; Ferré, S.; Zoli, M.; Agnati, L.F. Integrated events in central dopamine transmission as analyzed at multiple levels. Evidence for intramembrane adenosine A2A/dopamine D2 and adenosine A1/dopamine D1 receptor interactions in the basal ganglia. *Brain Res. Rev.* **1998**, *26*, 258–273. [[CrossRef](#)]
126. Diaz-Cabiale, Z.; Hurd, Y.; Guidolin, D.; Finnman, U.B.; Zoli, M.; Agnati, L.F.; Vanderhaeghen, J.J.; Fuxe, K.; Ferré, S. Adenosine A2A agonist CGS 21680 decreases the affinity of dopamine D2 receptors for dopamine in human striatum. *Neuroreport* **2001**, *12*, 1831–1834. [[CrossRef](#)]
127. Cervetto, C.; Venturini, A.; Passalacqua, M.; Guidolin, D.; Genedani, S.; Fuxe, K.; Borroto-Escuela, D.O.; Cortelli, P.; Woods, A.; Maura, G.; et al. A2A-D2 receptor–receptor interaction modulates gliotransmitter release from striatal astrocyte processes. *J. Neurochem.* **2016**, *140*, 268–279. [[CrossRef](#)] [[PubMed](#)]
128. Borroto-Escuela, D.; Romero-Fernandez, W.; Tarakanov, A.; Ciruela, F.; Agnati, L.; Fuxe, K. On the existence of a possible A2A-D2-beta-arrestin2 complex: A2A agonist modulation of D2 agonist-induced beta-arrestin2 recruitment. *J. Mol. Biol.* **2011**, *406*, 687–699. [[CrossRef](#)] [[PubMed](#)]
129. Agnati, L.F.; Ferré, S.; Genedani, S.; Leo, G.; Guidolin, D.; Filaferrero, M.; Carriba, P.; Casado, V.; Lluis, C.; Franco, R.; et al. Allosteric modulation of dopamine D2 receptors by homocysteine. *J. Proteome Res.* **2006**, *5*, 3077–3083. [[CrossRef](#)] [[PubMed](#)]

130. Cervetto, C.; Venturini, A.; Guidolin, D.; Maura, G.; Passalacqua, M.; Tacchetti, C.; Cortelli, P.; Genedani, S.; Candiani, S.; Ramoino, P.; et al. Homocysteine and A2A-D2 receptor-receptor interaction at striatal astrocyte processes. *J. Mol. Neurosci.* **2018**, *65*, 456–466. [[PubMed](#)]
131. Borgkvist, A.; Marcellino, D.; Fuxe, K.; Greengard, P.; Fisone, G. Regulation of DARPP-32 phosphorylation by Delta(9)-tetrahydrocannabinol. *Neuropharmacology* **2008**, *54*, 31–35. [[CrossRef](#)]
132. Lukaszewicz, S.; Błasiak, E.; Szafran-Pilch, K.; Dziedzicka-Wasylewska, M. Dopamine D2 and serotonin 5-HT1A receptor interaction in the context of the effects of antipsychotics—In Vitro studies. *J. Neurochem.* **2016**, *137*, 549–560. [[CrossRef](#)]
133. Fuxe, K.; Marcellino, D.; Rivera, A.; Diaz-cabiale, Z.; Filip, M.; Gago, B.; Roberts, D.C.S.; Lange, U.; Genedani, S.; Ferraro, L.; et al. Receptor-receptor interactions within receptor mosaics. Impact on neuropharmacology. *Brain Res. Rev.* **2008**, *58*, 415–452.
134. Ferré, S.; Sarasola, L.I.; Quiroz, C.; Ciruela, F. Presynaptic adenosine receptor heteromers as key modulators of glutamatergic and dopaminergic neurotransmission in the striatum. *Neuropharmacology* **2023**, *223*, 109329.
135. Liu, F.; Wan, Q.; Pristupa, Z.B.; Yu, X.M.; Wang, Y.T.; Niznik, H.B. Direct protein-protein coupling enables cross-talk between dopamine D5 and gamma-aminobutyric acid A receptors. *Nature* **2000**, *403*, 274–280. [[CrossRef](#)]
136. McNeill, J.; Rudyk, C.; Hildebrand, M.E.; Salmaso, N. Ion channels and electrophysiological properties of astrocytes: Implications for emergent stimulation technologies. *Front. Cell. Neurosci.* **2021**, *15*, 644126.
137. Fitzner, D.; Bader, J.M.; Penkert, H.; Bergner, C.G.; Su, M.; Weil, M.-T.; Surma, M.A.; Mann, M.; Klose, C.; Simons, M. Cell-type- and brain-region-resolved mouse brain lipidome. *Cell Rep.* **2020**, *32*, 108132. [[PubMed](#)]
138. Lee, J.; Patel, D.S.; Stähle, J.; Park, S.-J.; Kern, N.R.; Kim, S.; Lee, J.; Cheng, X.; Valvano, M.A.; Holst, O.; et al. CHARMM-GUI Membrane Builder for Complex Biological Membrane Simulations with glycolipids and lipoglycans. *Chem. Comput.* **2019**, *15*, 775–786. [[CrossRef](#)] [[PubMed](#)]
139. Kuriata, A.; Gierut, A.M.; Oleniecki, T.; Ciemny, M.P.; Kolinski, A.; Kurcinski, M.; Kmiecik, S. CABS-Flex 2.0: A Web Server for fast simulations of flexibility of protein structures. *Nucleic Acids Res.* **2018**, *46*, W338–W343. [[CrossRef](#)]
140. Borroto-Escuela, D.O.; Carlsson, J.; Ambrogini, P.; Narváez, M.; Wydra, K.; Tarakanov, A.D.; Li, X.; Millón, C.; Ferraro, L.; Cuppini, R.; et al. Understanding the role of GPCR heteroreceptor complexes in modulating the brain networks in health and disease. *Front. Cell. Neurosci.* **2017**, *11*, 37. [[PubMed](#)]
141. Guidolin, D.; Tortorella, C.; Marcoli, M.; Cervetto, C.; Maura, G.; Agnati, L.F. Receptor–Receptor Interactions and Glial Cell Functions with a Special Focus on G Protein-Coupled Receptors. *Int. J. Mol. Sci.* **2021**, *22*, 8656. [[CrossRef](#)] [[PubMed](#)]
142. Perreault, M.; Hasbi, A.; O’Dowd, B.F.; George, S.R. heteromeric dopamine receptor signaling complexes: Emerging neurobiology and disease relevance. *Neuropsychopharmacology* **2014**, *39*, 156–168.
143. Lindvall, O.; Björklund, A. The organization of the ascending catecholamine neuron systems in the rat brain as revealed by the glyoxylic acid fluorescence method. *Acta Physiol. Scand.* **1974**, *412*, 1–48.
144. Gerfen, C.R. The neostriatal mosaic: Multiple levels of compartmental organization. *Trends Neurosci.* **1992**, *15*, 133–139.
145. Liu, Y.-J.; Chen, J.; Li, X.; Zhou, X.; Hu, Y.-M.; Chu, S.-F.; Peng, Y.; Chen, N.-H. Research progress on adenosine in central nervous system diseases. *CNS Neurosci. Ther.* **2019**, *25*, 899–910.
146. Ferré, S.; Popoli, P.; Gimenez-Llort, L.; Finnman, U.B.; Martinez, E.; Scotti de Carolis, A.; Fuxe, K. Postsynaptic antagonistic interaction between adenosine A1 and dopamine D1 receptors. *Neuroreport* **1994**, *6*, 73–76. [[CrossRef](#)]
147. Emmi, A.; Antonini, A.; Sandre, M.; Baldo, A.; Conran, M.; Macchi, V.; Guidolin, D.; Porzionato, A.; De Caro, R. Topography and distribution of adenosine A2A and dopamine D2 receptors in the human subthalamic nucleus. *Front. Neurosci.* **2022**, *16*, 945574. [[CrossRef](#)] [[PubMed](#)]
148. Cotzias, G.C. Levodopa in the treatment of Parkinsonism. *JAMA* **1971**, *218*, 1903–1908. [[CrossRef](#)] [[PubMed](#)]
149. Marsden, C.D. Problems with long-term levodopa therapy for Parkinson’s disease. *Clin. Neuropharmacol.* **1994**, *17*, S32–S544. [[CrossRef](#)] [[PubMed](#)]
150. Fuxe, K.; Guidolin, D.; Agnati, L.F.; Borroto-Escuela, D.O. Dopamine heteroreceptor complexes as therapeutic targets in Parkinson’s disease. *Exp. Opin. Ther. Targets* **2015**, *19*, 377–398.
151. Chen, J.F.; Cunha, R.A. The belated US FDA approval of the adenosine A2A receptor antagonist istradefylline for treatment of Parkinson’s disease. *Purinergic Signal.* **2020**, *16*, 167–174.
152. Marcellino, D.; Carriba, P.; Filip, M.; Borgkvist, A.; Frankowska, M.; Bellido, I.; Tanganelli, S.; Muller, C.E.; Roberts, D.C.; Fisone, G.; et al. Antagonistic cannabinoid CB1/dopamine D2 receptor interactions in striatal CB1/D2 heteromers. A combined neurochemical and behavioural analysis. *Neuropharmacology* **2008**, *54*, 815–828. [[CrossRef](#)]
153. Gilgun-Sherki, Y.; Melamed, E.; Mechoulam, R.; Offen, D. The CB1 cannabinoid receptor agonist, HU-210, reduces levodopa-induced rotations in 6-hydroxydopamine-lesioned rats. *Pharmacol. Toxicol.* **2003**, *93*, 66–70. [[CrossRef](#)]
154. Haber, S.N. Neuroanatomy of reward: A view from the ventral striatum. In *Neurobiology of Sensation and Reward*; Gottfried, J.A., Ed.; CRC Press/Taylor and Francis: Boca Raton, FL, USA, 2011.
155. Fuxe, K.; Dahlstrom, A.; Jonsson, G.; Marcellino, D.; Guescini, M.; Dam, M.; Manger, P.; Agnati, L. The discovery of central monoamine neurons gave volume transmission to the wired brain. *Prog. Neurobiol.* **2010**, *90*, 82–100.
156. Borroto-Escuela, D.O.; Ferraro, L.; Narvaez, M.; Tanganelli, S.; Beggiato, S.; Liu, F.; Rivera, A.; Fuxe, K. Multiple adenosine-dopamine (A2A-D2 like) heteroreceptor complexes in the brain and their role in schizophrenia. *Cells* **2020**, *9*, 1077. [[CrossRef](#)]
157. Borroto-Escuela, D.O.; Pintsuk, J.; Schäfer, T.; Friedland, K.; Ferraro, L.; Tanganelli, S.; Liu, F.; Fuxe, K. Multiple D2 heteroreceptor complexes: New targets for treatment of schizophrenia. *Ther. Adv. Psychopharmacol.* **2016**, *6*, 77–94. [[CrossRef](#)]

158. Edwards, S.; Whisler, K.N.; Fuller, D.C.; Orsulak, P.J.; Self, D.W. Addiction-related alterations in D1 and D2 dopamine receptor behavioral responses following chronic cocaine self-administration. *Neuropsychopharmacology* **2007**, *32*, 354–366. [[CrossRef](#)] [[PubMed](#)]
159. Briand, L.A.; Flagel, S.B.; Garcia-Fuster, M.J.; Watson, S.J.; Akil, H.; Sarter, M.; Robinson, T.E. Persistent alterations in cognitive function and prefrontal dopamine D2 receptors following extended, but not limited, access to self-administered cocaine. *Neuropsychopharmacology* **2008**, *33*, 2969–2980. [[PubMed](#)]
160. Wydra, K.; Suder, A.; Borroto-Escuela, D.O.; Filip, M.; Fuxe, K. On the role of A(2)A and D(2) receptors in control of cocaine and food-seeking behaviors in rats. *Psychopharmacology* **2015**, *232*, 1767–1778. [[CrossRef](#)] [[PubMed](#)]
161. Romieu, P.; Phan, V.L.; Martin-Fardon, R.; Maurice, T. Involvement of the sigma1 receptor in cocaine-induced conditioned place preference: Possible dependence on dopamine uptake blockade. *Neuropsychopharmacology* **2002**, *26*, 444–455. [[CrossRef](#)]
162. Canales, J.J. Stimulant-induced adaptations in neostriatal matrix and striosome systems: Transiting from instrumental responding to habitual behavior in drug addiction. *Neurobiol. Learn. Mem.* **2005**, *83*, 93–103. [[CrossRef](#)]
163. Seeman, P. Targeting the dopamine D2 receptor in schizophrenia. *Expert Opin. Ther. Targets* **2006**, *10*, 515–531. [[CrossRef](#)]
164. Carlsson, A. The current status of the dopamine hypothesis of schizophrenia. *Neuropsychopharmacology* **1988**, *1*, 179–186. [[CrossRef](#)]
165. Andersen, M.B.; Fuxe, K.; Werge, T.; Gerlach, J. The adenosine A2A receptor agonist CGS 21680 exhibits antipsychotic-like activity in Cebus apella monkeys. *Behav. Pharmacol.* **2022**, *13*, 639–644. [[CrossRef](#)]
166. Caldwell, H.K.; Stephens, S.L.; Young, W.S. III. Oxytocin as a natural antipsychotic: A study using oxytocin knockout mice. *Mol. Psychiatry* **2009**, *14*, 190–196. [[CrossRef](#)]
167. Feifel, D. Oxytocin as a potential therapeutic target for schizophrenia and other neuropsychiatric conditions. *Neuropsychopharmacology* **2012**, *37*, 304–305. [[CrossRef](#)]
168. Hökfelt, T.; Fuxe, K.; Johansson, O.; Ljungdahl, Å. Pharmaco-histochemical evidence of the existence of dopamine nerve terminals in the limbic cortex. *Eur. J. Pharmacol.* **1974**, *25*, 108–112. [[CrossRef](#)] [[PubMed](#)]
169. Berger, B.; Gaspar, P.; Verney, C. Dopaminergic innervation of the cerebral cortex: Unexpected differences between rodents and primates. *Trends Neurosci.* **1991**, *14*, 21–27. [[CrossRef](#)] [[PubMed](#)]
170. Malenka, E.J.; Nestler, S.E.; Hyman, R.C. *Molecular Neuropharmacology: A Foundation for Clinical Neuroscience*, 2nd ed.; McGraw-Hill Medical: New York, NY, USA, 2009; Chapter 13; p. 318.
171. Casadó-Anguera, V.; Moreno, E.; Sánchez-Soto, M.; Cai, N.S.; Bonaventura, J.; Homar-Ruano, P.; Rubinstein, M.; Cortés, A.; Canela, E.I.; Ferré, S.; et al. Heteromerization between a_{2A} adrenoceptors and different polymorphic variants of the dopamine D₄ receptor determines pharmacological and functional differences. implications for impulsive-control disorders. *Pharmacol. Res.* **2021**, *170*, 105745. [[CrossRef](#)] [[PubMed](#)]
172. LaHoste, G.J.; Swanson, J.M.; Wigal, S.B.; Glabe, C.; Wigal, T.; King, N.; Kennedy, J. Dopamine D4 receptor gene polymorphism is associated with attention deficit hyperactivity disorder. *Mol. Psychiatry* **1996**, *1*, 121–124.
173. Sancho, L.; Contreras, M.; Allen, N.J. Glia as sculptors of synaptic plasticity. *Neurosci. Res.* **2021**, *167*, 17–29.
174. Allen, N.J.; Barres, B.A. Signaling between glia and neurons: Focus on synaptic plasticity. *Curr. Opin. Neurobiol.* **2005**, *15*, 542–548. [[CrossRef](#)]
175. Fellin, T.; Carmignoto, G. Neurone-to-astrocyte signalling in the brain represents a distinct multifunctional unit. *J. Physiol.* **2004**, *559*, 3–15.
176. Cervetto, C.; Maura, G.; Guidolin, D.; Amato, S.; Ceccoli, C.; Agnati, L.F.; Marcoli, M. Striatal astrocytic A2A-D2 receptor-receptor interactions and their role in neuropsychiatric disorders. *Neuropharmacology* **2023**, *237*, 109636.
177. Ren, C.; He, K.J.; Hu, H.; Zhang, J.B.; Dong, L.G.; Li, D.; Chen, J.; Mao, C.J.; Wang, F.; Liu, C.F. Induction of parkinsonian-like changes via targeted downregulation of astrocytic glutamate transporter GLT-1 in the striatum. *J. Park. Dis.* **2022**, *12*, 295–314.
178. Martín, R.; Bajo-Grañeras, R.; Moratalla, R.; Perea, G.; Araque, A. Circuit-specific signaling in astrocyte-neuron networks in basal ganglia pathways. *Science* **2015**, *349*, 730–734. [[CrossRef](#)]
179. Huang, G.; Dragan, M.; Freeman, D.; Wilson, J.X. Activation of catechol-O-methyltransferase in astrocytes stimulates homocysteine synthesis and export to neurons. *Glia* **2005**, *51*, 47–55. [[CrossRef](#)] [[PubMed](#)]
180. Matos, M.; Shen, H.Y.; Augusto, E.; Wang, Y.; Wei, C.J.; Wang, Y.T.; Agostinho, P.; Boison, D.; Cunha, R.A.; Chen, J.F. Deletion of adenosine A2A receptors from astrocytes disrupts glutamate homeostasis leading to psychomotor and cognitive impairment: Relevance to schizophrenia. *Biol. Psychiatr.* **2015**, *78*, 763–774.
181. Kruyer, A.; Kalivas, P.W.; Scofield, M.D. Astrocyte regulation of synaptic signaling in psychiatric disorders. *Neuropsychopharmacology* **2023**, *48*, 21–36. [[PubMed](#)]
182. Berretta, S. Extracellular matrix abnormalities in schizophrenia. *Neuropharmacology* **2012**, *62*, 1584–1597. [[PubMed](#)]
183. Wardas, J. Potential role of adenosine A2A receptors in the treatment of schizophrenia. *Front. Biosci.* **2008**, *13*, 4071–4096. [[CrossRef](#)]
184. Valle-Léon, M.; Casajuana-Martin, N.; del Torrent, C.L.; Argerich, J.; Gómez-Acero, L.; Sahlholm, K.; Ferré, S.; Pardo, L.; Ciruela, F. Unique effect of clozapine on adenosine A2A-dopamine D2 receptor heteromerization. *Biomed. Pharmacother.* **2023**, *160*, 114327.
185. Kruyer, A.; Kalivas, P.W. Astrocytes as cellular mediators of cue reactivity in addiction. *Curr. Opin. Pharmacol.* **2021**, *56*, 1–6. [[CrossRef](#)]

186. Alexander, G.E.; Crutcher, M.D. Functional architecture of basal ganglia circuits: Neural substrates of parallel processing. *Trends Neurosci.* **1990**, *13*, 266–271.
187. Daniels, D.J.; Lenard, N.R.; Etienne, C.L.; Law, P.-Y.; Roerig, S.C.; Portoghese, P.S. Opioid-induced tolerance and dependence in mice is modulated by the distance between pharmacophores in a bivalent ligand series. *Proc. Natl. Acad. Sci. USA* **2005**, *102*, 19208–19213. [[CrossRef](#)]

Disclaimer/Publisher’s Note: The statements, opinions and data contained in all publications are solely those of the individual author(s) and contributor(s) and not of MDPI and/or the editor(s). MDPI and/or the editor(s) disclaim responsibility for any injury to people or property resulting from any ideas, methods, instructions or products referred to in the content.