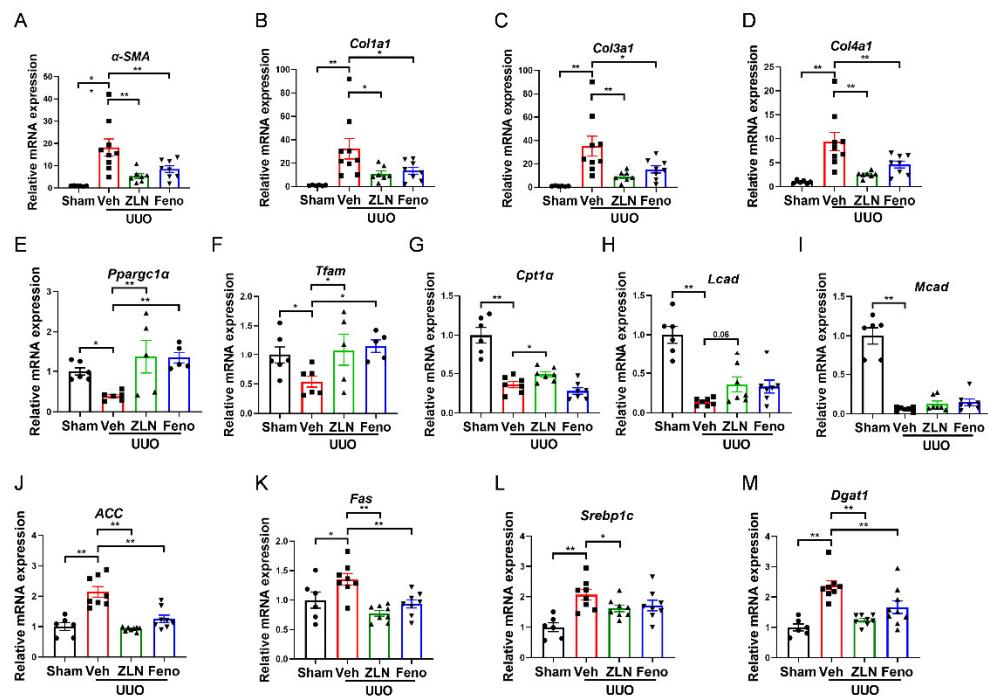
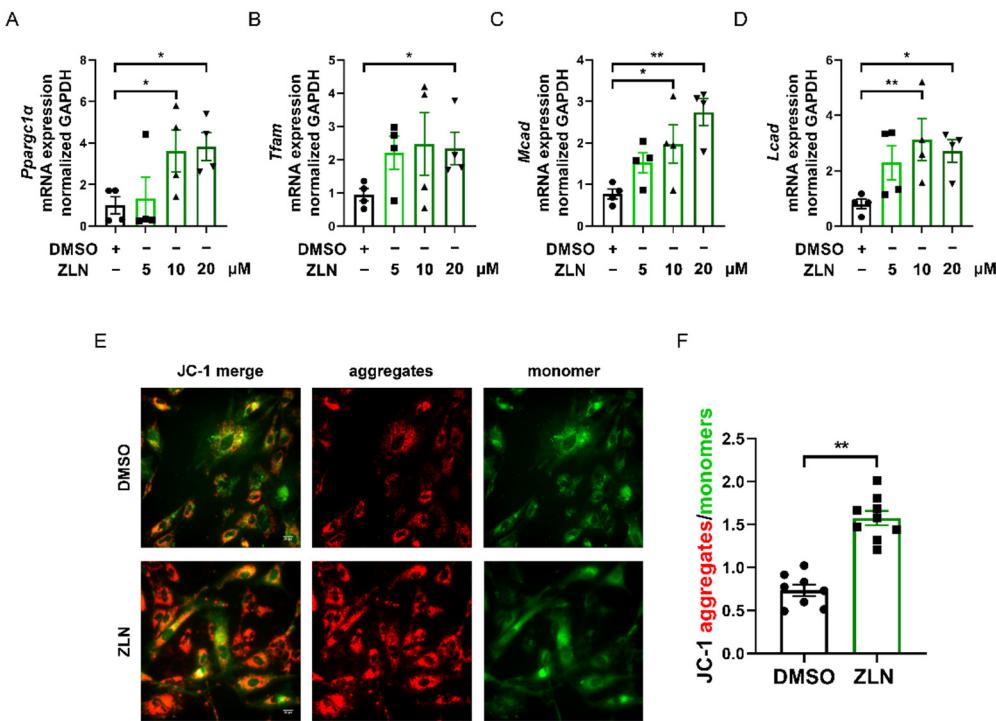




**Supplementary Materials:** The following supporting information can be downloaded at: [www.mdpi.com/xxx/s1](http://www.mdpi.com/xxx/s1).



**Figure S1.** mRNA expression of  $\alpha$ -SMA (A), *Col1a1* (B), *Col1a3* (C) and *Col1a4* (D) in kidney tissue were examined by Quantitative RT-PCR. Data are expressed as the mean  $\pm$  SEM. (n=6~9). \*P<0.05, \*\*P<0.01. Level of *Ppargc1 $\alpha$*  (E), *Tfam* (F), *Cpt1 $\alpha$*  (G), *Lcad* (H) and *Mcad* (I) mRNA in kidney tissue were detected by Quantitative RT-PCR. Data are expressed as the mean  $\pm$  SEM. (n=5~6). \*P<0.05, \*\*P<0.01. The mRNA level of *ACC* (J), *Fas* (K), *Srebp1c* (L) and *Dgat1* (M) were measured. Data are expressed as the mean  $\pm$  SEM. (n=6~8). \*P<0.05, \*\*P<0.01. ZLN005 is abbreviated to ZLN and Fenofibrate is abbreviated to Feno.



**Figure S2.** ZLN005 alone treatment on HKC cells for 48 h. The mRNA level of *Ppargc1α* (A), *Tfam* (B), *Mcad*(C) and *Lcad* (D) were measured. Data are expressed as the mean  $\pm$  SEM. (n=4). \*P<0.05, \*\*P<0.01. One hour treatment of ZLN005 on HKC cells. (E) Representative images of JC-1 staining showing red fluorescence of JC-1 aggregates and green signal of monomers. Bar=20  $\mu$ m. (F) Quantification of the ratio of red to green fluorescence. Data are expressed as the mean  $\pm$  SEM. (n=8~9). \*\*P<0.01. ZLN005 is abbreviated to ZLN.

**Table 1:** Primer sequences used in the experiments.

	Forward Primer	Reverse Primer
<b>QPCR primers</b>		
<i>mGAPDH</i>	TCTCCTGCGACTTCAACA	TGGTCCAGGGTTCTTACT
<i>mPpargc1α</i>	ACTGAGCTACCCTGGGATG	TAAGAATTCCGGTGGTGACA
<i>mα-SMA</i>	CTGACAGAGGCACCACTGAA	AGAGGCATAGAGGGACAGCA
<i>mCol1a1</i>	GCTCTTTTAGATACTGTGGTGAGGAA	GTTTCCACGTCTCACCATTG
<i>mCol3a1</i>	ACAGCTGGTGAACCTGGAAG	ACCAGGAGATCCATCTCGAC
<i>mCol4a1</i>	GACAGCCAGGTTGACAGGT	GGCAGCTCTCCTTCTGA
<i>mil1β</i>	GCAACTGTTCTGAACCTCAACT	ATCTTTGGGTCCGTCAACT
<i>mil6</i>	TAGTCCTCCTACCCAATTICC	TTGGTCCTTAGCCACTCCTTC
<i>miNos</i>	GAGACAGGGAAGTCTGAAGCAC	CCAGCAGTAGTTGCTCCTCTTC
<i>mTnfa</i>	GAAGTTCCCAAATGGCCTCC	TTGTCACTCGAATTGAGAAGATG
<i>mTfam</i>	GAGGCAAAGGATGATTGGCTC	CGAACCTATCATCTTAGCAAGC
<i>mACC</i>	TCTACGGCAGCAGTTACACCACAT	TCTCTCATTACCTCAATCTCAGCATAG
<i>mFas</i>	TGGGTTCTAGCCAGCAGAGT	TACCACCAGAGACCGTTATGC
<i>mSrebp1c</i>	CGACTACATCCGCTTCTGCAG	CCTCCATAGACACATCTGTGCC

<i>mDgat1</i>	GGAGACCGCGAGTTCTACAG	CTCATGGAAGAAGGCTGAGG
<i>mCpt1α</i>	ATGACGGCTATGGTGTTC	TGTCCATCATGGCTTGTCTC
<i>mLcad</i>	TCACCAACCGTGAAGCTCGA	CCAAAAGAGGCTAATGCCATG
<i>mMcad</i>	AGCTGCTAGTGGAGCACCAAG	TCGCCATTCTGCGAGC
<i>hGAPDH</i>	GCTCTCTGCTCCTCCTGTC	ATGGTGTCTGAGCGATGTGG
<i>hPpargc1α</i>	CCCTGTCTGCTCTGTGGACT	GCTCCAAGCTACTGTGGTGA
<i>hα-SMA</i>	ATCACCAACTGGGACGACAT	GGCAACACGAAGCTCATTG
<i>hFibronectin</i>	CCCTGGTGTACAGAGGCTA	TGTATATTGGTCCCGGTT
<i>hCol1a1</i>	GCGGACTTTGTTGCTGCTTGCAG	ATCTCCGGCTGGGCCCTTCTT
<i>hCol4a1</i>	CAAGGGCTGCCGGTTCTG	CCGGTGTACCACGACTGCC
<i>hTGFβ</i>	TACCTGAACCGTGTTGCTCTC	GTTGCTGAGGTATGCCAGGAA

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*Continued*

	Forward Primer	Reverse Primer
<i>hTfam</i>	GTGGTTTCATCTGCTTGGCAAG	TTCCCTCCAACGCTGGCAATT
<i>hLcad</i>	GTTTGGACTCCGCCACTGCTTG	GGCTGAACCTGGCATCCACAT
<i>hMcad</i>	AGAACCTGGAGCAGGCTCTGAT	GGATCTGGATCAGAACGTGCCA
<i>18s rRNA</i>	CTACCACATCCAAGGAAGCA	TTTTCGTCACTACCTCCCCG
<i>16s rRNA</i>	GCCTTCCCCGTAAATGATA	TTATGCGATTACCGGGCTCT

*Ubiquitin*

GCCCAGTGTACCACCAAGAAG

GCTTTTAGATACTGTGGTGAGGAA

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