

A Flexible Portable Glucose Sensor Based on Hierarchical Arrays of Au@Cu(OH)₂ Nanograss

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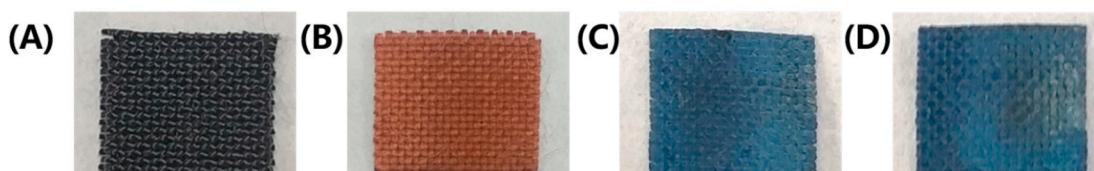


Figure S1. Optical images of (A) CFC, (B) Cu/CFC, (C) Cu(OH)₂/CFC (D) Au@Cu(OH)₂/CFC.

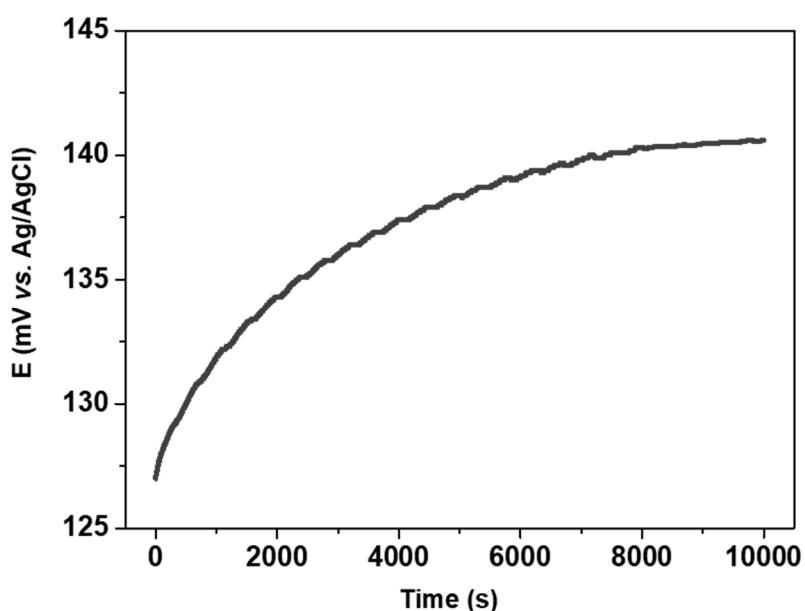


Figure S2. Open circuit potential (OCP) between deposited Ag/AgCl/CFC electrode and a commercial Ag/AgCl (3 M KCl) reference electrode in electrolyte solution with 0.1 M KOH and 0.01 M KCl.

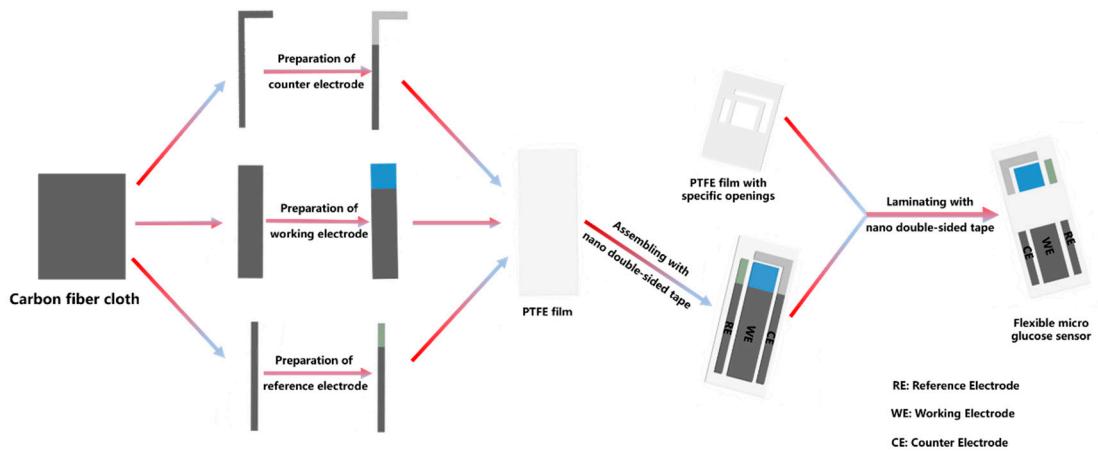


Figure S3. Schematic illustration of the fabrication process for the flexible micro glucose sensor.

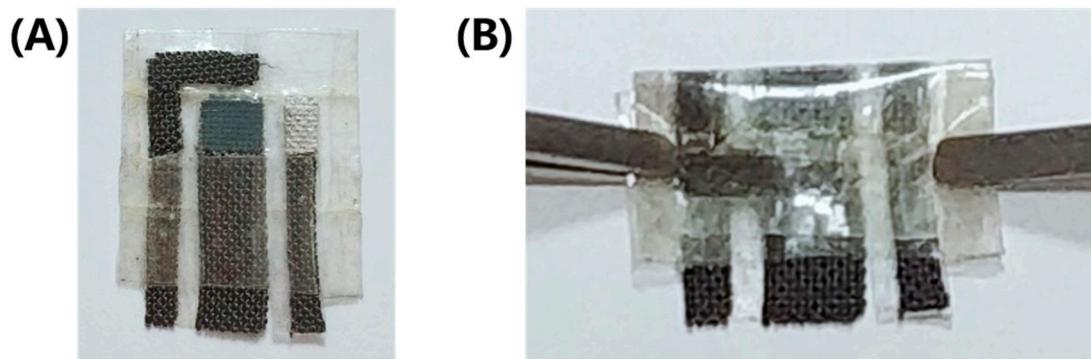


Figure S4. Optical images of the (A) original and (B) folded flexible micro glucose sensor.

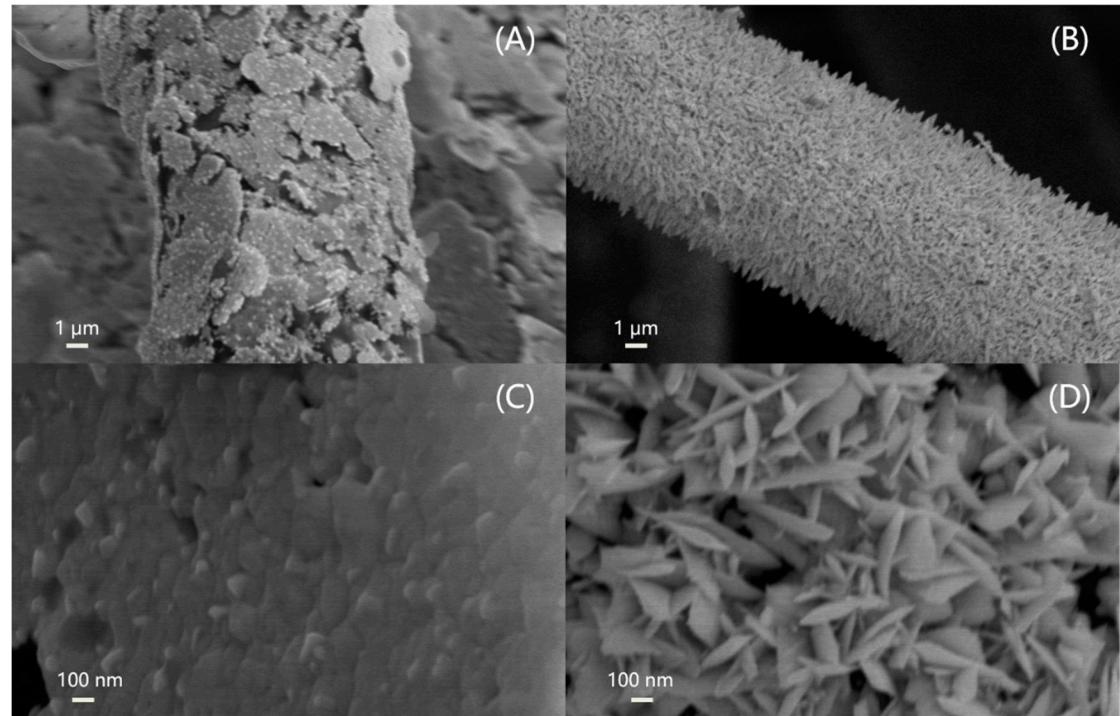


Figure S5. Low-magnification SEM images of (A) Ag/AgCl/CFC and (B) Pt/CFC, and High-magnification SEM images of (C) Ag/AgCl/CFC and (D) Pt/CFC.

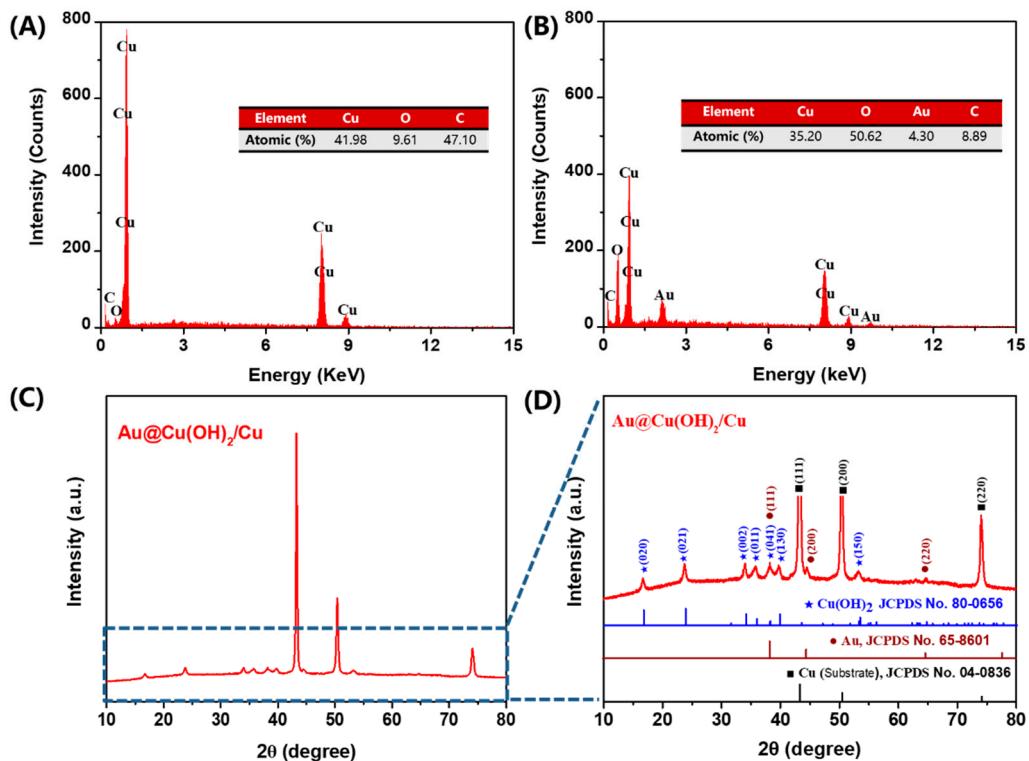


Figure S6. EDS patterns of (A) Cu/CFC and (B) Au@Cu(OH)₂/CFC, XRD patterns of (C) Au@Cu(OH)₂/Cu plate and (D) The magnification of (C).

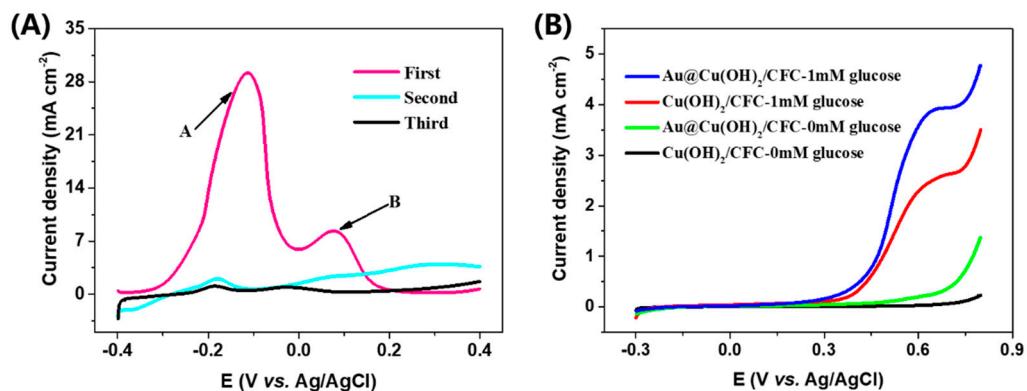


Figure S7. LSV curves of (A) fabricating Cu(OH)₂ samples, from -0.4 V to 0.4 V with 3 mV s^{-1} sweeping rate in 1 M KOH resolution for three times, (B) the Cu(OH)₂/CFC and Au@Cu(OH)₂/CFC sensors in 0.1 M KOH and 0.01 M KCl solution without and with 1 mM glucose .

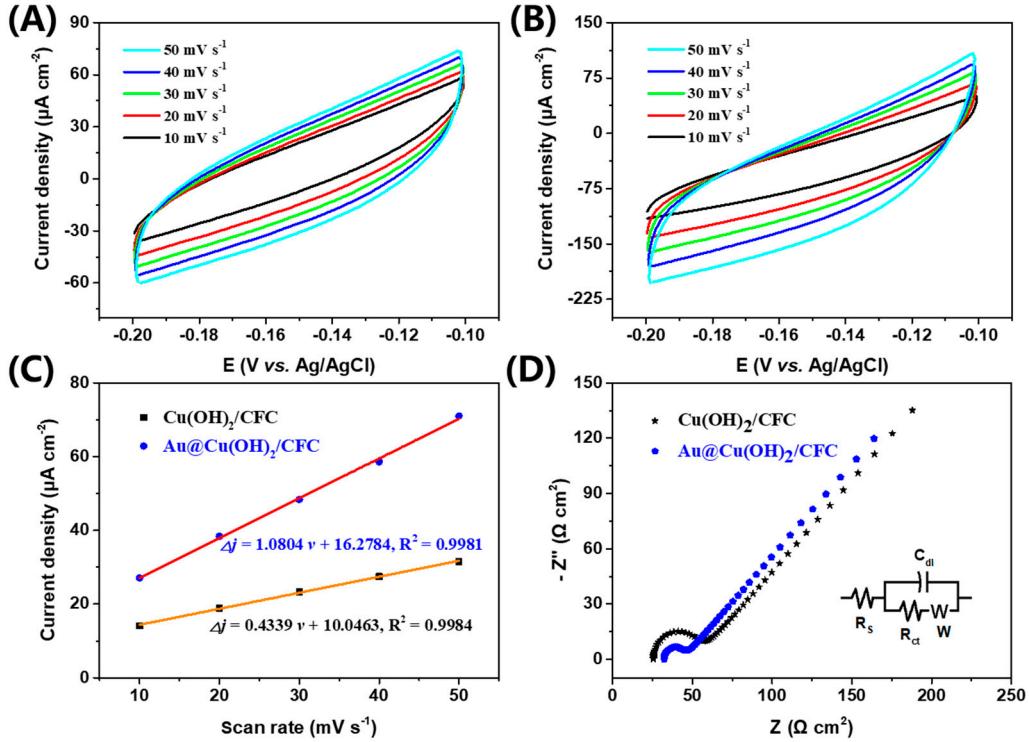


Figure S8. CVs for (A) Cu(OH)₂/CFC and (B) Au@Cu(OH)₂/CFC sensors with different scan rates(10–50 mV s⁻¹) in 0.1 M KOH and 0.01 M KCl solution, (C) The capacitive current densities–scan rates calibration plots of the two sensors at -0.15 V ($\Delta j_{-0.15V} = (j_a - j_c)/2$), (D) Nyquist plots of Cu(OH)₂/CFC and Au@Cu(OH)₂/CFC sensors in 0.1 M KCl electrolyte with 5 mM K₃[Fe(CN)₆] and 5 mM K₄[Fe(CN)₆], inset is equivalent Randle circuit for the two cases.

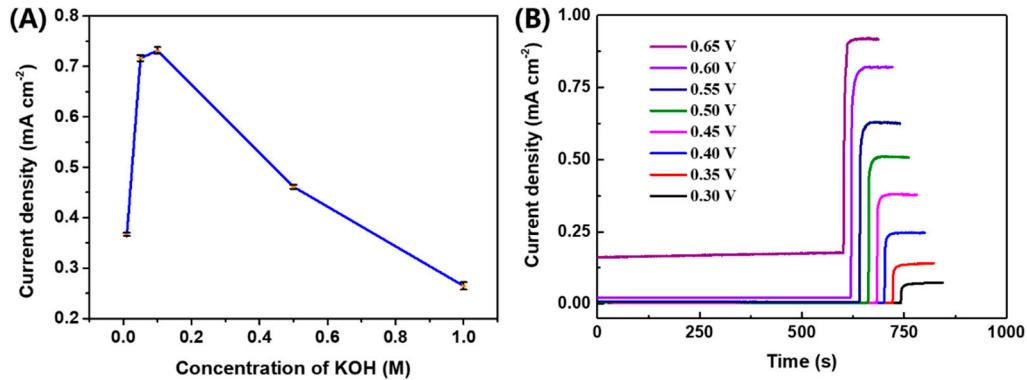


Figure S9. (A) The line chart of chronoamperometric current response of Au@Cu(OH)₂/CFC towards 0.10 mM glucose in different concentration KOH (0.01 to 1.00 M) and 0.01 M KCl solution at 0.60V. The maximum RSD is 2.8%. (B) The chronoamperometric current response of Au@Cu(OH)₂/CFC towards 0.10 mM glucose in 0.1 M KOH and 0.01 M KCl solution at different potentials (ranging from 0.30 to 0.65 V).

Table S1. The comparison of glucose sensing performances based on various composites which reported previously. SPE: screen printed electrode, GCE: glassy carbon electrode, rGO: reduced graphene oxide, PGF: porous graphene foam, MWCNTs: multi-walled carbon nanotubes array.

Electrode	Detection Method	Response time (s)	Sensitivity	Detection limit (μM)	Linear range (mM)	Cost	Refs
GQDs	Optical	~	0.012*	3000	4–40	Very high	[1]
polymer optode	Optical	900 ~	0.2*	1000	1–19	Very high	[2]
nанопорous pigments	Colorimetric	30	-	1000	2–50	high	[3]
Polymer Gel	Colorimetric	10^4 ~	-	5000	5–8	high	[4]
Au-CuO/rGO/SPE	LSV	10	2.36 [#]	0.100	0.0001–12	low	[5]
CuO/GCE-Nafion	Amperometry	5	0.47 [#]	0.016	1–10	low	[6]
Cu ₂ O/graphene nanosheets	Amperometry	5	0.29	3.300	0.3–3.3	low	[7]
Cu(OH) ₂ nanotubes	Amperometry	5	0.42 [#]	0.500	up to 3	low	[8]
CuO/rGO/GCE	Amperometry	5	1.36 [#]	0.700	0.002–4	low	[9]
CuO/carbon spheres	Amperometry	5	2.98 [#]	0.100	0.0005–2.3	low	[10]
CuO-MWCNTs	Amperometry	2	2.19 [#]	0.800	0.2–3	low	[11]
Cu(OH) ₂ /PGF	Amperometry	10	3.36 [#]	1.200	0.00012–6	low	[12]
CuO arrays/nanoporous Cu	Amperometry	3	1.62 [#]	0.200	0.0005–5.0	low	[13]
Au@Cu(OH) ₂ /CFC	Amperometry	5	7.35 [#]	0.027	0.1–3.3	low	This work

* mM per percent change of fluorescence intensity, [#] mA mM⁻¹ cm⁻².

Table S2. Influence of common interfering species on the determination of glucose with Au@Cu(OH)₂/CFC in 0.1 M KOH and 0.01 M KCl solution.

Label	Analyte	Tested Concentration(mM)	Current Response (%)
-	Glucose	0.1	100
a	Maltose	0.01	3.46
b	Fructose	0.01	3.79
c	Uric acid (UA)	0.01	1.13
d	Dopamine (DA)	0.01	1.93
e	Ascorbic acid (AA)	0.01	3.83
f	Cysteine	0.01	2.29
g	Acetaminophen	0.01	0.04
h	KCl	1	1.80
i	Na ₂ SO ₄	1	1.56

References

1. Shehab, M.; Ebrahim, S.; Soliman, M. Graphene quantum dots prepared from glucose as optical sensor for glucose. *J Lumin* **2017**, *184*, 110–116.
2. Billingsley, K.; Balaconis, M.K.; Dubach, J.M.; Zhang, N.; Lim, E.; Francis, K.P.; Clark, H.A.; Ester, G.B. Fluorescent Nano-Optodes for Glucose Detection. *Anal Chem* **2010**, *82*, 3707–3713.
3. Lim, S.H.; Musto, C.J.; Park, E.; Zhong, W.; Suslick, K.S. A Colorimetric Sensor Array for Detection and Identification of Sugars. *Org Lett* **2009**, *10*, 4405–4408.
4. Honda, M.; Kataoka, K.; Seki, T.; Takeoka, Y. Confined Stimuli-Responsive Polymer Gel in Inverse Opal Polymer Membrane for Colorimetric Glucose Sensor. *Langmuir* **2009**, *25*, 8349–8356.
5. Dhara, K.; Ramachandran, T.; Nair, B.G.; Satheesh Babu, T.G. Single step synthesis of Au-CuO nanoparticles decorated reduced graphene oxide for high performance disposable nonenzymatic glucose sensor. *J Electroanal Chem* **2015**, *743*, 1–9.
6. Baloach, Q.A.; Tahira, A.; Mallah, A.B.; Abro, M.I.; Uddin, S.; Willander, M.; Ibupoto, Z.H. A robust, enzyme-free glucose sensor based on lysine-assisted CuO nanostructures. *Sensors* **2016**, *16*, 1878–1888.
7. Liu, M.; Liu, R.; Chen, W. Graphene wrapped Cu₂O nanocubes: Non-enzymatic electrochemical sensors for the detection of glucose and hydrogen peroxide with enhanced stability. *Biosens Bioelectron* **2013**, *45*, 206–212.
8. Zhou, S.; Feng, X.; Shi, H.; Chen, J.; Zhang, F.; Song, W. Direct growth of vertically aligned arrays of Cu (OH)₂ nanotubes for the electrochemical sensing of glucose. *Sensors Actuators B Chem* **2013**, *177*, 445–452.
9. Luo, L.; Zhu, L.; Wang, Z. Nonenzymatic amperometric determination of glucose by CuO nanocubes – graphene nanocomposite modified electrode. *Bioelectrochemistry* **2012**, *88*, 156–163.
10. Zhang, J.; Ma, J.; Zhang, S.; Wang, W.; Chen, Z. A highly sensitive nonenzymatic glucose sensor based on CuO nanoparticles decorated carbon spheres. *Sensors Actuators B Chem* **2015**, *211*, 385–391.
11. Yang, J.; Jiang, L.; Zhang, W.; Gunasekaran, S. A highly sensitive non-enzymatic glucose sensor based on a simple two-step electrodeposition of cupric oxide (CuO) nanoparticles onto multi-walled carbon nanotube arrays. *Talanta* **2010**, *82*, 25–33.
12. Shackery, I.; Patil, U.; Pezeshki, A.; Shinde, N.M.; Kang, S.; Im, S.; Chan, S. Copper Hydroxide Nanorods Decorated Porous Graphene Foam Electrodes for Non-enzymatic Glucose Sensing. *Electrochim Acta* **2016**, *191*, 954–961.
13. Chen, H.; Fan, G.; Zhao, J.; Qiu, M.; Sun, P.; Fu, Y.; Han, D.; Cui, G. A portable micro glucose sensor based on copper-based nanocomposite structure. *New J Chem* **2019**, *43*, 7806–7813.