

Article

Seagrass Blue Carbon Stock and Air–Sea CO₂ Fluxes in the Karimunjawa Islands, Indonesia during Southeast Monsoon Season

Nurul Latifah ^{1,2}, Nining Sari Ningsih ^{3,*}, Aditya Rakhmat Kartadikaria ³, Anindya Wirasatriya ⁴, Sigit Febrianto ², Novi Susetyo Adi ⁵ and Faisal Hamzah ⁶

¹ Doctoral Program in Earth Sciences, Faculty of Earth Sciences and Technology, Institut Teknologi Bandung, Bandung 40132, Indonesia; nurul.latifah@live.undip.ac.id

² Department of Aquatic Sciences, Faculty of Fisheries and Marine Science, Universitas Diponegoro, Semarang 50275, Indonesia; febriantosigit40@gmail.com

³ Research Group of Oceanography, Faculty of Earth Sciences and Technology, Institut Teknologi Bandung, Bandung 40132, Indonesia; a.r.kartadikaria@itb.ac.id

⁴ Department of Oceanography, Faculty of Fisheries and Marine Science, Universitas Diponegoro, Semarang 50275, Indonesia; aninosi@yahoo.co.id

⁵ Center for Marine Research, Ministry of Marine Affairs and Fisheries, Jakarta 14430, Indonesia; novisusetyoadi@gmail.com

⁶ The National Research and Innovation Agency, Jakarta Pusat 10340, Indonesia; faisal.hamzah@brin.go.id

* Correspondence: nsningsih@itb.ac.id

Abstract: Research focusing on seagrass ecosystems as carbon storage has been conducted in various Indonesian waters. However, an essential aspect that remains unexplored is the simultaneous analysis of blue carbon storage in seagrass alongside carbon dioxide (CO₂) flux values, particularly within Karimunjawa waters. This study aims to assess the organic carbon stock and sea–air CO₂ flux in seagrass ecosystems in Karimunjawa. Our hypothesis posits that although seagrass ecosystems release CO₂ into the water, their role as blue carbon ecosystems enables them to absorb and accumulate organic carbon within seagrass biomass and sediments. This investigation took place in Karimunjawa waters, encompassing both vegetated (seagrass meadows) and unvegetated (non-seagrass meadows) areas during August 2019, 2020, and 2022. Over this period, the organic carbon stock in seagrass and sediment displayed an increase, rising from 28.90 to 35.70 gC_{org} m⁻² in 2019 and from 37.80 to 45.25 gC_{org} m⁻² in 2022. Notably, the expanse of seagrass meadows in Karimunjawa dwindled by 328.33 ha from 2019 to 2022, resulting in a total carbon stock reduction of the seagrass meadows of 452.39 tC to 218.78 tC. Sediment emerges as a pivotal element in the storage of blue carbon in seagrass, with sedimentary organic carbon outweighing seagrass biomass in storage capacity. The conditions in Karimunjawa, including a high A:B ratio, low dry bulk density, and elevated water content, foster a favorable environment for sediment carbon absorption and storage, facilitated by the waters' CO₂ emission during the southeast monsoon season. Notably, our findings reveal that CO₂ release within vegetated areas is lower compared to unvegetated areas. This outcome underscores how seagrass ecosystems can mitigate CO₂ release through their adeptness at storing organic carbon within biomass and sediment. However, the presence of inorganic carbon in the form of calcium carbonate introduces a nuanced dynamic. This external source, stemming from allochthonous origins like mangroves, brown algae like *Padina pavonica*, and calcareous epiphytes, leads to an increase in sedimentary organic carbon stock of 53.2 ± 6.82 gC_{org} m⁻². Moreover, it triggers the release of CO₂ into the atmosphere, quantified at 83.4 ± 18.26 mmol CO₂ m⁻² d⁻¹.

Keywords: blue carbon; seagrass; CO₂ fluxes; Karimunjawa



Citation: Latifah, N.; Ningsih, N.S.; Kartadikaria, A.R.; Wirasatriya, A.; Febrianto, S.; Adi, N.S.; Hamzah, F. Seagrass Blue Carbon Stock and Air–Sea CO₂ Fluxes in the Karimunjawa Islands, Indonesia during Southeast Monsoon Season. *Diversity* **2023**, *15*, 978. <https://doi.org/10.3390/d15090978>

Academic Editors: Zhongqiao Li and Alexander Polukhin

Received: 30 June 2023

Revised: 27 August 2023

Accepted: 27 August 2023

Published: 29 August 2023



Copyright: © 2023 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (<https://creativecommons.org/licenses/by/4.0/>).

1. Introduction

The global concentration of atmospheric carbon dioxide ($\text{CO}_{2\text{atm}}$) has increased from 340.00 ppm in 1980 to 413.25 ppm as of 2020, attributed to human activities lacking environmental friendliness [1–5]. This increase in $\text{CO}_{2\text{atm}}$ subsequently amplifies the presence of CO_2 within aquatic environments. Through the solubility pump mechanism, the ocean absorbs $\text{CO}_{2\text{atm}}$, transforming it into Dissolved Inorganic Carbon (DIC), which encompassing aqueous CO_2 , carbonic acid (H_2CO_3), bicarbonate (HCO_3^-), and carbonate (CO_3^{2-}) forms [6,7]. However, the equilibrium of the solubility pump was disrupted following the industrial revolution, leading to an unequal balance between CO_2 influx and efflux. Remarkably, even a minor rise of 1 part per million (ppm) in $\text{CO}_{2\text{atm}}$ triggers the oceans to absorb an additional 0.97 ± 0.40 GtC of CO_2 [8,9]. As a carbon reservoir, the oceans have absorbed approximately 30% of $\text{CO}_{2\text{atm}}$, thereby inducing alterations in the oceanic carbonate system [2,10,11]. Referring to [12], as per the Global Carbon Project (GCP), 55% of anthropogenic carbon remains in the atmosphere, while 27% becomes absorbed by the oceans.

The open sea functions as a carbon reservoir, playing a pivotal role in reducing anthropogenic CO_2 emissions [10]. Beyond the open sea, there exist coastal habitats collectively referred to as blue carbon ecosystems, encompassing mangroves, salt marshes, and seagrass beds, which substantially contribute to carbon absorption and the mitigation of anthropogenic CO_2 within the atmosphere [13,14]. Despite their relatively sparse coverage, comprising less than 0.2% of the Earth's oceans, these habitats are estimated to sequester around 10% of the annual organic carbon [15]. According to [13], seagrass ecosystems, occupying less than 0.2% of the global oceanic expanse, have the remarkable capacity to store $27.4 \text{ TgC year}^{-1}$, amounting to 10% of the ocean's annual carbon capture. A portion of the CO_2 absorbed by the oceans is integrated into living biomass and sediment deposits, with the majority remaining in an inorganic state [16]. Coastal ecosystems hold a pivotal role as intermediary regions between land and sea, functioning as vital carbon sinks [17]. As highlighted by [14,15], the sediment substrate within seagrass ecosystems hosts a significant organic carbon stock, which could be halved in the absence of seagrass meadows. Further emphasized by [18], the decline and loss of seagrass ecosystems could instigate the release of CO_2 into the atmosphere, diminishing carbon absorption and exacerbating global warming. Natural perturbations such as wave energy, eutrophication, and turbidity, coupled with human interventions like the direct release of pond waste into water bodies and the use of trawl nets, can lead to the deterioration of seagrass canopies. This, in turn, directly or indirectly exposes organic carbon sediments to oxygen-rich conditions, fostering the decomposition of organic matter [19,20]. Post-disturbance, the rate of organic carbon conversion in oxic environments is estimated at $0.0005/\text{day}$, potentially removing 70–80% of sedimentary organic carbon over a span of 40 years [21].

Functioning as a blue carbon ecosystem, a seagrass meadow serves a multifaceted purpose beyond carbon sequestration within its biomass and sediment. Its significance extends to the generation of inorganic carbon through processes like respiration or the calcification of calcium carbonate, which can notably influence air–sea CO_2 fluxes. As elucidated by [17], the air–sea CO_2 flux constitutes a pivotal mechanism determining whether an ecosystem functions as a CO_2 sink or a CO_2 source. Due to intensified anthropogenic activities, the degradation of blue carbon ecosystems, including seagrass meadows, has become a concerning reality. Under these circumstances, the ecosystem's role in regulating carbon within the waters might shift from that of a CO_2 sink to a CO_2 source. The authors of [22] conducted insightful research on CO_2 flux within seagrass ecosystems, discovering that they can act as sources of CO_2 , emitting between 0.015 and $0.346 \text{ mmol CO}_2 \text{ m}^{-2} \text{ d}^{-1}$ in Iromote Island, Japan. Similarly, [23] carried out a comprehensive investigation on CO_2 flux along Gilimanuk Bay in Bali, revealing that seagrass ecosystems act as a source of CO_2 , releasing approximately $2.5 \pm 3.4 \text{ mmol CO}_2 \text{ m}^{-2} \text{ d}^{-1}$.

Indonesia boasts an extensive expanse of seagrass ecosystems, encompassing approximately 3,000,000 ha, which accounts for 44.48% of the total seagrass area across Southeast Asia [24]. This habitat demonstrates a noteworthy diversity in terms of species, with an

estimated range of 12 to 15 seagrass species present [24]. A more detailed study by [25] has identified 13 distinct seagrass species within Indonesian waters, spanning across 22 out of the 38 provinces. Notably, the regions of Gorontalo (16.51 TgC ha⁻¹), West Sumatra (13.66 TgC ha⁻¹), North Sulawesi (11.38 TgC ha⁻¹), and Central Java (4.40 TgC ha⁻¹) exhibit the highest levels of carbon stocks and absorption. In particular, the province of Central Java emerges as a standout, boasting elevated seagrass carbon stocks and absorption rates surpassing the national average carbon uptake of Indonesia (which stands at 3.16 TgC ha⁻¹) [26]. Referring to Indonesia's seagrass distribution map, the preponderance of seagrass beds in Central Java is notably clustered within the aquatic expanse of Karimunjawa, situated within the Java Sea [24–26].

Previous studies [27,28] have primarily focused on assessing the organic carbon stock within seagrass biomass in Karimunjawa waters. Carbon sequestration evaluations conducted at Kartini Harbor and Pancuran Beach in Karimunjawa range from 0.5 to 0.73 TgC ha⁻¹ [27], whereas at the Pokemon Beaches within the same region, this range extends from 0.13 to 0.23 TgC ha⁻¹ [28]. Parallel research investigating CO₂ flux within seagrass ecosystems of Karimunjawa waters was undertaken by [29], revealing that these waters function as sources of CO₂, with emissions measuring around 8.55–13.27 mmol CO₂ m⁻² d⁻¹. This aligns with the findings of [30], who showcased the role of Karimunjawa waters as a CO₂ source, registering values of 1.79–21.64 mmol CO₂ m⁻² d⁻¹, and particularly identified two stations in the north of Kemujan Island as CO₂ sinks, ranging from −4.41 to −3.69 mmol CO₂ m⁻² d⁻¹. In the broader context, the Java Sea acts as a CO₂ source, contributing to the release of CO₂ into the atmosphere [31,32]. In light of the Karimunjawa waters acting as a CO₂ source and the seagrass bed's capacity to store carbon, comprehending the pivotal role of seagrass beds in curtailing CO₂ release within the Karimunjawa waters becomes of paramount importance. The present study stands as the inaugural endeavor to concurrently explore the organic carbon stock (OCS) within seagrass meadows and air–sea CO₂ flux in both vegetated and unvegetated areas of Karimunjawa waters. Our hypothesis posits that although the CO₂ source status of waters within the seagrass ecosystem is unknown, seagrass beds, as integral blue carbon ecosystems, can effectively absorb and retain organic carbon within their biomass and sedimentary layers.

2. Materials and Methods

2.1. Study Sites

Karimunjawa, situated in the Java Sea about 83 km northwest of Jepara, Indonesia (Figure 1), is an archipelago composed of 27 islands. Within the Karimunjawa waters, the prevailing currents exhibit a range of velocities spanning from 0.015 to 0.05 m s⁻¹, encompassing an average depth that varies from 0 to 54 m [33]. Enveloping a multitude of ecosystems, the Karimunjawa Islands showcase a diverse landscape that encompasses lowland rainforests, mangrove forests, coral reefs, algae, and seagrass meadows [34].

2.2. Data Collection

2.2.1. Primary Data

Over the course of three years, specifically during the southeast monsoon season of August in 2019, 2020, and 2022, we meticulously collected primary data encompassing CO₂ fluxes and organic carbon (C_{org}) stocks within sediments and seagrass. To comprehensively investigate the seagrass ecosystem's pivotal role in carbon storage and CO₂ fluxes, we partitioned our sampling stations into two categories: vegetated and unvegetated. The former represents stations adorned with flourishing seagrass meadows, while the latter symbolizes stations lacking such meadows. In the initial phase, encompassing August 2019, our research included six vegetated stations alongside nine unvegetated stations (Figure 1). Progressing to August 2020, our focus shifted to ten vegetated stations and five unvegetated stations (Figure 1). By August 2022, our study featured seven vegetated stations and a corresponding seven unvegetated stations (Figure 1).

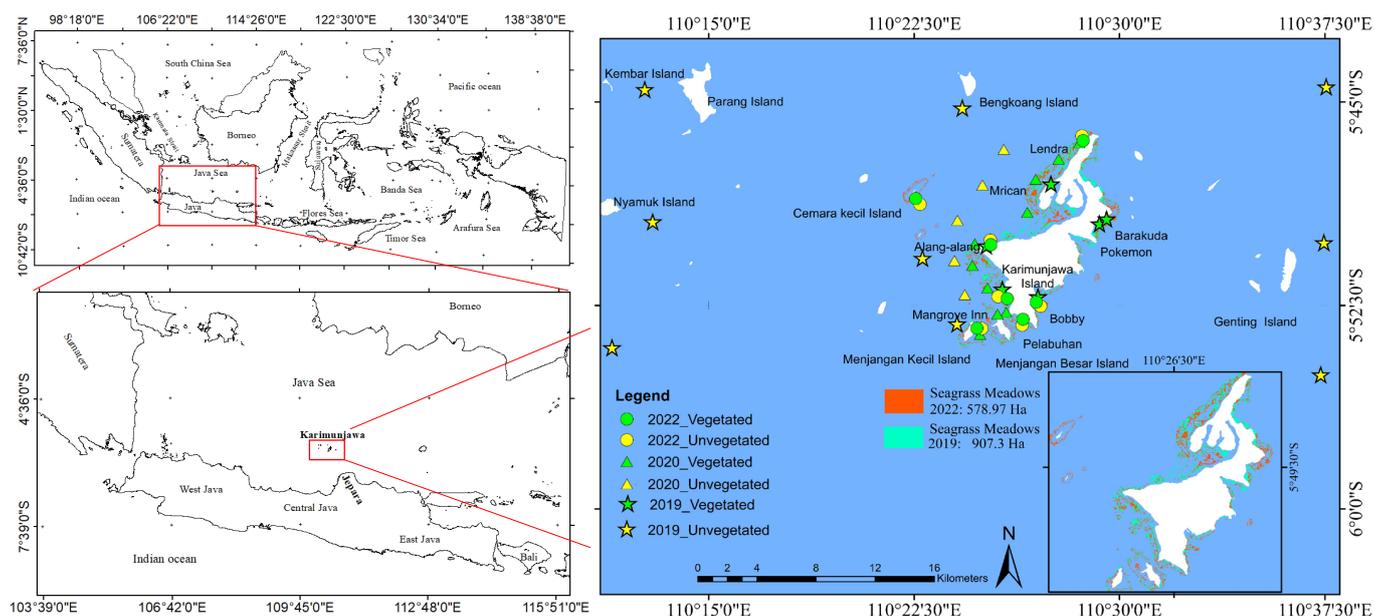


Figure 1. The research stations in Karimunjawa waters. The 2019 stations are indicated by stars (6 stations: vegetated and 9 stations: unvegetated). The 2020 stations are indicated by triangles (10 stations: vegetated and 5 stations: unvegetated). Meanwhile, the 2022 stations are indicated by circles (7 stations: vegetated and 7 stations: unvegetated). Seagrass and sediment carbon measurements were carried out in both 2019 and 2022, while measurements of air–sea CO₂ flux were taken across the years 2019, 2020, and 2022.

Vegetated (unvegetated) stations were located at depths of 0–1.3 m (2–10 m). Throughout the field survey, we diligently measured essential parameters such as sea surface temperature (SST), pH, dissolved oxygen (DO), and sea surface salinity (SSS). Additionally, our research involved the collection of a total of 94 water samples. These samples were divided into two categories: 100 mL for the meticulous analysis of nutrients (including nitrates, phosphates, and silicates) and 1500 mL for the thorough examination of chlorophyll-a levels (as outlined in Table 1).

Table 1. Parameters (water quality), measuring instruments, and analysis methods.

Parameters	Unit	Procedure	Tools/Methods
Temperature	°C	In situ measurement	Thermometer and logger
pH	-	In situ measurement	pH meter
DO	mg L ⁻¹	In situ measurement	DO meters
CO ₂ atm	ppm	In situ measurement	CO ₂ meters
Salinity	‰	In situ measurement	Refractometer
Nitrate	ppm	Laboratory measurements (100 mL sample)	Spectrophotometer/Spectrophotometry with a wavelength of 500 nm, nitra-ver 5 nitrate reagent powder pillow [35]
Silicate	ppm	Laboratory measurements (100 mL sample)	Spectrophotometer/Spectrophotometry with a wavelength of 452 nm, molybdate reagent powder pillows [36]
Total phosphate	ppm	Laboratory measurements (100 mL sample)	Spectrophotometer/Spectrophotometry with a wavelength of 890 nm, phos-ver 3 phosphate reagent powder pillow [37]
Chlorophyll-a	mg m ⁻³	Laboratory measurements (1500 mL sample)	Spectrophotometer/Spectrophotometry with a wavelength of 630, 647, 664, and 750 nm [36]

2.2.2. Secondary Data

As for the secondary data, we employed sea level pressure and wind speed from the European Centre for Medium-Range Weather Forecasts (ECMWF) Reanalysis v5 (ERA5), characterized by a spatial resolution of $0.25^\circ \times 0.25^\circ$. Moreover, we utilized xCO2atm data sourced from the Atmospheric Infrared Sounder (AIRS)/Aqua, which features a spatial resolution of $2^\circ \times 2.5^\circ$, to enrich our current study.

2.3. Estimation of Carbon Stock and Storage

At each station depicted in Figure 1, we established 2–3 transect lines, and each of these lines is equipped with five quadrats. The study utilized quadrats measuring 50×50 cm ($n = 136$; depicted in Figure 2A) to facilitate the assessment of various parameters, including shoot density, organic matter (%), seagrass biomass (gDW shoot^{-1}), organic carbon content (%), and organic carbon stock ($\text{gC}_{\text{org}} \text{m}^{-2}$). The determination of shoot density involved a meticulous count of all shoots within each quadrat. Prior to measurement, samples were meticulously cleansed of epiphytes and sediments and subsequently categorized into aboveground (AG), encompassing leaves, and belowground (BG), encompassing rhizome and root.

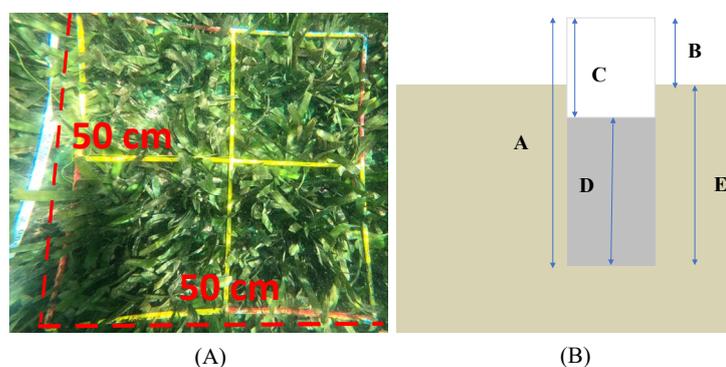


Figure 2. (A) Quadrat and (B) Sediment corer. Description: A = the length of pipe; B = the length of pipe outside the sediment; C = the length of inside pipe; D = length of sediment sample; E = depth of core (Source: [38]).

For the analysis of seagrass samples ($n = 332$ samples), a drying process at 105 – 110 °C was executed until a constant weight was achieved. In the laboratory setting, approximately 1 g of the subsample was weighed and then subjected to an oven at 105 – 110 °C for 4–8 h, followed by cooling in a desiccator for 15 min. This process was repeated until a constant sample weight was obtained. The water content was then calculated based on these measurements. In order to determine the ash content, around 1 g of the subsample was weighed, then subjected to furnace drying at 400 – 600 °C for 4–6 h, and subsequently cooled in a desiccator for 15 min. This step yielded essential parameters, namely pre-combustion mass, post-combustion mass (also referred to as mass after furnace), and dry weight (g DW).

With these parameters in hand, computations were performed to derive organic matter (using Equation (1)), organic carbon content (using Equations (2) and (3)), aboveground biomass (AGB), belowground biomass (BGB), aboveground OCS (AG-OCS), belowground OCS (BG-OCS), and organic carbon stock within the seagrass (OCS-Sg). The average total organic carbon stocks in living seagrass biomass ($\text{Mg C}_{\text{org}} \text{ha}^{-1}$) were calculated by using the average OCS-Sg ($\text{g C}_{\text{org}} \text{m}^{-2}$), which were subsequently converted to a per-hectare basis.

The collected seagrass samples were divided into AG and BG portions before undergoing analysis through the Loss on Ignition (LOI) method:

$$\%LOI = \frac{(\text{mass before combustion} - \text{mass after combustion})}{\text{mass before combustion}} \times 100, \quad (1)$$

the following formula was used to calculate organic carbon content (OCC) [13]:

$$\%C_{org} = 0.4 \times \%LOI - 0.21, \quad (2)$$

$$\%C_{org} = 0.43 \times \%LOI - 0.33. \quad (3)$$

OCS-Sg was calculated by multiplying the biomass of each seagrass species with the corresponding percentage of OCC [38]. Sediment sampling was carried out in 2019 and 2022 through the utilization of a sediment corer (Figure 2B). The sediment corer, comprising subsampling ports with a diameter of 5 cm, was covered during the collection of samples. Employing a manual hammer, the sediment corer was carefully driven into the sediment, aiming for a depth of 5–20 cm. Measurements of B (representing the length of the pipe outside the sediment) and C (indicating the length of the inside pipe) were executed to derive essential parameters. These measurements facilitated the computation of critical factors, including a compaction correction (CCF), a correction for core length (H'), and the acquisition of a volumetric sample. This volumetric sample was subsequently employed in the analysis of pivotal sediment characteristics. These characteristics encompass dry bulk density (calculated using Equation (4)), water content, organic matter (evaluated via Equation (1)), sediment organic carbon content (SOCC, determined using Equations (2) and (3)), and organic carbon stock within the sediment (OCS-S, derived from Equations (6) and (7)).

Analysis of organic matter in sediments ($n = 150$ samples) was carried out using the LOI method. Dry bulk density is calculated based on the following formula [38]:

$$\text{Dry bulk density (DBD)} \left(\text{g cm}^{-3} \right) = \frac{\text{Mass of dry sediment (g)}}{\text{Original volume sample (cm}^{-3})}, \quad (4)$$

$$\text{Original Volume sample (cm}^{-3}) = 3.14 \times r^2 \times \text{length of corrected sample}, \quad (5)$$

OCS-S is obtained by the multiplication of DBD with SOCC [38]:

$$\text{OCS-S (gC}_{org}\text{cm}^{-3}) = \text{DBD} \times \text{SOCC}, \quad (6)$$

$$\text{OCS-S (gC}_{org}\text{m}^{-2}) = \text{OCS-S (gC}_{org}\text{cm}^{-3}) \times \text{length of corrected sample} \times 10^4. \quad (7)$$

2.4. Estimation of Sea–Air CO₂ Fluxes

The calculation of CO₂ fluxes, representing the exchange of CO₂ gas flow, is calculated using the following formula [31,32,39–44]:

$$\text{CO}_2\text{Fluxes} = k_{wa} \times K_H \times (\text{pCO}_{2\text{sea}} - \text{pCO}_{2\text{atm}}), \quad (8)$$

where CO₂ fluxes is designated to represent the net carbon dioxide flux (mmol m⁻² d⁻¹), k_{wa} signifies the gas transfer velocity (m s⁻¹), and K_H stands for the solubility of CO₂ in seawater (mol m⁻³ atm⁻¹). When the CO₂ flux exhibits a positive value, the water body functions as a CO₂ source; conversely, a negative CO₂ flux indicates the water body is acting as a CO₂ sink.

The K_H value is calculated through the interplay of temperature and salinity functions [32,45]:

$$\ln K_H = -58.0931 + \frac{90.5069}{\left(\frac{T}{100}\right)} + 22.2940 \times \ln \left(\frac{T}{100}\right) + \text{SSS} \times (0.02776 - 0.02588 \times \left(\frac{T}{100}\right) + 0.0050578 \times \left(\frac{T}{100}\right)^2), \quad (9)$$

where T represents the SST (in K) and SSS denotes sea surface salinity (psu). The calculation of the k_{wa} value follows the W92 formula [46], which has been applied in numerous studies; nevertheless, this formula is most suitable for offshore sites. Given the coastal context of this study, it is important to note that utilizing the W92 formula could potentially lead to an underestimation of the flux. Thus, to address this, RC01 [47] and B04 formulas [48] were implemented for estimating estuarine CO₂ fluxes:

$$k_{wa} = 0.31 \times U_{10}^2 \times \left(\frac{660}{Sc}\right)^{0.5} \quad (\text{W92}), \quad (10)$$

$$k_{wa} = 1.91 \times e^{(0.35 \times U_{10})} \times \left(\frac{660}{Sc}\right)^{0.5} \quad (\text{RC01}), \quad (11)$$

$$k_{wa} = 5.141 \times U_{10}^{0.758} \times \left(\frac{660}{Sc}\right)^{0.5} \quad (\text{B04}), \quad (12)$$

in these formulas, U_{10} corresponds to the wind speed at a height of 10 m (m s^{-1}), while Sc represents the *Schmidt* number, and the coefficient 660 is applied to Sc for SSS (35 psu) and SST (2 °C). In this study, the k_{wa} value employed in the calculation of CO₂ fluxes, as indicated in Equation (8), represents the average of k_{wa} values derived from Equations (10) through to (12).

Furthermore, the partial pressure of atmospheric CO₂ can be calculated using the following formula [31,32,41,49]:

$$p\text{CO}_{2\text{atm}} = x\text{CO}_{2\text{atm}} \times (p_b - p\text{H}_2\text{O}), \quad (13)$$

where $x\text{CO}_{2\text{atm}}$ represents the molar fraction of carbon dioxide (ppm), p_b is sea level pressure (Pa), and $p\text{H}_2\text{O}$ denotes the saturation vapor pressure of seawater in the atmosphere. Subsequently, the calculation of $p\text{CO}_{2\text{sea}}$ (partial pressure of oceanic carbon dioxide) was carried out utilizing both the SST (in °C) and chlorophyll-a (*Chla*) approaches [32,49]:

$$p\text{CO}_{2\text{sea}} = (6.31 \times \text{SST}^2) + (61.9 \times \text{Chla}^2) - 365.85 \text{ SST} - 94.41 \text{ Chla} + 5715.94, \quad (14)$$

$$\ln p\text{H}_2\text{O} = 24.4543 - 67.4509 \times \left(\frac{100}{T}\right) - 4.8489 \times \ln\left(\frac{T}{100}\right) - 0.000544 \times \text{SSS}. \quad (15)$$

2.5. Statistical Analyses

The results are presented as the mean \pm standard error (SE). In order to satisfy the assumptions of parametric methodologies, all collected data underwent assessments for both homogeneity and normality. One-way analysis of variance was used to evaluate disparities in seagrass carbon attributes (including AGB, BGB, above- to belowground (A:B) ratio, shoot density, AG-OCS, BG-OCS, and OCS-Sg), as well as sediment carbon parameters (comprising dry bulk density, water content, %LOI, and OCS-S). Whenever statistically significant variations ($\alpha = 0.05$) were observed, subsequent pairwise comparisons of means were conducted through the Student–Newman–Keuls (SNK) test to pinpoint the specific sources of divergence [50–52]. The interplay between the dependent variable (C_{org}) and the biological as well as sediment characteristics (including shoot density, seagrass biomass, A:B ratio, dry bulk density, and water content) was scrutinized via Principal Component Analysis (PCA) within the Rstudio software framework.

3. Results

3.1. Organic Carbon Stock in the Seagrass Ecosystem

The present study reveals the presence of seven distinct seagrass species within the Karimunjawa waters. These species include *Enhalus acoroides* (Ea), *Thalassia hemprichii* (Th), *Cymodocea rotundata* (Cr), *Oceana serrulata* (Os), *Halophila ovalis* (Ho), *Syringodium*

isoetifolium (Si), and *Halodule uninervis* (Hu). An analysis of OCS-Sg among these seagrass species demonstrated statistically significant differences ($p < 0.001$) at a 95% confidence level. The seagrass species were ordered based on their organic carbon stock content ($\text{gC}_{\text{org}} \text{m}^{-2}$), with Th exhibiting the highest value (19.4 ± 1.8), followed by Ea (11.0 ± 1.5), Cr (8.1 ± 1.3), Os (5.1 ± 1.1), Ho (3.5 ± 1.0), Hu (3.0 ± 1.7), and Si (0.9 ± 0.5), as depicted in Figure 3. Th and Ea were observed across nearly all stations, while Cr, Os, Hu, and Si were found in less than 25% of the sampling locations. Interestingly, when these less frequently encountered species were present in the sample quadrat, their OCS values could range from 20 to 50 $\text{gC}_{\text{org}} \text{m}^{-2}$. Consequently, the box plots shown in Figure 3 for Cr, Os, Ho, Hu, and Si display numerous data points extending beyond their upper extremes, owing to the inclusion of instances where species were absent throughout the study area. In order to achieve uniformity in the measurement of OCS for both above- and belowground components, we integrated zero counts for species that were not detected in particular sampling locations. This strategic approach plays a pivotal role in constructing a more precise depiction of the variations in OCS.

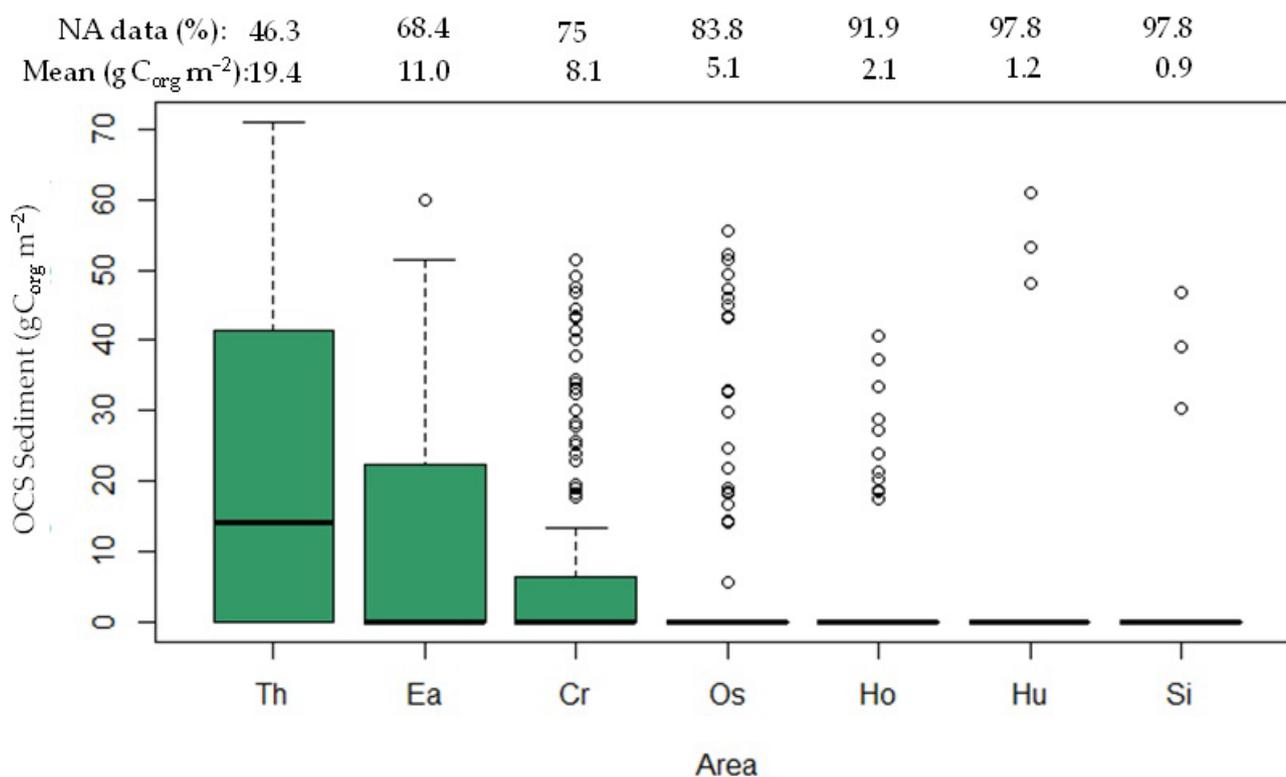


Figure 3. OCS-Sg in Karimunjawa waters. The notation NA (%) signifies data unavailability due to the absence of certain seagrass species within the quadrat.

Our data underscores a consistent pattern, revealing that the majority of AG-OCS values were notably lower than the corresponding BG-OCS values. Specifically, the average measurements for AG-OCS and BG-OCS were $15.9 \pm 0.9 \text{ gC}_{\text{org}} \text{m}^{-2}$ and $17.4 \pm 0.8 \text{ gC}_{\text{org}} \text{m}^{-2}$, respectively. Importantly, this disparity was statistically significant ($p = 0.02$) at a 95% confidence level. These AG-OCS and BG-OCS values are susceptible to diminishment due to the ongoing annual reduction in the extent of seagrass meadows within Karimunjawa waters.

Over successive years, this area has experienced a concerning decrease. Specifically, in 2019, seagrass meadows spanned 907.3 ha, but by 2022, this area had contracted to 578.97 ha (Figure 1). This trend is particularly evident when translating OCS measurements into the context of the seagrass meadow area. This conversion demonstrates a clear reduction in the overall OCS-Sg within the Karimunjawa study area. Notably, this metric declined from 262.66 Mg C_{org} in 2019 to 214.59 Mg C_{org} in 2022.

A projection based on the hypothetical scenario of maintaining the 2022 seagrass area equivalent to that of 2019 yields significant insights. It suggests that the total OCS-Sg could have doubled by 2022. Tragically, the actual scenario reflects the loss of half of the seagrass meadow areas, although the cumulative OCS-Sg remains above the 200 Mg C_{org} threshold. These findings underscore the critical role played by seagrass meadows in preserving the cumulative OCS-Sg. The sustainability of these meadows emerges as a paramount concern in this context.

Among the three monitoring stations that gathered data in both 2019 and 2022 (Mangrove Inn, Alang–Alang, and Bobby, as shown in Figure 1), a noteworthy pattern emerged. Two of these stations exhibited an increasing trend in OCS-Sg levels. Specifically, at the Mangrove Inn and Alang–Alang sites, OCS-Sg experienced an increase from 34.5 to 43.3 gC_{org} m⁻² and from 26.6 to 42.6 gC_{org} m⁻², respectively (Table 2). However, a different trend was observed at the Bobby station, where OCS-Sg decreased from 40.17 gC_{org} m⁻² in 2019 to 28.5 gC_{org} m⁻² in 2022. The augmentation in shoot density played a pivotal role in driving the rise in OCS-Sg at these locations. Additionally, a notable surge in the number of seagrass species, more than doubling in count, contributed significantly to the observed OCS increase at the Mangrove Inn and Alang–Alang sites. Taken together, the data amassed from 2019 to 2022 showcases a distinct shift. This shift is statistically significant ($p = 0.002$) at a 95% confidence level, reflecting an average OCS-Sg increase from 28.9 ± 1.33 gC_{org} m⁻² to 37.8 ± 2.29 gC_{org} m⁻².

The expanse of seagrass meadow in the Karimunjawa waters has been progressively diminishing over the years. This decline can be attributed to the conversion of land formerly occupied by mangrove forests into shrimp ponds, a trend that continues to intensify annually. In 2015, the area dedicated to shrimp ponds spanned 21.93 ha, but by 2021, it had expanded significantly to 47.46 ha (Figure 4). The transition of shrimp pond operations from traditional to high-density, intensive methods has resulted in the discharge of untreated effluent directly into the ocean. This discharge poses a concerning threat to aquatic ecosystems, including seagrass beds, due to potential contamination.

Given that seagrass meadows constitute vital blue carbon habitats capable of mitigating atmospheric CO₂ levels through absorption and storage within their biomass and sediments, it is imperative to prioritize their long-term viability. Sediment substrates housing vegetation (seagrass-vegetated sediments) exhibit a notably higher OCS (41.5 ± 2.01 gC_{org} m⁻²) compared to unvegetated substrates (unvegetated sediments) (32.4 ± 2.9 gC_{org} m⁻²), with a statistically significant difference ($p = 0.018$) at a 95% confidence level. Remarkably, the highest OCS-S is observed in vegetated stations, particularly at Mangrove Inn in the western sector (63.6 ± 10.4 gC_{org} m⁻²), Lendra in the northern sector (54.6 ± 2.9 gC_{org} m⁻²), and Bobby in the eastern sector (47.3 ± 1.7 gC_{org} m⁻²) (Table 3). This emphasizes the pivotal role of seagrass vegetation in bolstering carbon storage and underscores the importance of addressing its preservation.

Table 2. Seagrass species, shoot density, tissue carbon content, biomass, A:B ratio, and OCS at the study sites along the coast of Karimunjawa (mean ± SE).

Study Sites	Seagrass Species	Shoot Density (Shoots m ⁻²)	Tissue Carbon Content/OCC (%)		Biomass (gDW m ⁻²)		A:B Ratio	OCS (gC _{org} m ⁻²)		OCS-Sg (gC _{org} m ⁻²)
			AG	BG	AG	BG		AG	BG	
East										
Bobby										
2019 (n = 22)	Ea, Th, Cr, Ho, Hu	446.5 ± 172.7	6.5 ± 0.8	7.6 ± 1.0	2.8 ± 0.3	4.5 ± 0.7	0.7 ± 0.0	15.6 ± 1.4	24.6 ± 3.9	40.17 ± 4.8
2022 (n = 32)	Ea, Th, Cr, Os, Ho	212.7 ± 40.2	4.9 ± 1.2	8.3 ± 1.4	3.6 ± 0.5	2.2 ± 0.5	1.9 ± 0.3	16.0 ± 2.9	15.2 ± 3.3	28.5 ± 6.2
Pokemon										
2019 (n = 18)	Ea, Th, Cr, Ho	278.2 ± 85.3	4.9 ± 0.0	5.6 ± 0.0	1.3 ± 0.2	2.1 ± 0.4	0.5 ± 0.1	9.3 ± 0.5	14.8 ± 0.8	18.7 ± 3.6
Barakuda										
2019 (n = 18)	Ea, Th, Cr, Hu	158.2 ± 51.2	5.0 ± 0.0	5.6 ± 0.0	1.4 ± 0.0	2.6 ± 0.2	0.6 ± 0.0	7.8 ± 0.3	14.4 ± 1.3	22.2 ± 1.4
West										
Mangrove Inn										
2019 (n = 18)	Ea, Th	39.6 ± 4.5	4.9 ± 0.0	5.6 ± 0.0	2.3 ± 0.2	3.8 ± 0.3	0.6 ± 0.1	13.2 ± 0.9	21.3 ± 1.8	34.5 ± 2.1
2022 (n = 34)	Ea, Th, Cr, Os, Ho	74.8 ± 16.5	7.9 ± 1.5	9.7 ± 1.8	4.4 ± 0.4	3.5 ± 1.1	2.1 ± 0.5	21.1 ± 2.4	21.5 ± 6.6	43.3 ± 8.1
Alang–Alang										
2019 (n = 22)	Ea, Th, Cr, Os	156.4 ± 40.1	5.4 ± 1.0	6.2 ± 1.1	2.1 ± 0.2	2.6 ± 0.2	0.8 ± 0.1	12.0 ± 1.1	14.6 ± 1.2	26.6 ± 1.8
2022 (n = 40)	Ea, Th, Cr, Os, Ho	188.8 ± 33.4	7.8 ± 1.1	11.5 ± 1.8	4.7 ± 0.8	3.5 ± 0.5	1.3 ± 0.1	22.3 ± 4.8	20.3 ± 3.3	42.6 ± 6.6
Mrican										
2019 (n = 20)	Ea, Th, Cr, Os	107.6 ± 12.7	5.4 ± 0.5	6.1 ± 0.6	1.8 ± 0.2	3.0 ± 0.4	0.6 ± 0.1	10.1 ± 1.1	16.1 ± 2.1	26.1 ± 2.7
Cemara Kecil										
2022 (n = 18)	Th	205.2 ± 29.0	4.5 ± 0.5	5.3 ± 0.6	1.6 ± 0.3	1.7 ± 0.2	0.9 ± 0.2	10.2 ± 1.4	10.8 ± 0.5	18.9 ± 2.4
North										
Lendra										
2022 (n = 36)	Ea, Th, Cr, Os, Ho	250 ± 35.0	9.2 ± 1.0	10.4 ± 1.2	3.1 ± 0.5	3.3 ± 0.4	0.9 ± 0.1	18.4 ± 3.5	19.3 ± 2.3	37.7 ± 5.2
South										
Pelabuhan										
2022 (n = 46)	Ea, Th, Cr, Ho, Si	339.7 ± 37.9	8.2 ± 0.9	9.2 ± 1.0	4.3 ± 0.6	2.7 ± 0.3	1.7 ± 0.2	26.7 ± 3.4	16.0 ± 1.8	42.7 ± 4.8
Menjangan kecil										
2022 (n = 63)	Ea, Th, Cr, Os, Ho	273.6 ± 28.9	9.2 ± 1.3	11.0 ± 1.5	3.9 ± 0.6	4.1 ± 0.5	0.9 ± 0.1	18.8 ± 3.5	22.0 ± 3.0	41.0 ± 6.1

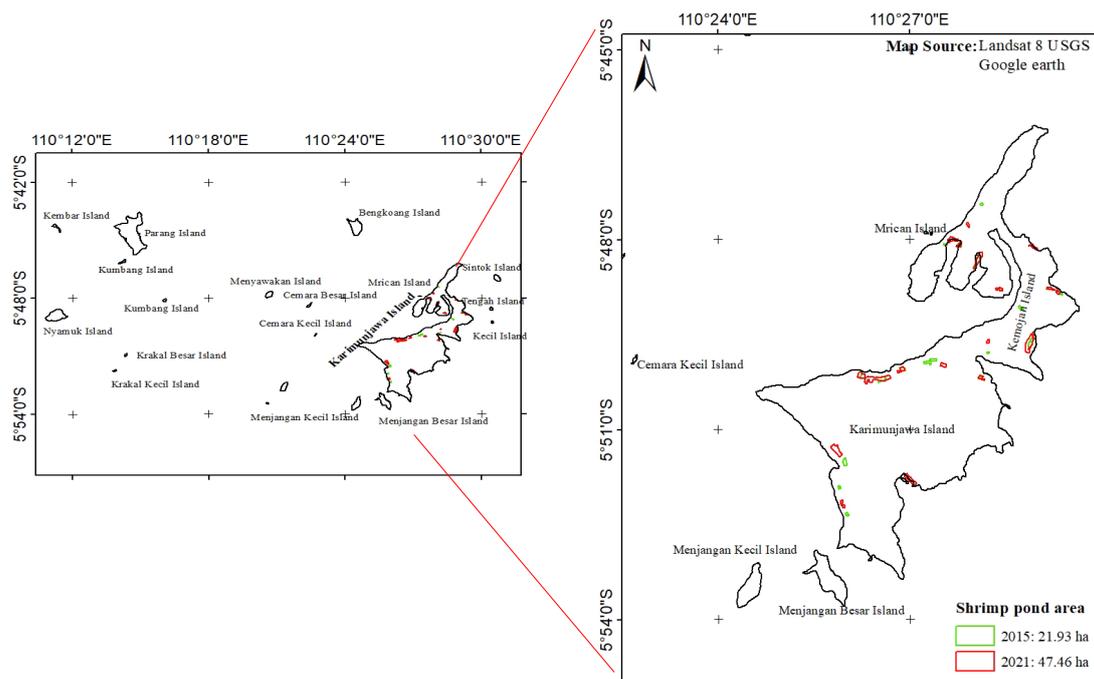


Figure 4. The shrimp pond areas within the Karimunjawa waters for the years 2015 and 2021.

Table 3. OCS-S at the study sites along the coast of Karimunjawa (mean ± SE).

Study Sites	Water Content (%)	Dry Bulk Density (g cm ⁻³)	%LOI	SOCC (%)	OCS-S (gC _{org} m ⁻²)
East					
Bobby					
2019 (vegetated; n = 9)	18.7 ± 0.8	0.0051 ± 0.0005	23.0 ± 0.7	9.5 ± 0.3	39.9 ± 0.9
2022 (vegetated; n = 12)	22.7 ± 1.7	0.0048 ± 0.0004	28.7 ± 1.7	12.0 ± 0.7	47.3 ± 1.7
2022 (unvegetated; n = 2)	41.7 ± 8.4	0.0047 ± 0.0006	34.7 ± 1.9	14.6 ± 0.8	37.8 ± 3.6
Pokemon					
2019 (vegetated; n = 9)	15.8 ± 0.6	0.0053 ± 0.0005	20.0 ± 0.7	8.3 ± 0.3	35.3 ± 1.0
Barakuda					
2019 (vegetated; n = 9)	15.0 ± 0.7	0.0049 ± 0.0002	19.5 ± 0.7	8.0 ± 0.3	34.8 ± 1.0
West					
Mangrove Inn					
2019 (vegetated; n = 9)	21.1 ± 0.8	0.0035 ± 0.0002	25.3 ± 0.8	10.6 ± 0.4	42.3 ± 1.0
2022 (vegetated; n = 10)	19.0 ± 1.4	0.0036 ± 0.0002	36.0 ± 5.2	15.2 ± 2.2	63.6 ± 10.4
2022 (unvegetated; n = 2)	16.3 ± 2.2	0.0070 ± 0.0002	27.4 ± 0.1	11.4 ± 0.0	48.8 ± 1.1
Alang–Alang					
2019 (vegetated; n = 9)	17.8 ± 1.1	0.0043 ± 0.0004	22.0 ± 1.1	9.1 ± 0.5	38.1 ± 0.3
2022 (vegetated; n = 10)	18.6 ± 1.2	0.0041 ± 0.0004	27.0 ± 1.6	11.3 ± 0.7	47.1 ± 3.3
2022 (unvegetated; n = 2)	14.1 ± 0.4	0.0072 ± 0.0000	22.6 ± 0.3	9.4 ± 0.1	41.2 ± 0.3
Mrican					
2019 (vegetated; n = 9)	11.1 ± 1.7	0.0046 ± 0.0004	13.7 ± 1.8	5.6 ± 0.8	24.7 ± 2.9
Cemara Kecil					
2022 (vegetated; n = 10)	31.9 ± 3.5	0.0033 ± 0.0004	33.2 ± 3.3	13.9 ± 1.4	48.2 ± 4.8
2022 (unvegetated; n = 2)	21.6 ± 0.6	0.0059 ± 0.0006	17.9 ± 2.2	7.4 ± 0.9	29.5 ± 3.9
North					
Lendra					
2022 (vegetated; n = 10)	19.6 ± 1.3	0.0046 ± 0.0002	31.7 ± 1.4	13.3 ± 0.6	54.6 ± 3.0
2022 (unvegetated; n = 2)	21.1 ± 1.9	0.0048 ± 0.0000	9.7 ± 3.4	3.8 ± 1.5	15.7 ± 6.3
South					
Pelabuhan					
2022 (vegetated; n = 15)	16.5 ± 1.0	0.0045 ± 0.0003	22.1 ± 1.0	9.2 ± 0.4	38.9 ± 1.8
2022 (unvegetated; n = 2)	25.0 ± 2.7	0.0063 ± 0.0003	20.2 ± 2.2	8.3 ± 0.9	31.8 ± 2.5
Menjangan kecil					
2022 (vegetated; n = 15)	27.3 ± 2.6	0.0039 ± 0.0004	26.7 ± 1.2	11.1 ± 0.5	41.1 ± 2.4
2022 (unvegetated; n = 2)	27.0 ± 0.7	0.0061 ± 0.0000	27.4 ± 0.2	6.9 ± 0.1	25.6 ± 0.0

The combined effect of various predictors accounted for 55.6% of the variability observed in OCS-S (Figure 5). The outcomes of the PCA analysis revealed that the A:B ratio and water content exerted a positive influence on the OCS-S, while dry bulk density had a negative impact (Figure 5). These findings align with prior research, as noted in [19,53]. In general, a higher shoot density and greater BGB correlate positively with higher OCS-S, owing to their capacity to capture particles and accumulate C_{org} in the sediment, a concept substantiated by [52]. The elevated OCS-S can be attributed to the intricate canopy structure’s elevated complexity, as discussed in [19,54].

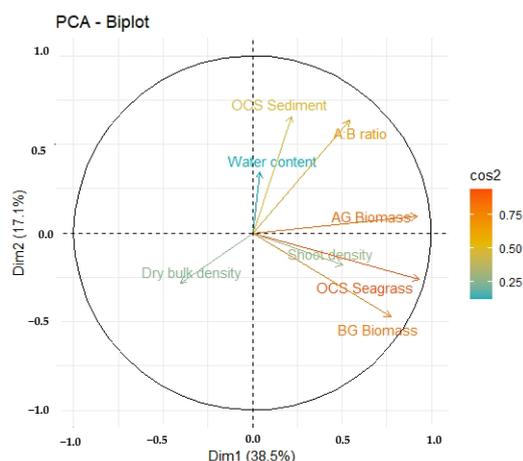


Figure 5. The results of the PCA analysis conducted on carbon stock and storage in seagrass and sediments.

3.2. Air–Sea CO₂ Fluxes

During the southeast monsoon season in Karimunjawa’s waters, a noticeable disparity in CO₂ flux values came to light between vegetated and unvegetated areas. Specifically, the vegetated regions exhibited a significantly higher average CO₂ flux value of 24.4 ± 4.55 mmol CO₂ m⁻² d⁻¹. In contrast, the unvegetated locations displayed a somewhat lower average CO₂ flux value, approximately 20.0 ± 4.34 mmol CO₂ m⁻² d⁻¹. This discrepancy amounted to a notable 4.4 mmol CO₂ m⁻² d⁻¹ reduction in CO₂ concentration when compared to the CO₂ flux values observed in vegetated zones.

In 2022, there was a notable elevation in CO₂ flux, particularly evident in the northern part of vegetated stations (Lendra: 156.96 mmol CO₂ m⁻² d⁻¹) and the western region of the vegetated station (Mangrove Inn: 97.05 mmol CO₂ m⁻² d⁻¹). This surge in CO₂ flux can be attributed to elevated SST during the sampling period at Lendra Beach and Mangrove Inn. In these areas, the SST reached a relatively high 33.5 °C within the vegetated area and 32.1 °C within the unvegetated zone, in contrast to other stations where the average SST in Karimunjawa was about 29.93 °C. The heightened temperatures prompted a concurrent rise in pCO_{2sea} values, with Lendra recording a pCO_{2sea} value of 529.745 µatm, and Mangrove Inn showing pCO_{2sea} values of 493.697 µatm in 2019 and 486.091 µatm in 2022. Additionally, stronger winds contributed to the CO₂ flux. This trend is evident in Figure 6, which highlights a strong positive Pearson correlation between CO₂ flux and several variables: pCO_{2sea} ($r = 0.975$ with $p < 0.001$), SST ($r = 0.790$ with $p < 0.001$), and k_{wa} ($r = 0.580$ with $p < 0.001$) (see Figure 6).

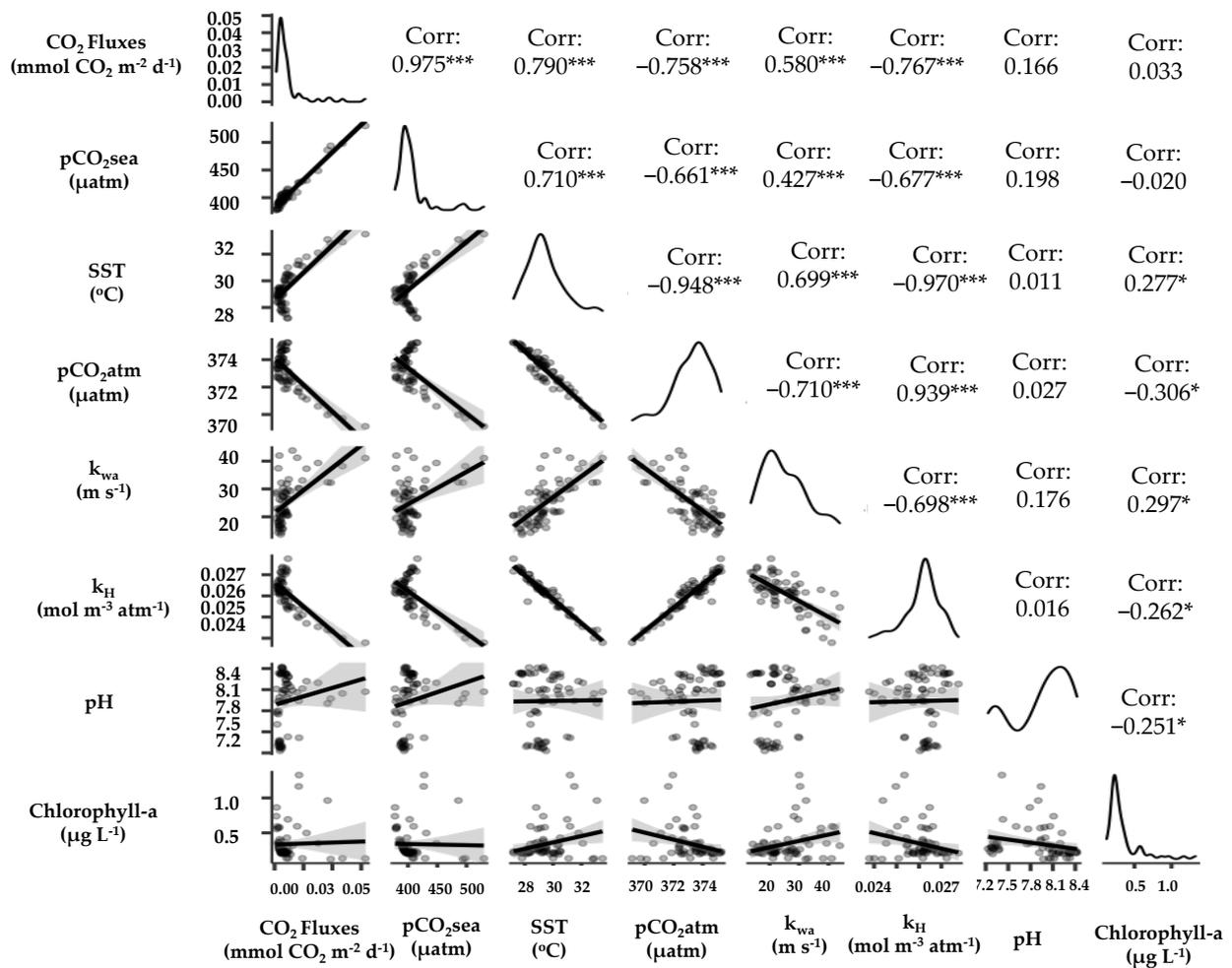


Figure 6. The correlations among various variables, including CO₂ fluxes, pCO_{2sea}, SST, pCO_{2atm}, k_{wa} , k_H , pH, and chlorophyll-a. The graphs visually represent the linear relationships between these

variables along the x and y axes. Statistical significance is denoted by asterisks (*), with * and *** indicating p -values of <0.05 , and <0.001 , respectively. The data points are represented as dots, which serve to display the spread of the data. A more tightly clustered distribution of data around a sloping straight line indicates a stronger correlation, whereas a broader dispersion suggests a weaker one. The correlation coefficient (Corr) and the orientation of the straight line together convey the strength and direction of the relationship between the variables. A positive Corr value, as well as a straight line that slopes upwards to the right and another that slopes downwards to the left, all indicate a positive relationship. Conversely, a negative Corr value, in conjunction with a straight line sloping downwards to the right and another sloping upwards to the left, signifies a negative relationship.

3.3. Organic Carbon Stock and Air–Sea CO₂ Flux

In the context of investigating the interplay between seagrass and CO₂ flux in Karimunjawa waters, a distinct separation was made between vegetated stations enriched with additional carbonate sources (vegetated + source carbonate in 2022) and other seagrass-covered stations (vegetated in 2022) (depicted in Figure 7A). Drawing insights from Figure 7A, it becomes evident that three stations—Lendra, Mangrove Inn, and Menjangan Kecil—stand out due to their notably elevated levels of OCS-S and CO₂ flux.

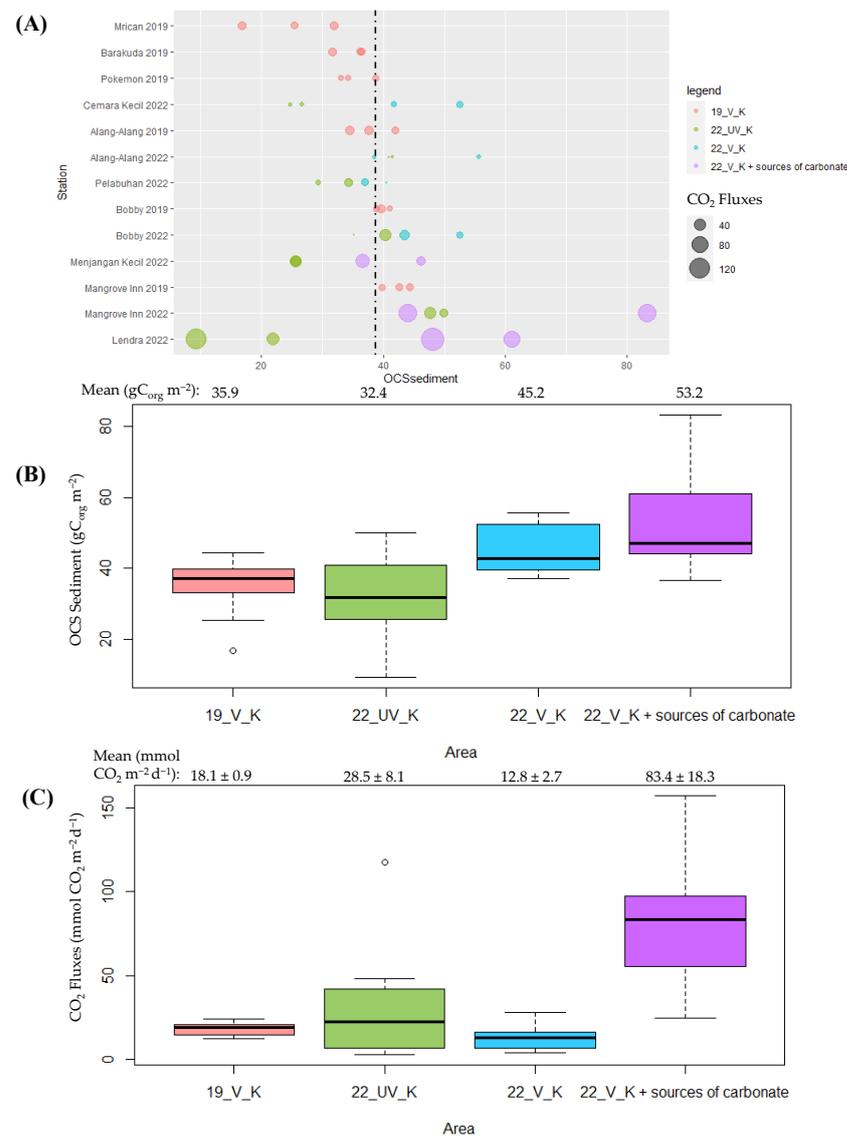


Figure 7. (A) The correlation between OCS-S and air–sea CO₂ fluxes; (B) the distribution of OCS-S per unit area; and (C) the distribution of CO₂ fluxes per unit area at the study site located along the

coast of Karimunjawa. The stations are labeled as follows: 22_V_K + source carbonate (depicted in purple) represents vegetated stations with additional source carbonate in 2022; 22_V_K (in blue) signifies vegetated stations in 2022; 22_UV_K (in green) represents unvegetated stations in 2022; and 19_V_K (in pink) represents vegetated stations from 2019.

Of these stations, those characterized by seagrass vegetation along with supplemental carbonate sources (represented in purple) exhibit the highest content of sediment organic carbon ($53.2 \pm 6.8 \text{ gC}_{\text{org}} \text{ m}^{-2}$). This reading surpasses the organic carbon content found in the vegetated stations of 2022 (blue, $45.2 \pm 2.6 \text{ gC}_{\text{org}} \text{ m}^{-2}$), the vegetated stations of 2019 (pink, $35.9 \pm 1.6 \text{ gC}_{\text{org}} \text{ m}^{-2}$), and even the least unvegetated zones of 2022 (green, $32.4 \pm 2.9 \text{ gC}_{\text{org}} \text{ m}^{-2}$) (as shown in Figure 7B). This observation underscores the pivotal role that seagrass plays in facilitating the accumulation of organic carbon stock within the sediment layers.

In contrast, the highest recorded air–sea CO_2 flux value was observed at the vegetated station featuring an additional carbonate source, registering a mean of $83.4 \pm 18.3 \text{ mmolCO}_2 \text{ m}^{-2} \text{ d}^{-1}$ (depicted in Figure 7C). These stations with supplemental carbonate sources are derived from three distinct origins: (1) Mangrove Inn (fed by allochthonous inputs from mangrove, as shown in Figure 8A); (2) Menjangan Kecil (where *Padina pavonica* constitutes more than 50% of the seagrass meadow composition, as depicted in Figure 8B); and (3) Lendra (enriched with calcareous epiphytes, as seen in Figure 8C).

(A). Mangrove Inn



(B). Menjangan Kecil



(C). Lendra

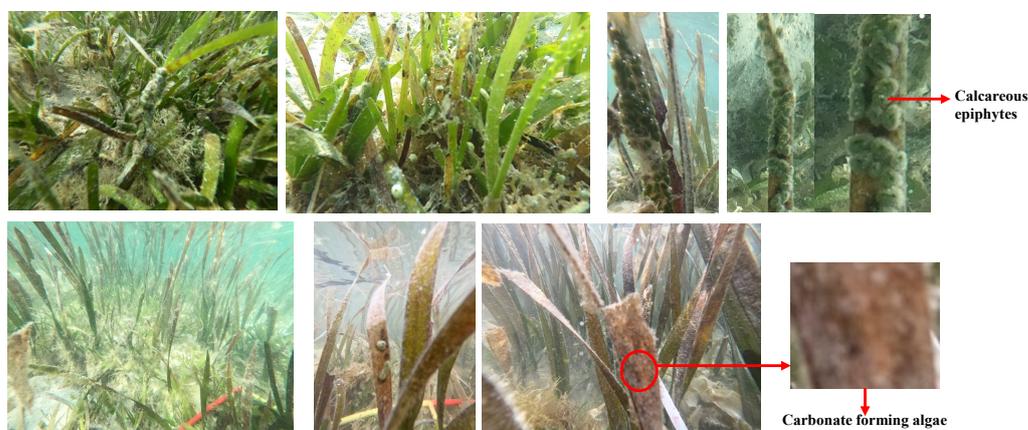


Figure 8. The state of seagrass within the Karimunjawa study area in 2022: (A) Supplementary allochthonous carbon input originating from mangroves; (B) the presence of calcifying brown algae, *Padina pavonica*; and (C) the existence of calcareous epiphytes and algae contributing to carbonate formation.

When comparing this value to vegetated waters devoid of additional carbonate sources, a noticeable disparity emerges with lower values of $18.1 \pm 0.9 \text{ mmol CO}_2 \text{ m}^{-2} \text{ d}^{-1}$ (in 2019) and $12.8 \pm 2.7 \text{ mmol CO}_2 \text{ m}^{-2} \text{ d}^{-1}$ (in 2022). This highlights how the presence of supplementary carbonate sources within vegetated waters enhances their inorganic carbon content, thereby elevating their $\text{pCO}_{2\text{sea}}$ levels and leading to an augmented release of CO_2 into the atmosphere. Furthermore, when evaluating the sea–air CO_2 flux values between vegetated and unvegetated waters, a notable difference is evident, with the CO_2 flux of unvegetated waters surpassing that of vegetated waters by $28.5 \pm 8.1 \text{ mmol CO}_2 \text{ m}^{-2} \text{ d}^{-1}$. This observation underscores the role of seagrass vegetation in reducing the quantity of inorganic carbon within the water, resulting in lower $\text{pCO}_{2\text{sea}}$ values and, consequently, a reduced flow of CO_2 into the atmosphere.

4. Discussion

Building upon the findings from [55], it was determined that Th exhibited the highest Gross Primary Production (GPP) ($9.95 \pm 7.25 \text{ mmol O}_2 \text{ g DW}^{-1} \text{ d}^{-1}$) among a diverse range of 15 seagrass species, taking into consideration the seagrass biomass ratio within the seagrass meadow. Conversely, as elucidated by [56], smaller seagrass species such as Ho displayed elevated OCS-Sg due to their thriving presence within sediment environments. In contrast, smaller-sized seagrass varieties like Si might not support as substantial OCS-Sg as Ho. These earlier investigations align with our own finding that Th boasts the most significant standing OCS-Sg at $19.4 \pm 1.8 \text{ gC}_{\text{org}} \text{ m}^{-2}$. Meanwhile, Si exhibited the smallest OCS-Sg, registering at $0.9 \pm 0.5 \text{ gC}_{\text{org}} \text{ m}^{-2}$. Ho, surpassing Si, demonstrated an OCS-Sg of $3.5 \pm 1.0 \text{ gC}_{\text{org}} \text{ m}^{-2}$.

In 2022, the total OCS within the study area registered a decrease, totaling 218.78 tC, in stark contrast to the reading of 452.39 tC recorded in 2019. This decline can be attributed to a substantial reduction of 328.33 ha in seagrass coverage. Notably, certain regions have undergone transformation, transitioning into shrimp ponds and fishing zones equipped with trawl nets. Such alterations in land use have direct repercussions on the viability of seagrass and lead to the contraction of the seagrass meadow. As cited in [57], global statistics indicate that a considerable 29% of the overall seagrass expanse has been lost, consequently compromising the carbon sink capacity of these ecosystems. The ongoing trend of land conversion to pond setups bears the potential for further shrinkage of seagrass areas. Consequently, this continuous decline in seagrass territory could subsequently result in an annual reduction in carbon uptake by these ecosystems. Given the imperatives of climate mitigation, prioritizing seagrass conservation and restoration within the waters of Karimunjawa becomes paramount. This is especially crucial in regions with limited species diversity and scant seagrass coverage.

The OCS-Sg in this study (ranging from 0.05 to 1.03 tC ha^{-1}) falls within Indonesia's range of $0.94\text{--}1.15 \text{ tC ha}^{-1}$ [25]. Notably, a significant portion of organic carbon, about 58.10%, is stored in sediments, while the remaining portion is distributed between above- and belowground seagrass biomass, accounting for 41.90%. These findings align well with comparable studies indicating a prevalence of sediment $\text{C}_{\text{org}} >$ seagrass C_{org} , both globally ($87.63\% > 12.37\%$) [13], in Indonesia ($97.99\% > 2.01\%$) [58], and on the Korean Peninsula ($96\text{--}99\% > 0.67\text{--}3.33\%$) [52].

The substantial proportion of seagrass C_{org} highlights the pivotal role of seagrass in carbon absorption, underscoring the high potential of Karimunjawa waters' sediments to sequester even more carbon in the future. Additionally, this study reveals that the content of OCS in vegetated sediments ($31.6 \pm 3.15 \text{ gC}_{\text{org}} \text{ m}^{-2}$) surpasses that of unvegetated sediments ($16.2 \pm 3.44 \text{ gC}_{\text{org}} \text{ m}^{-2}$), an observation supported by previous findings [52,59]. Furthermore, it is worth noting that blue carbon ecosystems, as mentioned by [52,59], exhibit remarkably high sediment carbon accumulation rates (CARs), which are 30–50 times greater than those found in terrestrial forests.

The elevated OCS-S value observed in vegetated areas in comparison to unvegetated sites underscores the significant contribution of seagrass to both the deposition and long-

term preservation of OCS-S. This phenomenon can be attributed to the synergy of factors, including the protracted decomposition of organic matter under anoxic sediment conditions, in conjunction with the abundant C_{org} content and elevated Carbon-Nitrogen-Phosphorus (CNP) ratios associated with seagrass habitats [52]. Remarkably, the heightened CARs observed in seagrass ecosystems are closely linked to the gradual degradation of organic materials within anoxic sediment environments, along with the substantial proportions of C_{org} and the elevated CNP ratios characteristic of seagrass habitats [52]. Furthermore, there is a well-documented prevalence of elevated C_{org} deposition rates within seagrass beds that are relatively sheltered, experience low hydrodynamic conditions, and receive moderate to low nutrient inputs [60].

Large seagrass species (such as Ea and Th) exhibit a heightened capacity to attenuate ocean currents through their dense canopies, thereby proving effective in facilitating sediment deposition [61]. Notably, sediment attributes, including water content, are acknowledged to exert a positive impact on OCS-S. Conversely, attributes like dry bulk density are recognized for their adverse influence on OCS-S. Similar conclusions have been drawn by other studies, affirming that diminished sediment density allows for mineral organic particles to outweigh it in composition [61], while concurrently displaying elevated water content [62] that contributes to heightened OCS-S. Based on the findings derived, it is deducible that the waters around Karimunjawa possess a substantial potential for harboring abundant OCS. This is attributed to the pronounced influence of robust seagrass biomass, a notable A:B ratio, diminished dry bulk density, and elevated water content.

The air–sea CO_2 flux results observed in the Karimunjawa waters during the southeast monsoon (ranging from 2.99 to 156.96 $mmol CO_2 m^{-2} d^{-1}$) surpassed those of prior studies conducted by [32] in the Java Sea, which ranged from 0.1 to 20 $mmol CO_2 m^{-2} d^{-1}$. Notably, all monitoring stations in this study were identified as contributors to CO_2 exchange with the atmosphere. Evidently, areas featuring seagrass meadows (vegetated zones) displayed a positive CO_2 flux value, signifying a CO_2 source. Amidst relatively uniform wind conditions across the study sites, these vegetated regions exhibited elevated CO_2 flux, primarily due to ocean temperature effects.

The challenge of CO_2 dissolution in warm tropical waters is underscored by elevated sea temperatures, as indicated by previous research [59,63,64]. Consequently, CO_2 tends to be liberated back into the atmosphere, rendering tropical waters a predominant source of CO_2 [29,31,32,65–67]. Conversely, in subtropical waters characterized by lower temperatures, the increased solubility of CO_2 facilitates oceanic absorption (CO_2 sink) [68–72]. This aligns with research focused on CO_2 flux within the Northeast Atlantic Ocean during summer, where the presence of a high pCO_{2sea} ($426.77 \pm 0.83 \mu atm$) coincided with maximum temperatures ($22.93 \pm 0.05 ^\circ C$) [72].

The air–sea CO_2 flux within vegetated regions is also subject to the influence of inorganic carbon, which can elevate pCO_{2sea} , thereby leading to heightened air–sea CO_2 exchange. This phenomenon is vividly depicted in Figure 9A, wherein the presence of calcium carbonate reservoirs within seagrass vegetation yields a notable impact on bolstering the organic carbon stores in the aquatic environment. As a result, the air–sea CO_2 flux reaches an elevated level of $83.4 \pm 18.3 mmol CO_2 m^{-2} d^{-1}$. This relationship is mirrored in the sediment domain as well, with organic carbon levels reaching $53.2 \pm 6.8 gC_{org} m^{-2}$. Various studies have asserted that the supplementary inorganic carbon within seagrass beds is contributed by sources such as mangroves [23], macroalgae [73–76], and land-based pollutants, which can lead to the colonization of epiphytes (macrophytes) [77]. Notably, additional sources of carbonate (inorganic carbon) can be attributed to the presence of mangroves in the Mangrove Inn (depicted in Figure 8A). The introduction of allochthonous carbon from upstream sources stems from contributions originating from rivers and mangroves. As elucidated by the findings of [23], the allochthonous influx of mangroves, primarily in the form of groundwater input (measured at $18.1 \pm 5.8 mmol CO_2 m^{-2} d^{-1}$), emerges as a significant factor driving the escalation of pCO_{2sea} . This underscores the pivotal role of water in acting as a substantial source of CO_2 within mangrove ecosystems.

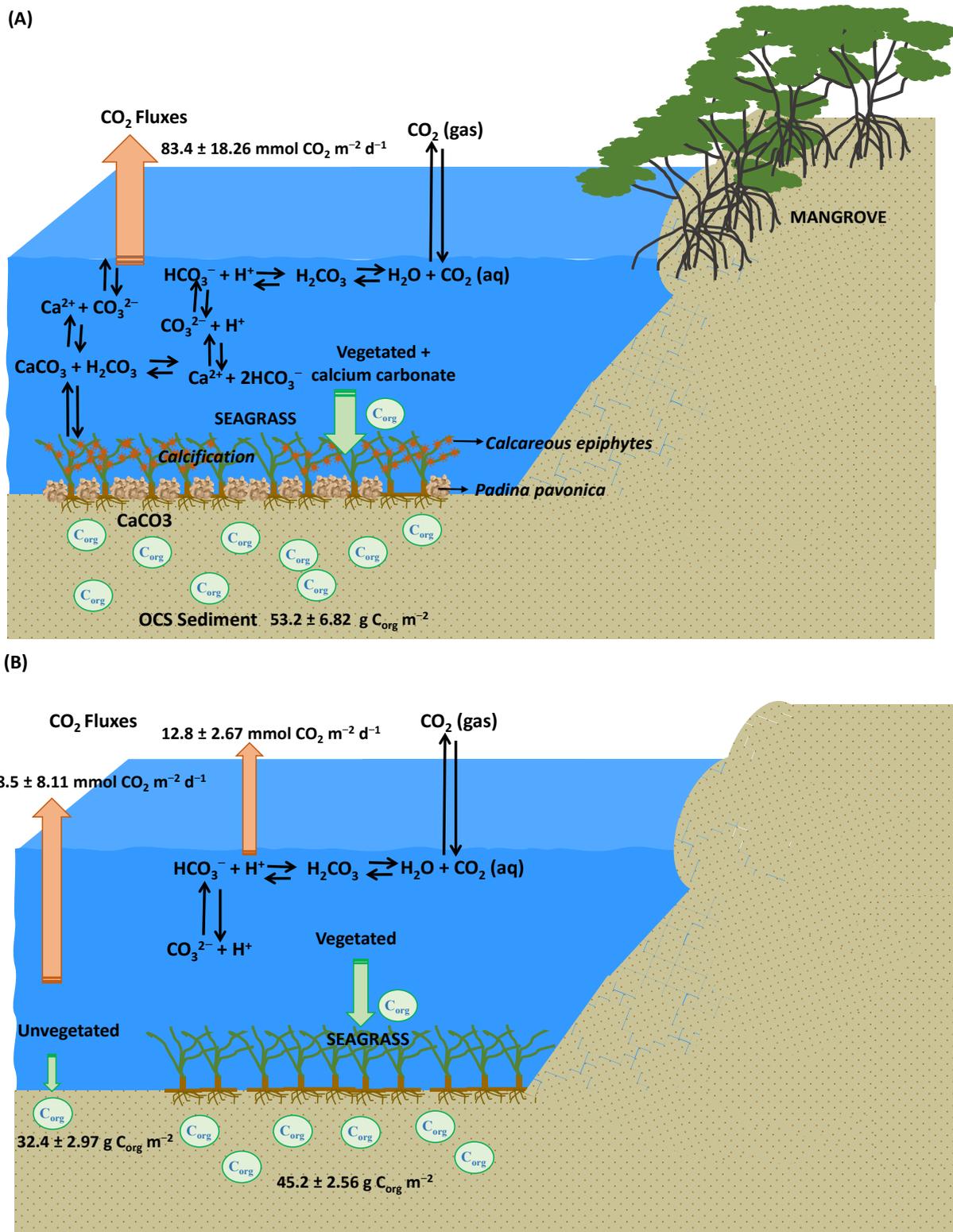


Figure 9. A schematic diagram illustrating the following: **(A)** The presence of vegetated seagrass and alongside source carbonates (*Padina pavonica*, calcareous epiphytes, and allochthonous mangrove), leading to an augmentation of OCS as well as an elevated release of CO₂ into the atmosphere; **(B)** a comparison between vegetated and unvegetated seagrass scenarios, highlighting that vegetated seagrass not only exhibits a higher capacity for storing OCS but also contributes to a comparatively lower release of CO₂ into the atmosphere.

Ecosystems of seagrass, encompassing algae and epiphytes, exhibit an increase in their inorganic carbon content. This increase is notably attributed to the calcification process within the macroalgae *Padina pavonica*, belonging to the Phaeophyceae class, as observed in Menjangan Kecil (Figure 8B). Likewise, calcareous epiphytes contribute to this trend, notably in Lendra (Figure 8C). In seagrass meadows enriched with substantial nutrient concentrations, the presence of green and red algae predominates, whereas seagrass beds characterized by lower nutrient levels are predominantly populated by brown algae, such as *Padina pavonica* [78]. This investigation reveals that both Mangrove Inn and Lendra have elevated nutrient levels (measuring 0.78 and 0.85 mg L⁻¹, respectively), where green calcareous epiphytes are prevalent. On the other hand, Menjangan Kecil displays lower nutrient levels (0.66 mg L⁻¹) and hosts *Padina pavonica* (brown algae), constituting over 50% of the seagrass meadow composition. Notably, the calcification processes in both the macroalgae and the green calcareous epiphytes lead to the generation of calcium carbonate (CaCO₃), thereby establishing the waters as a source of CO₂, resulting in positive CO₂ fluxes. These intricate interactions highlight the dynamic relationship between seagrass ecosystems, algae, and epiphytes, as well as their roles in influencing the inorganic carbon content and subsequent CO₂ fluxes within the aquatic environment.

The process of photosynthesis engenders CO₂ consumption, leading to a rise in water pH and consequent enhancement of carbonate saturation (CO₃²⁻), a phenomenon pivotal for supporting CaCO₃ precipitation (as depicted in the equation in Figure 9A). The subsequent calcification of CaCO₃ engenders an upsurge in carbonate (CO₃²⁻) within the water, contributing to an increase in pCO_{2,sea} and, subsequently, a significant return of CO₂ to the atmosphere (as illustrated in Figure 9A).

In accordance with [74], calcium carbonate takes form in three crystalline structures: calcite, aragonite, and magnesium calcite. Among the Phaeophyceae class, only two genera, *Padina* spp. and *Newhousia imbricata*, are recognized for their calcification capabilities, precipitating aragonite, a compound with distinct solubility characteristics in seawater, further augmenting inorganic carbon [74]. Earlier research [14] posits that seagrass meadows within tropical regions serve as vital hubs for calcium carbonate production. The calcium carbonate content notably depends on seagrass species marked by larger sizes and prolonged leaf lifespans. These factors contribute to heightened calcareous epiphyte presence, facilitating an elevated accumulation of particulate inorganic carbon (PIC) and saturation state of CO₃²⁻ [79,80]. Furthermore, they play a role in CO₂ removal from water by actively participating in CaCO₃ precipitation [75].

The occurrence of calcium carbonate precipitation brings about transformations in the inorganic carbon system, thereby amplifying pCO_{2,sea}, a process that prompts the liberation of dissolved CO₂ from seawater into the atmosphere. This phenomenon has been substantiated by previous studies [14,74,81–86]. Collectively, these outcomes underscore that organic carbon stocks alone cannot be deemed the sole, determinant factor in delineating the status and role of seagrass-dominated waters [83,84,87]. The intricate interplay of various processes, including CO₂ consumption, carbonate saturation, and CaCO₃ precipitation, collectively contribute to the intricate balance of the inorganic carbon system within these ecosystems.

On the contrary, this study has revealed that aquatic environments with abundant vegetation have the capacity to effectively sequester a larger quantity of organic carbon within their sediments. For instance, in comparison to sediments devoid of vegetation, which contained approximately 1840 ± 666 gC_{org} m⁻² of organic carbon, a previous investigation conducted in the Baltic Sea demonstrated that sediment enriched with *Zostera marina* vegetation harbored a substantial 7785 ± 679 gC_{org} m⁻² of organic carbon [88]. Consequently, while vegetated regions might act as sources of CO₂, they also contribute significantly to the mitigation of CO₂ emissions into the atmosphere. In scenarios devoid of vegetation, CO₂ emissions could escalate, potentially even reaching double the emissions observed in vegetated environments.

The findings of this study underscore the pivotal role of seagrass ecosystems in effectively sequestering organic carbon within both their biomass and sediment layers. Notably, the carbon content within organic sediments present in vegetated areas (seagrass sediments) surpasses that of their non-vegetated counterparts (non-seagrass sediments), as depicted in Figure 9B. This disparity in carbon storage indirectly influences the CO₂ flux within the surrounding waters; the existence of seagrass vegetation correlates with a diminished CO₂ flux, as contrasted with areas where seagrass is absent (unvegetated), as illustrated in Figure 9B.

Nonetheless, it is important to acknowledge that various factors can augment CO₂ flux within seagrass ecosystems. Notably, the introduction of carbonate sources (inorganic carbon) from allochthonous mangroves, *Padina pavonica*, and calcareous epiphytes, as depicted in Figure 9A, can intensify CO₂ flux. Regrettably, this study did not encompass measurements of inorganic carbon and calcium carbonate calcification. Consequently, safeguarding the health and carbon storage capacity of seagrass demands a proactive approach, which includes mitigating detrimental human activities like the direct disposal of pond waste into the waters and the use of trawl nets. Furthermore, the imperative for seagrass conservation and restoration within Karimunjawa waters becomes evident, particularly in areas with limited species diversity and sparse seagrass coverage.

5. Conclusions

This research stands as the inaugural account of blue carbon within the context of seagrass and sea–air CO₂ flux in Karimunjawa waters. Our study illuminates the paramount significance of seagrass ecosystems in the Karimunjawa region, elucidating that vegetated areas (seagrass meadows) boast substantial sediment organic carbon stocks. Notably, these vegetated areas contribute to the reduction of CO₂ release into the atmosphere, as evidenced by their lower air–sea CO₂ flux values in comparison to unvegetated regions (non-seagrass meadows). These findings underscore the pivotal role of seagrass meadows in carbon sequestration and the storage of carbon within both seagrass biomass and sediment layers, potentially curtailing the release of CO₂ into the atmosphere.

However, it is worth acknowledging that the presence of a calcium carbonate source in the form of inorganic carbon can amplify sediment organic carbon stocks and correspondingly escalate the release of CO₂ into the atmosphere. The inclusion of inorganic carbon (calcium carbonate) originating from sources like allochthonous mangroves in Mangrove Inn, calcification processes in brown algae *Padina pavonica* at Menjangan Kecil, and calcareous epiphytes at Lendra was not encompassed within this study's scope. Consequently, future research endeavors should delve into specific investigations to calculate the contribution of inorganic carbon (carbonate) alongside organic carbon, especially across different seasons (such as the northwest monsoon). Such studies would shed light on the intricate dynamics of water–air CO₂ flux within seagrass meadows and the net carbon absorption potential of seagrass ecosystems.

Author Contributions: Conceptualization, N.L., A.R.K. and A.W.; methodology, N.L., A.R.K. and A.W.; software, N.L. and S.F.; formal analysis, N.L., A.R.K., A.W. and F.H.; investigation, N.L. and S.F.; resources, N.S.N., N.L. and N.S.A.; writing—original draft preparation, N.L.; writing—review and editing, N.S.N., A.R.K. and A.W.; visualization, N.L., S.F. and F.H.; supervision, N.S.N., A.R.K. and A.W.; project administration, N.S.N.; funding acquisition, N.S.N. All authors have read and agreed to the published version of the manuscript.

Funding: This research was generously funded through multiple sources. First and foremost, the Indonesian Ministry of Education, Culture, Research, and Technology (Kemendikbudristek) provided financial support under the research program known as Penelitian Disertasi Doktor (PDD) 2023, with the Research Grant Contract (RGC) Number: 110/E5/PG.02.00.PL/2023. Furthermore, additional support was received through partial funding from the Institute for Research and Community Services at Universitas Diponegoro, as evidenced by the RGC Numbers: 385-31/UN7.P4.3/PP/2019 and 385-31/UN7.6.1/PP/2020. Finally, we express our sincere appreciation for the invaluable support

extended by Universitas Diponegoro, exemplified by Grant Number: 1276/UN7.P/HK/2021, which also granted a scholarship to facilitate the pursuit of this doctoral study program.

Institutional Review Board Statement: Not applicable.

Data Availability Statement: The sea level pressure data and wind speed were obtained from <https://www.ecmwf.int/en/forecasts/datasets/browse-reanalysis-datasets> (accessed on 30 September 2022). The molar fraction of carbon dioxide was available at https://disc.gsfc.nasa.gov/datasets/AIRX3C2M_V005/summary (accessed on 30 September 2022).

Acknowledgments: The authors are very thankful for the support of Kemendikbudristek, Institut Teknologi Bandung (ITB), Marine Technology Cooperation Research Center (MTCRC), Balai Taman Nasional (BTNKJ) Karimunjawa, Laboratorium Pengelolaan Sumberdaya Ikan dan Lingkungan Universitas Diponegoro, and Laboratorium Ilmu dan Nutrisi Pakan Universitas Diponegoro. We would also like to gratefully acknowledge data support from NASA and ECMWF.

Conflicts of Interest: The authors declare no conflict of interest.

References

1. Takahashi, T.; Olafsson, J.; Goddard, J.G.; Chipman, D.W.; Sutherland, S.C. Seasonal Variation of CO₂ and Nutrients in the High-Latitude Oceans: A Comparative Study. *Glob. Biogeochem. Cycles* **1993**, *7*, 843–878. [\[CrossRef\]](#)
2. Hoegh-Guldberg, O.; Danie, J.; Michael, T.; Marco, B.; Sally, B.; Ines, C.; Arona, D.; Riyanti, D. Chapter 3: Impacts of 1.5 °C Global Warming on Natural and Human Systems. In *Global Warming of 1.5 °C. An IPCC Special Report on the Impacts of Global Warming of 1.5 °C above Preindustrial Levels and Related Global Greenhouse Gas Emission Pathways*; Cambridge University Press: Cambridge, UK; New York, NY, USA, 2018; pp. 175–312. [\[CrossRef\]](#)
3. IPCC. Summary for Policymakers. In *IPCC Special Report on the Ocean and Cryosphere in a Changing Climate. A Special Report of the Intergovernmental Panel on Climate Change*; Pörtner, H.O., Roberts, D.C., Masson-Delmotte, V., Zhai, P., Tignor, M., Poloczanska, E., Mintenbeck, K., Nicolai, M., Okem, A., Petzold, J., et al., Eds.; Cambridge University Press: Cambridge, UK; New York, NY, USA, 2019.
4. Friedlingstein, P.; O’Sullivan, M.; Jones, M.; Andrew, R.; Hauck, J.; Olsen, A.; Peters, G.; Peters, W.; Pongratz, J.; Sitch, S.; et al. Global Carbon Budget 2020. *Earth Syst. Sci. Data Discuss.* **2020**, *12*, 3269–3340. [\[CrossRef\]](#)
5. Gulev, S.K.; Thorne, P.W.; Ahn, J.; Dentener, F.J.; Domingues, C.M.; Gerland, S.; Gong, D.; Kaufman, D.S.; Nnamchi, H.C.; Quaas, J.; et al. Changing State of the Climate System. In *Climate Change 2021: The Physical Science Basis. Contribution of Working Group I to the Sixth Assessment Report of the Intergovernmental Panel on Climate Change*; Masson-Delmotte, V., Zhai, P., Pirani, A., Connors, S.L., Péan, C., Berger, S., Caud, N., Chen, Y., Goldfarb, L., Gomis, M.I., et al., Eds.; Cambridge University Press: Cambridge, UK; New York, NY, USA, 2021; pp. 287–422. [\[CrossRef\]](#)
6. Borges, A.V.; Morana, C.; Bouillon, S.; Servais, P.; Descy, J.P.; Darchambeau, F. Carbon Cycling of Lake Kivu (East Africa): Net Autotrophy in the Epilimnion and Emission of CO₂ to the Atmosphere Sustained by Geogenic Inputs. *PLoS ONE* **2014**, *9*, e109500. [\[CrossRef\]](#)
7. Tye, A.M.; Williamson, J.L.; Jarvie, H.P.; Dise, N.B.; Lapworth, D.J.; Monteith, D.; Sanders, R.; Mayor, D.J.; Bowes, M.J.; Bowes, M.; et al. Dissolved Inorganic Carbon Export from Rivers of Great Britain: Spatial Distribution and Potential Catchment-Scale Controls. *J. Hydrol.* **2022**, *615*, 128677. [\[CrossRef\]](#)
8. Arora, V.K.; Jones, C.D.; Williams, R.; Katavouta, A.; Friedlingstein, P.; Brovkin, V.; Cadule, P.; Hajima, T.; Seferian, R.; Joetzer, E.; et al. Carbon-Concentration and Carbon-Climate Feedbacks in CMIP6 Models. *J. Clim.* **2019**. *preprint*.
9. Christianson, A.B.; Cabré, A.; Bernal, B.; Baez, S.K.; Leung, S.; Pérez-Porro, A.; Poloczanska, E. The Promise of Blue Carbon Climate Solutions: Where the Science Supports Ocean-Climate Policy. *Front. Mar. Sci.* **2022**, *9*, 1–16. [\[CrossRef\]](#)
10. Otero, P.; Padin, X.A.; Ruiz-Villarreal, M.; García-García, L.M.; Ríos, A.F.; Pérez, F.F. Net Sea-Air CO₂ Flux Uncertainties in the Bay of Biscay Based on the Choice of Wind Speed Products and Gas Transfer Parameterizations. *Biogeosciences* **2013**, *10*, 2993–3005. [\[CrossRef\]](#)
11. Zhang, M.; Cheng, Y.; Bao, Y.; Zhao, C.; Wang, G.; Zhang, Y.; Song, Z.; Wu, Z.; Qiao, F. Seasonal to Decadal Spatiotemporal Variations of the Global Ocean Carbon Sink. *Glob. Chang. Biol.* **2022**, *28*, 1786–1797. [\[CrossRef\]](#)
12. Le Quéré, C.; Andres, R.J.; Boden, T.; Conway, T.; Houghton, R.A.; House, J.I.; Marland, G.; Peters, G.P.; Van Der Werf, G.R.; Ahlström, A.; et al. The Global Carbon Budget 1959–2011. *Earth Syst. Sci. Data* **2013**, *5*, 165–185. [\[CrossRef\]](#)
13. Fourqurean, J.W.; Duarte, C.M.; Kennedy, H.; Marbà, N.; Holmer, M.; Mateo, M.A.; Apostolaki, E.T.; Kendrick, G.A.; Krause-Jensen, D.; McGlathery, K.J.; et al. Seagrass Ecosystems as a Globally Significant Carbon Stock. *Nat. Geosci.* **2012**, *5*, 505–509. [\[CrossRef\]](#)
14. Mazarrasa, I.; Marbà, N.; Lovelock, C.E.; Serrano, O.; Lavery, P.S.; Fourqurean, J.W.; Kennedy, H.; Mateo, M.A.; Krause-Jensen, D.; Steven, A.D.L.; et al. Seagrass Meadows as a Globally Significant Carbonate Reservoir. *Biogeosciences* **2015**, *12*, 4993–5003. [\[CrossRef\]](#)
15. Bayley, D.; Marengo, I.; Pelembe, T. *Giant Kelp ‘Blue Carbon’ Storage and Sequestration Value in the Falkland Islands*; Joint Nature Conservation Committee: Peterborough, UK, 2017; p. 31. [\[CrossRef\]](#)

16. Egea, L.G.; Jiménez-Ramos, R.; Hernández, I.; Bouma, T.J.; Brun, F.G. Effects of Ocean Acidification and Hydrodynamic Conditions on Carbon Metabolism and Dissolved Organic Carbon (DOC) Fluxes in Seagrass Populations. *PLoS ONE* **2018**, *13*, e0192402. [[CrossRef](#)]
17. Watanabe, K.; Kuwae, T. How Organic Carbon Derived from Multiple Sources Contributes to Carbon Sequestration Processes in a Shallow Coastal System? *Glob. Chang. Biol.* **2015**, *21*, 2612–2623. [[CrossRef](#)] [[PubMed](#)]
18. Otero, M.M.; Walton, M. *Manual for the Creation of Blue Carbon Projects in Europe and the Mediterranean*; IUCN, International Union for Conservation of Nature Center for Mediterranean Cooperation: Malaga, Spain, 2021.
19. Samper-Villarreal, J.; Lovelock, C.E.; Saunders, M.I.; Roelfsema, C.; Mumby, P.J. Organic Carbon in Seagrass Sediments Is Influenced by Seagrass Canopy Complexity, Turbidity, Wave Height, and Water Depth. *Limnol. Oceanogr.* **2016**, *61*, 938–952. [[CrossRef](#)]
20. Salinas, C.; Duarte, C.M.; Lavery, P.S.; Masque, P.; Arias-Ortiz, A.; Leon, J.X.; Callaghan, D.; Kendrick, G.A.; Serrano, O. Seagrass Losses since Mid-20th Century Fuelled CO₂ Emissions from Soil Carbon Stocks. *Glob. Chang. Biol.* **2020**, *26*, 4772–4784. [[CrossRef](#)] [[PubMed](#)]
21. Lovelock, C.E.; Fourqurean, J.W.; Morris, J.T. Modeled CO₂ Emissions from Coastal Wetland Transitions to Other Land Uses: Tidal Marshes, Mangrove Forests, and Seagrass Beds. *Front. Mar. Sci.* **2017**, *4*, 1–11. [[CrossRef](#)]
22. Akhand, A.; Watanabe, K.; Chanda, A.; Tokoro, T.; Chakraborty, K.; Moki, H.; Tanaya, T.; Ghosh, J.; Kuwae, T. Lateral Carbon Fluxes and CO₂ Evasion from a Subtropical Mangrove-Seagrass-Coral Continuum. *Sci. Total Environ.* **2021**, *752*, 142190. [[CrossRef](#)]
23. Macklin, P.A.; Suryaputra, I.G.N.A.; Maher, D.T.; Murdiyarso, D.; Santos, I.R. Drivers of CO₂ along a Mangrove-Seagrass Transect in a Tropical Bay: Delayed Groundwater Seepage and Seagrass Uptake. *Cont. Shelf Res.* **2019**, *172*, 57–67. [[CrossRef](#)]
24. Thorhaug, A.; Gallagher, J.B.; Kiswara, W.; Prathep, A.; Huang, X.; Yap, T.K.; Dorward, S.; Berlyn, G. Coastal and Estuarine Blue Carbon Stocks in the Greater Southeast Asia Region: Seagrasses and Mangroves per Nation and Sum of Total. *Mar. Pollut. Bull.* **2020**, *160*, 111168. [[CrossRef](#)]
25. Wahyudi, A.J.; Rahmawati, S.; Irawan, A.; Hadiyanto, H.; Prayudha, B.; Hafizt, M.; Afdal, A.; Adi, N.S.; Rustam, A.; Hernawan, U.E.; et al. Assessing Carbon Stock and Sequestration of the Tropical Seagrass Meadows in Indonesia. *Ocean Sci. J.* **2020**, *55*, 85–97. [[CrossRef](#)]
26. Wahyudi, A.J.; Febriani, F. Country-Specific Emission Factor for Developing a Tier 3 System of Indonesia's Seagrass Carbon Inventory. *IOP Conf. Ser. Earth Environ. Sci.* **2021**, *944*, 012058. [[CrossRef](#)]
27. Ganefiani, A.; Suryanti, S.; Latifah, N. Potensi Padang Lamun sebagai Penyerap Karbon di Perairan Pulau Karimunjawa, Taman Nasional Karimunjawa. *SAINTEK Perikan. Indones. J. Fish. Sci. Technol.* **2019**, *14*, 115. [[CrossRef](#)]
28. Ramadona, D.N.; Ain, C.; Febrianto, S.; Suryanti, S.; Latifah, N. Carbon Storage Potential of Seagrass Meadows in Pokemon Beach, Karimunjawa. *J. Ilmu Teknologi. Kelautan Tropis.* **2021**, *13*, 319–332. [[CrossRef](#)]
29. Latifah, N.; Febrianto, S.; Wirasatriya, A.; Endrawati, H.; Zainuri, M.; Suryanti, S.; Hidayat, A.N. Air-Sea Flux of CO₂ in the Waters of Karimunjawa Island, Indonesia. *Saintek Perikan. Indones. J. Fish. Sci. Technol.* **2020**, *16*, 171–178. [[CrossRef](#)]
30. Latifah, N.; Endrawati, H.; Febrianto, S. Distribusi Spasial Fluks Karbon Dioksida Di Perairan Karimunjawa, Indonesia. *J. Ilmu Teknol. Kelaut. Trop.* **2019**, *11*, 357–368. [[CrossRef](#)]
31. Kartadikaria, A.R.; Watanabe, A.; Nadaoka, K.; Adi, N.S.; Prayitno, H.B.; Soemorumekso, S.; Muchtar, M.; Triyulianti, I.; Setiawan, A.; Suratno, S.; et al. CO₂ Sink/Source Characteristics in the Tropical Indonesian Seas. *J. Geophys. Res. Ocean.* **2015**, *120*, 7842–7856. [[CrossRef](#)]
32. Wirasatriya, A.; Sugianto, D.N.; Maslukah, L.; Ahkam, M.F.; Wulandari, S.Y.; Helmi, M. Carbon Dioxide Flux in the Java Sea Estimated from Satellite Measurements. *Remote Sens. Appl. Soc. Environ.* **2020**, *20*, 100376. [[CrossRef](#)]
33. Aryetti, F.; Pratomo, D.G.; Pribadi, C.B. Utilization of Bathymetry and Sea Surface Current Data for Coral Reef Habitats in The Karimunjawa Islands. *IOP Conf. Ser. Earth Environ. Sci.* **2023**, *1127*, 012041. [[CrossRef](#)]
34. Wungo, G.L.; Mussadun, M.; Ma'rif, S. Edukasi Penerapan Konsep Ecotourism Di Kepulauan Karimunjawa. *J. Pasopati* **2020**, *2*, 142–149.
35. Hach Company. *Scope and Application: For Water, Wastewater and Seawater. Nitrate*; Hach Company: Loveland, CO, USA, 2014; Volume 584.
36. Hach Company. *DR/2010 Spectrophotometer Procedures Manual*; Hach Company: Loveland, CO, USA, 2010.
37. Hach Company. *Scope and Application: For Water, Wastewater and Seawater. Phosphorus, Reactive (Orthophosphate)*; Hach Company: Loveland, CO, USA, 2014; Volume 584.
38. Rahmawati, S.; Hernawan, U.E.; Rustam, A. The Seagrass Carbon Content of 0.336 of Dry Weight Can Be Applied in Indonesian Seagrasses. *AIP Conf. Proc.* **2019**, *2120*, 030012. [[CrossRef](#)]
39. Wanninkhof, R.; McGill, W. A Cubic Relationship between Air-Sea CO₂ Exchange and Wind Speed. *Geophys. Res. Lett.* **1999**, *26*, 1889–1892. [[CrossRef](#)]
40. Takahashi, T.; Sutherland, S.C.; Sweeney, C.; Poisson, A.; Metz, N.; Tilbrook, B.; Bates, N.; Wanninkhof, R.; Feely, R.A.; Sabine, C.; et al. Global Sea-Air CO₂ Flux Based on Climatological Surface Ocean PCO₂, and Seasonal Biological and Temperature Effects. *Deep. Res. Part II Top. Stud. Oceanogr.* **2002**, *49*, 1601–1622. [[CrossRef](#)]
41. Zhu, Y.; Shang, S.; Zhai, W.; Dai, M. Satellite-Derived Surface Water PCO₂ and Air-Sea CO₂ Fluxes in the Northern South China Sea in Summer. *Prog. Nat. Sci.* **2009**, *19*, 775–779. [[CrossRef](#)]

42. Du, Y.; Zhang, Y.; Feng, M.; Wang, T.; Zhang, N.; Wijffels, S. Decadal Trends of the Upper Ocean Salinity in the Tropical Indo-Pacific since Mid-1990s. *Sci. Rep.* **2015**, *5*, 16050. [[CrossRef](#)] [[PubMed](#)]
43. Sisma-Ventura, G.; Bialik, O.M.; Yam, R.; Herut, B.; Silverman, J. PCO₂ Variability in the Surface Waters of the Ultra-Oligotrophic Levantine Sea: Exploring the Air–Sea CO₂ Fluxes in a Fast Warming Region. *Mar. Chem.* **2017**, *196*, 13–23. [[CrossRef](#)]
44. Yan, H.; Yu, K.; Shi, Q.; Lin, Z.; Zhao, M.; Tao, S.; Liu, G.; Zhang, H. Air-Sea CO₂ Fluxes and Spatial Distribution of Seawater PCO₂ in Yongle Atoll, Northern-Central South China Sea. *Cont. Shelf Res.* **2018**, *165*, 71–77. [[CrossRef](#)]
45. Weiss, R.F. Carbon Dioxide in Water and Seawater: The Solubility of a Non-Ideal Gas. *Mar. Chem.* **1974**, *2*, 203–215. [[CrossRef](#)]
46. Wanninkhof, R. Relationship between Wind Speed and Gas Exchange over the Ocean. *J. Geophys. Res.* **1992**, *97*, 7373–7382. [[CrossRef](#)]
47. Raymond, P.A.; Cole, J.J. Technical Notes and Comments: Gas Exchange in Rivers and Estuaries: Choosing a Gas Transfer Velocity. *Estuaries* **2001**, *24*, 312–317. [[CrossRef](#)]
48. Borges, A.V.; Vanderborcht, J.P.; Schiettecatte, L.S.; Gazeau, F.; Ferrón-Smith, S.; Delille, B.; Frankignoulle, M. Variability of the Gas Transfer Velocity of CO₂ in a Macrotidal Estuary (the Scheldt). *Estuaries* **2004**, *27*, 593–603. [[CrossRef](#)]
49. Dickson, A.G.; Sabine, C.L.; Christian, J.R. *Guide to Best Practices for Ocean CO₂ Measurements*; North Pacific Marine Science Organization: Sidney, Australia, 2007; Volume 3, ISBN 1-897176-07-4.
50. Williams, L.J. Newman-Keuls Test and Tukey Test 1 Pairwise Comparisons. *Encyclopedia Res. Des.* **2010**, *1*, 897–902.
51. Park, S.R.; Moon, K.; Kim, S.H.; Lee, K.S. Growth and Photoacclimation Strategies of Three *Zostera* Species Along a Vertical Gradient: Implications for Seagrass Zonation Patterns. *Front. Mar. Sci.* **2021**, *8*, 594779. [[CrossRef](#)]
52. Kim, S.H.; Suonan, Z.; Qin, L.Z.; Kim, H.; Park, J.I.; Kim, Y.K.; Lee, S.; Kim, S.G.; Kang, C.K.; Lee, K.S. Variability in Blue Carbon Storage Related to Biogeochemical Factors in Seagrass Meadows off the Coast of the Korean Peninsula. *Sci. Total Environ.* **2022**, *813*, 152680. [[CrossRef](#)] [[PubMed](#)]
53. Gullstrom, M.; Lyimo, L.D.; Dahl, M.; Eggertsen, M.; Anderberg, E.; Rasmusson, L.M.; Mtwana, L.; Linderholm, H.W.; Knudby, A.; Bjo, M. Blue Carbon Storage in Tropical Seagrass Meadows Relates to Carbonate Stock Dynamics, Plant–Sediment Processes, and Landscape Context: Insights from the Western Indian Ocean. *Ecosystems* **2018**, *21*, 551–566. [[CrossRef](#)]
54. Ricart, A.M.; Pérez, M.; Romero, J. Landscape Configuration Modulates Carbon Storage in Seagrass Sediments. *Estuar. Coast. Shelf Sci.* **2017**, *185*, 69–76. [[CrossRef](#)]
55. Duarte, C.M.; Marbà, N.; Gacia, E.; Fourqurean, J.W.; Beggins, J.; Barrón, C.; Apostolaki, E.T. Seagrass Community Metabolism: Assessing the Carbon Sink Capacity of Seagrass Meadows. *Glob. Biogeochem. Cycles* **2010**, *24*, 1–8. [[CrossRef](#)]
56. Lavery, P.S.; Mateo, M.Á.; Serrano, O.; Rozaimi, M. Variability in the Carbon Storage of Seagrass Habitats and Its Implications for Global Estimates of Blue Carbon Ecosystem Service. *PLoS ONE* **2013**, *8*, e73748. [[CrossRef](#)]
57. Waycott, M.; Duarte, C.M.; Carruthers, T.J.B.; Orth, R.J.; Dennison, W.C.; Olyarnik, S.; Calladine, A.; Fourqurean, J.W.; Heck, K.L.; Hughes, A.R.; et al. Accelerating Loss of Seagrasses across the Globe Threatens Coastal Ecosystems. *Proc. Natl. Acad. Sci. USA* **2009**, *106*, 12377–12381. [[CrossRef](#)]
58. Alongi, D.M.; Murdiyarto, D.; Fourqurean, J.W.; Kauffman, J.B.; Hutahaeen, A.; Crooks, S.; Lovelock, C.E.; Howard, J.; Herr, D.; Fortes, M.; et al. Indonesia’s Blue Carbon: A Globally Significant and Vulnerable Sink for Seagrass and Mangrove Carbon. *Wetl. Ecol. Manag.* **2015**, *23*, 3–13. [[CrossRef](#)]
59. McLeod, E.; Chmura, G.L.; Bouillon, S.; Salm, R.; Björk, M.; Duarte, C.M.; Lovelock, C.E.; Schlesinger, W.H.; Silliman, B.R. A Blueprint for Blue Carbon: Toward an Improved Understanding of the Role of Vegetated Coastal Habitats in Sequestering CO₂. *Front. Ecol. Environ.* **2011**, *9*, 552–560. [[CrossRef](#)]
60. Jiang, Z.; Liu, S.; Zhang, J. Eutrophication Indirectly Reduced Carbon Sequestration in a Tropical Seagrass Bed Eutrophication Indirectly Reduced Carbon Sequestration in a Tropical Seagrass Bed. *Plant Soil* **2018**, *426*, 135–152. [[CrossRef](#)]
61. Asplund, M.E.; Dahl, M.; Ismail, R.O.; Arias-Ortiz, A.; Deyanova, D.; Franco, J.N.; Hammar, L.; Hoamby, A.I.; Linderholm, H.W.; Lyimo, L.D.; et al. Dynamics and Fate of Blue Carbon in a Mangrove–Seagrass Seascape: Influence of Landscape Configuration and Land-Use Change. *Landsc. Ecol.* **2021**, *36*, 1489–1509. [[CrossRef](#)]
62. Avnimelech, Y.; Ritvo, G.; Meijer, L.E. Water Content, Organic Carbon and Dry Bulk Density in Flooded Sediments Water Content, Organic Carbon and Dry Bulk Density in Flooded Sediments. *Aquac. Eng.* **2001**, *25*, 25–33. [[CrossRef](#)]
63. Susana, T. Karbon Dioksida. *Oseana* **1988**, *XIII*, 1–11.
64. Curbelo-Hernández, D.; Santana-Casiano, J.M.; González, A.G.; González-Dávila, M. Air-Sea CO₂ Exchange in the Strait of Gibraltar. *Front. Mar. Sci.* **2021**, *8*, 745304. [[CrossRef](#)]
65. Fitrianti, B.A.; Juliandri, D.; Herunadi, B. Potensi Pelepasan dan Penyerap CO₂ Kaitannya Dengan Suhu Dan Salinitas Di Perairan Teluk Banten. *J. Akuatika* **2013**, *2*, 174–182.
66. Basah, P.B.; Bontang, K.; Timur, K. Distribusi Karbon Anorganik Dan Fluks CO₂. *Lingkungan Tropis* **2016**, *6*, 149–158.
67. Afdal. Fluks CO₂ Di Perairan Pesisir Pulau Lombok, Nusa Tenggara Barat CO₂ Flux in the Coastal Waters of Lombok, West Nusa Tenggara. *Oceanologi Limnol.* **2016**, *1*, 91–103.
68. Cai, W.J.; Dai, M.; Wang, Y. Air-Sea Exchange of Carbon Dioxide in Ocean Margins: A Province-Based Synthesis. *Geophys. Res. Lett.* **2006**, *33*, 2–5. [[CrossRef](#)]
69. Ito, R.G.; Tavano, V.M.; Mendes, C.R.B.; Garcia, C.A.E. Sea-Air CO₂ Fluxes and PCO₂ Variability in the Northern Antarctic Peninsula during Three Summer Periods (2008–2010). *Deep. Res. Part II Top. Stud. Oceanogr.* **2018**, *149*, 84–98. [[CrossRef](#)]

70. Tian, H.; Xu, R.; Canadell, J.G.; Thompson, R.L.; Winiwarter, W.; Suntharalingam, P.; Davidson, E.A.; Ciais, P.; Jackson, R.B.; Janssens-Maenhout, G.; et al. A Comprehensive Quantification of Global Nitrous Oxide Sources and Sinks. *Nature* **2020**, *586*, 248–256. [[CrossRef](#)]
71. Zheng, Z.; Luo, X.; Wei, H.; Zhao, W.; Qi, D. Analysis of the Seasonal and Interannual Variations of Air-Sea CO₂ Flux in the Chukchi Sea Using A Coupled Ocean-Sea Ice-Biogeochemical Model. *J. Geophys. Res. Ocean.* **2021**, *126*, e2021JC017550. [[CrossRef](#)]
72. Curbelo-Hernández, D.; González-Dávila, M.; González, A.G.; González-Santana, D.; Santana-Casiano, J.M. CO₂ Fluxes in the Northeast Atlantic Ocean Based on Measurements from a Surface Ocean Observation Platform. *Sci. Total Environ.* **2021**, *775*, 145804. [[CrossRef](#)]
73. Semesi, I.S.; Beer, S.; Björk, M. Seagrass Photosynthesis Controls Rates of Calcification and Photosynthesis of Calcareous Macroalgae in a Tropical Seagrass Meadow. *Mar. Ecol. Prog. Ser.* **2009**, *382*, 41–47. [[CrossRef](#)]
74. Hofmann, L.C.; Bischof, K. Ocean Acidification Effects on Calcifying Macroalgae. *Aquat. Biol.* **2014**, *22*, 261–279. [[CrossRef](#)]
75. Kalokora, O.J.; Buriyo, A.S.; Asplund, M.E.; Gullström, M.; Mtolera, M.S.P.; Björk, M. An Experimental Assessment of Algal Calcification as a Potential Source of Atmospheric CO₂. *PLoS ONE* **2020**, *15*, e0231971. [[CrossRef](#)] [[PubMed](#)]
76. Ismail, R.O.; Asplund, M.E.; Gullström, M.; George, R.; Dahl, M.; Buriyo, A.S.; Mtolera, M.S.P.; Björk, M. Effects of Calcification on Air-Water CO₂ Fluxes in Tropical Seagrass Meadows: A Mesocosm Experiment. *J. Exp. Mar. Biol. Ecol.* **2023**, *561*, 151864. [[CrossRef](#)]
77. Banerjee, K.; Panerselvam, A.; Ramachandran, P.; Ganguly, D.; Singh, G.; Ramesh, R. Seagrass and Macrophyte Mediated CO₂ and CH₄ Dynamics in Shallow Coastal Waters. *PLoS ONE* **2018**, *13*, e0249253. [[CrossRef](#)]
78. Cui, L.; Jiang, Z.; Huang, X.; Chen, Q.; Wu, Y.; Liu, S.; Li, J.; Macreadie, P.I. Eutrophication Reduces Seagrass Contribution to Coastal Food Webs. *Ecosphere* **2021**, *12*, e03626. [[CrossRef](#)]
79. Duarte, C.M. Allometric Scaling of Seagrass Form and Productivity. *Mar. Ecol. Prog. Ser.* **1991**, *77*, 289–300. [[CrossRef](#)]
80. Cebrián, J.; Duarte, C.M.; Marbà, N.; Enríquez, S. Magnitude and Fate of the Production of Four Co-Occurring Western Mediterranean Seagrass Species. *Mar. Ecol. Prog. Ser.* **1997**, *155*, 29–44. [[CrossRef](#)]
81. Ware, J.R.; Smith, S.V.; Reaka-Kudla, M.L. Coral Reefs: Sources or Sinks of Atmospheric CO₂? *Coral Reefs* **1991**, *11*, 127–130. [[CrossRef](#)]
82. Frankignoulle, M.; Canon, C.; Gattuso, J.-P. Marine Calcification as a Source of Carbon Dioxide: Positive Feedback of Increasing Atmospheric CO₂. *Limnol. Oceanogr.* **1994**, *39*, 458–462. [[CrossRef](#)]
83. Macreadie, P.I.; Serrano, O.; Maher, D.T.; Duarte, C.M.; Beardall, J. Addressing Calcium Carbonate Cycling in Blue Carbon Accounting. *Limnol. Oceanogr. Lett.* **2017**, *2*, 195–201. [[CrossRef](#)]
84. Chou, W.-C.; Fan, L.-F.; Hung, C.-C.; Shih, Y.-Y.; Huang, W.-J.; Lui, H.-K.; Chen, T.-Y. Dynamics of O₂ and PCO₂ in a Southeast Asia Seagrass Meadow: Metabolic Rates and Carbon Sink Capacity. *Front. Mar. Sci.* **2023**, *10*, 1076991. [[CrossRef](#)]
85. Chou, W.C.; Fan, L.F.; Yang, C.C.; Chen, Y.H.; Hung, C.C.; Huang, W.J.; Shih, Y.Y.; Soong, K.; Tseng, H.C.; Gong, G.C.; et al. A Unique Diel Pattern in Carbonate Chemistry in the Seagrass Meadows of Dongsha Island: The Enhancement of Metabolic Carbonate Dissolution in a Semienclosed Lagoon. *Front. Mar. Sci.* **2021**, *8*, 717685. [[CrossRef](#)]
86. Van Dam, B.R.; Lopes, C.C.; Polsenaere, P.; Price, R.M.; Rutgersson, A.; Fourqurean, J.W. Water Temperature Control on CO₂ Flux and Evaporation over a Subtropical Seagrass Meadow Revealed by Atmospheric Eddy Covariance. *Limnol. Oceanogr.* **2021**, *66*, 510–527. [[CrossRef](#)]
87. Saderne, V.; Galdi, N.R.; Macreadie, P.I.; Maher, D.T.; Middelburg, J.J.; Serrano, O.; Almahasheer, H.; Arias-Ortiz, A.; Cusack, M.; Eyre, B.D.; et al. Role of Carbonate Burial in Blue Carbon Budgets. *Nat. Commun.* **2019**, *10*, 1106. [[CrossRef](#)] [[PubMed](#)]
88. Stevenson, A.; Ó Corcora, T.C.; Hukriede, W.; Schubert, P.R.; Reusch, T.B.H. Substantial Seagrass Blue Carbon Pools in the Southwestern Baltic Sea Include Relics of Terrestrial Peatlands. *Front. Mar. Sci.* **2022**, *9*, 1–17. [[CrossRef](#)]

Disclaimer/Publisher’s Note: The statements, opinions and data contained in all publications are solely those of the individual author(s) and contributor(s) and not of MDPI and/or the editor(s). MDPI and/or the editor(s) disclaim responsibility for any injury to people or property resulting from any ideas, methods, instructions or products referred to in the content.