

# Two New Edible *Lyophyllum* Species from Tibetan Areas, China

Shuhong Li <sup>1,2</sup>, Songming Tang <sup>2</sup> , Jun He <sup>2</sup> and Dequn Zhou <sup>1,\*</sup>

<sup>1</sup> Faculty of Environmental Sciences and Engineering, Kunming University of Science and Technology, Kunming 650500, China; lsh@yaas.org.cn

<sup>2</sup> Biotechnology and Germplasm Resources Institute, Yunnan Academy of Agricultural Sciences, Kunming 650223, China; tang202205@gmail.com (S.T.); junhe219@163.com (J.H.)

\* Correspondence: dqzhou2009@gmail.com

**Abstract:** Two new species, *Lyophyllum yiqunyang* and *L. heimogu*, that belong to the section *Difformia* of the genus *Lyophyllum*, are described based on collections from Tibetan areas, China. The two species are delicious edible low-temperature mushrooms and are widely collected and eaten by local people. *Lyophyllum yiqunyang* sp. nov. is saprotrophic and has medium-sized basidiomata, olive-grey pileus, cheilocystidia, absent pleurocystidia, globose to subglobose basidiospores (6.12–6.31 × 6.02–6.23 μm) and clamp connections at the pileus context, hymenophoral trama and stipe. *Lyophyllum heimogu* sp. nov. is saprotrophic and has a dark grey to olive pileus, medium-sized basidiomata and globose to subglobose basidiospores (5.31–5.63 × 5.22–5.41 μm). In the phylogenetic analyses, our two new species formed distinct clades that are well supported by posterior probabilities and bootstrap proportions. Detailed descriptions, colour photos, illustrations and a phylogenetic tree to show the positions of the two new species are presented.

**Keywords:** ITS; Lyophyllaceae; morphology; novel taxa; phylogeny; taxonomy



**Citation:** Li, S.; Tang, S.; He, J.; Zhou, D. Two New Edible *Lyophyllum* Species from Tibetan Areas, China. *Diversity* **2023**, *15*, 1027.

<https://doi.org/10.3390/d15091027>

Academic Editor: Ipek Kurtboke

Received: 18 August 2023

Revised: 18 September 2023

Accepted: 18 September 2023

Published: 21 September 2023



**Copyright:** © 2023 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (<https://creativecommons.org/licenses/by/4.0/>).

## 1. Introduction

The genus *Lyophyllum* P. Karst is classified under the family Lyophyllaceae, order Agaricales, <http://www.indexfungorum.org/Names/Names.asp> accessed on 10 June 2023, and is mainly characterised by basidiomata colour that is unchanging or changes to dark when injured; scattered, gregarious or solitary growth; a smooth pileus surface; a solid cylindrical or upwards-tapering stipe; basidiospores with variable shape (globose, ellipsoid or broadly fusiform); and saprophytic and symbiotic habits [1,2].

There is a complex without blackening when injured and with caespitose-growing basidiomata in the section *Difformia* [3]. *Lyophyllum* sect. *Difformia* includes 14 species worldwide, most of these species from Europe (*L. brunneum* Dähncke, Contu & Vizzini; *L. calabrum* Lavorato & Contu; *L. cistophilum* Vila & Llimona; *L. decastes* (Fr.) Singer; *L. lanzonii* Candusso; *L. pergamenum* [Sacc. & P. Syd.] Horniček; *L. pseudoloricatum* Dähncke, Contu & Vizzini; *L. subglobisporum* Consiglio & Contu; *L. soniae* Picillo & Contu) and a few from North/South America (*L. multifforme* [Peck] H.E. Bigelow and *L. tucumanense* Singer) and Asia (*L. shimeji* [Kawam.] Hongo and two novel species, *L. yiqunyang* and *L. heimogu*) [4–11]. *Lyophyllum* consists of 60 species worldwide [12].

Dai 1979 [13] identified the first *Lyophyllum* species in China, *L. cinerascens* (Bull.) Konrad & Maubl., which is now *L. decastes*. Since then, 20 additional *Lyophyllum* species have been recorded in China [14]. These species were identified based on phenotypic similarities to European *Lyophyllum* and lacked detailed descriptions and molecular data.

Many *Lyophyllum* species names have been combined and changed with the advancement of molecular biology methods. To date, 16 *Lyophyllum* species have been reported in China, *viz.* *L. atrofusum* S.W. Wei, Q. Wang & Yu Li (Tibet); *L. decastes* (Yunnan, Sichuan, Qinghai, Liaoning and Fujian, etc.); *L. deqinense* Y.H. Ma, W.M. Chen & Y.C. Zhao (Yunnan);

*L. immundum* (Berk.) Kühner (not clear); *L. infumatum* (Bres.) Kühner (not clear); *L. loriscatum* (Fr.) Kühner (Tibet); *L. macrosporum* Singer (not clear); *L. ochraceum* (R. Haller Aar.) Schwöbel & Reutter (Sichuan); *L. pallidofumosum* (Yunnan); *L. rhombisporum* Shu H. Li & Y.C. Zhao (Yunnan); *L. semitale* (Fr.) Kühner (Yunnan, Tibet, Qinghai, Heilongjiang, Shanxi); *L. subalpinarum* S.W. Wei, Q. Wang & Yu Li (Tibet); *L. subdecastes* S.W. Wei, Q. Wang & Yu Li (Gansu); *L. trigonosporum* (Bres.) Kühner (Yunnan, Tibet); *L. transforme* (Lapl.) Singer (Yunnan, Tibet, Qinghai, Liaoning, Heilongjiang); and *L. shimeji* (Yunnan) [14–18].

Most species of *Lyophyllum* have been described as well-known edible mushrooms, such as *L. shimeji*, *L. decastes* and *L. fumosum*. Currently, the classification of this genus and the cultivation of *L. shimeji* have been studied extensively in Italy, Japan, Sweden and Switzerland [19].

During a survey of *Lyophyllum* in China, two saprotrophic new species, viz. *L. yiqunyang* and *L. heimogu*, belonging to sect. *Difformia* were collected and identified based on morphological features and molecular data.

## 2. Materials and Methods

### 2.1. Site Description

Jiuzhaigou and Bomi are important and well-preserved nature sites on the edge of the Tibetan Plateau. The Tibetan area is made up of high-altitude hydrological and tectonic activity. The rock strata are mostly carbonate rocks such as dolomite and tufa, as well as some sandstone and shales. *Lyophyllum yiqunyang* and *L. heimogu* were found growing on soil in mixed coniferous and broad-leaved forests dominated by *Abies* spp., *Picea* spp. and *Salix cupularis*, with an elevation of 2000–3000 m a.s.l., a temperate climate and annual rainfall of 550 mm.

### 2.2. Morphological Studies

Five specimens were photographed in situ and collected from Tibetan areas in China. After collecting, samples were wrapped in aluminium foil and placed in a collection box until they were examined. Macro-morphological features were recorded from fresh collections, and colour was determined following Korerup and Wanscher [20]. Specimens were dried at 50 °C in a food drier, stored in sealed plastic bags and deposited at the Biotechnology and Germplasm Resources Institute, Yunnan Academy of Agricultural Sciences, Kunming and Herbarium of Cryptogams at the Kunming Institute of Botany, Chinese Academy of Sciences (HKAS). Micromorphological features were observed under a Y-TV55 light microscope (Nikon, Tokyo, Japan) from the dried material that was studied. All tissues were revived in 5–10% KOH and mounted in Congo Red. Scanning electron microscopy (SEM) images were captured using Regulus 8100 (Hitachi, Tokyo, Japan). Twenty basidia and more than fifty basidiospores were measured and photographed using a Nikon Eclipse 80i microscope at magnifications up to  $\times 1000$ . The notation [x/y/z] specified that measurements were made on 'x' basidiospores from 'y' basidiomata and 'z' collections. Basidiospore dimensions were given as '(a) b–n–c (d)'. Where 'a' and 'd' refer to the lower and upper extremes of all measurements, respectively, 'b–c' refers to the range of 95% of the measured basidiospores, 'n' refers to the average dimension, 'Q' is the length/width ratio of basidiospores and 'Q<sub>m</sub>' is the average Q of all basidiospores.

### 2.3. DNA Extraction, PCR Amplification and Sequencing

Molecular analyses were performed at the Yunnan Academy of Agricultural Sciences, China. Genomic DNA was extracted from dry specimens using the Ezup Column Fungi Genomic DNA Extraction Kit (Sangon Biotech, Shanghai, China) following the manufacturer's protocol. Primer pairs (forward/reverse) used for PCR cycling of the ITS regions were ITS1/ITS4 [21]. PCR was carried out using a C1000 Thermal Cycler (Bio-Rad, Beijing, China) with a cycling program as follows. ITS: initial denaturation at 95 °C for 5 min, 35 cycles of denaturation at 95 °C for 30 s, annealing at 55 °C for 30 s, extension at 72 °C for 90 s and a final extension at 72 °C for 10 min [21].

PCR products were visualised via UV light after electrophoresis on 1% agarose gels stained with ethidium bromide. The PCR amplicons were sent to Sangon Biotech (Shanghai, China) for Sanger sequencing in both directions using the PCR primers.

#### 2.4. Sequence Alignment and Phylogenetic Analyses

The sequences of new *Lyophyllum* identified in this study were submitted to the NCBI database. The Basic Local Alignment Search Tool for the GenBank database was used to check whether the newly generated sequences were amplified contaminant DNA and examine clusters with closely related sequences (see Table 1). DNA sequences were retrieved and assembled using SeqMan version 5.0. The sequence alignments were aligned using MAFFT version 7 (<https://mafft.cbrc.jp/alignment/server/>, accessed on 10 June 2023) [22], and each gene was analysed using BioEdit v. 7 [23]. Maximum likelihood (ML) analysis was performed using RAxML-HPC2 v. 8.2.12 [24] as implemented in the CIPRES (<https://www.phylo.org/portal2/login!input.action>, accessed on 10 June 2023) portal [25] using the GTR+G+I model and 1000 rapid bootstrap (BS) replicates for all genes. A reciprocal 70% bootstrap support approach was used to check for conflicts between the tree topologies from individual genes. Since there was no significant incongruence in topology between the ML trees, the ITS was partitioned for phylogenetic analyses. For Bayesian inference (BI), the best substitution model for each partition was determined by MrModeltest 2.2 [26]. The results suggested ITS1: JC+I, 5.8S: GTR+G+I, ITS2: K80+I+G. Bayesian analysis was performed using MrBayes ver. 3.2.7a [27] in the CIPRES portal. Four parallel runs, each consisting of one cold and three heated chains, were performed for 10 million generations with sampling every 100 generations for the single gene trees and 50 million generations with sampling every 1000 generations. Parameter convergence > 200 was verified in Tracer v. 1.7 [28]. Phylogenetic clades were strongly supported if bootstrap support value (BS) was  $\geq 70\%$  and/or posterior probability (PP) was  $\geq 0.95$ .

**Table 1.** Names, specimen numbers, origin, references and corresponding GenBank accession numbers of *Lyophyllum* sequences used in the phylogenetic analyses. Newly generated sequences are in bold black, and a "\*" before taxa indicates holotypes.

| Taxon Name                  | Specimen Number | Origin      | ITS      | Reference |
|-----------------------------|-----------------|-------------|----------|-----------|
| <i>Calocybe gambosa</i>     | HC78/64         | Switzerland | AF357027 | [7]       |
| <i>C. carnea</i>            | CBS552.50       | Switzerland | AF357028 | [7]       |
| <i>C. persicolor</i>        | HC80/99         | Switzerland | AF357026 | [7]       |
| <i>Hypsizygus marmoreus</i> | V.1611          | Germany     | AJ494834 | [11]      |
| <i>H. marmoreus</i>         | V.3133          | China       | FJ609271 | [11]      |
| <i>H. marmoreus</i>         | HMW2            | Malaysia    | HM561971 | [11]      |
| <i>H. marmoreus</i>         | HZND-1          | China       | JX046028 | [11]      |
| <i>H. marmoreus</i>         | 1-1             | Korea       | KF192813 | [11]      |
| <i>H. tessulatus</i>        | AFTOL-ID 1898   | USA         | DQ917653 | [11]      |
| <i>H. tessulatus</i>        | L2              | China       | FJ467372 | [11]      |
| <i>H. ulmarius</i>          | CBS 286.77      | Korea       | AY265850 | [29]      |
| <i>H. ulmarius</i>          | DUKE-JM/HW      | USA         | EF421105 | [30]      |
| <i>L. ambustum</i>          | CBS452.87       | Switzerland | AF357057 | [7]       |
| <i>L. anthracophilum</i>    | HC79/132        | Switzerland | AF357055 | [7]       |
| <i>L. atratum</i>           | CBS709.87       | Switzerland | AF357053 | [7]       |
| <i>L. atrofusum</i>         | HMJAU63461      | China       | OP605493 | [16]      |
| <i>L. atrofusum</i>         | HMJAU63456 *    | China       | OP605494 | [16]      |
| <i>L. caerulescens</i>      | HC80-140        | Switzerland | AF357052 | [7]       |
| <i>L. caerulescens</i>      | V.15759         | USA         | JF908339 | [11]      |
| <i>L. crassifolium</i>      | V.5077          | Italy       | JF908331 | [11]      |
| <i>L. decastes</i>          | dd08054         | China       | FJ810160 | [11]      |
| <i>L. decastes</i>          | Ld418           | China       | HM119485 | [11]      |
| <i>L. deqinense</i>         | YAASM6949 *     | China       | OQ418117 | [18]      |
| <i>L. deqinense</i>         | YAASM6948       | China       | OQ418116 | [18]      |
| <i>L. deliberatum</i>       | V.15032         | Slovenia    | JF908338 | [11]      |

Table 1. Cont.

| Taxon Name                 | Specimen Number | Origin       | ITS             | Reference         |
|----------------------------|-----------------|--------------|-----------------|-------------------|
| <i>L. favrei</i>           | BSI94cp2        | Switzerland  | AF357035        | [7]               |
| <i>L. favrei</i>           | V.6334          | Italy        | JF908333        | [11]              |
| <i>L. fumosum</i>          | SJ02/006        | Sweden       | HM572539        | [11]              |
| <i>L. fumosum</i>          | LAS00/144       | Sweden       | HM572541        | [11]              |
| <i>L. fumosum</i>          | V.16077         | Italy        | JF908340        | [11]              |
| <i>L. fumosum</i>          | LfumNlf24       | Japan        | JN983977        | [11]              |
| <i>L. fumosum</i>          | L2010512371     | China        | JX966310        | [11]              |
| <i>L. fumosum</i>          | YAASM6215       | China        | ON681708        | [17]              |
| <i>L. fumosum</i>          | YAASM6340       | China        | ON681709        | [17]              |
| <i>L. gangraenosum</i>     | V.12332         | Italy        | JF908335        | [11]              |
| <b><i>L. heimogu</i></b>   | <b>L3026 *</b>  | <b>China</b> | <b>KY434100</b> | <b>This study</b> |
| <b><i>L. heimogu</i></b>   | <b>L3033</b>    | <b>China</b> | <b>KY434101</b> | <b>This study</b> |
| <b><i>L. heimogu</i></b>   | <b>L3035</b>    | <b>China</b> | <b>KY434102</b> | <b>This study</b> |
| <i>L. infumatum</i>        | V.10152         | Italy        | JF908334        | [11]              |
| <i>L. leucophaeatum</i>    | Hae251.97       | Switzerland  | AF357032        | [7]               |
| <i>L. littoralis</i>       | CA20091210      | Italy        | JX280410        | [11]              |
| <i>L. lorricatum</i>       | V.13175         | USA          | JF908336        | [11]              |
| <i>L. lorricatum</i>       | CA20090202.03   | Italy        | JX280406        | [11]              |
| <i>L. lorricatum</i>       | 01.12.09        | Italy        | JX280407        | [11]              |
| <i>L. moncalvoanum</i>     | PDD 96328 *     | New Zealand  | NR_137615       | [2]               |
| <i>L. moncalvoanum</i>     | PDD 102581      | New Zealand  | KJ461912        | [2]               |
| <i>L. ochraceum</i>        | BSI94.cp1       | Switzerland  | AF357033        | [7]               |
| <i>L. ochraceum</i>        | V.537           | Italy        | JF908329        | [11]              |
| <i>L. rhombisporum</i>     | L1762*          | China        | JX966307        | [11]              |
| <i>L. rhombisporum</i>     | L2082           | China        | JX966308        | [11]              |
| <i>L. semitale</i>         | HC85/13         | Switzerland  | AF357049        | [7]               |
| <i>L. semitale</i>         | EL187-09        | Sweden       | HM572552        | [8]               |
| <i>L. shimeji</i>          | Olsen821006     | Sweden       | HM572530        | [8]               |
| <i>L. shimeji</i>          | NZ4Q88          | New Zealand  | JN983985        | [11]              |
| <i>L. shimeji</i>          | L2010512377     | China        | JX966311        | [11]              |
| <i>L. sp.</i>              | PBM 2688        | USA          | DQ182502        | [11]              |
| <i>L. sp.</i>              | SB102           | China        | FJ687273        | [11]              |
| <i>L. sp.</i>              | Aase811014      | Sweden       | HM572550        | [8]               |
| <i>L. sp.</i>              | TO-2011         | Italy        | JF908337        | [11]              |
| <i>L. sp.</i>              | JN001           | China        | FJ687270        | [11]              |
| <i>L. sp.</i>              | O73586          | Netherlands  | GU234137        | [11]              |
| <i>L. sp.</i>              | Cultivar Jpn    | Sweden       | HM572551        | [8]               |
| <i>L. sp.</i>              | SL-2013         | China        | JX966308        | [11]              |
| <i>L. sykosporum</i>       | IFO30978        | Switzerland  | AF357050        | [7]               |
| <i>L. subalpinarum</i>     | HMJAU63447 *    | China        | OP605490        | [16]              |
| <i>L. subalpinarum</i>     | HMJAU63453      | China        | OP605491        | [16]              |
| <i>L. subdecastes</i>      | HMJAU63470      | China        | OP605488        | [16]              |
| <i>L. subdecastes</i>      | HMJAU63467 *    | China        | OP605489        | [16]              |
| <i>L. turcicum</i>         | KATO-2971 *     | Turkey       | KJ158159        | [11]              |
| <b><i>L. yiqunyang</i></b> | <b>L4206</b>    | <b>China</b> | <b>KY434104</b> | <b>This study</b> |
| <b><i>L. yiqunyang</i></b> | <b>L2989 *</b>  | <b>China</b> | <b>KY434103</b> | <b>This study</b> |
| <i>Tephroclybe boudier</i> | BSI96/84        | USA          | DQ825427        | [11]              |

### 3. Result

#### 3.1. Phylogenetic Analysis

Five newly generated and 64 retrieved sequences from GenBank were used as the ingroup. Three sequences of *Calocybe gambosa*, *C. carnea* and *C. persicolor* retrieved from GenBank were used as the outgroup [30]. The ITS was 575 characters in length, of which 284 characters were constant, 291 were variable but parsimony-uninformative and 231 were parsimony-informative. Estimated base frequencies were as follows: A = 0.231384, C = 0.218313, G = 0.226647, T = 0.323655, substitution rates AC = 1.548998, AG = 4.813675,

AT = 2.464671, CG = 0.797610, CT = 7.018015, GT = 1.000000; gamma distribution shape parameter  $\alpha = 0.394616$ .

ML and BI analyses generated nearly identical tree topologies with minimal variation in statistical support values. Thus, only the ML tree is displayed (Figure 1). Phylogenetic data and thorough morphological analysis (see below) showed that the two newly described taxa in this study are significantly distinguished from other known *Lyophyllum* species.



**Figure 1.** Phylogram generated from maximum likelihood (RAxML) analysis based on combined sequence data of ITS1–5.8S–ITS2 alignment of *Lyophyllum*. *Calocybe persicolor*, *C. carnea* and *C. gambosa* were used as the outgroup. ML bootstrap support values/Bayesian posterior probability greater than 70%/0.95 are indicated. Species names in red represent new species.

In our phylogeny, our species *L. yiqunyang* (L4206 and L2989) and *L. heimogu* (L3026, L3033 and L3035) formed independent branches. Comparing the ITS sequences, there were 12/575 (2.09%), 18/575 (3.13%), 16/575 (2.78%) and 6/575 (1.04%) differences between *L. yiqunyang* (L2989, holotype) and *L. heimogu* (L3026, holotype), *L. loricatum* (V.13175), *L. littoralis* (CA20091210) and *L. subdecastes* (HMJAU 63467 holotype), respectively, as well as 15/575 (2.78%), 15/575 (2.78%), 13/575 (2.26%) and 9/575 (1.57%) ITS sequence differences

between *L. heimogu* (L3026, holotype) and *L. lorricatum* (V.13175), *L. littoralis* (CA20091210), *L. subdecastes* (HMJAU63467 holotype) and *L. decastes* (Ld418), respectively.

### 3.2. Taxonomy

*Lyophyllum yiqunyang* Shu H. Li sp. nov. Figure 2, Figure 3 and Figure 6.



**Figure 2.** Fresh basidiomata of *Lyophyllum yiqunyang* ((a) YAAS L2989; (b) YAAS L4206). Scale bars: (a,b) = 1 cm.

MycoBank: 849794.

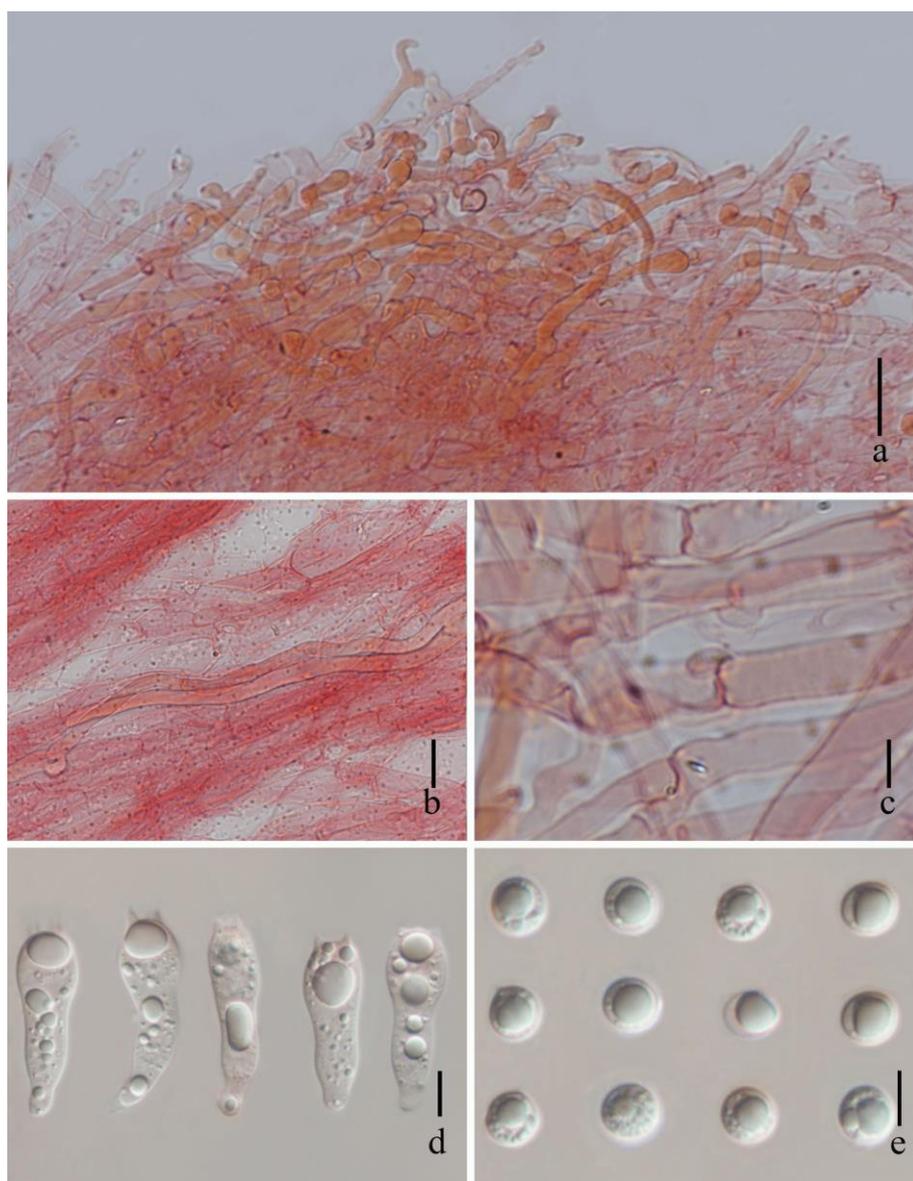
Holotype: China, Sichuan Province, Jiuzhaigou County, 103°54'37" E, 33°10'34" N, elev. 3251 m, in a forest dominated *Abies* spp., *Picea* spp. and *Salix cupularis*, Shu–Hong Li, 10 August 2014. (YAAS L2989, holotype!).

Etymology: “yiqunyang” refers to the common name used in mushroom markets in China.

Basidiomata mid-sized (Figure 2), Pileus 3.0–6.0 cm diameter, fleshy, fragile, variable in shape according to growth conditions, hemispherical when young, becoming convex with age; surface smooth, dry, olive-grey (2E-F), unchanging, without umbo, deflexed aspects of margin; pileus context thick, 0.1–0.3 cm wide, white (1A1), watery soaked in wet weather. Lamellae moderately close together, ventricose to broadly ventricose, adnate to narrowly adnate, broad, white (1A1) to pale grey (1B1), non-discolouring when bruised; edge even or entire. Stipe 3.0–7.0 × 0.5–1.5 cm, cylindrical to clavate, white or light grey, pruinose at the base, sometimes tapered towards at the base, often twisted, white (1A1) to pale grey (1B1), smooth, sometimes squamous on the surface, unchanging in colour when injured. Odour and taste not distinctive.

Basidiospores [120/2/2] 6.1–6.3 × 6.0–6.2 μm (Figure 3 and Figure 6), Q = 1.1–1.3, Q<sub>m</sub> = 1.18 ± 0.35, av. 6.21 ± 0.12 × 6.15 ± 0.24 μm, globose, subglobose to broadly ellipsoid, hyaline, smooth. Basidia 31.0–42.0 × 7.8–9.0 μm (N = 20), av. 38.1 ± 3.4 × 8.3 ± 1.28 μm, four-spored, sometime with basal clamp connections, clavate, siderophilous granulation. Subhymenium made up of moderately thin-walled hyphae, 5–15 μm across. Hymenophoral trama regular, consisting of thin and hyaline hyphae, some with clamp connection. Marginal cells absent. Pileipellis an interwoven trichodermium to a subcutis composed of almost hyaline interwoven filamentous hyphae, terminal cells 20–83 × 3–8 μm, almost cylindrical to subcylindrical, occasional hyphal tips flexuous and sometimes inflate, some with clamp connections. Stipitipellis a cutis of elongate hyphae. Clamp connection present at some septa. Thromboplerous hyphae present on the hymenophoral trama, 5–8 μm wide.

Ecology and distribution: saprotrophic (growing on fallen wood or soil); when growing on the soil, the soil's upper layer has a relatively thick humus layer; gregarious in forests of *Abies* spp., *Picea* spp. or *Salix cupularis*. Summer to autumn (August to September). Known only from Sichuan Province, China.



**Figure 3.** *Lyophyllum yiqunyang* (YAAS L2989). (a) Pileipellis; (b) Thromboplerous hyphae; (c) Clamp connection; (d) Basidia; (e) Basidiospores. Scale bars: (a) = 20  $\mu\text{m}$ , (b–e) = 5  $\mu\text{m}$ .

Additional specimens examined: China, Sichuan province, Jiuzhaigou County, elev. 3200 m, latitude 103°54'37" E and longitude 33°10'34" N, in a forest dominated by *Abies* spp. and *Salix cupularis*, Shu-Hong Li, 8 August 2015, (L4206).

Notes: *Lyophyllum yiqunyang* is quite similar to *L. deqinense* in having an olive-greyish to greyish-orange pileus and subglobose to globose basidiospores, the *L. deqinense* basidia are shorter (24.1–33.8  $\times$  7.3–9.9  $\mu\text{m}$ ). The pileipellis of *L. deqinense* is a cutis composed of parallel elements [18].

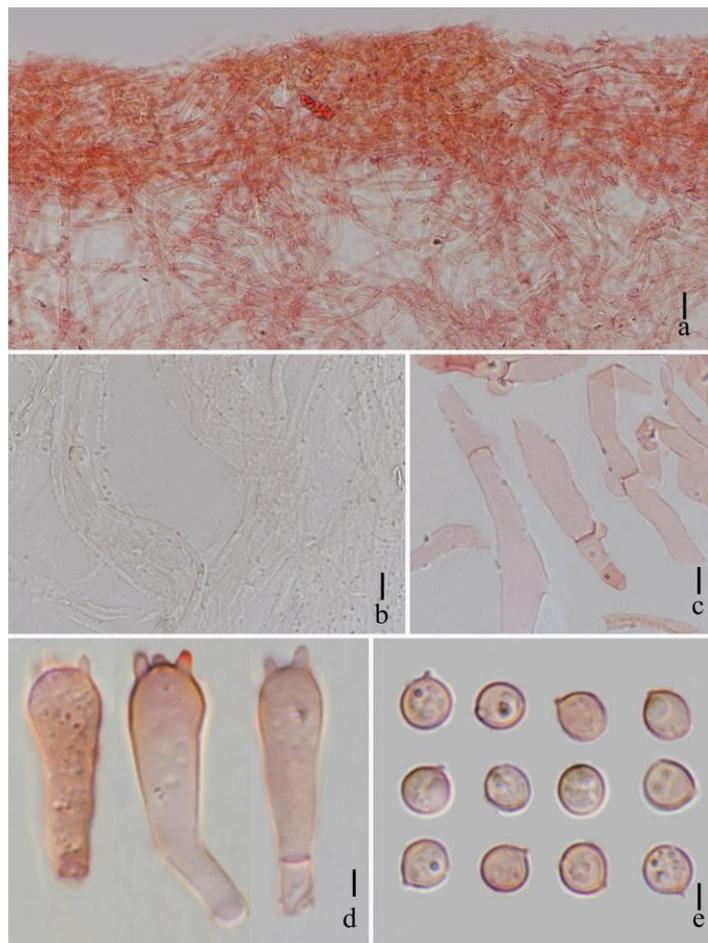
*Lyophyllum yiqunyang* is also similar to *L. subalpinarum* in having a greyish-yellow to olive-greyish pileus. *Lyophyllum subalpinarum* stipe is hollow, whitish-greyish to dark in the middle. The lamellae of *L. subalpinarum* change to black when touched or injured and present rounded-cylindrical basidiospores (6.9–8.7  $\times$  4.3–5.1  $\mu\text{m}$ ) [16].

*Lyophyllum yiqunyang* is related to *L. subdecastes*; however, *L. subdecastes* has a yellowish-brown, brown, greyish-red pileus, relatively small basidiospores (3.9–5.0  $\times$  3.7–5.0  $\mu\text{m}$ ) and larger basidia (36.7–50.6  $\times$  8.4–10.9  $\mu\text{m}$ ) [16].

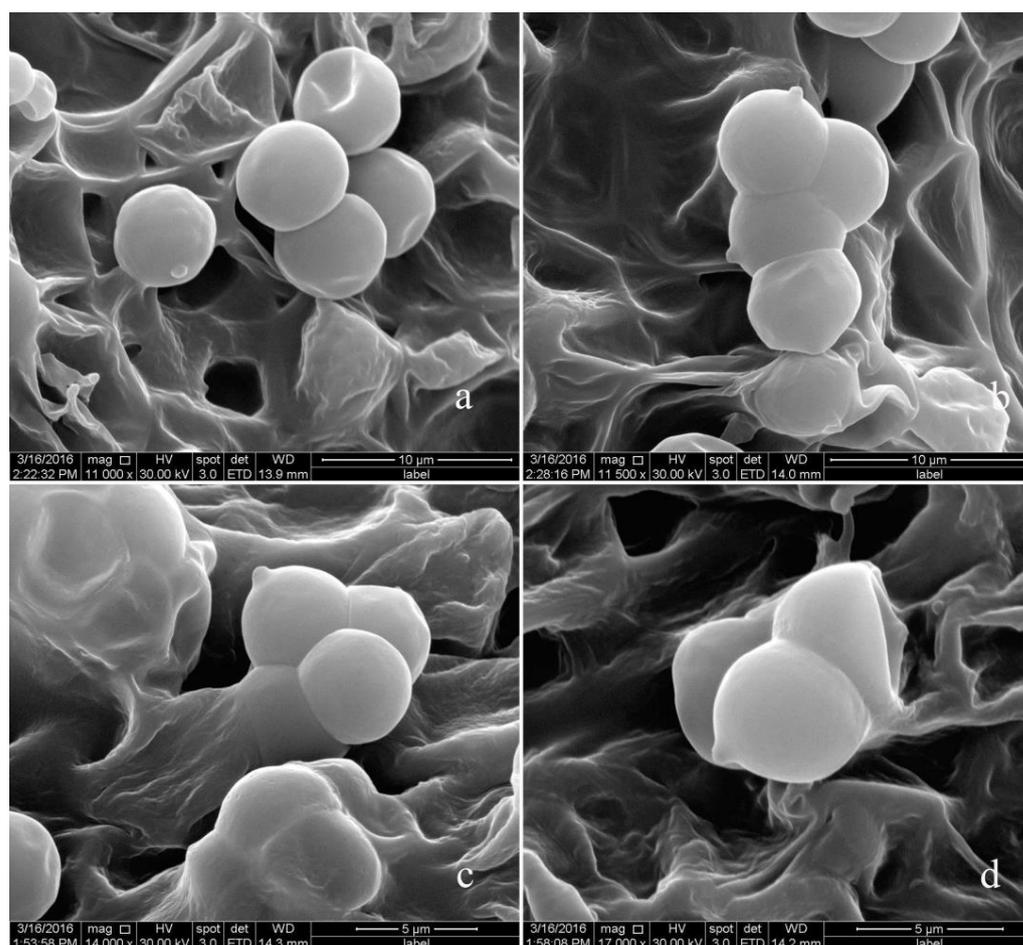
*Lyophyllum heimogu* S. H. Li sp. nov. Figures 4–6.



**Figure 4.** Fresh basidiomata of *Lyophyllum heimogu* ((a) YAAS L3026, holotype; (b) YAAS L3035; (c) YAAS L3533; (d) YAAS L3033). Scale bars: (a–d) = 1 cm.



**Figure 5.** *Lyophyllum heimogu* (YAAS L3026, holotype) (a) Pileipellis; (b) Thromboplerous hyphae (c) Clamp connection; (d) Basidia; (e) Basidiospores. Scale bars: (a) = 20  $\mu\text{m}$ , (b–e) = 5  $\mu\text{m}$ .



**Figure 6.** SEM photo of *Lyophyllum* Basidiospores. (a,b) *Lyophyllum heimogu* (YAAS L3026), (c,d) *Lyophyllum yiqunyang* (YAAS L2989).

Mycobank: 849795.

Holotype: China, Tibet Autonomous Region, Bomi County, elev. 2717 m, 95°43'06'' E, 29°51'41'' N, in a forest dominated by *Abies* spp. and *Picea* spp., Shu-Hong Li, 3 October 2013. (YAAS L3026, holotype!).

Etymology: “heimogu” refers to the common name used by local people.

Basidiomata mid-size (Figure 4), Pileus 3.0–6.5 cm wide, fleshy, fragile, hemispherical when young, becoming convex with age; surface smooth, dark grey (1F1) at the centre, becoming olive (1E-F3) towards the margin with an indistinctly striated margin, umbo papilla, dark (1F1), involute aspects of margin; pileus context thick, 0.1–0.2 cm wide, white (1A1). Lamellae moderately close together, subventricose to ventricose, free to adnexed, lamella edge even or entire, white when young, becoming yellowish-white with age. Context 0.3–0.8 cm thick, white to pale white, unchanging when injured. Stipe 2.5–7.0 × 0.5–2.0 cm, yellowish-brown (5E4-5), central, cylindrical to clavate, sometime bulbous at the base, solid, often twisted, smooth. Odour and taste not distinctive.

Basidiospores [120/2/2] 5.31–5.63 × 5.22–5.41 µm (Figures 5 and 6), Q = 1.1–1.2, Qm = 1.12 ± 0.03, av. 5.5 ± 0.24 × 5.3 ± 0.36 µm, globose to subglobose, hyaline, smooth. Basidia 28.5–32.0 × 7.1–8.3 µm (N = 20), av. 30.2 ± 3.51 × 7.5 ± 0.82 µm, four-spored, sometime with basal clamp connections, clavate, siderophilous granulation. Subhymenium made up of moderately thin-walled hyphae, 4–10 µm across. Hymenophoral trama regular, consisting of thin and hyaline hyphae, some with clamp connection. Marginal cells absent. Pileipellis an interwoven trichodermium to a subcutis composed of almost hyaline interwoven filamentous hyphae, terminal cells 25–100 × 2–5 µm, almost cylindrical to subcylindrical, occasional hyphal tips flexuous and sometimes inflate, some with clamp

connections. Stipitipellis a cutis of elongate hyphae. Clamp connection present at some septa. Thromboplerous hyphae present on the hymenophoral trama, 3–7 µm wide.

Ecology and distribution: saprotrophic (growing on fallen wood or soil); when growing on the soil, the soil has a relatively thick humus layer; clusters or gregarious in forests dominated by *Abies* or *Picea*. Summer to autumn (August to October). Known only from Tibet Autonomous Region, China.

Additional specimens examined: China, Tibet Autonomous Region, Bomi County, alt. 2700 m, in forest of *Abies* spp. and *Picea* spp., Shu–Hong Li, 95°43′15″ E, 29°51′52″ N, 18 September 2014 (YAAS L3035); *ibid*, Shu–Hong Li, 19 August 2014, YAAS L3533; *ibid*, Shu–Hong Li, 20 August 2014, YAAS L3033.

Notes: *Lyophyllum heimogu* is similar to *L. subdecastes*, having medium-sized basidiomata, brown to dark pileus and globose to subglobose basidiospores. However, *L. subdecastes* basidiospores are smaller (av. = 4.47 × 4.25 µm) and present fusoid-ventricose to broadly fusoid-ventricose pleurocystidia [16].

In our phylogenetic analyses, the phylogenetic positions of *L. heimogu* and *L. decastes* within *Lyophyllum* were well supported (80/1.00) as monophyletic clades. However, *L. decastes* was found in Poland and has a broad pileus, ellipsoid basidiospores (5.0–7.0 µm) [4]; ITS sequence differences between *L. heimogu* (L3026, holotype) and *L. decastes* (Bengtsson 19910929, HM572545 from Sweden) were 12/616 (1.95%).

#### 4. Discussion

From a morphological perspective, *L. yiqunyang* and *L. heimogu* are very similar to *L. decastes* and *L. shimeji* but differ in their trophic modes. While *L. yiqunyang* and *L. heimogu* are saprotrophic, *L. decastes* and *L. shimeji* are symbiotic. Molecularly, *L. yiqunyang* and *L. heimogu* are closely related to *L. decastes*. However, the original description of *L. decastes* from Sweden in 1818 [4] was shorter. When comparing the ITS sequences of *L. heimogu* (L3026, holotype) and *L. yiqunyang* (L2989, holotype) with *L. decastes* (Bengtsson 19910929, HM572545 from Sweden), the differences are 12/616 (1.95%) and 13/616 (2.11%), respectively. *Lyophyllum yiqunyang* and *L. heimogu* are considered the most delicious mushrooms that can be stored at low temperatures, making them potential candidates for commercialisation. Detailed comparisons of the diagnostic characteristics of *L. heimogu* and *L. yiqunyang* with similar species are mentioned in Table 2.

Species of sect. *Difformia* are characterised by their caespitose growth [31], which is seen in *L. decastes*, *L. fumosum* and *L. shimeji* belonging to sect. *Difformia*. In our phylogenetic analyses (Figure 1), these three species formed a strongly supported group (90/100) in the sect. *Difformia*. Therefore, we believe that sect. *Difformia* is monophyletic with *L. ambustum*, *L. decastes*, *L. fumosum*, *L. shimeji*, *L. loricatum*, *L. littoralis*, *L. subdecastes* and the two new species studied here, *L. yiqunyang* and *L. heimogu*.

Most species of *Lyophyllum* are delicious edible mushrooms, such as *L. decastes*, *L. shimeji* and *L. subdecastes*. Cases of poisoning associated with *Lyophyllum* species have not been reported. Wu et al. [32] have indicated the diversity of Chinese macrofungi, including seven edible *Lyophyllum* species that are distributed in China. Furthermore, some species of *Lyophyllum* have been used in traditional medicine, for example, *L. decastes*, which has been used to treat hypoglycaemia [33].

*Lyophyllum* species are mainly distributed in temperate to subtropical regions [11,31,34]. Most species are saprotrophic, with few being symbiotic [4,11,16,31]. An important characteristic for identifying *Lyophyllum* species is the shape of their basidiospores. In most species of *Lyophyllum*, basidiospores are globose to subglobose (e.g., *L. fumosum*, *L. heimogu*, *L. subdecastes*, *L. shimeiji*, *L. loricatum* and *L. yiqunyang*, among others) [3,11,16,31,33,35,36]. However, in rare cases, species may have irregular rhombus basidiospores (e.g., *L. subalpinarum* and *L. atrofusum* among others) [16].

**Table 2.** Comparison of the diagnostic characteristics of *Lyophyllum heimogu* and *L. yiqunyang* with similar species.

| Taxa                  | Pileus  | Stipe  | Spores   | Basidia                       | Reference            |
|-----------------------|---|--|--|-------------------------------|----------------------|
| <i>L. decastes</i>    | Greyish-brown to yellowish-brown or brown, usually darker when young, 4–12 cm | Whitish-greyish, 5–10 cm   | Broadly ellipsoid, 5–7 × 5–7 µm                        | -                             | [4,33,34]            |
| <i>L. fumosum</i>     | Dark to light grey, brown, 2–10 cm  | Cream-coloured to brown, 2.5–10 cm                                 | Globose to subglobose, 5.5–7 × 5–7 µm                  | 40–45 × 8–10 µm               | [11]                 |
| <i>L. heimogu</i>     | <b>Dark grey to olive, 3.0–6.5 cm</b>   | <b>Yellowish-brown, 2.5–7.0 × 0.5–2.0 cm</b><br>Orange-white,      | <b>Globose to subglobose, 5.30–5.60 × 5.20–5.40 µm</b> | <b>28.5–32.0 × 7.1–8.3 µm</b> | <b>Present study</b> |
| <i>L. subdecastes</i> | Yellowish-brown, brown to greyish-red   | reddish-grey to greyish-red<br>2.7–6.6 × 0.5–1.5 cm                | Globose to subglobose, 3.9–5.0 × 3.7–5.0 µm            | 36.7–50.6 × 8.4–10.9 µm       | [16]                 |
| <i>L. shimeiji</i>    | Dark grey to grey, brown, 2–8 cm  | White, 3–8 cm  | Globose to subglobose, 4.0–6.0 × 4.0–6.0 µm            | -                             | [11]                 |
| <i>L. lorcatum</i>    | Reddish-brown to chestnut brown<br>3–12 cm                                    | Cream to pale brownish, grey-brown when old,<br>3.5–9 × 0.7–1.5 cm | Globose to subglobose, 5.0–6.0 × 4.5–5.3 µm            | 28.0–32.0 × 7.0–8.0 µm        | [4]                  |
| <i>L. littoralis</i>  | Grey to brownish-grey, 5–15 cm  | Grey, 1.5–4 × 0.4–1.5 cm   | Globose to subglobose, 4.5–5.5 × 4.5–5.5 µm            | -                             | [36]                 |
| <i>L. yiqunyang</i>   | <b>Olive-grey, 3.0–6.0 cm</b>   | <b>White or light grey, 3.0–7.0 × 0.5–1.5 cm</b>                   | <b>Globose to subglobose, 6.1–6.3 × 6.0–6.2 µm</b>     | <b>31.0–42.0 × 7.8–9.0 µm</b> | <b>Present study</b> |

**Author Contributions:** S.L. wrote the manuscript and phylogenetic analyses; S.T. and J.H. prepared the samples; D.Z. designed the article structure and correct manuscript. All authors have read and agreed to the published version of the manuscript.

**Funding:** This research was funded by the National Natural Science Foundation of China (Project ID: 32060006, 31560009, 31160010), China Agriculture Research System (Project ID: CARS-20), Central guidance for local scientific and technological development funds (202307AB110001).

**Institutional Review Board Statement:** Not applicable.

**Informed Consent Statement:** Not applicable.

**Data Availability Statement:** Not applicable.

**Acknowledgments:** We would like to thank the three anonymous reviewers for giving us comments that have improved our manuscript.

**Conflicts of Interest:** The authors declare no conflict of interest.

## References

- Dähncke, R.M.; Contu, M.; Vizzini, A. Two new species of *Lyophyllum* s.l. (Basidiomycota, Agaricomycetes) from La Palma (Canary Islands, Spain). *Mycotaxon* **2011**, *115*, 65–71. [[CrossRef](#)]
- Cooper, J.A. New species and combinations of some New Zealand agarics belonging to *Clitopilus*, *Lyophyllum*, *Gerhardtia*, *Clitocybe*, *Hydnangium*, *Mycena*, *Rhodocollybia* and *Gerronema*. *Mycosphere* **2014**, *5*, 263–288. [[CrossRef](#)]
- Bon, M. *Flore Mycologique d'Europe. Les collybiomarasmioides et Ressemblants*; Doc Mycol Mémoire Hors-Série. n. 5; Association d'Ecologie et de Mycologie: Amiens, France, 1999.
- Breitenbach, J. *Fungi of Switzerland. Volume 3, Boletes and Agarics 1*; Verlag Mycologia: Luzern, Switzerland, 1991.
- Cléménçon, H.; Smith, A.H. New species of *Lyophyllum* (Agaricales) from North America and a key to the known staining species. *Mycotaxon* **1983**, *18*, 379–437.
- Cléménçon, H. Schwarzende *Lyophyllum*-Arten Europas. *Z. Mykol.* **1986**, *52*, 61–84.
- Hofstetter, V.; Cléménçon, H.; Vilgalys, R.; Moncalvo, J.M. Phylogenetic analyses of the *Lyophylleae* (Agaricales, Basidiomycota) based on nuclear and mitochondrial rDNA sequences. *Mycol. Res.* **2002**, *106*, 1043–1059. [[CrossRef](#)]
- Larsson, E.; Sundberg, H. *Lyophyllum shimeiji*, a species associated with lichen pine forest in northern Fennoscandia. *Mycoscience* **2011**, *52*, 289–295. [[CrossRef](#)]

9. Vizzini, A.; Contu, M. *Lyophyllum rosae-mariae* sp. nov. (Basidiomycota, Agaricomycetes) from La Palma (Canary Islands, Spain). *Mycosphere* **2010**, *1*, 83–86.
10. Yamada, A.; Ogura, T.; Ohmasa, M. Cultivation of mushrooms of edible ectomycorrhizal fungi associated with *Pinus densiflora* by in vitro mycorrhizal synthesis: I. Primordium and basidiocarp formation in open-pot culture. *Mycorrhiza* **2001**, *11*, 59–66. [[CrossRef](#)]
11. Sesli, E.; Vizzini, A.; Contu, M. *Lyophyllum turcicum* (Agaricomycetes: Lyophyllaceae), a new species from Turkey. *Turk. J. Bot.* **2015**, *39*, 512–519. [[CrossRef](#)]
12. He, M.Q.; Zhao, R.L.; Kevin, D.H.; Dominik, B.; Martin, K.; Andrey, Y.; Eric, H.C.; Olivier, R.; Makoto, K.; Santiago, S.; et al. Notes, outline and divergence times of Basidiomycota. *Fungal Divers.* **2019**, *99*, 105–367. [[CrossRef](#)]
13. Fanglan, D. *A Summary of Chinese Fungi*; Science Press: Beijing, China, 1979.
14. Li, X.; Li, Y. Research progress on fungus of the genus *Lyophyllum* in China. *J. Edible Fungi* **2009**, *16*, 75–79.
15. Wang, X.Q.; Zhou, D.Q.; Zhao, Y.C.; Zhang, X.L.; Li, L.; Li, S.H. *Lyophyllum rhombisporum* sp. nov. from China. *Mycotaxon* **2013**, *123*, 473–477. [[CrossRef](#)]
16. Wei, S.W.; Lu, B.Y.; Wang, Y.; Dou, W.J.; Wang, Q.; Li, Y. Morphology and Phylogeny of Lyophylloid Mushrooms in China with Description of Four New Species. *J. Fungi* **2023**, *9*, 77. [[CrossRef](#)]
17. Ma, Y.H.; Liu, P.; Zhao, Z.Y.; Chen, W.M.; Zhao, Y.C. *Lyophyllum pallidofumosum* sp. nov. (Lyophyllaceae, Agaricales), from southwestern China. *Phytotaxa* **2022**, *576*, 173–183. [[CrossRef](#)]
18. Ma, Y.H.; Liu, P.; Zhao, Z.Y.; Chen, W.M.; Zhao, Y.C. *Lyophyllum deqinense* (Lyophyllaceae, Agaricales), a new species from southwestern China. *Phytotaxa* **2023**, *598*, 219–228. [[CrossRef](#)]
19. Visnovsky, S.B.; Cummings, N.; Guerin-Laguette, A.; Wang, Y.; Yamada, A.; Kobayashi, H.; Kawai, M.; Pitman, A.R. Detection of the edible ectomycorrhizal fungus *Lyophyllum shimeji* colonising seedlings of cultivated conifer species in New Zealand. *Mycorrhiza* **2014**, *24*, 453–463. [[CrossRef](#)] [[PubMed](#)]
20. Kornerup, A.; Wanscher, J.H. *Methuen Handbook of Colour*, 3rd ed.; Eyre Methuen: London, UK, 1978.
21. White, T.J.; Bruns, T.D.; Lee, S.; Taylor, J.W. Amplification and direct sequencing of fungal ribosomal RNA genes for phylogenetics science direct. *PCR Protoc.* **1990**, *18*, 315–322.
22. Katoh, K.; Standley, D.M. MAFFT multiple sequence alignment software version 7, improvements in performance and usability. *Mol. Biol. Evol.* **2013**, *30*, 772–780. [[CrossRef](#)]
23. Hall, T. BioEdit v7. 2007. Available online: <http://www.mbio.ncsu.edu/BioEdit/BioEdit.html> (accessed on 10 June 2023).
24. Stamatakis, A. RAxML version 8, a tool for phylogenetic analysis and post-analysis of large phylogenies. *Bioinformatics* **2014**, *30*, 1312–1313. [[CrossRef](#)]
25. Miller, M.A.; Pfeiffer, W.; Schwartz, T. Creating the CIPRES Science Gateway for inference of large phylogenetic trees. In Proceedings of the Gateway Computing Environments Workshop (GCE), New Orleans, LA, USA, 14 November 2010.
26. Nylander, J.A.A. *MrModeltest*, v. 2.2.; Program Distributed by the Author; Uppsala University, Department of Systematic Zoology: Uppsala, Sweden, 2004.
27. Ronquist, F.; Huelsenbeck, J.; Teslenko, M. MrBayes Version 3.2 Manual: Tutorials and Model Summaries. 2011. Available online: <https://brahms.biology.rochester.edu/software.html> (accessed on 10 June 2023).
28. Rambaut, A.; Drummond, A.J.; Xie, D.; Baele, G.; Suchard, M.A. Posterior summarization in Bayesian phylogenetics using Tracer 1.7. *Syst. Biol.* **2018**, *67*, 901–904. [[CrossRef](#)]
29. Ravikumar, G.; Kalaiselvi, M.; Gomathi, D.; Vidhya, B.; Devaki, K.; Uma, C. Effect of laccase from *Hypsizygus ulmarius* in decolorization of different dyes. *J. Appl. Pharm. Sci.* **2013**, *3*, 150–152. [[CrossRef](#)]
30. Hofstetter, V.; Redhead, S.A.; Kauff, F.; Moncalvo, J.M.; Matheny, P.B.; Vilgalys, R. Taxonomic revision and examination of ecological transitions of the Lyophyllaceae (Basidiomycota, Agaricales) based on a multigene phylogeny. *Cryptogam. Mycol.* **2014**, *35*, 399–425. [[CrossRef](#)]
31. Singer, R. *The Agaricales in Modern Taxonomy*, 4th ed.; Koeltz Scientific Books: Koenigstein, Germany, 1986.
32. Wu, F.; Zhou, L.W.; Yang, Z.L.; Bau, T.; Li, T.H.; Dai, Y.C. Resource diversity of Chinese macrofungi: Edible, medicinal and poisonous species. *Fungal Divers.* **2019**, *98*, 1–76. [[CrossRef](#)]
33. Trudell, S.; Ammirati, J. Mushrooms of the Pacific Northwest. In *Timber Press Field Guides*; Timber Press: Portland, OR, USA, 2009; pp. 112–113.
34. Davis, R.; Sommer, R.; John, A. *Field Guide to Mushrooms of Western North America*; University of California Press: Berkeley, CA, USA, 2012; p. 139.
35. Miura, T.; Kubo, M.; Itoh, Y.; Iwamoto, N.; Kato, M.; Park, S.R.; Yuuichi, U.; Yukio, K.; Ikukatsu, S.; Suzuki, I. Antidiabetic activity of *Lyophyllum decastes* in genetically type 2 diabetic mice. *Biol. Pharm. Bull.* **2002**, *2*, 1234–1237. [[CrossRef](#)] [[PubMed](#)]
36. Ballero, M.; Contu, M. A new species of *Calocybe* (Agaricales, Lyophylleae) from litoral pine woods of Sardinia (Italy). *Mycotaxon* **1990**, *39*, 473–476.

**Disclaimer/Publisher’s Note:** The statements, opinions and data contained in all publications are solely those of the individual author(s) and contributor(s) and not of MDPI and/or the editor(s). MDPI and/or the editor(s) disclaim responsibility for any injury to people or property resulting from any ideas, methods, instructions or products referred to in the content.