

## Article

# Diversity of Seed Flavan-3-Ols in Croatian Native Grapevine Cultivars (*Vitis vinifera* L.) Grown in Coastal Region

Željko Andabaka <sup>1</sup> , Iva Šikuten <sup>1,2,\*</sup> , Ivana Tomaz <sup>1,2</sup> , Domagoj Stupić <sup>1</sup>, Zvezdana Marković <sup>1,2</sup>,  
Jasminka Karoglan Kontić <sup>1,2</sup>, Edi Maletić <sup>1,2</sup> and Darko Preiner <sup>1,2</sup> 

<sup>1</sup> Department of Viticulture and Enology, Faculty of Agriculture, University of Zagreb, 10 000 Zagreb, Croatia

<sup>2</sup> Centre of Excellence for Biodiversity and Molecular Plant Breeding, Faculty of Agriculture, University of Zagreb, 10 000 Zagreb, Croatia

\* Correspondence: isikuten@agr.hr; Tel.: +385-01-4627-974

**Abstract:** Seed extracts are becoming more important due to their beneficial biological activities. The main constituents of seed extracts are flavan-3-ols, compounds important in winemaking. The coastal region in Croatia is rich in native grapevine varieties, which are used in wine production. The aim of the research was to analyze the flavan-3-ol profiles of 20 native varieties, and to evaluate the potential use of grape seeds as a source of flavan-3-ols. The flavan-3-ols from seeds were analyzed by HPLC. The predicted yield of flavan-3-ols was calculated using the analyzed profiles. In total, eight compounds were identified, with the most abundant compounds being catechin, epicatechin, and procyanidin B2. In general, the red grape varieties had higher content of flavan-3-ols than the white varieties, which was confirmed by PCA. The coastal region could potentially yield up to 73.97 kg/ha of flavan-3-ols, depending on the variety. The results show the diversity of flavan-3-ol profiles among Croatian varieties and their potential usage as a source of valuable nutraceuticals.

**Keywords:** grape seeds; flavan-3-ols; grapevine varieties; Croatia



**Citation:** Andabaka, Ž.; Šikuten, I.; Tomaz, I.; Stupić, D.; Marković, Z.; Kontić, J.K.; Maletić, E.; Preiner, D. Diversity of Seed Flavan-3-Ols in Croatian Native Grapevine Cultivars (*Vitis vinifera* L.) Grown in Coastal Region. *Diversity* **2022**, *14*, 667. <https://doi.org/10.3390/d14080667>

Academic Editor: Michael Wink

Received: 7 July 2022

Accepted: 16 August 2022

Published: 17 August 2022

**Publisher's Note:** MDPI stays neutral with regard to jurisdictional claims in published maps and institutional affiliations.



**Copyright:** © 2022 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (<https://creativecommons.org/licenses/by/4.0/>).

## 1. Introduction

Croatia is a home to more than 120 native grapevine varieties grown in four different climate zones. The coastal region of Croatia is very important for its wealth of indigenous varieties, and about a hundred different varieties are grown in this area [1]. The Dalmatian subregion is characterized by a Mediterranean climate and karst relief, which enable the production of premium quality wines from indigenous *Vitis vinifera* varieties, such as Plavac mali, Grk, Posip, Babic, and Malvasija dubrovacka.

However, the quality of wines depends not only on the basic chemical parameters, such as total soluble solids, organic acids, and pH, but also on the secondary metabolites, polyphenolic, and volatile compounds. The polyphenolic compounds are important secondary metabolites, influencing the grape and wine quality, color, and sensory properties, such as astringency and bitterness [2]. Furthermore, these compounds are becoming more important as compounds with a beneficial influence on human health, due to their antioxidant, anti-cancer, and antimicrobial properties [3,4]. The polyphenols in grapes are distributed between skin (28–35%), seeds (60–70%), and pulp (<10%) [5], thus, the seeds represent a valuable source of polyphenolic compounds. The flavan-3-ols are present in grapes as monomers and polymers. The main monomers are catechin, epicatechin, epigallocatechin, and epicatechin-gallate [6]. The polymers are represented by procyanidins or condensed tannins, which are built from monomeric forms of flavan-3-ols. The procyanidins B1–B4 differ in the arrangement of catechin and epicatechin starter and extension units, and are under strict enzymatic control [7]. In grape seeds, the most abundant are procyanidins, followed by catechin, epicatechin, and epicatechin-gallate [8]. The flavan-3-ols, like other classes of polyphenolic compounds, also showed beneficial biological activities,

such as antioxidant, antidiabetic, cardioprotection, which were extensively reviewed by several groups of authors [9–11]. Moreover, the phenolics in the grape seeds are important in improving the wine structure and enhancing the aging potential, as well as protecting the wine against oxidation and stabilizing wine color [12]. Only a few pieces of research on seed polyphenols were conducted that included the Croatian varieties. Curko et al. [13] investigated seed flavan-3-ols from Plavac mali and Babic, where the dominant compounds were catechin and epicatechin. Similar results were presented for Vranac seeds, where catechin was also the dominant compound [14].

During vine cultivation and winemaking, high amounts of waste are produced, including pomace, seeds, stems, prunings, yeast and bacteria lees, organic acids, CO<sub>2</sub>, and wastewater [15]. The seeds are a relevant part of the waste, and represent 20–25% of the biomass generated by the wine industry, which in many countries is considered as a disposable material [16]. Furthermore, the conventional treatments of the wastes are expensive, as they require considerable amounts of economic and energy resources for their safe discharge into the environment [17]. Besides the polyphenols, the seeds contain beneficial polyunsaturated fatty acids and oils [16,18,19], thus representing a promising source of nutraceuticals. These physical–chemical properties of the waste material determine its subsequent use in order to minimize the ecological footprint [15]. Thus, these compounds used as grape seed extracts can demonstrate antioxidant and antimicrobial activities [20–23], which enables their usage in the food industry as natural additives, for increasing product quality, safety, and shelf life [24].

Since the research on the seed polyphenols is scarce, the aim of this research was to analyze polyphenolic compounds from the seeds of important native grapevine cultivars grown in the Dalmatia subregion. Another aim of this study was to evaluate the potential use of seeds as a valuable source of flavan-3-ols in commercial production. Thus, we calculated the potential yield of individual compounds based on the analyzed profiles of flavan-3-ols.

## 2. Materials and Methods

### 2.1. Grape Samples

The grape samples were collected at the germplasm collection of the Institute for Adriatic Crops and Karst Reclamation in Split (Dalmatia, Croatia) and belong to the wine-growing region of Central and South Dalmatia. In total, nineteen grape varieties (10 red varieties and 9 white varieties) were collected during two consecutive years, 2011 and 2012. All of the varieties were sampled in both years and subjected to the analysis of seed flavan-3-ols. In Table 1, the list of the collected *Vitis vinifera* cultivars is shown. The cultivars are important native varieties grown in Dalmatia and are included in the revitalization program. All of the grapevine cultivars are of the same age and pruning system (bilateral cordon training), grafted onto 1013 Paulsen with planting spacing  $2.2 \times 1.1$  m. The samples of each cultivar consisted of five bunches randomly picked from five different vines.

**Table 1.** The list of sampled grapevine cultivars.

Red Cultivars	White Cultivars
Babica	Bogdanusa
Babic	Cetinka
Dobricic	Debit
Lasina	Gegic
Ljutun	Grk
Nincusa	Malvazija dubrovacka
Plavac mali	Posip
Plavina	Prc
Tribidrag	
Vranac	Zilavka

The picked bunches were used to record cluster weight, the number of berries per bunch, and the weight of 100 berries. Each cluster for every variety was weighed, and then the berries were removed from each cluster while counting them. The three samples of 100 berries were randomly counted and weighed. These batches of 100 berries were used for counting the seed number per berry, and the given numbers were then used to calculate the percentage of seeds in the berry and the bunch. In Table 2 are shown the mean values of the seed content in the analyzed varieties.

**Table 2.** Mean values of seed content in analyzed grape varieties.

Variety	Color	Mass Cluster (g)	Average Mass 1 Berry (g)	% Seeds in Berry	% Seeds in Bunch
Babica	Red	329.62 ± 91.66 abcd *	2.49 ± 0.65 abcde	2.00 ± 0.71 d	1.95 ± 0.63 d
Babic	Red	422.66 ± 92.83 abc	3.65 ± 1.15 a	2.00 ± 0.70 d	1.90 ± 0.56 d
Dobricic	Red	235.52 ± 86.38 bcd	1.77 ± 0.22 cdefg	2.90 ± 1.41 bcd	2.80 ± 1.41 bcd
Lasina	Red	302.86 ± 85.63 abcd	2.35 ± 0.35 abcde	8.00 ± 3.26 a	7.70 ± 3.48 a
Ljutun	Red	165.53 ± 27.52 d	1.34 ± 0.09 efg	2.50 ± 0.84 cd	2.40 ± 0.84 bcd
Nincusa	Red	343.06 ± 32.31 abcd	3.27 ± 0.61 ab	2.10 ± 0.56 d	2.05 ± 0.63 d
Plavac mali	Red	174.91 ± 49.83 cd	1.99 ± 0.52 abcdefg	2.10 ± 0.55 d	2.10 ± 0.55 d
Plavina	Red	177.33 ± 50.54 cd	1.59 ± 0.08 defg	5.60 ± 1.27 ab	5.30 ± 1.27 ab
Tribidrag	Red	166.86 ± 31.39 d	1.09 ± 0.33 fg	7.40 ± 2.26 a	7.15 ± 2.19 a
Vranac	Red	232.04 ± 55.04 bcd	1.68 ± 0.73 defg	5.35 ± 0.35 abc	5.10 ± 0.28 abc
Bogdanusa	White	294.33 ± 80.76 abcd	3.10 ± 1.12 abc	2.45 ± 0.91 cd	2.30 ± 0.84 cd
Cetinka	White	477.25 ± 190.77 a	2.93 ± 0.53 abcd	3.15 ± 0.77 bcd	3.10 ± 0.84 bcd
Debit	White	433.91 ± 162.02 ab	2.32 ± 0.33 abcdef	2.75 ± 0.49 bcd	2.65 ± 0.49 bcd
Gegic	White	276.06 ± 94.15 abcd	1.80 ± 0.41 cdefg	2.20 ± 0.28 d	2.10 ± 0.28 d
Grk	White	349.79 ± 32.31 abcd	2.28 ± 0.53 abcde	1.93 ± 0.56 d	0.58 ± 0.32 d
M. dubrovacka	White	143.07 ± 25.88 d	0.83 ± 0.26 g	7.00 ± 2.83 a	6.45 ± 2.47 a
Posip	White	244.40 ± 2.19 bcd	2.19 ± 0.37 abcdef	2.65 ± 1.20 bcd	2.55 ± 1.20 bcd
Prc	White	218.99 ± 37.75 cd	2.06 ± 0.65 abcdefg	2.20 ± 0.42 d	2.10 ± 0.42 d
Zilavka	White	223.38 ± 54.90 cd	1.98 ± 0.79 bcdefg	2.85 ± 0.49 bcd	2.75 ± 0.35 bcd
Pr > F		0.036	0.025	0.001	0.001

\* Means with different letters in the same column differ significantly ( $p \leq 0.05$ ).

## 2.2. Sample Preparation and Extraction

The grape seeds were manually removed from the pulp and skin, and air dried. To obtain the powder, the dried seeds were ground using a coffee-grinder (Tefal, France). The analysis for each sample was carried out in triplicate. The extraction procedure was carried out as described by Tomaz et al. [25]. In brief, the solid–liquid extraction was performed using 125 mg of grape seed powder and 10 mL of extraction solvent, comprising of acetonitrile: water: formic acid (20:79:1,  $v/v/v$ ). The extraction was completed on a magnetic stirrer (RTC basic, IKA, Staufen, Germany) at 400 rpm and temperature of 50 °C. The extraction time was one hour. After the extraction was completed, the extracts were filtered with a Phenex-PTFE (polytetrafluorethylene) 0.20 µm, syringe filter (Phenomenex, Torrance, CA, USA), and analyzed by HPLC.

## 2.3. HPLC Analysis

The HPLC separation, identification and quantification of polyphenolic compounds were performed on an Agilent 1100 Series system (Agilent, Germany), according to the method described by Tomaz and Maslov [26]. The separation was performed with a reversed-phase column LunaPhenyl-Hexyl (4.6 × 250 mm; 5 µm particle; Phenomenex, Torrance, CA, USA). The solvents were water: phosphoric acid (99.5:0.5,  $v/v$ , eluent A), and acetonitrile: water: phosphoric acid (50:49.5:0.5,  $v/v/v$ , eluent B). The flavan-3-ols were detected by FLD detector at  $\lambda_{ex} = 225$  nm and  $\lambda_{em} = 320$  nm and by DAD detector at 280 nm. The individual compounds in the berry seeds' extracts were identified by matching the retention time of each chromatographic peak with external standards. The individual peaks of the flavan-3-ols were quantified using a calibration curve of the corresponding standard compound which was based on the peak area. All of the used standard compounds were

obtained from Extrasynthese (Genay, France) and were analytical standard grade with a purity greater than 98%. The results are expressed in mg/kg.

#### 2.4. Statistical Analysis

The individual polyphenolic compounds from the grape seeds were analyzed, using one-way ANOVA and the differences between the given means were evaluated by Duncan's multiple range test at a confidence level of 95% ( $p < 0.05$ ). The data reported in all of the tables are the average of triplicate observations. The mean values were used for principal component analysis (PCA). The statistical analysis was carried out using XLSTAT program (Addinsoft, 2020, New York, NY, USA).

### 3. Results and Discussion

#### 3.1. Seed Content

The varieties differed significantly in all of the analyzed parameters, the most important ones being the seed share per berry and per bunch. The seed share per berry and per bunch were similar and ranged 1.9–7.4%. The highest seed content both in the berry and in the bunch was recorded for Tribidrag and Malvasija dubrovacka, followed by Plavina and Vranac. The varieties Cetinka, Dobricic, Zilavka, Debit had a seed content of around 3%. The other varieties had a seed content around 2%. Although there was no statistically significant differences, the Grk variety had the smallest seed content both in berry (1.92%) and in bunch (0.57%). It is worth noting that Grk is the only analyzed variety with a female flower, resulting in two types of berries in the bunch, fully developed seeded and undersized seedless [27]. Thus, this is probably one of the reasons why the seed content in the bunches was under 1%.

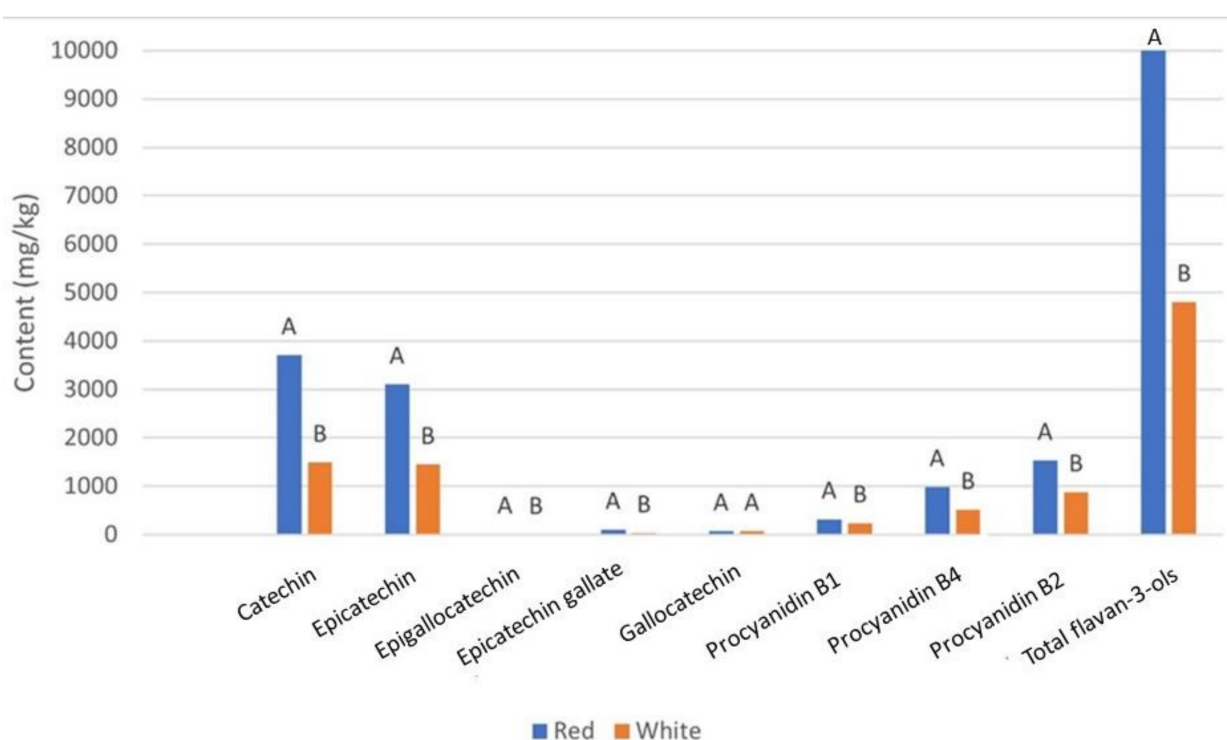
#### 3.2. Flavan-3-Ols in Grape Seeds

The flavan-3-ols are one of the main classes of polyphenolic compounds, influencing the sensory properties of the wine, contributing to the overall astringency, bitterness, and the perception of dryness in the wine [28,29]. However, due to their positive biological activities, these compounds have become increasingly popular, confirmed by the number of functional foods combining them in various formulations [30]. Moreover, grape seeds contain 60–70% of the extractable phenols present in grapes [5], making them a valuable source of polyphenolic compounds. In Table 3 the mean values of the analyzed polyphenolic compounds for all of the cultivars are shown. A total of eight individual compounds of flavan-3-ols were identified and quantified in the grape seeds, and those were catechin, epicatechin, epigallocatechin, epicatechin gallate, gallocatechin, and procyanidins B1, B2, B4.

The analyzed grape cultivars differed significantly in the content of the flavan-3-ols. The total content of the flavan-3-ols ranged from 1723.66 mg/kg to 17,966.52 mg/kg. The most abundant compound was catechin, comprising 35.25% of the total flavan-3-ols, and followed by epicatechin (30.89%), procyanidins B2 (16.19%), B4 (9.92%), and B1 (3.70%). The other compounds were represented with less than 1% in the total content of flavan-3-ols.

[illegible]

The red cultivars had significantly higher content of all of the flavan-3-ols than the white cultivars (Figure 1). The exception was galocatechin, the content of which was similar between the red and white cultivars. Furthermore, the epigallocatechin was not found in the white cultivars, as well as in some red cultivars. The red cultivars in which epigallocatechin was identified (Table 3) were Plavac mali (53.68 mg/kg), Babica (38.31), Lasina (12.11 mg/kg), and Vranac (7.21 mg/kg). Pantelic et al. [31] also identified epigallocatechin in the red varieties, Cabernet Sauvignon, Merlot, Cabernet Franc, Sangiovese, Pinot noir, and Prokupac, ranging from 3.73 mg/kg up to 39.29 mg/kg. Furthermore, the authors reported the presence of epigallocatechin in the white varieties, Sauvignon Blanc, Welschriesling, and Chardonnay. Among the red cultivars, the highest content of total flavan-3-ols (Table 3) was found in Plavac mali and Babica, while the lowest content was found in Nincusa, Tribidrag, and Vranac. Among the white cultivars, the highest amount of total flavan-3-ols was recorded for cultivars Cetinka, Grk, and Posip, while the lowest amount was recorded for Gegic and Debit.



**Figure 1.** Mean values of total content of flavan-3-ols (mg/kg) in red and white cultivars. Different letters above individual compounds represent statistically significant difference ( $p \leq 0.05$ ).

Regarding the most abundant compound, catechin, the highest content among the red cultivars was found in Plavac mali and Babica, while Nincusa, Vranac, and Tribidrag showed the lowest content. However, Nincusa and Tribidrag had an epicatechin content higher than catechin. In the case of Nincusa, the epicatechin content was almost two-fold higher than catechin. The results reported for Touriga Nacional and Touriga Francesa also showed a higher content of epicatechin than catechin [32]. Among the white varieties, the highest content of catechin was recorded for Cetinka and Grk. An epicatechin content higher than catechin was observed for Posip and Zilavka.

Among the analyzed procyanidins, the most abundant was B2, comprising 15.32% and 18.01% of the total flavan-3-ols in the red and white varieties. Even though the red varieties had a higher content of procyanidin B2, the white varieties had a higher share of this compound in the total content of flavan-3-ols (18.01% for the white and 15.32% for the red varieties). The highest content was recorded for the varieties Plavac mali, Babica and Babic for red, and Posip and Grk for white varieties.



The main constituents of the seed flavan-3-ols in both the red and the white varieties were catechin and epicatechin, while epicatechin gallate occurred in small amounts. This is in accordance with the reported results in Babic, Plavac mali [13], Pinot noir, Teroldego, and Cabernet Sauvignon [33] for the red varieties, and in Chardonnay, Sauvignon Blanc, Pinot Blanc, Riesling, Viognier, and Veltliner [34,35] for the white varieties. Regarding the differences between the red and white varieties, in our study, the red varieties had considerably higher content of all of the flavan-3-ols, which is in accordance with the results reported by Popov et al. [35]. On the other hand, in a study by Montealegre et al. [34] the white varieties had a higher content of flavan-3-ols than the red varieties. However, the author highlighted that the white varieties were less ripe than the red, which can influence the content of polyphenols in seeds. After catechin and epicatechin, the most abundant compound was procyanidin B2, in both the red and white cultivars, followed by procyanidin B4. Our results are in accordance with results reported in Merlot, Cabernet Sauvignon [36,37], Gamay, Chardonnay [37], Ugni Blanc, and Semillon [38].

### 3.3. Prediction of Flavan-3-Ols Yield in Conventional Production

The amount of grape pomace, which includes seeds, generated from winemaking is dependent on the grape cultivar and the pressing process, as well as the fermentation steps [39]. Traditionally, the grape pomace has been used as a fertilizer, animal feed, or to produce distillates. However, over the last decade, other approaches have been proposed for the production and utilization of grape pomace derivatives [40], mainly dietary fibers and polyphenolic compounds. Since there is an increasing demand for healthy and natural food preservation substances that can replace synthetic antioxidants and food preservation substances [41], the grape seed polyphenols are becoming an ingredient in novel quality products with health benefits.

In Table 4, the expected yields of flavan-3-ols are shown, if the grape yield is 10 t/ha. If the grape yield is 10 t/ha, the yield of flavan-3-ols, depending on the variety, would range from 0.18 kg/ha up to 73.98 kg/ha. The highest yield of total flavan-3-ols would come from Lasina, Plavina, and Plavac mali for the red varieties, and M. dubrovacka and Cetinka for the white varieties. Catechin, as the most abundant compound in grape seeds, would range 0.06–25.81 kg/ha. The highest yield would come from the varieties Plavina, Lasina, and Plavac mali. The smallest catechin yield would come from Grk and Gegic. After catechin, the highest yield would be epicatechin, ranging 0.07–25.74 kg/ha. The highest yield would come from the varieties Lasina, Plavina, and Tribidrag. The procyanidins, namely B1, B2, and B4, have a positive impact on human health. They possess antioxidant, anti-inflammatory, anticarcinogenic, and antibacterial activity [3]. The procyanidins make up 56.35% (Gegic) of the total flavan-3-ol yield, while the total procyanidin yield ranged from 0.18 (Grk) to 73.98 (Lasina) kg/ha. The most abundant procyanidin was procyanidin B2, with a yield of 21.59 kg/ha (Lasina), followed by procyanidin B4 with 6.67 kg/ha (Lasina). In general, the red grape varieties had significantly higher procyanidin yields than the white grape varieties. Besides Lasina, among the red varieties, Plavina (59.45 kg/ha) and Plavac mali (37.73 kg/ha) had the highest yield of procyanidins, while among the white varieties, the highest yield was found in M. dubrovacka (26.58 kg/ha), followed by Cetina (24.6 kg/ha), and Posip (16.28 kg/ha).

In the Dalmatia subregion, the area under vineyards is 5 978 hectares, representing 31% of the total vineyard area in Croatia. Moreover, the grape seed extracts' global market value was estimated to be 392.1 million USD in 2015 [42], thus making grape seeds a source of added value.

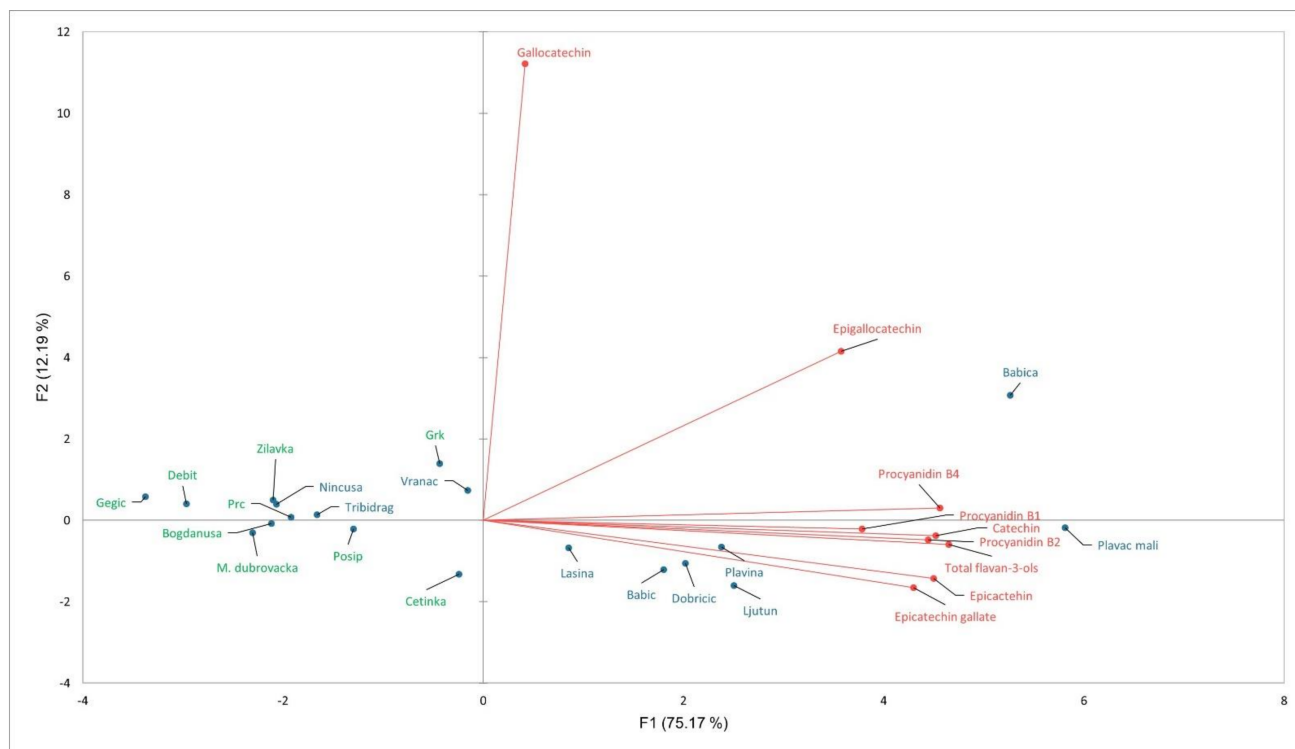
**Table 4.** Prediction of flavan-3-ols yield (kg/ha) when grape yield is 10 t/ha. Means with different letters in the same column differ significantly ( $p \leq 0.05$ ).

[illegible]



### 3.4. PCA Analysis

To evaluate the variability in the seed flavan-3-ols of 20 varieties, principal component analysis (PCA) was performed. The results of the PCA are shown in Figure 2, which represents the distribution of the varieties in the space defined with the first two canonical factors, which explained 87.36% of variability (75.17% for F1 and 12.19% for F2). The Figure also shows the vectors which explain the direction of the variables in the same space.



**Figure 2.** The scatter plot of PCA analysis. Blue shows the red varieties, green shows the white varieties.

Half of the varieties are located near the x axis in the quadrants II and III, and most of these varieties are white varieties. A few of the red varieties are situated near white varieties, which are Nincusa, Tribidrag, and Vranac. When looking at their flavan-3-ol profile, these varieties have a similar profile to the white varieties. This means a low content of total flavan-3-ols comparing to the analyzed red varieties. The rest of the red varieties are situated in the IV quadrant. Two more varieties that are separated from the rest of the red varieties are Plavac mali and Babica. When looking at the vectors, these varieties have a higher content of all of the individual compounds, as well as the total flavan-3-ols. These results confirm the overall profile given in Table 3.

### 4. Conclusions

This research gives insight into the phenolic profiles of the seeds from Croatian grapevine varieties. The flavan-3-ol profiles varied among the varieties, with a higher content recorded for the red varieties. The most abundant compounds were catechin, epicatechin, and procyanidin B2. Based on the analyzed profiles, the potential yield of flavan-3-ols was calculated. Depending on the variety, the potential yield of flavan-3-ols grown in the coastal region could reach up to 73.97 kg/ha. This shows that the usage of grape seeds after vinification could bring an added value to the winemaking industry.

**Author Contributions:** Conceptualization, Ž.A.; methodology, I.T.; formal analysis, I.T. and I.Š.; investigation, Ž.A., D.S. and Z.M.; resources, Ž.A. and E.M.; writing—original draft preparation, I.Š. and Ž.A.; writing—review and editing, D.P. and J.K.K.; supervision, D.P.; funding acquisition, E.M. All authors have read and agreed to the published version of the manuscript.

**Funding:** This research was funded by the European Structural and Investment Funds and the Croatian Ministry of Science, grant number KK.01.1.1.04.0031 project “New Start for Croatian Grapevine Cultivars—CroVitiRestart”.

**Institutional Review Board Statement:** Not applicable.

**Data Availability Statement:** Not applicable.

**Conflicts of Interest:** The authors declare no conflict of interest.

## References

1. Maletic, E.; Karoglan Kontic, J.; Pejic, I.; Preiner, D.; Zdunic, G.; Bubola, M.; Stupic, D.; Andabaka, Z.; Markovic, Z.; Simon, S.; et al. *Green Book; Indigenous Grapevine Varieties of Croatia*; Ministarstvo Zaštite Okoliša i Prirode, Državni Zavod za Zaštitu Prirode: Zagreb, Croatia, 2015.
2. Soares, S.; Brandao, E.; Mateus, N.; de Freitas, V. Sensorial properties of red wine polyphenols: Astringency and bitterness. *Crit. Rev. Food Sci. Nutr.* **2017**, *57*, 937–948. [\[CrossRef\]](#) [\[PubMed\]](#)
3. Šikuten, I.; Štambuk, P.; Andabaka, Ž.; Tomaz, I.; Marković, Z.; Stupić, D.; Maletić, E.; Kontić, J.K.; Preiner, D. Grapevine as a Rich Source of Polyphenolic Compounds. *Molecules* **2020**, *25*, 5604. [\[CrossRef\]](#) [\[PubMed\]](#)
4. Xia, E.Q.; Deng, G.F.; Guo, Y.J.; Li, H.B. Biological Activities of Polyphenols from Grapes. *Int. J. Mol. Sci.* **2010**, *11*, 622–646. [\[CrossRef\]](#) [\[PubMed\]](#)
5. Shi, J.; Yu, J.; Pohoryl, J.E.; Kakuda, Y. Polyphenolics in Grape Seeds—Biochemistry and Functionality. *J. Med. Food* **2003**, *6*, 291–299. [\[CrossRef\]](#)
6. Zerbib, M.; Mazauric, J.P.; Meudec, E.; Le Guerneve, C.; Lepak, A.; Nidetzky, B.; Cheynier, V.; Terrier, N.; Saucier, C. New flavanol O-glycosides in grape and wine. *Food Chem.* **2018**, *266*, 441–448. [\[CrossRef\]](#)
7. Dixon, R.A.; Xie, D.Y.; Sharma, S.B. Proanthocyanidins—A final frontier in flavonoid research? *New Phytol.* **2005**, *165*, 9–28. [\[CrossRef\]](#)
8. Trad, M.; Le Bourvellec, C.; Ben Hamda, H.; Renard, C.; Harbi, M. Flavan-3-ols and procyanidins in grape seeds: Biodiversity and relationships among wild and cultivated vines. *Euphytica* **2017**, *213*, 12. [\[CrossRef\]](#)
9. Burton-Freeman, B.; Brzezinski, M.; Park, E.; Sandhu, A.; Xiao, D.; Edirisinghe, I. A Selective Role of Dietary Anthocyanins and Flavan-3-ols in Reducing the Risk of Type 2 Diabetes Mellitus: A Review of Recent Evidence. *Nutrients* **2019**, *11*, 841. [\[CrossRef\]](#)
10. Campos, E.M.; Jakobs, L.; Simon, M.C. Antidiabetic Effects of Flavan-3-ols and Their Microbial Metabolites. *Nutrients* **2020**, *12*, 1592. [\[CrossRef\]](#)
11. Rodriguez-Perez, C.; Garcia-Villanova, B.; Guerra-Hernandez, E.; Verardo, V. Grape Seeds Proanthocyanidins: An Overview of In Vivo Bioactivity in Animal Models. *Nutrients* **2019**, *11*, 2435. [\[CrossRef\]](#)
12. Rousserie, P.; Rabot, A.; Geny-Denis, L. From Flavanols Biosynthesis to Wine Tannins: What Place for Grape Seeds? *J. Agric. Food Chem.* **2019**, *67*, 1325–1343. [\[CrossRef\]](#)
13. Curko, N.; Ganic, K.K.; Gracin, L.; Dapic, M.; Jourdes, M.; Teissedre, P.L. Characterization of seed and skin polyphenolic extracts of two red grape cultivars grown in Croatia and their sensory perception in a wine model medium. *Food Chem.* **2014**, *145*, 15–22. [\[CrossRef\]](#)
14. Pajović-Šćepanović, R.; Wendelin, S.; Forneck, A.; Eder, R. Suitability of flavan-3-ol analysis to differentiate grapes from Vranac, Kratošija and Cabernet Sauvignon (*Vitis vinifera* L.) grown in Montenegro. *Aust. J. Grape Wine Res.* **2019**, *25*, 376–383. [\[CrossRef\]](#)
15. Maicas, S.; Mateo, J.J. Sustainability of Wine Production. *Sustainability* **2020**, *12*, 559. [\[CrossRef\]](#)
16. Coelho, J.P.; Filipe, R.M.; Robalo, M.P.; Stateva, R.P. Recovering value from organic waste materials: Supercritical fluid extraction of oil from industrial grape seeds. *J. Supercrit. Fluids* **2018**, *141*, 68–77. [\[CrossRef\]](#)
17. Baiano, A. An Overview on Sustainability in the Wine Production Chain. *Beverages* **2021**, *7*, 15. [\[CrossRef\]](#)
18. Dimic, I.; Teslic, N.; Putnik, P.; Kovacevic, D.B.; Zekovic, Z.; Sojic, B.; Mrkonjic, Z.; Colovic, D.; Montesano, D.; Pavlic, B. Innovative and Conventional Valorizations of Grape Seeds from Winery By-Products as Sustainable Source of Lipophilic Antioxidants. *Antioxidants* **2020**, *9*, 568. [\[CrossRef\]](#)
19. Lucarini, M.; Durazzo, A.; Kiefer, J.; Santini, A.; Lombardi-Boccia, G.; Souto, E.B.; Romani, A.; Lampe, A.; Nicoli, S.F.; Gabrielli, P.; et al. Grape Seeds: Chromatographic Profile of Fatty Acids and Phenolic Compounds and Qualitative Analysis by FTIR-ATR Spectroscopy. *Foods* **2020**, *9*, 10. [\[CrossRef\]](#)
20. Amin, R.A.; Edris, S.N. Grape Seed Extract as Natural Antioxidant and Antibacterial in Minced Beef. *PSM Biol. Res.* **2017**, *2*, 89–96.
21. Aybastier, O.; Dawbaa, S.; Demir, C. Investigation of antioxidant ability of grape seeds extract to prevent oxidatively induced DNA damage by gas chromatography-tandem mass spectrometry. *J. Chromatogr. B* **2018**, *1072*, 328–335. [\[CrossRef\]](#)
22. Libera, J.; Latoch, A.; Wojciak, K.M. Utilization of Grape Seed Extract as a Natural Antioxidant in the Technology of Meat Products Inoculated with a Probiotic Strain of LAB. *Foods* **2020**, *9*, 103. [\[CrossRef\]](#)
23. Rajakumari, R.; Volova, T.; Oluwafemi, O.S.; Kumar, S.R.; Thomas, S.; Kalarikkal, N. Grape seed extract-soluplus dispersion and its antioxidant activity. *Drug Dev. Ind. Pharm.* **2020**, *46*, 1219–1229. [\[CrossRef\]](#)

24. Martin, M.E.; Grao-Cruces, E.; Millan-Linares, M.C.; Montserrat-de la Paz, S. Grape (*Vitis vinifera* L.) Seed Oil: A Functional Food from the Winemaking Industry. *Foods* **2020**, *9*, 1360. [\[CrossRef\]](#)
25. Tomaz, I.; Maslov, L.; Stupic, D.; Preiner, D.; Asperger, D.; Kontic, J.K. Solid-liquid Extraction of Phenolics from Red Grape Skins. *Acta Chim. Slov.* **2016**, *63*, 287–297. [\[CrossRef\]](#) [\[PubMed\]](#)
26. Tomaz, I.; Maslov, L. Simultaneous Determination of Phenolic Compounds in Different Matrices using Phenyl-Hexyl Stationary Phase. *Food Anal. Meth.* **2016**, *9*, 401–410. [\[CrossRef\]](#)
27. Preiner, D.; Kontić, J.K.; Šimon, S.; Marković, Z.; Stupić, D.; Maletić, E. Intravarietal Agronomic Variability in Croatian Native *Vitis vinifera* L. Cultivar Grk with Female Flower and Seedless Berries. *Am. J. Enol. Vitic.* **2012**, *63*, 291–295. [\[CrossRef\]](#)
28. McRae, J.M.; Schulkin, A.; Kassara, S.; Holt, H.E.; Smith, P.A. Sensory Properties of Wine Tannin Fractions: Implications for In-Mouth Sensory Properties. *J. Agric. Food Chem.* **2013**, *61*, 719–727. [\[CrossRef\]](#) [\[PubMed\]](#)
29. Watrelot, A.A.; Heymann, H.; Waterhouse, A.L. Red Wine Dryness Perception Related to Physicochemistry. *J. Agric. Food Chem.* **2020**, *68*, 2964–2972. [\[CrossRef\]](#)
30. Unusan, N. Proanthocyanidins in grape seeds: An updated review of their health benefits and potential uses in the food industry. *J. Funct. Food.* **2020**, *67*, 11. [\[CrossRef\]](#)
31. Pantelic, M.M.; Dabic Zagorac, D.; Davidovic, S.M.; Todric, S.R.; Beslic, Z.S.; Gasic, U.M.; Tesic, Z.L.; Natic, M.M. Identification and quantification of phenolic compounds in berry skin, pulp, and seeds in 13 grapevine varieties grown in Serbia. *Food Chem.* **2016**, *211*, 243–252. [\[CrossRef\]](#)
32. Mateus, N.; Marques, S.; Gonçalves, A.C.; Machado, J.M.; De Freitas, V. Proanthocyanidin Composition of Red *Vitis vinifera* Varieties from the Douro Valley during Ripening: Influence of Cultivation Altitude. *Am. J. Enol. Vitic.* **2001**, *52*, 115–121.
33. Mattivi, F.; Vrhovsek, U.; Masuero, D.; Trainotti, D. Differences in the amount and structure of extractable skin and seed tannins amongst red grape varieties. *Aust. J. Grape Wine Res.* **2009**, *15*, 27–35. [\[CrossRef\]](#)
34. Montealegre, R.R.; Peces, R.R.; Vozmediano, J.L.C.; Gascuena, J.M.; Romero, E.G. Phenolic compounds in skins and seeds of ten grape *Vitis vinifera* varieties grown in a warm climate. *J. Food Compos. Anal.* **2006**, *19*, 687–693. [\[CrossRef\]](#)
35. Popov, M.; Hejtmankova, A.; Kotikova, Z.; Stralkova, R.; Lachman, J. Content of flavan-3-ol monomers and gallic acid in grape seeds by variety and year. *Vitis* **2017**, *56*, 45–48. [\[CrossRef\]](#)
36. De Freitas, V.A.P.; Glories, Y.; Monique, A. Developmental changes of procyanidins in grapes of red *Vitis vinifera* varieties and their composition in respective wines. *Am. J. Enol. Vitic.* **2000**, *51*, 397–403.
37. Fuleki, T.; Ricardo-da-Silva, J.M. Catechin and procyanidin composition of seeds from grape cultivars grown in Ontario. *J. Agric. Food Chem.* **1997**, *45*, 1156–1160. [\[CrossRef\]](#)
38. De Freitas, V.A.P.; Glories, Y. Concentration and compositional changes of procyanidins in grape seeds and skin of white *Vitis vinifera* varieties. *J. Sci. Food Agric.* **1999**, *79*, 1601–1606. [\[CrossRef\]](#)
39. Beres, C.; Costa, G.N.S.; Cabezudo, I.; da Silva-James, N.K.; Teles, A.S.C.; Cruz, A.P.G.; Mellinger-Silva, C.; Tonon, R.V.; Cabral, L.M.C.; Freitas, S.P. Towards integral utilization of grape pomace from winemaking process: A review. *Waste Manag.* **2017**, *68*, 581–594. [\[CrossRef\]](#)
40. Bordiga, M.; Travaglia, F.; Locatelli, M. Valorisation of grape pomace: An approach that is increasingly reaching its maturity—A review. *Int. J. Food Sci. Technol.* **2019**, *54*, 933–942. [\[CrossRef\]](#)
41. Antonić, B.; Jančíková, S.; Dordević, D.; Tremlová, B. Grape Pomace Valorization: A Systematic Review and Meta-Analysis. *Foods* **2020**, *9*, 1627. [\[CrossRef\]](#)
42. Zhao, D.; Simon, J.E.; Wu, Q. A critical review on grape polyphenols for neuroprotection: Strategies to enhance bioefficacy. *Crit. Rev. Food Sci. Nutr.* **2020**, *60*, 597–625. [\[CrossRef\]](#)