



Article Relations between Benthic Diatom Community and Characteristics of Karst Ponds in the Alpine Region of Slovenia

Katarina Novak ^{1,2} and Igor Zelnik ^{1,*}

- ¹ Department of Biology, Biotechnical Faculty, University of Ljubljana, Jamnikarjeva 101, 1000 Ljubljana, Slovenia; novakkatarina7@gmail.com
- ² Environmental Agency of the Republic of Slovenia, Vojkova 1b, 1000 Ljubljana, Slovenia
- Correspondence: igor.zelnik@bf.uni-lj.si; Tel.: +386-1-3203339

Abstract: The aim of this research was to investigate the structure of the benthic diatom community and its relations to selected environmental parameters. We collected samples in 16 karst ponds in the alpine region of Slovenia, where the Alpine karst is found. Since the predominating substrate in these ponds was clay, the epipelic community was analyzed. Hydromorphological characteristics, and physical and chemical conditions were also measured at each site. We found 105 species of diatoms, which belonged to 32 genera. The most frequent taxa were *Gomphonema parvulum* (Kützing) Kützing, *Navicula cryptocephala* Kützing, *Sellaphora pupula* (Kützing) Mereschkowsky (species group) and *Achnanthidium pyrenaicum* (Hustedt) Kobayasi. The pond with the lowest diversity was found at the highest altitude, while, on the other hand, the most species-rich pond was found at the lowest altitude. Regarding the ecological types, the most common were motile species. We confirmed a positive correlation between the number of diatom species and the saturation of water with oxygen, while correlation between species richness and NH₄-N was negative. The content of NO₃-N and NH₄-N explained almost 20% of the total variability of diatom community. Unlike our expectations, we calculated a negative correlation between the diversity of macroinvertebrates and diatoms, which is probably a consequence of different responses to environmental conditions.

Keywords: epipelon; diatoms; karst ponds; wetlands; southeast Alps

1. Introduction

Ponds are water bodies ranging from 1 m² to 2 hectares, of natural or anthropogenic origin, with permanent or seasonal water [1]. Researchers used to treat them as lakes, but ponds differ from lakes due to several characteristics [2]: (a) smaller surface area and depth, (b) smaller ratio between the volume of water and the shore area, and therefore more direct contact with the terrestrial environment making them more susceptible to various influences; (c) smaller drainage basin and therefore bigger isolation [1]; (d) relatively small volume and water intake, which increases the connection between the sediments and water column and a more significant impact of sediment on the nutrient content in water, (e) due to the low water depth, the surface of the entire waterbody could be covered with macrophytes [3,4]. This is also the main reason why we consider ponds as a type of wetland. It is characteristic that their conditions change faster than in larger water bodies [5], which is reflected in large daily and seasonal fluctuations [1,5].

Ponds as a habitat have been neglected in ecological studies [6]. Today, we recognize them as an important carbon sink, pollution filter, and source of biodiversity, hosting several specialized and rare species [2,6]. For organisms living in the aquatic environment, ponds are refuges in degraded and inhospitable areas [1,7].

Karst ponds were made in areas with no surface water bodies (e.g., Karst), where people had problems with water supply [8]. Although they were used to water livestock and gardens, they lost their importance when water pipelines were constructed. However, today they represent an important source of biodiversity, like all other types of



Citation: Novak, K.; Zelnik, I. Relations between Benthic Diatom Community and Characteristics of Karst Ponds in the Alpine Region of Slovenia. *Diversity* **2021**, *13*, 531. https://doi.org/10.3390/d13110531

Academic Editor: Michael Wink

Received: 28 September 2021 Accepted: 21 October 2021 Published: 25 October 2021

Publisher's Note: MDPI stays neutral with regard to jurisdictional claims in published maps and institutional affiliations.



Copyright: © 2021 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (https:// creativecommons.org/licenses/by/ 4.0/). ponds [1,7,9,10]. Smol and Stoermer [11] suggest that Karstic aquatic habitats are the most interesting environments in which to study algae, especially diatoms.

With their distribution, they form a network of aquatic ecosystems, which increases γ diversity [1,8]. Biodiversity and abundance of the biota in Alpine ponds significantly correlate with altitude—with it, the average air temperature decreases, the amount of local precipitation increases, and UV radiation is more intense. In addition, the organisms in these environments face high daily and annual temperature differences and have a short period suitable for growth, which gives cold stenothermic species a better chance of survival [12–14].

The substrate consisting of clay and silt mostly covers the entire bottom of these ponds. On such a substrate, an epipelic biofilm develops, which is dominated by diatoms constituting the basal trophic levels for extensive food webs [15]. Diatoms are present in different aquatic environments and their sensitivity to various environmental factors, makes them a good bioindicator of water quality [16]. Recent studies have highlighted the high level of cryptic diversity of diatoms [17]. The diatom community is influenced by several factors such as water chemistry (pH, nutrient concentration, and organic load), physical (electrical conductivity, temperature, light) hydromorphological characteristics (substrate, water regime), and biotic pressures such as grazing, competition, and parasitism [17–20].

Benthic diatoms are important primary producers in shallow waters where light penetrates to the bottom [21]. On a fine substrate, a specific epipelic diatom community usually forms, which is adapted to low light conditions, consisting mainly of motile taxa that can move through interstitial waters to avoid newly deposited sediments [22]. Due to their location between substrate and water, they play a fundamental role in various biogeochemical cycles and dynamics of aquatic ecosystems [23].

The biological characteristics of diatoms, such as cell size class and ecological types, give us information about the structure of the community [17,24], as well as environmental conditions. Low-profile diatoms are well adapted to physical disturbances and are more abundant in waters with low nutrient content [17,24,25]. For high-profile diatoms, the formation of colonies allows exploiting nutrients that are not available to other groups but are therefore more exposed to grazing [24,25]. Motile diatoms are fast-growing species. Their abundance increases with a higher concentration of nutrients and organic load. They are also well adapted to high physical disorders [24]. Planktic species are present in lentic water, where they float in the water column [25], but due to sinking they can also be abundant in phytobenthos [26].

Despite their import roles, karst ponds are disappearing due to the abandonment of their original use. In addition to natural processes such as overgrowing with plants, they are also threatened by anthropogenic factors, especially intensification of agriculture, abandonment of livestock farming, backfilling, the input of non-native species and chemical pollution [1–3]. Pollutants cannot be sufficiently diluted [27], and nutrients are retained and potentially recycled by internal processes, which is difficult in the affected ecosystem [4]. All this can be significantly reflected in the structure of the diatom community.

However, we have not found any published work on the epipelic diatom community in karst ponds. Even the studies of periphytic diatom communities in ponds are rare, which had been discovered by Šumberova et al. [28]. In central Europe, we have found one paper about epipelic diatoms in ponds [29], while in southern Europe there are some papers that analyze epipelic diatoms (e.g., [16,30–33]).

We measured physical, hydromorphological, and chemical factors in 16 ponds at various locations in the Alpine region and sampled the epipelon. In this paper, we focused primarily on their response to various environmental characteristics. The study aimed to determine the species composition of the benthic diatom community in the Alpine karst ponds, determine the relationships between the structure of diatom community and the studied parameters, and find out the significant correlation between them.

We hypothesized that: (a) the diatom's species diversity correlates with the diversity of macroinvertebrates; (b) the diversity of species will decline with altitude and declining

of ponds size; (c) the species composition will be significantly affected by the pH and electrical conductivity of the water and the land use in the drainage basin.

2. Materials and Methods

2.1. Study Sites and Sampling

We chose 16 karst ponds in the alpine region of Slovenia, which is a part of the South— Eastern calcareous Alps. Since limestone and dolomite are predominating rocks in this area, the Alpine karst is found there [34]. These water bodies are found in the area of the Julian Alps (Pokljuka, Jelovica, Ratitovec) and the Kamnik Alps (Krvavec, Velika planina, and Menina) (Figure 1). During the sample preparation we realized, that there were almost no frustules in samples from four ponds.





Mountain climate prevails in the area, where the average temperature of the coldest month is lower than -3 °C, and the average temperature of the warmest month depends on the altitude and location [35]. Macrophyte and macroinvertebrate communities were studied before in the same ponds and results were published in Zelnik et al. [10].

Sampling took place in August of 2016, during the peak pasture season. Argilal and clay, respectively, was the only type of substrate present in all sites, so we decided to sample epipelon. Since we experienced difficulties with cleaning the samples from four ponds as well as very poor presence of diatom frustules in them, the samples from 12 sites were studied only (Table 1).

Basic physical and chemical factors were measured with a portable multimeter (EU-TECH, PCD 650). For each pond, we measured the pH and T of water (°C), electrical conductivity (μ S/cm), total dissolved solids (mg/L), saturation with O₂ (%), and O₂ concentration (mg/L). For laboratory analyzes, a water sample (1 L) was taken at each site.

In the laboratory, the concentrations of NO_3 -N (LCK 339), NH₄-N (LCK 304), TN (LCK 138), and orthophosphates (LCK 349) were determined using HACH Lange cuvette tests. Values were measured in individual samples with a HACH Lange LT 200 spectropho-

tometer. Dry mass and total suspended solids content (TSS) were determined by filtration and drying at 105 $^{\circ}$ C.

	Kanal Dan 1	Altitude	Gauß-Krüge	r Coordinates	Precipitation per Year	
Code	Karst Pond	[m]	Y	X	[mm]	
POK1	Pokljuka 1	1201	425202	134889	2200	
POK2	Pokljuka 2	1302	424023	133737	2200	
JEL1	Jelovica 1	1129	431399	125787	1900	
JEL2	Jelovica 2	1138	430695	127923	1900	
KRV1	Krvavec 1	1724	464378	128300	1650	
	Krvavec 2	1509	463564	128355	1600	
KRV3	Krvavec 3	1445	464227	127589	1600	
RAT1	Ratitovec 1	1577	430192	122130	2100	
RAT2	Ratitovec 2	1620	430104	121849	2100	
	Velika planina 1	1434	475035	128689	1700	
	Velika planina 2	1481	474750	128275	1700	
VEL3	Velika planina 3	1454	474958	128408	1700	
MEN1	Menina 1	1318	488084	122280	1250	
MEN2	Menina 2	1403	487335	123639	1500	
	Menina 3	1360	487473	123194	1500	
MEN4	Menina 4	1419	487053	123695	1500	

Table 1. Information about sampling sites.

2.2. Biotic Analyses

Due to the absence of a firm substrate, diatom samples were taken from the surface of the loamy substrate. We scraped the top layer of argilal with an area of approximately 2 cm^2 , with a spoon, at a 20–25 cm water depth. The samples were placed into bottles and 37% formaldehyde was added for fixation, in a ratio of 1:9.

Each sample was first homogenized with magnetic stirrer at a rate of 1200 rpm. We put 2 mL of the sample into a test tube and added 2.5 mL of 65% nitric (V) acid (HNO₃). The samples were heated over a fire until the smoke turned white to remove organic matter from the sample. After cooling the tube contents were centrifuged with a SIGMA 2-16PK centrifuge, 4 min at 4000 rpm, and the supernatant was discarded. The sample was further washed with distilled water. The resulting pellet was added to 2 mL of distilled water and mixed. We put single drops onto slides, dried them, and fixed them with Naphrax[®] mountant.

The prepared preparations were examined with an Olympus CX41 microscope under $1000 \times$ magnification, and the first 400 frustules of each sample were determined. Identification was performed using the keys of Hoffman et al. [36], Lange-Bertalot et al. [37], and in some cases Krammer and Lange-Bertalot [38–41].

2.3. Data Analysis

Correlation analysis was performed with PAST program [42]. Some data (land use, number of habitat types, turbidity) were of the interval type and thus not normally distributed, so we used Kendall correlation coefficients (tau).

Similarity in taxonomic composition of diatom community between the ponds was calculated using Sørensen similarity index. Diversity was calculated as Shannon-Wiener diversity index (S-WI) and Margalef diversity index. The trophic index (TI) was calculated according to Rott et al. [43].

The influence of individual factors on the composition of the diatom community was checked by direct gradient analyzes. First, we performed a detrended correspondence analysis (DCA) to determine whether the distribution of the diatom species along potential gradients is unimodal or linear. We found that the mentioned distribution was unimodal (Length of gradient: 9.7 S.D.), so we used Canonical Correspondence Analysis (CCA). All analyzes were performed with the Canoco 4.5 software package [44].

Environmental parameters were grouped into spatial variables (coordinates, altitude, annual precipitation, a distance from the next pond or road), substrate (inorganic and organic), chemical and physical variables, hydromorphological data, drainage basin etc.

We used the method of forward selection to check the effect of individual environmental factors on the taxonomic composition. The program made 999 permutations in each round, three rounds were performed. In each next round, we considered only factors with p less than 0.1. In the last round, we considered the two most statistically significant factors, that were in fact marginally significant (p = 0.06 and 0.07). Based on two factors that had a marginally statistically significant effect on the structure of the diatom community, we also created an ordination diagram in which the ponds are distributed along gradients of environmental factors.

3. Results

3.1. Structure of the Benthic Diatom Community

A total of 105 species of diatoms were identified in 12 ponds (Table A1). Of these, most species-rich was JEL1 (43 species) and POK1 (30 species) (Figure 2). The pond with the lowest number of species was KRV1 (14 species). Dominant species and their proportions vary significantly between ponds (Table 2). *Navicula cryptocephala* Kützing was the most dominant in four ponds (POK1, JEL2 MEN2 and MEN4), and it was also present in a large proportion in RAT1. The pioneer complex *Achnanthidium minutissimum* (Kützing) Czarnecki was the most common taxon in three ponds (JEL1, RAT2 and MEN2). The highest dominance index is in POK2 and KRV1, where two dominant taxa represent 77% of the identified species (Table 2).



Figure 2. Number of diatom species in individual karst ponds.

Table 2. Dominance index (proportion in %) of the two most common species (highlighted in gray) in studied ponds. Diatoms that are not dominant in the sample but have a proportion $\geq 10\%$ are also shown.

Species	POK1	POK2	JEL1	JEL2	KRV1	KRV3	RAT1	RAT2	VEL3	MEN1	MEN2	MEN4
Achnanthidium minutissimum			13					24			35	
Achnanthidium pyrenaicum				13				38			18	
Craticula accomoda						16						
Eucoconeis alpestris										10		
Eunotia bilunaris										19		
Eunotia tenella		41										
Gomphonema angustum									10	13		

Species	POK1	POK2	JEL1	JEL2	KRV1	KRV3	RAT1	RAT2	VEL3	MEN1	MEN2	MEN4
Gomphonema parvulum				11					13			19
Navicula cryptocephala	26			40			14				19	14
Navicula exilis							45					
Nitzschia acicularis					53							
Nitzschia adamata						16						
Nitzschia palea												
Nitzschia perminuta										16		
Nitzschia supralitorea					19							
Pinnularia interrupta		36										
Sellaphora pseudopupula				10								
Sellaphora pupula	28						17					10
Tabellaria flocculosa			15									
Dominance index	53.3	77.3	27.5	52.5	71.5	32.4	61.3	62.7	23.1	34.9	53	32.6

Table 2. Cont.

Ponds with the highest similarity of diatom community are POK1 and VEL3, although the huge distance between them (see Figure 1). On the other side there was POK2, which stood out the most in rare species—with four ponds (KRV1, KRV3, MEN2, and MEN4) had no species in common (Table 3).

Table 3. Similarity of diatom community between the studied ponds according to Sørenson index. The similarity indices >0.5 are in bold.

POK1	POK2	JEL1	JEL2	KRV1	KRV3	RAT1	RAT2	VEL3	MEN1	MEN2	MEN4	
	0.29	0.44	0.46	0.23	0.30	0.26	0.40	0.62	0.43	0.47	0.44	POK1
		0.13	0.11	0	0	0.06	0.05	0.23	0.18	0	0	POK2
			0.33	0.25	0.27	0.24	0.41	0.44	0.32	0.40	0.31	JEL1
				0.31	0.49	0.29	0.56	0.37	0.27	0.46	0.29	JEL2
					0.22	0.27	0.26	0.15	0.20	0.39	0.33	KRV1
						0.26	0.38	0.29	0.25	0.40	0.31	KRV3
							0.39	0.34	0.19	0.30	0.31	RAT1
								0.36	0.28	0.52	0.39	RAT2
									0.55	0.52	0.44	VEL3
										0.37	0.48	MEN1
											0.55	MEN2

Figure 3 shows the proportion of diatoms according to their ecological type. Motile and high-profile diatoms are present in all samples. Low-profile diatoms are absent in one pond, while in four ponds (KRV3, RAT1, MEN1, and MEN4) they are very rare. Their largest proportion is in RAT2 (68%) and MEN2 (52%). Planktic diatoms are present with a negligible proportion (JEL1, KRV3, and RAT1), except for KRV1, representing half of the specimens. The most common are motile diatoms. In POK1, JEL2, KRV3, RAT1, VEL3 and MEN4, they represent the majority proportion of diatoms.

Figure 4 shows the size classes of diatoms. The most common size class is 3, followed by 2 and 4. Members of size classes 1 and 5 are infrequent. Smaller diatoms (size classes 1 and 2) are dominant in RAT2, POK2, and MEN2. Data for POK2 are not representative, as 77% of specimens were not determined a size class due to lack of data in the literature. There is also a considerable proportion of unknown size classes in KRV1 and KRV3 (22% and 28%).

3.2. Effects of Environmental Factors on the Diatom Community Composition

The concentration of NO₃-N and NH₄-N in water explains almost 20% of the total variability of the diatom community in ponds (Table 4). The concentration of NO₃-N explains 10% of the variability, and the NH₄-N concentration in water 9.6%. The content of these two nutrients or nitrogen species is probably mainly due to the higher load in ponds

and their basin area with livestock. The same shows the ordination diagram based on CCA (Figure 5), where ponds are arranged according to the diatom taxonomic composition along the gradients of NO₃-N and NH₄-N concentration in water.



Figure 3. Diatoms according to their ecological type [in %]. (PL—planktic, H-P—high-profile, L-P—low-profile).



Figure 4. Diatoms according to their size class [%].

Table 4. Results of Canonical correspondence analysis (CCA) and forward selection. (% TVE-proportion of the explained variability by specific variable).

Variable	Р	% TVE
NO ₃ -N	0.064	10.0
NH_4 -N	0.072	9.6



Figure 5. A CCA-based ordination diagram in which karst ponds are distributed along environmental gradients.

According to the S-WI index (Figure 6), the highest diversity is in JEL1, VEL 3 is next. The lowest diversity is in POK2, the lower diversity is also in KRV1 and RAT1. The Margalef index (Figure 7) showed a different assessment of diversity than S-WI.



Figure 6. Shannon-Wiener diversity index values of diatoms in karst ponds.

JEL1 still has the highest diversity value (7.01), but the ponds with the lowest diversity are RAT1 and MEN4.

3.3. Environmental Factors and Diversity of Diatom Community

Kendall correlation coefficients showed that the number of diatom species is in a statistically significant positive correlation with oxygen saturation and a negative correlation with the concentration of NH₄-N (Table 5). The Margalef index was also positively correlated with oxygen saturation and negatively with NH₄-N concentration. A negative statistically significant correlation (p = 0.05) was calculated between altitude and the Margalef index.



Figure 7. Margalef index values of diatoms in karst ponds.

Table 5. Kendall (tau) correlation coefficients between environmental factors and diversity parameters of diatom communities in ponds. Only statistically significant correlations (*—p < 0.05) and marginally statistically significant correlations (p = 0.05) are shown.

	No. of Diatom Species	Margalef Index
altitude [m]	n.s.	-0.431
O ₂ saturation [%]	0.531 *	0.543 *
NH_4 -N [mg/L]	-0.481 *	-0.492 *
SW_I h.taxa macoinvertebrates	-0.481 *	-0.492 *

We also found a negative correlation between the number of diatom species and S-WI and the Margalef index calculated based on the composition of the invertebrate community, which was contrary to our expectations.

Great differences in TI values were found between the ponds (Figure 8). The lowest TI value was in POK1 (ultraoligotrophic) and the highest in JEL2, KRV3, KRV1, POK1, and MEN4 (polytrophic).



Figure 8. Trophic index values for sampled karst ponds.

4. Discussion

4.1. Structure of the Benthic Diatom Community

In total, 105 diatom species belonging to 32 genera were identified. The most common taxa were *Gomphonema parvulum* (Kützing) Kützing, *Navicula cryptocephala* Kützing, species group *Sellaphora pupula* (Kützing) Mereschkowsky (present in 10 sites). Almost half of the species (52) were present in only one site, from which we can assume that the composition of diatom communities differs much between the ponds. The genera with the highest number of species were *Nitzschia, Pinnularia, Navicula* and *Neidium*. The highest number of species was identified in the JEL1, whereas in KRV1, we found the lowest number of species, of which *Nitzschia acicularis* (Kützing) W. Smith represented more than half of the identified frustules. We expected lower diversity as well as variability of epipelic diatom community, as karst ponds are small water bodies with frequent disturbances, which make the conditions unfavorable. The number of species varied from 14 to 43, which is much higher than 11–26 taxa from ponds in South-eastern Alps reported by Cantonati et al. [29]. However, the mentioned researchers studied different type of ponds in alpine region.

Among the ecological types, the motile diatoms were the most common. They dominated in four ponds (POK1, KRV3, RAT1, and MEN4) and were codominant in another four ponds (Figure 2). Sites where deposition occurs are advantageous for motile diatoms [45–47] as well as nutrient-rich sites [48–50]. Typical representatives from genera *Navicula*, *Nitzschia*, *Sellaphora*, and *Surirella* [24] were also present in our samples. However, we did not calculate any significant correlation between environmental factors and the share of motile species. In ponds with higher trophic index values motile species dominated, which are well adapted to higher nutrient content. We expected that high-profile (H-P) diatoms would also be present here with higher proportion. However, they were probably not present in such high proportion due to physical disturbances.

High-profile diatoms, which are also common in nutrient-rich water but with fewer disturbances [48] are less common in our samples. The proportion of H-P negatively correlated with TSS (p = 0.009), which negatively influence light conditions with turbidity and deposition. On the other hand, we calculated positive correlation between proportions of H-P diatoms and argilal (p = 0.029). The typical genera of this group, which were also present in our samples, were *Eunotia*, *Fragilaria* and *Gomphonema*. The proportions of H-P diatoms were lower than motile, except POK2, where H-P represent two-thirds of the community. Disturbances and grazing, made motile species more efficient than H-P ones.

Low-profile (L-P) diatoms were rare, but in two samples (RAT2 and MEN2) they were dominant. Both ponds are fenced, so with no access of the cattle. Proportions of L-P diatoms negatively correlated with NO₃-N (p = 0.023) and positively with habitat diversity in the catchment area (p = 0.039), which actually means low density of the cattle. Typical representatives are from the genera *Achnantes, Achnanthidium, Amphora, Cocconeis,* and *Meridion* [24]. *Achnantidium minutissiumum* (Kützing) Czarnecki was the most dominant taxon in RAT2 and MEN2, as well as in JEL1. It seems that cattle cause problem for L-P diatoms due to high input of nutrients to ponds, to which L-P species are not adapted [48]. In some samples (JEL1, KRV1, KRV3, and RAT1), planktic diatoms were also present.

In ponds with higher concentrations of orthophosphates, we find mainly motile and H-P diatoms adapted to higher concentrations of nutrients [24,51,52] (Figure 3). In POK2 (0.3 mg/L of ortophosphate), MEN1 (0.92 mg/L) and VEL3 (0.23 mg/L) motile and H-P diatoms represent almost the entire sample, L-P diatoms are almost absent. However, the significant correlation between P and ecological types was not calculated. There was also no correlation between P concentration and diatom size classes, which also report Lavoie et al. [53].

The concentration of NO₃-N and NH₄-N in water explained almost 20% of the total variability of diatom community (Table 4). The concentration of NO₃-N explains 10% of the variability of the diatom community, and the concentration of NH₄-N 9.6% (Table 4, Figure 5). The ponds are arranged according to the taxonomic composition of diatom communities along the gradients of NO₃-N and NH₄-N concentration in water.

The results did not show statistically significant correlations between the composition of diatom community and concentrations of either orthophosphate or TP as expected, which is consistent with Soininen et al. [54]. This is probably because absorption rate for phosphorus from the water column by epipelon is lower than in other groups of primary producers [55].

Haubois et al. [56] report that large and small species do coexist within the epipelon. We found that size-class three had the highest proportion in five ponds, while size-class 2 and 4 in three ponds each (Figure 4). However, most of the identified frustules belonged to the middle-size class (3), which also report Lavoie et al. [53]. In ponds with higher biodiversity (JEL1, KRV3, VEL3, MEN1, and MEN4), size-classes 4 and 3 dominated.

4.2. Diversity of Benthic Diatom Community and Environmental Factors

In general, altitude affects biota in ponds as it affects temperature, precipitation, and radiation [12]. The results showed a negative correlation between altitude and the Margalef index, which is in line with our hypothesis and with the general rules in ecology [57]. The diatom species richness did not correlate with altitude, but pond at the highest altitude (KRV1) had the lowest number of species, while pond at the lowest altitude (JEL1) had the highest diversity. On the contrary for mountain ponds in Spain Blanco et al. [31] report positive correlation of diatom diversity with altitude.

The water depth in these shallow ponds is important mainly because of poor light conditions in turbid water. One of the dominant species was also *Nitzschia perminuta* (Grunow) M. Peragallo, which dominates in low light conditions [58]. Due to shallowness, there is no stratification during the summer [59].

We calculated no significant correlation between pH and diversity indices. The most extreme values were measured at POK2 (pH = 3.8) and MEN1 (pH = 9.6) (Table A2). The first is located in a coniferous forest and is a dystrophic system. Therefore, diatom species in this pond differed from others the most (Table 3). As reported in DeNicola [60] and Della Bella [16], we found there mainly species from the genera *Neidium, Eunotia, Pinnularia, Stauroneis,* and *Sellaphora,* which occurred in small numbers or were absent in other ponds. Diatom community from this pond had no species in common with four other ponds. This pond was more similar to the shallow ponds on mires presented in [29,61]. The lowest value of the electrical conductivity was also measured there (16 μ S/cm), which coincides with the trophic index, which defines it as ultraoligotrophic.

We found a positive correlation between the number of diatom species and water saturation with oxygen and the Margalef index and water saturation with oxygen. The highest oxygen saturation was in MEN1 (almost 250%) due to intense photosynthetic activity of the phytoplankton, making the water very turbid.

In KRV1 and MEN4, a large proportion of N is in the form of NH₄-N, which can be explained by the high density of cattle in their catchments. Correlation coefficients showed a negative correlation between the Margalef index and the NH₄-N concentration. In ponds with a higher concentration (KRV1 and MEN4), the diversity was lower, while it was higher in ponds with lower NH₄-N concentrations (POK1, JEL1, RAT1, and VEL3). In contrast to NH₄-N concentrations, NO₃-N concentrations did not differ much between ponds. Values were 0.2–0.5 mg/L. NO₃-N and NH₄-N concentrations classify our ponds as eutrophic (POK2, JEL2, KRV3, RAT1), mesotrophic (POK1 and MEN1), or oligotrophic (JEL1, RAT2, and MEN2) [54]. In KRV1 and MEN4, the values of NH₄-H and NO₃-N were so high that they can be classified as hypereutrophic.

Cattle can have a substantial negative effect on the diversity of communities in ponds [62]. Trampling the bottom and the shore presents physical disturbances. In ponds with moderate intensity of trampling, the diatom diversity was higher than in those without trampling, which is consistent with the intermediate-disturbance hypothesis [63]. More important is the influence of the cattle as the source of nutrients and organic matter from their excrements. Smaller water bodies in the agricultural landscape are highly exposed to

influences from nearby agricultural areas, since they can be strongly affected by nutrient accumulation [4].

Based on the trophic index (TI), ponds vary from ultraoligotrophic to polytrophic. Della Bella et al. [30] report that trophic diatom index highly correlated with nutrient content, especially orthophosphate and NO₃-N in wetlands in central Italy. However, in our case orthophosphate concentrations were the highest where the TI values were low (POK2 and MEN1). According to TP concentrations and nutrient estimates for lakes [58], both ponds were hypertrophic, but TI classified them as ultraoligotrophic (POK2) and mesotrophic (MEN1). Due to the pH = 3.8, there were probably not enough basic ions in POK2, despite the high concentration of TP and NO₃⁻. Insufficient amount of HCO₃⁻ was present at pH = 9.6, which reduced primary production and thus nutrient uptake, which was probably the explanation for the condition in the MEN1.

4.3. Correlations between Diatoms and Macroinvertebrates

We found a negative correlation between the diatom species richness and the S-WI, and Margalef index calculated on the base of the macroinvertebrate community, which was contrary to our expectations. Similar findings report also Gascón et al. [64], which found out that different aquatic communities respond differently to the environmental factors, so we could not generalize relations between parameters and diversity patterns. Due to the larger size of macroinvertebrates, they might be more susceptible to physical destruction of the littoral zone, and loss of mesohabitats due to trampling of the bottom compared to diatoms, whereas diatoms, as primary producers, are particularly sensitive to water chemistry and light conditions [16,65]. Another reason is probably grazing [66]. We should not neglect the fact that on the same substrate on which diatoms thrive, Chironomidae dominate, which graze on epipelon.

5. Conclusions

We found a negative correlation between species-richness and diversity of the diatom community and diversity of the macroinvertebrate community (S-WI, Margalef index).

Despite relatively small differences in altitude, the results showed a marginal statistical correlation between altitude and Margalef Index. No effect of the pond size on the diversity of diatom community was observed.

We did not calculate significant correlations between pH and diversity. Half of the species in most acidic pond POK2 were present only in this pond. Correlations between electrical conductivity, land use, and diversity of diatom community were not significant.

Motile diatoms were most common. They are adapted to high nutrient concentrations and disturbances and can migrate to the site with sufficient light or nutrients when the re-suspended substrate is depositing.

We found a positive correlation between the number of diatom species and O_2 saturation and the Margalef index and O_2 saturation. The pond with the lowest oxygen saturation value (KRV1) had the lowest species diversity.

The results also showed a negative correlation between the number of diatoms and NH_4 -N concentration and the Margalef index and NH_4 -N concentration. NH_4 -N is probably present in the ponds due to the cattle grazing in the area in the summer. The concentrations of NO_3 -N and NH_4 -N explain almost 20% of the total variability of the diatom community.

Author Contributions: Conceptualization, I.Z.; methodology, I.Z. and K.N.; validation, K.N.; formal analysis, I.Z. and K.N.; investigation, K.N.; data curation, I.Z. and K.N.; writing—original draft preparation, K.N.; writing—review and editing, I.Z. and K.N.; visualization, I.Z. and K.N.; supervision, I.Z.; funding acquisition, I.Z. All authors have read and agreed to the published version of the manuscript.

Funding: This research and the APC were partly funded by the Slovenian Research Agency, Research program Biology of plants, grant number P1-0212.

Institutional Review Board Statement: Not applicable.

Informed Consent Statement: Not applicable.

Data Availability Statement: Data are stored within the documentation of Master Study program theses and P1-0212 Research program.

Acknowledgments: Authors thank to Matej Holcar for creation of the Figure 1.

Conflicts of Interest: The authors declare no conflict of interest.

Appendix A

Table A1. The list of the names of diatom taxa found in studied karst ponds.

Achnanthidium minutissimum (Kützing) Czarnecki Adlafa minuscula (Grunow) Lange-Bertalot var. minuscula Amphora copulata (Kützing) Schoeman et Archibald Amphora pediculus (Kützing) Grunow Brachysira neoscilis Lange-Bertalot Caloneis tenuis (Gregory) Krammer Chamaepinnularia mediocris (Krasske) Lange-Bertalot Chamaepinnularia muscicola (Petersen) Kulikovskiy, Lange-Bertalot et Witkowski Chamaepinnularia soehrensis (Krasske) Lange-Bertalot et Krammer Cocconeis pediculus Ehrenberg Craticula accomoda (Hustedt) D.G. Mann Craticula ambigua (Ehrenberg) D.G. Mann Craticula ambigua (Ehrenberg) D.G. Mann Craticula molestiformis (Hustedt) Lange-Bertalot Cyclotella stelligera Cleve & Grunow Cymbopleura amphicephala (Nägeli) Krammer Diploneis krammer Lange-Bertalot Europonema hebridicum Grunow x Cleve Encyonema himutum (Hilse) D.G. Mann Eucoconeis alpestris (Brustedt) D.G. Mann Eucoconeis alpestris (Bruno x Cleve Encyonema nimutum (Hilse) D.G. Mann Eucoconeis alpestris (Brun) Lange-Bertalot Eunotia arcus Ehrenberg Eunotia bilumaris (Ehrenberg) Schaarschmidt Eunotia ailesiacum (Bleisch) D.G. Mann Eucoconeis alpestris (Brun) Lange-Bertalot Eunotia arcus Ehrenberg Eunotia bilunaris (Ehrenberg) Schaarschmidt Eunotia pseudogroenlandica Lange-Bertalot Eunotia pseudogroenlandica Lange-Bertalot Eunotia arcus Ehrenberg Eunotia pseudogroenlandica Lange-Bertalot Eunotia arcus (Kützing) Grunow Eunotia pseudogroenlandica Lange-Bertalot Fragilaria radians (Kützing) Williams et Round Fragilaria radians (Kützing) Milams et Round Fragilaria radians (Kützing) Milams et Round Fragilaria radians (Kützing) Kabenhorst Gomphonema angustum (Kützing) Rabenhorst Gomphonema angustum (Kützing) Rabenhorst Gomphonema angustum (Kützing) Kabenhorst Gomphonema angustum (Kützi	Achnanthidium pyrenaicum (Hustedt) Kobayasi
Adlafia minuscula (Grunow) Lange-Bertalot var. minuscula Amphora copulata (Kützing) Schoeman et Archibald Amphora pediculus (Kützing) Grunow Brachysira neoexilis Lange-Bertalot Caloneis tenuis (Gregory) Krammer Chamaepinnularia mediocris (Krasske) Lange-Bertalot et Witkowski Chamaepinnularia soehrensis (Krasske) Lange-Bertalot et Witkowski Chamaepinnularia soehrensis (Krasske) Lange-Bertalot et Witkowski Chamaepinnularia soehrensis (Krasske) Lange-Bertalot et Krammer Cocconeis pediculus Ehrenberg Craticula accomoda (Hustedt) D.G. Mann Craticula anbigua (Ehrenberg) D.G. Mann Craticula anoestiformis (Hustedt) Lange-Bertalot Cyclotella stelligera Cleve & Grunow Cymbopleura ampticephala (Nägeli) Krammer Cymbopleura ampticephala (Nägeli) Krammer Diploneis krammeri Lange-Bertalot et Reichardt Encyonema hebridicum Grunow ex Cleve Encyonema silesiacum (Beisch) D.G. Mann Eucoconeis alpestris (Brun) Lange-Bertalot Eunotia arcus Ehrenberg Eunotia bilunaris (Ehrenberg) Schaarschmidt Eunotia arcus Glurow ex Cleve Eunotia paludosa Grunow Eunotia pseudogroenlandica Lange-Bertalot et Tagliaventi Eunotia pludosa Grunow Eunotia pseudogroenlandica Lange-Bertalot et Tagliaventi Eunotia pseudogroenlandica Lange-Bertalot et Tagliaventi Eunotia ariaus (Kützing) Grunow Eunotia pseudogroenlandica Lange-Bertalot et Tagliaventi Eunotia aradis (Kützing) Williams et Round Fragilaria radians (Kützing) Williams et Round Fragilaria radians (Kützing) Nilams et Round Fragilaria radians (Kützing) Nilams et Round Fragilaria radians (Kützing) Kutzing Rabenhorst Gomphonema acuminatum Ehrenberg Gomphonema acuminatum Ehrenberg Gomphonema acustum (Kützing) Rabenhorst Gomphonema acustum (Kützing) Rabenhorst Gomphonema acustum (Kützing) Rabenhorst Gomphonema acustum (Kützing) Katzing Gomphonema acustum (Kützing) Katzing Gomphonema acustum (Kützing) Katzing Gomphonema acustum (Kützing) Katzing Gomphonema acustum (Kützing) Kützing Gomphonema acustum (Kützing) Kützing Gomphonema acustum (Kützing) Lange-Bertalot Luticola mutica (Kützing) D.G. Mann Meridion	Achnanthidium minutissimum (Kützing) Czarnecki
Amphora copulata (Kützing) Schoeman et Archibald Amphora pediculus (Kützing) Grunow Brachysira neoexilis Lange-Bertalot Caloneis tenuis (Gregory) Krammer Chamaepinnularia mediocris (Krasske) Lange-Bertalot et Witkowski Chamaepinnularia sochrensis (Krasske) Lange-Bertalot et Witkowski Chamaepinnularia sochrensis (Krasske) Lange-Bertalot et Krammer Cocconeis pediculus Ehrenberg Craticula accomoda (Hustedt) D.G. Mann Craticula anbigua (Ehrenberg) D.G. Mann Craticula anore (Gunow) D.G. Mann Craticula nolesitjormis (Hustedt) Lange-Bertalot Cyclotella stelligera Cleve & Grunow Cymbopleura amphicephala (Nägeli) Krammer Diploneis krammeri Lange-Bertalot et Reichardt Encyonema hebridicum Grunow ex Cleve Encyonema nimutum (Hilse) D.G. Mann Encyonema niesiacum (Bleisch) D.G. Mann Encyonema niesiacum (Bleisch) D.G. Mann Encotia scipacu (Brebisson) Rabenhorst Eunotia arcus Ehrenberg Eunotia arcus Ehrenberg Eunotia pseudogroenlandica Lange-Bertalot et Tagliaventi Eunotia aninor (Kützing) Grunow Eunotia subarcuatoidas Alles, Nörpel et Lange-Bertalot Eunotia subarcuatoidas Alles, Nörpel et Lange-Bertalot Fragilaria renera (W. Smi	Adlafia minuscula (Grunow) Lange-Bertalot var. minuscula
Amphora pediculus (Kützing) Grunow Brachysira neoexilis Lange-Bertalot Caloneis tenuis (Gregory) Krammer Chamaepinnularia medicoris (Krasske) Lange-Bertalot et Witkowski Chamaepinnularia sochrensis (Krasske) Lange-Bertalot et Krammer Cocconeis pediculus Ehrenberg Craticula accomoda (Hustedt) D.G. Mann Craticula anabigua (Ehrenberg) D.G. Mann Craticula halophila (Grunow) D.G. Mann Craticula molestiformis (Hustedt) Lange-Bertalot Cyclotella stelligera Cleve & Grunow Cymbopleura amphicephala (Nägeli) Krammer Cymbopleura naviculiformis (Auerswald) Krammer Diploneis krammeri Lange-Bertalot et Reichardt Encyonema hebridicum Grunow ex Cleve Encyonema minutum (Hilse) D.G. Mann Encyonema silesiacum (Bleisch) D.G. Mann Eucocconeis alpestris (Brun) Lange-Bertalot Eunotia bilunaris (Ehrenberg) Eunotia bilunaris (Ehrenberg) Eunotia bilunaris (Ehrenberg) Eunotia bilunaris (Ehrenberg) Eunotia paludosa Grunow Eunotia paludosa Grunow Eunotia navicutati (Kützing) Grunow Eunotia navicutati (Kützing) Grunow Eunotia navicutati (Kützing) Grunow Eunotia navicutatoides Alles, Nörpel et Lange-Bertalot Fragilaria teneta (W. Smith) Lange-Bertalot Fragilaria radians (Kützing) Williams et Round Fragilaria teneta (W. Smith) Lange-Bertalot Eunotia comphonema acuminatum Ehrenberg Gomphonema angustum (Kützing) Rabenhorst Gomphonema angustum (Kützing) Rabenhorst Gomphonema acuifugum Lange-Bertalot et Reichardt Gomphonema acuifusum (Kützing) Nütizing Gomphonema sarcophagus Gregory Hantzschia abundans Lange-Bertalot Luticola mivalis (Ehrenberg) D.G. Mann Meridion circulare (Gréville) C. Agardh Navicula antonii Lange-Bertalot	Amphora copulata (Kützing) Schoeman et Archibald
Brachysira neoexilis Lange-Bertalot Caloneis tenuis (Gregory) Krammer Chamaepinnularia mediocris (Krasske) Lange-Bertalot et Witkowski Chamaepinnularia sochrensis (Krasske) Lange-Bertalot et Witkowski Chamaepinnularia sochrensis (Krasske) Lange-Bertalot et Krammer Cocconeis pediculus Ehrenberg Craticula accomoda (Hustedt) D.G. Mann Craticula anolestiformis (Hustedt) Lange-Bertalot Cyclotella stelligera Cleve & Grunow Cymbopleura amphicephala (Nägeli) Krammer Cymbopleura amphicephala (Nägeli) Krammer Cymbopleura anviculiformis (Auerswald) Krammer Diploneis krammeri Lange-Bertalot et Reichardt Encyonema heiridicum Grunow ex Cleve Encyonema heiridicum Grunow ex Cleve Encyonema nimutum (Hilse) D.G. Mann Eucocconeis alpestris (Brun) Lange-Bertalot Eunotia bilunaris (Ehrenberg) Elunotia bilunaris (Ehrenberg) Elunotia pseudogroen (Kützing) Grunow Eunotia pseudogroenlandica Ange-Bertalot et Tagliaventi Eunotia axigua (Brébisson) Rabenhorst Eunotia autoria grunow Eunotia pseudogroenlandica Lange-Bertalot et Tagliaventi Eunotia nimor (Kützing) Grunow Eunotia subarcuatoides Alles, Nörpel et Lange-Bertalot Eunotia forena (Grunow) Hustedt Fragilaria radians (Kützing) Williams et Round Fragilaria radians (Brébisson) Lange-Bertalot Erustulia crassinervia (Brébisson) Lange-Bertalot et Krammer Gomphonema acuminatum Ehrenberg Gomphonema acuminatum Ehrenberg Gomphonema acuminatum Ehrenberg Gomphonema angustum (Kützing) Rabenhorst Gomphonema angustum (Kützing) Rabenhorst Gomphonema acufiygum Lange-Bertalot et Reichardt Gomphonema acufiygum Lange-Bertalot et Reichardt Gomphonema angustum (Kützing) Rabenhorst Gomphonema acufiygum Lange-Bertalot et Reichardt Gomphonema angustum (Kützing) Rabenhorst Gomphonema angustum (Kützing) Rabenhorst Gomphonema angustum (Kützing) Rabenhorst Gomphonema angustum (Kützing) Rabenhorst Gomphonema angustum (Kützing) Ca. Mann Luticola mitais (Ehrenberg) D.G. Mann Luticola mitais (Ehrenberg) D.G. Mann Meridion circulare (Gréville) C. Agardh Navicula antonii Lange-Bertalot	Amphora pediculus (Kützing) Grunow
Caloneis tenuis (Gregory) Krammer Chamaepinnularia mediocris (Krasske) Lange-Bertalot Chamaepinnularia sochrensis (Krasske) Lange-Bertalot et Witkowski Chamaepinnularia sochrensis (Krasske) Lange-Bertalot et Krammer Cocconeis pediculus Ehrenberg Craticula accomoda (Hustedt) D.G. Mann Craticula ambigua (Ehrenberg) D.G. Mann Craticula anolesiformis (Hustedt) Lange-Bertalot Cyclotella stelligera Cleve & Grunow Cymbopleura amphicephala (Nägeli) Krammer Diploneis krammeri Lange-Bertalot te Reichardt Encyonema hebridicum Grunow ex Cleve Encyonema hieridicum Grunow ex Cleve Encyonema ninutum (Hilse) D.G. Mann Encyonema silesiacum (Bleisch) D.G. Mann Eucocconeis alpestris (Brun) Lange-Bertalot Eunotia bilunaris (Ehrenberg) Eunotia bilunaris (Ehrenberg) Eunotia bilunaris (Ehrenberg) Eunotia bilunaris (Ehrenberg) Eunotia bilunaris (Ehrenberg) Eunotia bilunaris (Ehrenberg) Schaarschmidt Eunotia arcus Ehrenberg Eunotia bilunaris (Ehrenberg) Schaarschmidt Eunotia narcus Ehrenberg Eunotia bilunaris (Ehrenberg) Schaarschmidt Eunotia narcus Ehrenberg Eunotia bilunaris (Ehrenberg) Schaarschmidt Eunotia narcus Ehrenberg Eunotia bilunaris (Ehrenberg) Schaarschmidt Eunotia ratus Ehrenberg Eunotia bilunaris (Ehrenberg) Schaarschmidt Eunotia ratus (Kützing) Grunow Eunotia paludosa Grunow Eunotia neula (Grunow) Hustedt Fragilaria radians (Kützing) Williams et Round Fragilaria radiams (Kützing) Williams et Round Fragilaria radiams (Kützing) Rabenhorst Gomphonema acuminatum Ehrenberg Gomphonema acuminatum Ehrenberg Gomphonema acufiyum Lange-Bertalot et Reichardt Gomphonema acifyugum Lange-Bertalot et Reichardt Gomphonema acifyugum Lange-Bertalot et Reichardt Gomphonema acifyugum Lange-Bertalot et Reichardt Gomphonema acifyugum Lange-Bertalot et Reichardt Gomphonema asarcophagus Gregory Hantzschia abundans Lange-Bertalot Luticola mitais (Ehrenberg) D.G. Mann Meridion circulare (Gréville) C. Agardh Navicula antonii Lange-Bertalot	Brachysira neoexilis Lange-Bertalot
Chamaepinnularia mediocris (Krasske) Lange-Bertalot Chamaepinnularia muscicola (Petersen) Kulikovskiy, Lange-Bertalot et Witkowski Chamaepinnularia soehrensis (Krasske) Lange-Bertalot et Krammer Cocconeis pediculus Ehrenberg Craticula acomoda (Hustedt) D.G. Mann Craticula ambigua (Ehrenberg) D.G. Mann Craticula molestiformis (Hustedt) Lange-Bertalot Cyclotella stelligera Cleve & Grunow Cymbopleura amphicephala (Nägeli) Krammer Cymbopleura naviculiformis (Auerswald) Krammer Diploneis krammeri Lange-Bertalot te Reichardt Encyonema hebridicum Grunow ex Cleve Encyonema nilesiacum (Bleisch) D.G. Mann Eucocconeis alpestris (Brun) Lange-Bertalot Eunotia arcus Ehrenberg Eunotia bilunaris (Ehrenberg) Schaarschmidt Eunotia acuigu (Brébisson) Rabenhorst Eunotia pseudogroenlandica Grunow Eunotia pseudogroenlandica Lange-Bertalot et Tagliaventi Eunotia pseudogroenlandica Lange-Bertalot et Tagliaventi Eunotia pseudogroenlandica Lange-Bertalot et Tagliaventi Eunotia fuenda (Kützing) Grunow Eunotia subarcuatoides Alles, Nörpel et Lange-Bertalot Fragilaria tenera (W. Smith) Lange-Bertalot Fragilaria tenera (W. Smith) Lange-Bertalot Gomphonema acuminatum Ehrenberg Gomphonema acuminatum Ehrenberg Gomphonema acuminatum Richizing) Rabenhorst Gomphonema acuminatum Richizing Rabenhorst Gomphonema acuifugum Lange-Bertalot et Reichardt Gomphonema acuifusum (Kützing) Rabenhorst Gomphonema acuifusum (Kützing) Rabenhorst Gomphonema acuifusum Keichardt et Lange-Bertalot Luticola nivalis (Ehrenberg) D.G. Mann Luticola nivalis (Ehrenberg) D.G. Mann Meridion circulare (Gréville) C. Agardh Navicula antonii Lange-Bertalot	Caloneis tenuis (Gregory) Krammer
Chamaepinnularia muscicola (Petersen) Kulikovskiy, Lange-Beralot et Witkowski Chamaepinnularia soehrensis (Krasske) Lange-Bertalot et Krammer Cocconeis pediculus Ehrenberg Craticula antigua (Ehrenberg) D.G. Mann Craticula antigua (Ehrenberg) D.G. Mann Craticula nalophila (Grunow) D.G. Mann Craticula nalophila (Grunow) D.G. Mann Craticula molestiformis (Hustedt) Lange-Bertalot Cyclotella stelligera Cleve & Grunow Cymbopleura amphicephala (Nägeli) Krammer Diploneis krammeri Lange-Bertalot et Reichardt Encyonema hebridicum Grunow ex Cleve Encyonema hebridicum Grunow ex Cleve Encyonema ninutum (Hilse) D.G. Mann Encyonema silesiacum (Bleisch) D.G. Mann Eucocconeis alpestris (Brun) Lange-Bertalot Eunotia bilunaris (Ehrenberg) Eunotia bilunaris (Ehrenberg) Eunotia bilunaris (Ehrenberg) Schaarschmidt Eunotia arcus Ehrenberg Eunotia bilunaris (Ehrenberg) Schaarschmidt Eunotia aninor (Kützing) Grunow Eunotia paludosa Grunow Eunotia paludosa Grunow Eunotia pseudogroenlandica Lange-Bertalot et Tagliaventi Eunotia tenella (Grunow) Hustedt Fragilaria tenera (W. Smith) Lange-Bertalot Fragilaria tenera (W. Smith) Lange-Bertalot Gomphonema agustum (Kützing) Rabenhorst Gomphonema accultum Reichardt et Lange-Bertalot trustulia crassinervia (Brébisson) Lange-Bertalot et Krammer Gomphonema agustum (Kützing) Rabenhorst Gomphonema agustum (Kützing) Rabenhorst Gomphonema accoltugum Lange-Bertalot et Reichardt Gomphonema accultum Reichardt et Lange-Bertalot <i>Gomphonema accultum</i> Reichardt et Lange-Bertalot <i>Gomphonema accophagus</i> Gregory Hantzschia abundans Lange-Bertalot Luticola nivalis (Ehrenberg) D.G. Mann <i>Meridion circulare</i> (Gréville) C. Agardh Navicula antonii Lange-Bertalot	Chamaepinnularia mediocris (Krasske) Lange-Bertalot
Chamaepinnularia soehrensis (Krasske) Lange-Bertalot et Krammer Cocconeis pediculus Ehrenberg Craticula accomoda (Hustedt) D.G. Mann Craticula anbigua (Ehrenberg) D.G. Mann Craticula halophila (Grunow) D.G. Mann Craticula molestiformis (Hustedt) Lange-Bertalot Cyclotella stelligera Cleve & Grunow Cymbopleura amphicephala (Nägell) Krammer Cymbopleura naviculiformis (Auerswald) Krammer Diploneis krammeri Lange-Bertalot et Reichardt Encyonema nielesiacum (Bleisch) D.G. Mann Encyonema silesiacum (Bleisch) D.G. Mann Eucocconeis alpestris (Brun) Lange-Bertalot Eunotia arcus Ehrenberg Eunotia bilunaris (Ehrenberg) Schaarschmidt Eunotia exigua (Brébisson) Rabenhorst Eunotia pludosa Grunow Eunotia pseudogroenlandica Lange-Bertalot et Tagliaventi Eunotia subarcuatoides Alles, Nörpel et Lange-Bertalot Fragilaria radians (Kützing) Williams et Round Fragilaria radians (Kützing) Williams et Krammer Gomphonema augustum (Kützing) Rabenhorst Gomphonema aculinatum Ehrenberg Gomphonema aculinatum Ehrenberg Gomphonema aculinatum Ehrenberg Gomphonema augustum (Kützing) Rabenhorst Gomphonema augustum (Kützing) Rabenhorst Gomphonema aculinatum Ehrenberg Gomphonema aculinatum Ehrenberg Gomphonema aculinatum Ehrenberg Gomphonema augustum (Kützing) Rabenhorst Gomphonema augustum (Kützing) Kützing Gomphonema aculitum Reichardt et Lange-Bertalot Luticola nivalis (Ehrenberg) D.G. Mann Luticola nivalis (Ehrenberg) D.G. Mann Meridion circulare (Gréville) C. Agardh Navicula antonii Lange-Bertalot	Chamaepinnularia muscicola (Petersen) Kulikovskiy, Lange-Beralot et Witkowski
Cocconeis pediculus Ehrenberg Craticula accomoda (Hustedt) D.G. Mann Craticula anbigua (Ehrenberg) D.G. Mann Craticula nolestiformis (Hustedt) Lange-Bertalot Cyclotella stelligera Cleve & Grunow Cymbopleura amphicephala (Nägeli) Krammer Cymbopleura naviculiformis (Auerswald) Krammer Diploneis krammeri Lange-Bertalot et Reichardt Encyonema hebridicum Grunow ex Cleve Encyonema hebridicum Grunow ex Cleve Encyonema ninutum (Hilse) D.G. Mann Encyonema silesiacum (Bleisch) D.G. Mann Eucoconeis alpestris (Brun) Lange-Bertalot Eunotia arcus Ehrenberg Eunotia bilunaris (Ehrenberg) Schaarschmidt Eunotia exigua (Brébisson) Rabenhorst Eunotia argus Ehrenberg Eunotia paludosa Grunow Eunotia paludosa Grunow Eunotia pseudogroenlandica Lange-Bertalot et Tagliaventi Eunotia tenella (Grunow) Hustedt Fragilaria radians (Kützing) Williams et Round Fragilaria tenera (W. Smith) Lange-Bertalot Eunotia resting (Brébisson) Lange-Bertalot Eunotia casismervia (Brébisson) Lange-Bertalot Eunotia tenella (Grunow) Hustedt Fragilaria radians (Kützing) Williams et Round Fragilaria tenera (W. Smith) Lange-Bertalot Eunota Gomphonema acuminatum Ehrenberg Gomphonema angustum (Kützing) Rabenhorst Gomphonema angustum (Kützing) Kützing Gomphonema angustum (Kützing) Kützing Gomphonema angustum (Kützing) Kützing Gomphonema paroulum (Kützing) Kützing Gomphonema paroulum (Kützing) Kützing Gomphonema paroulum (Kützing) Kützing Mann Luticola nitolis (Ehrenberg) D.G. Mann Meridion circulare (Gréville) C. Agardh Navicula antonii Lange-Bertalot	Chamaepinnularia soehrensis (Krasske) Lange-Bertalot et Krammer
Craticula acconoda (Hustedt) D.G. Mann Craticula ambigua (Ehrenberg) D.G. Mann Craticula molestiformis (Hustedt) Lange-Bertalot Cyclotella stelligera Cleve & Grunow Cymbopleura amplicephala (Nägeli) Krammer Cymbopleura naviculiformis (Auerswald) Krammer Diploneis krammeri Lange-Bertalot et Reichardt Encyonema hebridicum Grunow ex Cleve Encyonema hebridicum Grunow ex Cleve Encyonema silesiacum (Bleisch) D.G. Mann Eucocconeis alpestris (Brun) Lange-Bertalot Eunotia arcus Ehrenberg Eunotia bilunaris (Ehrenberg) Schaarschmidt Eunotia exigua (Brébisson) Rabenhorst Eunotia pseudogroenlandica Lange-Bertalot Eunotia pseudogroenlandica Lange-Bertalot Eunotia pseudogroenlandica Lange-Bertalot Eunotia tenella (Grunow) Eunotia tenella (Grunow) Hustedt Fragilaria radians (Kützing) Williams et Round Fragilaria tenera (W. Smith) Lange-Bertalot Erustulia crassimervia (Brébisson) Lange-Bertalot Eunotia tenella (Grunow) Hustedt Gomphonema angustum (Kützing) Rabenhorst Gomphonema acuminatum Ehrenberg Gomphonema acuminatum Ehr	Cocconeis pediculus Ehrenberg
Craticula ambigua (Ehrenberg) D.G. Mann Craticula halophila (Grunow) D.G. Mann Craticula molestiformis (Hustedt) Lange-Bertalot Cyclotella stelligera Cleve & Grunow Cymbopleura amphicephala (Nägeli) Krammer Cymbopleura naviculiformis (Auerswald) Krammer Diploneis krammeri Lange-Bertalot et Reichardt Encyonema hebridicum Grunow ex Cleve Encyonema hiebridicum Grunow ex Cleve Encyonema silesiacum (Bleisch) D.G. Mann Encocconeis alpestris (Brun) Lange-Bertalot Eunotia arcus Ehrenberg Eunotia bilunaris (Ehrenberg) Schaarschmidt Eunotia exigua (Brébisson) Rabenhorst Eunotia pseudogroenlandica Lange-Bertalot et Tagliaventi Eunotia pseudogroenlandica Lange-Bertalot et Tagliaventi Eunotia pseudogroenlandica Lange-Bertalot et Tagliaventi Eunotia tenella (Grunow) Hustedt Fragilaria radians (Kützing) Williams et Round Fragilaria tenera (W. Smith) Lange-Bertalot Erustulia crassimervia (Brébisson) Lange-Bertalot Eunotia tenella (Grunow) Hustedt Fragilaria tanians (Kützing) Rabenhorst Gomphonema angustum (Kützing) Rabenhorst Gomphonema angustum (Kützing) Rabenhorst Gomphonema calcifugum Lange-Bertalot et Krammer Gomphonema acuminatum Ehrenberg Gomphonema calcifugum Lange-Bertalot et Reichardt Gomphonema calcifugum Lange-Bertalot et Reichardt Gomphonema acuminatum (Kützing) Rabenhorst Gomphonema acuminatum (Kützing) Kützing Gomphonema acultum (Kützing) Kützing Gomphonema acultum (Kützing) Kützing Gomphonema acultum (Kützing) Kützing Mann Luticola nivalis (Ehrenberg) D.G. Mann Luticola nivalis (Ehrenberg) D.G. Mann Meridion circulare (Gréville) C. Agardh Navicula antoni Lange-Bertalot	Craticula accomoda (Hustedt) D.G. Mann
Craticula halophila (Grunow) D.G. Mann Craticula molestiformis (Hustedt) Lange-Bertalot Cyclotella stelligera Cleve & Grunow Cymbopleura amphicephala (Nägeli) Krammer Cymbopleura naviculiformis (Auerswald) Krammer Diploneis krammeri Lange-Bertalot et Reichardt Encyonema hebridicum Grunow ex Cleve Encyonema minutum (Hilse) D.G. Mann Encyonema silesiacum (Bleisch) D.G. Mann Encyonema silesiacum (Bleisch) D.G. Mann Eucocconeis alpestris (Brun) Lange-Bertalot Eunotia acus Ehrenberg Eunotia bilunaris (Ehrenberg) Schaarschmidt Eunotia exigua (Brébisson) Rabenhorst Eunotia aminor (Kützing) Grunow Eunotia paludosa Grunow Eunotia paludosa Grunow Eunotia subarcuatoides Alles, Nörpel et Lange-Bertalot Eunotia tenella (Grunow) Hustedt Fragilaria radians (Kützing) Williams et Round Fragilaria tenera (W. Smith) Lange-Bertalot Frustulia crassinervia (Brébisson) Lange-Bertalot Eunota acuminatum Ehrenberg Gomphonema agustum (Kützing) Rabenhorst Gomphonema acuminatum Ehrenberg Gomphonema acuifugum Lange-Bertalot et Reichardt Gomphonema acuifugum Lange-Bertalot et Reichardt Gomphonema acuifugum Lange-Bertalot et Reichardt Gomphonema acuifugum Cichardt et Lange-Bertalot Luticola nivalis (Ehrenberg) D.G. Mann Luticola nivalis (Ehrenberg) D.G. Mann Luticola mitica (Kützing) D.G. Mann Meridion circulare (Gréville) C. Agardh Navicula antonii Lange-Bertalot	Craticula ambigua (Ehrenberg) D.G. Mann
Craticula molestiformis (Hustedt) Lange-Bertalot Cyclotella stelligera Cleve & Grunow Cymbopleura amphicephala (Nägeli) Krammer Cymbopleura naviculiformis (Auerswald) Krammer Diploneis krammeri Lange-Bertalot et Reichardt Encyonema hebridicum Grunow ex Cleve Encyonema minutum (Hilse) D.G. Mann Encyonema silesiacum (Bleisch) D.G. Mann Eucocconeis alpestris (Brun) Lange-Bertalot Eunotia arcus Ehrenberg Eunotia bilunaris (Ehrenberg) Schaarschmidt Eunotia exigua (Brébisson) Rabenhorst Eunotia minor (Kützing) Grunow Eunotia paludosa Grunow Eunotia paludosa Grunow Eunotia subarcuatoides Alles, Nörpel et Lange-Bertalot Eunotia tenella (Grunow) Hustedt Fragilaria radians (Kützing) Williams et Round Fragilaria tenera (W. Smith) Lange-Bertalot Frustulia crassinervia (Brébisson) Lange-Bertalot Eunotia tenella (Grunow) Hustedt Gomphonema acuminatum Ehrenberg Gomphonema acustum (Kützing) Rabenhorst Gomphonema acustum (Kützing) Rabenhorst Gomphonema acustum (Kützing) Rabenhorst Gomphonema acustum Ehrenberg Gomphonema acultum Lange-Bertalot et Reichardt Gomphonema acleifugum Lange-Bertalot et Reichardt Gomphonema acustum (Kützing) Kützing Gomphonema parvulum (Kützing) Kützing Gomphonema parvulum (Kützing) Kützing Gomphonema parvulum (Kützing) Kützing Monhonema parvulum (Kützing) C. Mann Luticola mutica (Kützing) D.G. Mann Meridion circulare (Gréville) C. Agardh Navicula antonti Lange-Bertalot	Craticula halophila (Grunow) D.G. Mann
Cyclotella stelligera Cleve & Grunow Cymbopleura amphicephala (Nägeli) Krammer Cymbopleura naviculiformis (Auerswald) Krammer Diploneis krammeri Lange-Bertalot et Reichardt Encyonema hebridicum Grunow ex Cleve Encyonema niebridicum Grunow ex Cleve Encyonema silesiacum (Bleisch) D.G. Mann Encyonema silesiacum (Bleisch) D.G. Mann Eucocconeis alpestris (Brun) Lange-Bertalot Eunotia accus Ehrenberg Eunotia bilunaris (Ehrenberg) Schaarschmidt Eunotia exigua (Brébisson) Rabenhorst Eunotia vigua (Brébisson) Rabenhorst Eunotia ninor (Kützing) Grunow Eunotia pseudogroenlandica Lange-Bertalot et Tagliaventi Eunotia subarcuatoides Alles, Nörpel et Lange-Bertalot Eunotia tenella (Grunow) Hustedt Fragilaria radians (Kützing) Williams et Round Fragilaria tenera (W. Smith) Lange-Bertalot Frustulia crassinervia (Brébisson) Lange-Bertalot et Krammer Gomphonema acuminatum Ehrenberg Gomphonema acultum Lange-Bertalot et Reichardt Gomphonema acleifugum Lange-Bertalot et Reichardt Gomphonema acultum Kützing) Kützing Gomphonema acultum Reichardt et Lange-Bertalot Luticola nivalis (Ehrenberg) D.G. Mann Luticola nivalis (Ehrenberg) D.G. Mann Luticola mutica (Kützing) D.G. Mann Meridion circulare (Gréville) C. Agardh Navicula antonii Lange-Bertalot	Craticula molestiformis (Hustedt) Lange-Bertalot
Cymbopleura amphicephala (Nägeli) Krammer Cymbopleura naviculiformis (Auerswald) Krammer Diploneis krammeri Lange-Bertalot et Reichardt Encyonema hebridicum Grunow ex Cleve Encyonema ninutum (Hilse) D.G. Mann Encyonema silesiacum (Bleisch) D.G. Mann Eucoccneis alpestris (Brun) Lange-Bertalot Eunotia bilunaris (Ehrenberg Eunotia bilunaris (Ehrenberg) Schaarschmidt Eunotia exigua (Brébisson) Rabenhorst Eunotia paludosa Grunow Eunotia paludosa Grunow Eunotia subarcuatoides Alles, Nörpel et Lange-Bertalot Fragilaria radians (Kützing) Williams et Round Fragilaria radians (Kützing) Williams et Round Fragilaria radians (Kützing) Lange-Bertalot Frustulia Crassinervia (Brébisson) Lange-Bertalot Frustulia crassinervia (Brébisson) Lange-Bertalot Gomphonema acuminatum Ehrenberg Gomphonema acuminatum Ehrenberg Gomphonema aculitum (Kützing) Rabenhorst Gomphonema calcifugum Lange-Bertalot et Reichardt Gomphonema aculitum Reichardt et Lange-Bertalot Eunotia Grunow) Lange-Bertalot et Reichardt Gomphonema aculitum Ricenberg Gomphonema aculitum Ricenberg Gomphonema aculitum Ricenberg Gomphonema aculitum Ricenberg Gomphonema aculitum Stange-Bertalot et Reichardt Gomphonema aculitum Ricenberg Gomphonema aculitum Ricenberg Gomphonema aculitum Ricenberg Gomphonema aculitum Ricenberg Bertalot Luticola nivalis (Ehrenberg) C. Mann Luticola nivalis (Ehrenberg) D.G. Mann Meridion circulare (Gréville) C. Agardh Navicula antonii Lange-Bertalot	Cyclotella stelligera Cleve & Grunow
Cymbopleura naviculiformis (Auerswald) Krammer Diploneis krammeri Lange-Bertalot et Reichardt Encyonema hebridicum Grunow ex Cleve Encyonema silesiacum (Bleisch) D.G. Mann Encyonema silesiacum (Bleisch) D.G. Mann Eucocconeis alpestris (Brun) Lange-Bertalot Eunotia bilunaris (Ehrenberg) Eunotia bilunaris (Ehrenberg) Schaarschmidt Eunotia exigua (Brébisson) Rabenhorst Eunotia aminor (Kützing) Grunow Eunotia paludosa Grunow Eunotia subarcuatoides Alles, Nörpel et Lange-Bertalot Fragilaria radians (Kützing) Williams et Round Fragilaria radians (Kützing) Williams et Round Fragilaria radians (Kützing) Williams et Round Fragilaria tenera (W. Smith) Lange-Bertalot Frustulia crassinervia (Brébisson) Lange-Bertalot Eunotia tenera (W. Smith) Lange-Bertalot Frustulia crassinervia (Brébisson) Lange-Bertalot Eunotia tenera (W. Smith) Lange-Bertalot Frustulia crassinervia (Brébisson) Lange-Bertalot et Krammer Gomphonema acuminatum Ehrenberg Gomphonema angustum (Kützing) Rabenhorst Gomphonema aclifugum Lange-Bertalot et Reichardt Gomphonema acultum Reichardt et Lange-Bertalot Eunotia Gomphonema acultum Reichardt et Lange-Bertalot Luticola nivalis (Ehrenberg) D.G. Mann Luticola nivalis (Ehrenberg) D.G. Mann Luticola mutica (Kützing) D.G. Mann Meridion circulare (Gréville) C. Agardh Navicula antonii Lange-Bertalot	Cymbopleura amphicephala (Nägeli) Krammer
Diploneis krammeri Lange-Bertalot et Reichardt Encyonema hebridicum Grunow ex Cleve Encyonema silesiacum (Bleisch) D.G. Mann Encyonema silesiacum (Bleisch) D.G. Mann Eucocconeis alpestris (Brun) Lange-Bertalot Eunotia arcus Ehrenberg Eunotia bilunaris (Ehrenberg) Schaarschmidt Eunotia exigua (Brébisson) Rabenhorst Eunotia exigua (Brébisson) Rabenhorst Eunotia ninor (Kützing) Grunow Eunotia paludosa Grunow Eunotia pseudogroenlandica Lange-Bertalot et Tagliaventi Eunotia subarcuatoides Alles, Nörpel et Lange-Bertalot Eunotia tenella (Grunow) Hustedt Fragilaria radians (Kützing) Williams et Round Fragilaria tenera (W. Smith) Lange-Bertalot Frustulia crassinervia (Brébisson) Lange-Bertalot et Krammer Gomphonema angustum (Kützing) Rabenhorst Gomphonema angustum (Kützing) Rabenhorst Gomphonema calcifugum Lange-Bertalot et Reichardt Gomphonema acultum Reichardt et Lange-Bertalot Eunotia dendici (Grunow) Lange-Bertalot et Reichardt Gomphonema acultum Reichardt et Lange-Bertalot Eunotia dissimum (Grunow) Lange-Bertalot et Reichardt Gomphonema acultum Reichardt et Lange-Bertalot Eunotia dissimum (Grunow) Lange-Bertalot et Reichardt Gomphonema aryulum (Kützing) Kützing Gomphonema parvulum (Kützing) Kützing Gomphonema sarcophagus Gregory Hantzschia abundans Lange-Bertalot Luticola nivalis (Ehrenberg) D.G. Mann Meridion circulare (Gréville) C. Agardh Navicula antonii Lange-Bertalot	Cymbopleura naviculiformis (Auerswald) Krammer
 Encyonema hebridicum Grunow ex Cleve Encyonema minutum (Hilse) D.G. Mann Encyonema silesiacum (Bleisch) D.G. Mann Eucocconeis alpestris (Brun) Lange-Bertalot Eunotia arcus Ehrenberg Eunotia bilunaris (Ehrenberg) Schaarschmidt Eunotia exigua (Brébisson) Rabenhorst Eunotia minor (Kützing) Grunow Eunotia pseudogroenlandica Lange-Bertalot et Tagliaventi Eunotia tenella (Grunow) Hustedt Fragilaria radians (Kützing) Williams et Round Fragilaria radians (Kützing) Nabenhorst Gomphonema acuminatum Ehrenberg Gomphonema angustum (Kützing) Rabenhorst Gomphonema acultum Reichardt et Lange-Bertalot Gomphonema acultum Reichardt et Lange-Bertalot	Diploneis krammeri Lange-Bertalot et Reichardt
Encyonema minutum (Hilse) D.G. Mann Encyonema silesiacum (Bleisch) D.G. Mann Eucocconeis alpestris (Brun) Lange-Bertalot Eunotia arcus Ehrenberg Eunotia bilunaris (Ehrenberg) Schaarschmidt Eunotia exigua (Brébisson) Rabenhorst Eunotia exigua (Brébisson) Rabenhorst Eunotia minor (Kützing) Grunow Eunotia paludosa Grunow Eunotia paludosa Grunow Eunotia pseudogroenlandica Lange-Bertalot et Tagliaventi Eunotia subarcuatoides Alles, Nörpel et Lange-Bertalot Eunotia subarcuatoides Alles, Nörpel et Lange-Bertalot Eunotia tenella (Grunow) Hustedt Fragilaria radians (Kützing) Williams et Round Fragilaria tenera (W. Smith) Lange-Bertalot Frustulia crassinervia (Brébisson) Lange-Bertalot et Krammer Gomphonema acuminatum Ehrenberg Gomphonema angustum (Kützing) Rabenhorst Gomphonema calcifugum Lange-Bertalot et Reichardt Gomphonema occultum Reichardt et Lange-Bertalot Gomphonema acuminatum Ehrenberg Gomphonema acultum (Kützing) Kützing Mahar angustum (Kützing) Kützing Gomphonema sarcophagus Gregory Hantzschia abundans Lange-Bertalot Luticola nivalis (Ehrenberg) D.G. Mann Luticola mutica (Kützing) D.G. Mann Meridion circulare (Gréville) C. Agardh Navicula antonii Lange-Bertalot	Encyonema hebridicum Grunow ex Cleve
Encyonema silesiacum (Bleisch) D.G. Mann Eucocconeis alpestris (Brun) Lange-Bertalot Eunotia arcus Ehrenberg Eunotia bilunaris (Ehrenberg) Schaarschmidt Eunotia exigua (Brébisson) Rabenhorst Eunotia exigua (Brébisson) Rabenhorst Eunotia minor (Kützing) Grunow Eunotia pseudogroenlandica Lange-Bertalot et Tagliaventi Eunotia subarcuatoides Alles, Nörpel et Lange-Bertalot Eunotia subarcuatoides Alles, Nörpel et Lange-Bertalot Eunotia tenella (Grunow) Hustedt Fragilaria radians (Kützing) Williams et Round Fragilaria tenera (W. Smith) Lange-Bertalot et Krammer Gomphonema acuminatum Ehrenberg Gomphonema angustum (Kützing) Rabenhorst Gomphonema angustum (Kützing) Rabenhorst Gomphonema occultum Reichardt et Lange-Bertalot Gomphonema acucun keichardt et Lange-Bertalot Lange-Bertalot et Reichardt Gomphonema sarcophagus Gregory Hantzschia abundans Lange-Bertalot Luticola nivalis (Ehrenberg) D.G. Mann Luticola mutica (Kützing) D.G. Mann Meridion circulare (Gréville) C. Agardh Navicula antonii Lange-Bertalot	Encyonema minutum (Hilse) D.G. Mann
Eucocconeis alpestris (Brun) Lange-Bertalot Eunotia arcus Ehrenberg Eunotia bilunaris (Ehrenberg) Schaarschmidt Eunotia exigua (Brébisson) Rabenhorst Eunotia exigua (Brébisson) Rabenhorst Eunotia paludosa Grunow Eunotia pseudogroenlandica Lange-Bertalot et Tagliaventi Eunotia subarcuatoides Alles, Nörpel et Lange-Bertalot Eunotia subarcuatoides Alles, Nörpel et Lange-Bertalot Eunotia tenella (Grunow) Hustedt Fragilaria radians (Kützing) Williams et Round Fragilaria tenera (W. Smith) Lange-Bertalot Frustulia crassinervia (Brébisson) Lange-Bertalot et Krammer Gomphonema acuminatum Ehrenberg Gomphonema angustum (Kützing) Rabenhorst Gomphonema calcifugum Lange-Bertalot et Reichardt Gomphonema occultum Reichardt et Lange-Bertalot Gomphonema parvulum (Kützing) Kützing Gomphonema parvulum (Kützing) Kützing Gomphonema sarcophagus Gregory Hantzschia abundans Lange-Bertalot Luticola nivalis (Ehrenberg) D.G. Mann Luticola mutica (Kützing) D.G. Mann Meridion circulare (Gréville) C. Agardh Navicula antonii Lange-Bertalot	Encyonema silesiacum (Bleisch) D.G. Mann
Eunotia arcus Ehrenberg Eunotia bilunaris (Ehrenberg) Schaarschmidt Eunotia exigua (Brébisson) Rabenhorst Eunotia minor (Kützing) Grunow Eunotia paludosa Grunow Eunotia pseudogroenlandica Lange-Bertalot et Tagliaventi Eunotia subarcuatoides Alles, Nörpel et Lange-Bertalot Eunotia tenella (Grunow) Hustedt Fragilaria radians (Kützing) Williams et Round Fragilaria tenera (W. Smith) Lange-Bertalot et Krammer Gomphonema acuminatum Ehrenberg Gomphonema angustum (Kützing) Rabenhorst Gomphonema aclifugum Lange-Bertalot et Reichardt Gomphonema occultum Reichardt et Lange-Bertalot Gomphonema parvulum (Kützing) Kützing Gomphonema sarcophagus Gregory Hantzschia abundans Lange-Bertalot Luticola nivalis (Ehrenberg) D.G. Mann Luticola mutica (Kützing) D.G. Mann Meridion circulare (Gréville) C. Agardh Navicula antonii Lange-Bertalot	Eucocconeis alpestris (Brun) Lange-Bertalot
Eunotia bilunaris (Ehrenberg) Scharschmidt Eunotia exigua (Brébisson) Rabenhorst Eunotia minor (Kützing) Grunow Eunotia paludosa Grunow Eunotia pseudogroenlandica Lange-Bertalot et Tagliaventi Eunotia subarcuatoides Alles, Nörpel et Lange-Bertalot Eunotia radians (Kützing) Williams et Round Fragilaria radians (Kützing) Williams et Round Fragilaria tenera (W. Smith) Lange-Bertalot Frustulia crassinervia (Brébisson) Lange-Bertalot et Krammer Gomphonema angustum (Kützing) Rabenhorst Gomphonema angustum (Kützing) Rabenhorst Gomphonema angustum (Kützing) Kübenhorst Gomphonema angustum (Kützing) Rabenhorst Gomphonema angustum (Kützing) Rabenhorst Gomphonema angustum (Kützing) Kübenhorst Gomphonema acalcifugum Lange-Bertalot et Reichardt Gomphonema occultum Reichardt et Lange-Bertalot Gomphonema parvulum (Kützing) Kützing Gomphonema sarcophagus Gregory Hantzschia abundans Lange-Bertalot Luticola nivalis (Ehrenberg) D.G. Mann Luticola mutica (Kützing) D.G. Mann Meridion circulare (Gréville) C. Agardh Navicula antonii Lange-Bertalot	Eunotia arcus Ehrenberg
Eunotia exigua (Brébisson) Rabenhorst Eunotia minor (Kützing) Grunow Eunotia paludosa Grunow Eunotia pseudogroenlandica Lange-Bertalot et Tagliaventi Eunotia subarcuatoides Alles, Nörpel et Lange-Bertalot Eunotia tenella (Grunow) Hustedt Fragilaria radians (Kützing) Williams et Round Fragilaria tenera (W. Smith) Lange-Bertalot Frustulia crassinervia (Brébisson) Lange-Bertalot et Krammer Gomphonema acuminatum Ehrenberg Gomphonema angustum (Kützing) Rabenhorst Gomphonema calcifugum Lange-Bertalot et Reichardt Gomphonema exilissimum (Grunow) Lange-Bertalot et Reichardt Gomphonema aculum Reichardt et Lange-Bertalot Gomphonema aculum Reichardt et Lange-Bertalot Lange-Bertalot et Reichardt Gomphonema parvulum (Kützing) Kützing Gomphonema sarcophagus Gregory Hantzschia abundans Lange-Bertalot Luticola nivalis (Ehrenberg) D.G. Mann Luticola mutica (Kützing) D.G. Mann Meridion circulare (Gréville) C. Agardh Navicula antonii Lange-Bertalot	Eunotia bilunaris (Ehrenberg) Schaarschmidt
Eunotia minor (Kützing) Grunow Eunotia paludosa Grunow Eunotia pseudogroenlandica Lange-Bertalot et Tagliaventi Eunotia subarcuatoides Alles, Nörpel et Lange-Bertalot Eunotia tenella (Grunow) Hustedt Fragilaria radians (Kützing) Williams et Round Fragilaria tenera (W. Smith) Lange-Bertalot Frustulia crassinervia (Brébisson) Lange-Bertalot et Krammer Gomphonema acuminatum Ehrenberg Gomphonema angustum (Kützing) Rabenhorst Gomphonema calcifugum Lange-Bertalot et Reichardt Gomphonema exilissimum (Grunow) Lange-Bertalot et Reichardt Gomphonema occultum Reichardt et Lange-Bertalot Gomphonema parvulum (Kützing) Kützing Gomphonema parvulum (Kützing) Kützing Gomphonema sarcophagus Gregory Hantzschia abundans Lange-Bertalot Luticola nivalis (Ehrenberg) D.G. Mann Luticola mutica (Kützing) D.G. Mann Meridion circulare (Gréville) C. Agardh Navicula antonii Lange-Bertalot	Eunotia exigua (Brébisson) Rabenhorst
Eunotia paludosa Grunow Eunotia pseudogroenlandica Lange-Bertalot et Tagliaventi Eunotia subarcuatoides Alles, Nörpel et Lange-Bertalot Eunotia tenella (Grunow) Hustedt Fragilaria radians (Kützing) Williams et Round Fragilaria tenera (W. Smith) Lange-Bertalot Frustulia crassinervia (Brébisson) Lange-Bertalot et Krammer Gomphonema acuminatum Ehrenberg Gomphonema angustum (Kützing) Rabenhorst Gomphonema calcifugum Lange-Bertalot et Reichardt Gomphonema exilissimum (Grunow) Lange-Bertalot et Reichardt Gomphonema occultum Reichardt et Lange-Bertalot Gomphonema parvulum (Kützing) Kützing Gomphonema parvulum (Kützing) Kützing Gomphonema sarcophagus Gregory Hantzschia abundans Lange-Bertalot Luticola nivalis (Ehrenberg) D.G. Mann Luticola mutica (Kützing) D.G. Mann Meridion circulare (Gréville) C. Agardh Navicula antonii Lange-Bertalot	Eunotia minor (Kützing) Grunow
Eunotia pseudogroenlandica Lange-Bertalot et Tagliaventi Eunotia subarcuatoides Alles, Nörpel et Lange-Bertalot Eunotia tenella (Grunow) Hustedt Fragilaria radians (Kützing) Williams et Round Fragilaria tenera (W. Smith) Lange-Bertalot Frustulia crassinervia (Brébisson) Lange-Bertalot et Krammer Gomphonema acuminatum Ehrenberg Gomphonema angustum (Kützing) Rabenhorst Gomphonema calcifugum Lange-Bertalot et Reichardt Gomphonema exilissimum (Grunow) Lange-Bertalot et Reichardt Gomphonema occultum Reichardt et Lange-Bertalot Gomphonema parvulum (Kützing) Kützing Gomphonema parvulum (Kützing) Kützing Gomphonema sarcophagus Gregory Hantzschia abundans Lange-Bertalot Luticola nivalis (Ehrenberg) D.G. Mann Luticola mutica (Kützing) D.G. Mann Meridion circulare (Gréville) C. Agardh Navicula antonii Lange-Bertalot	Eunotia paludosa Grunow
Eunotia subarcuatoides Alles, Nörpel et Lange-Bertalot Eunotia tenella (Grunow) Hustedt Fragilaria radians (Kützing) Williams et Round Fragilaria tenera (W. Smith) Lange-Bertalot Frustulia crassinervia (Brébisson) Lange-Bertalot et Krammer Gomphonema acuminatum Ehrenberg Gomphonema angustum (Kützing) Rabenhorst Gomphonema calcifugum Lange-Bertalot et Reichardt Gomphonema exilissimum (Grunow) Lange-Bertalot et Reichardt Gomphonema occultum Reichardt et Lange-Bertalot Gomphonema parvulum (Kützing) Kützing Gomphonema parvulum (Kützing) Kützing Gomphonema sarcophagus Gregory Hantzschia abundans Lange-Bertalot Luticola nivalis (Ehrenberg) D.G. Mann Luticola mutica (Kützing) D.G. Mann Meridion circulare (Gréville) C. Agardh Navicula antonii Lange-Bertalot	Eunotia pseudogroenlandica Lange-Bertalot et Tagliaventi
Eunotia tenella (Grunow) Hustedt Fragilaria radians (Kützing) Williams et Round Fragilaria tenera (W. Smith) Lange-Bertalot Frustulia crassinervia (Brébisson) Lange-Bertalot et Krammer Gomphonema angustum (Kützing) Rabenhorst Gomphonema angustum (Kützing) Rabenhorst Gomphonema calcifugum Lange-Bertalot et Reichardt Gomphonema exilissimum (Grunow) Lange-Bertalot et Reichardt Gomphonema occultum Reichardt et Lange-Bertalot Gomphonema parvulum (Kützing) Kützing Gomphonema parvulum (Kützing) Kützing Gomphonema sarcophagus Gregory Hantzschia abundans Lange-Bertalot Luticola nivalis (Ehrenberg) D.G. Mann Luticola mutica (Kützing) D.G. Mann Meridion circulare (Gréville) C. Agardh Navicula antonii Lange-Bertalot	Eunotia subarcuatoides Alles, Nörpel et Lange-Bertalot
 Fragilaria radians (Kützing) Williams et Round Fragilaria tenera (W. Smith) Lange-Bertalot Frustulia crassinervia (Brébisson) Lange-Bertalot et Krammer Gomphonema acuminatum Ehrenberg Gomphonema angustum (Kützing) Rabenhorst Gomphonema calcifugum Lange-Bertalot et Reichardt Gomphonema exilissimum (Grunow) Lange-Bertalot et Reichardt Gomphonema occultum Reichardt et Lange-Bertalot Gomphonema parvulum (Kützing) Kützing Gomphonema sarcophagus Gregory Hantzschia abundans Lange-Bertalot Luticola nivalis (Ehrenberg) D.G. Mann Luticola mutica (Kützing) D.G. Mann Meridion circulare (Gréville) C. Agardh Navicula antonii Lange-Bertalot 	Eunotia tenella (Grunow) Hustedt
Fragilaria tenera (W. Smith) Lange-Bertalot Frustulia crassinervia (Brébisson) Lange-Bertalot et Krammer Gomphonema acuminatum Ehrenberg Gomphonema angustum (Kützing) Rabenhorst Gomphonema calcifugum Lange-Bertalot et Reichardt Gomphonema exilissimum (Grunow) Lange-Bertalot et Reichardt Gomphonema occultum Reichardt et Lange-Bertalot Gomphonema parvulum (Kützing) Kützing Gomphonema parvulum (Kützing) Kützing Gomphonema sarcophagus Gregory Hantzschia abundans Lange-Bertalot Luticola nivalis (Ehrenberg) D.G. Mann Luticola mutica (Kützing) D.G. Mann Meridion circulare (Gréville) C. Agardh Navicula antonii Lange-Bertalot	Fragilaria radians (Kützing) Williams et Round
Frustulia crassinervia (Brébisson) Lange-Bertalot et Krammer Gomphonema acuminatum Ehrenberg Gomphonema angustum (Kützing) Rabenhorst Gomphonema calcifugum Lange-Bertalot et Reichardt Gomphonema exilissimum (Grunow) Lange-Bertalot et Reichardt Gomphonema occultum Reichardt et Lange-Bertalot Gomphonema parvulum (Kützing) Kützing Gomphonema parvulum (Kützing) Kützing Gomphonema sarcophagus Gregory Hantzschia abundans Lange-Bertalot Luticola nivalis (Ehrenberg) D.G. Mann Luticola mutica (Kützing) D.G. Mann Meridion circulare (Gréville) C. Agardh Navicula antonii Lange-Bertalot	Fragilaria tenera (W. Smith) Lange-Bertalot
Gomphonema acuminatum Ehrenberg Gomphonema angustum (Kützing) Rabenhorst Gomphonema calcifugum Lange-Bertalot et Reichardt Gomphonema exilissimum (Grunow) Lange-Bertalot et Reichardt Gomphonema occultum Reichardt et Lange-Bertalot Gomphonema parvulum (Kützing) Kützing Gomphonema sarcophagus Gregory Hantzschia abundans Lange-Bertalot Luticola nivalis (Ehrenberg) D.G. Mann Luticola mutica (Kützing) D.G. Mann Meridion circulare (Gréville) C. Agardh Navicula antonii Lange-Bertalot	Frustulia crassinervia (Brébisson) Lange-Bertalot et Krammer
Gomphonema angustum (Kützing) Rabenhorst Gomphonema calcifugum Lange-Bertalot et Reichardt Gomphonema exilissimum (Grunow) Lange-Bertalot et Reichardt Gomphonema occultum Reichardt et Lange-Bertalot Gomphonema parvulum (Kützing) Kützing Gomphonema sarcophagus Gregory Hantzschia abundans Lange-Bertalot Luticola nivalis (Ehrenberg) D.G. Mann Luticola mutica (Kützing) D.G. Mann Meridion circulare (Gréville) C. Agardh Navicula antonii Lange-Bertalot	Gomphonema acuminatum Ehrenberg
Gomphonema calcifugum Lange-Bertalot et Reichardt Gomphonema exilissimum (Grunow) Lange-Bertalot et Reichardt Gomphonema occultum Reichardt et Lange-Bertalot Gomphonema parvulum (Kützing) Kützing Gomphonema sarcophagus Gregory Hantzschia abundans Lange-Bertalot Luticola nivalis (Ehrenberg) D.G. Mann Luticola mutica (Kützing) D.G. Mann Meridion circulare (Gréville) C. Agardh Navicula antonii Lange-Bertalot	Gomphonema angustum (Kützing) Rabenhorst
Gomphonema exilissimum (Grunow) Lange-Bertalot et Reichardt Gomphonema occultum Reichardt et Lange-Bertalot Gomphonema parvulum (Kützing) Kützing Gomphonema sarcophagus Gregory Hantzschia abundans Lange-Bertalot Luticola nivalis (Ehrenberg) D.G. Mann Luticola mutica (Kützing) D.G. Mann Meridion circulare (Gréville) C. Agardh Navicula antonii Lange-Bertalot	Gomphonema calcifugum Lange-Bertalot et Reichardt
Gomphonema occultum Reichardt et Lange-Bertalot Gomphonema parvulum (Kützing) Kützing Gomphonema sarcophagus Gregory Hantzschia abundans Lange-Bertalot Luticola nivalis (Ehrenberg) D.G. Mann Luticola mutica (Kützing) D.G. Mann Meridion circulare (Gréville) C. Agardh Navicula antonii Lange-Bertalot	Gomphonema exilissimum (Grunow) Lange-Bertalot et Reichardt
Gomphonema parvulum (Kützing) Kützing Gomphonema sarcophagus Gregory Hantzschia abundans Lange-Bertalot Luticola nivalis (Ehrenberg) D.G. Mann Luticola mutica (Kützing) D.G. Mann Meridion circulare (Gréville) C. Agardh Navicula antonii Lange-Bertalot	Gomphonema occultum Reichardt et Lange-Bertalot
Gomphonema sarcophagus Gregory Hantzschia abundans Lange-Bertalot Luticola nivalis (Ehrenberg) D.G. Mann Luticola mutica (Kützing) D.G. Mann Meridion circulare (Gréville) C. Agardh Navicula antonii Lange-Bertalot	Gomphonema parvulum (Kützing) Kützing
Hantzschia abundans Lange-Bertalot Luticola nivalis (Ehrenberg) D.G. Mann Luticola mutica (Kützing) D.G. Mann Meridion circulare (Gréville) C. Agardh Navicula antonii Lange-Bertalot	Gomphonema sarcophagus Gregory
Luticola nivalis (Ehrenberg) D.G. Mann Luticola mutica (Kützing) D.G. Mann Meridion circulare (Gréville) C. Agardh Navicula antonii Lange-Bertalot	Hantzschia abundans Lange-Bertalot
Luticola mutica (Kützing) D.G. Mann Meridion circulare (Gréville) C. Agardh Navicula antonii Lange-Bertalot	Luticola nivalis (Ehrenberg) D.G. Mann
Meridion circulare (Gréville) C. Agardh Navicula antonii Lange-Bertalot	Luticola mutica (Kützing) D.G. Mann
Navicula antonii Lange-Bertalot	Meridion circulare (Gréville) C. Agardh
	Navicula antonii Lange-Bertalot

Table A1. Cont.

Navicula cryptocephala Kützing Navicula cryptotenella Lange-Bertalot Navicula exilis Kützing Navicula menisculus Schumann Navicula reichardtiana Lange-Bertalot Navicula trivialis Lange-Bertalot Navicula veneta Kützing Navicula wildii Lange-Bertalot Neidium affine (Ehrenberg) Pfitzer Neidium alpinum Hustedt Neidium ampliatum (Ehrenberg) Krammer Neidium bergii (Cleve-Euler) Krammer Neidium binodeforme Krammer Neidium bisulcatum (Lagerstedt) Cleve var. bisulcatum Neidium dubium (Ehrenberg) Cleve Neidium iridis (Ehrenberg) Cleve Neidium productum (W. Smith) Cleve Nitzschia acicularis (Kützing) W. Smith Nitzschia adamata Hustedt Nitzschia angustata (W. Smith) Grunow Nitzschia communis Rabenhorst Nitzschia dissipata (Kützing) Grunow ssp. dissipata Nitzschia fonticola Grunow Nitzschia gisela Lange-Bertalot Nitzschia palea (Kützing) W. Smith Nitzschia perminuta (Grunow) M. Peragallo Nitzschia pura Hustedt Nitzschia pusilla Grunow Nitzschia supralitorea Lange-Bertalot Nitzschia umbonata (Ehrenberg) Lange-Bertalot Pinnularia borealis Ehrenberg Pinnularia gibba Ehrenberg Pinnularia grunowii Krammer Pinnularia interupta W. Smith Pinnularia marchica I. Schönfelder ex Krammer Pinnularia microstauron (Ehrenberg) Cleve Pinnularia rupestris Hantzsch Pinnularia sinistra Krammer Pinnularia subcapitata Gregory var. subcapitata Pinnularia viridiformis Krammer Placoneis ignorata (Schimanski) Lange-Bertalot Placoneis paraelginensis Lange-Bertalot Planothidium lanceolatum (Brébisson ex Kützing) Lange-Bertalot Psammothidium grischunum (Wunthrich) Bukhtiyarova et Round Psammothidium helveticum (Hustedt) Bukhtiyarova & Round Sellaphora pseudopupula (Krasske) Lange-Bertalot Sellaphora pupula (Kützing) Mereschkowsky (species group) Sellaphora stroemii (Hustedt) D.G.Mann Sellaphora verecundiae Lange-Bertalot Stauroneis acidoclinata Lang-Bertalot et Werum Stauroneis anceps Ehrenberg Stauroneis gracilis Ehrenberg Stauroneis kriegeri Patrick Stauroneis smithii Grunow Stauroneis thermicola (Petersen) Lund Stephanodiscus alpinus Hustedt Surirella angusta Kützing Surirella minuta Brébisson ex Kützing Tabellaria flocculosa (Roth) Kützing

Sample	POK1	POK2	JEL1	JEL2	KRV1	KRV3	RAT1	RAT2	VEL3	MEN1	MEN2	MEN4
date	23.8.	23.8.	23.8.	23.8.	19.8.	19.8.	23.8.	23.8.	19.8.	18.8.	18.8.	18.8.
pН	5.9	3.8	6.5	6.4	6.7	8.3	7.4	6.5	5.9	9.6	7.2	6.2
T ^{[°} C]	17.5	12.2	14.1	9.8	14.9	15.3	7.7	10.2	17.3	17.7	17.9	16.0
Conductivity [µS/cm]	37	16	149	47	242	92	95	256	36	158	55	90
O ₂ saturation [%]	75	53	56	62	10	69	56	74	100	244	90	25
$O_2 [mg/L]$	6.6	4.7	5.0	6.0	0.9	5.9	4.9	7.5	8.1	19.4	7.3	2.0
Secchi depth [cm]	25 *	30 *	60 *	55 *	30 *	13.0	20 *	30 *	35	10	56	36
depth [cm]	25	30	60	55	30	100	20	30	40	20	100	48
Turbidity [1–3]	1	1	1	3	3	3	1	1	3	3	1	3
Clay, silt [%]	100	100	100	90	80	5	100	100	100	95	-	100
Sand, gravel [%]	0	0	0	10	20	65	0	0	0	0	-	0
Pebbles [%]	0	0	0	+	0	30	0	0	0	5	-	0
Stones [%]	0	0	+	0	0	0	0	0	0	0	-	0
CPOM [%]	0	20	0	0	+	5	+	+	+	0	1	0
FPOM [%]	0	80	0	0	0	1	100	80	100	100	80	0
[%] of trampled shore	1	1	0	45	70	70	20	0	50	100	0	80
Intensity of												
trampled shores (0–5)	1	1	0	3	5	2	3	0	4	5	0	4
TP [mg/L]	0.17	0.34	0.03	0.05	0.28	0.07	0.07	0.06	0.23	0.92	0.08	0.15
PO_4^{3-} [mg/L]	0.17	0.30	0.02	0.02	0.07	0.03	0.01	0.001	0.23	0.92	0.05	0.02
TN [mg/L]	1.35	0.82	0.59	0.84	5.91	1.21	1.62	0.56	1.53	6.56	0.95	16.0
NO ₃ -N [mg/L]	0.39	0.52	0.30	0.34	0.42	0.26	0.41	0.30	0.32	0.40	0.21	0.42
NH_4 -N [mg/L]	0.08	0.14	0.03	0.51	4.0	0.73	0.28	0.07	0.06	0.21	0.03	3.08
TDS [mg/l]	72	70	96	50	120	78	94	58	80	226	74	92
TSS [mg/L]	3	8	17	58	151	98	49	93	30	201	257	39

Table A2. Characteristics of karst ponds in the year 2016. * Secchi depth in most transparent ponds is the same as water depth; the bottom of the pond MEN2 was covered with plastic layer on which fine substrate deposited. + represents presence of substrate, cover <5%.

References

- 1. Biggs, J.; Williams, P.; Whitfield, M.; Nicolet, P.; Weatherby, A. 15 years of pond assessment in Britain: Results and lessons learned from the work of Pond Conservation. *Aquat. Conserv. Mar. Freshw. Ecosyst.* **2005**, *6*, 693–714. [CrossRef]
- 2. Oertli, B.; Biggs, J.; Céréghino, R.; Grillas, P.; Joly, P.; Lachavanne, J.-B. Conservation and monitoring of pond biodiversity: Introduction. *Aquat. Conserv. Mar. Freshw. Ecosyst.* **2005**, *6*, 535–540. [CrossRef]
- Declerck, S.; De Bie, T.; Ercken, D.; Hampel, H.; Schrijvers, S.; Van Wichelen, J.; Gillard, V.; Mandiki, R.; Losson, B.; Bau-wens, D.; et al. Ecological characteristics of small farmland ponds: Associations with land use practices at multiple spatial scales. *Biol. Conserv.* 2006, 131, 523–532. [CrossRef]
- 4. Søndergaard, M.; Jeppesen, E.; Jensen, J.P. Pond or lake: Does it make any difference? *Fundam. Appl. Limnol.* 2005, 162, 143–165. [CrossRef]
- 5. Davies, B.; Biggs, J.; Williams, P.; Whitfield, M.; Nicolet, P.; Sear, D.; Bray, S.; Maund, S. Comparative biodiversity of aquatic habitats in the European agricultural landscape. *Agric. Ecosyst. Environ.* **2008**, *125*, 1–8. [CrossRef]
- 6. Hassall, C.; Hollinshead, J.; Hull, A. Environmental correlates of plant and invertebrate species richness in ponds. *Biodivers. Conserv.* **2011**, *20*, 3189–3222. [CrossRef]
- Céréghino, R.; Biggs, J.; Oertli, B.; Declerck, S. The ecology of European ponds: Defining the characteristics of a neglected freshwater habitat. *Hydrobiologia* 2008, 597, 1–6. [CrossRef]
- 8. Čelik, T.; Zelnik, I.; Babij, V.; Vreš, B.; Pirnat, A.; Seliškar, A.; Drovenik, B. Inventory of karstic ponds and their importance for biotic diversity. In *Kras: Water and Life in a Rocky Landscape*; Mihevc, A., Ed.; ZRC: Ljubljana, Slovenia, 2005; pp. 72–82.
- 9. Zelnik, I.; Potisek, M.; Gaberščik, A. Environmental Conditions and Macrophytes of Karst Ponds. *Pol. J. Environ. Stud.* 2012, 21, 1911–1920.
- 10. Zelnik, I.; Gregorič, N.; Tratnik, A. Diversity of macroinvertebrates positively correlates with diversity of macrophytes in karst ponds. *Ecol. Eng.* **2018**, *117*, 96–103. [CrossRef]
- 11. Smol, J.P.; Stoermer, E.F. (Eds.) *The Diatoms: Applications for the Environmental and Earth Sciences*, 2nd ed.; Cambridge University Press: Cambridge, UK, 2010; ISBN 978-1-107-56496-1.
- 12. Hinden, H.; Oertli, B.; Menetrey, N.; Sager, L.; Lachavanne, J.-B. Alpine pond biodiversity: What are the related environmental variables? *Aquat. Conserv. Mar. Freshw. Ecosyst.* **2005**, *15*, 613–624. [CrossRef]

- 13. Ilg, C.; Oertli, B. How can we conserve cold stenotherm communities in warming Alpine ponds? *Hydrobiologia* **2014**, 723, 53–62. [CrossRef]
- 14. Frisbie, M.P.; Lee, R.E. Inoculative Freezing and the Problem of Winter Survival for Freshwater Macroinvertebrates. J. N. Am. Benthol. Soc. 1997, 16, 635–650. [CrossRef]
- 15. Ocón, C.S.; López van Oosterom, M.V.; Munoz, M.I.; Rodrigues-Capítulo, A. Macroinvertebrate trophic responses to nutrient addition in a temperate stream in South America. *Arch. Hydrobiol.* **2013**, *182*, 17–30. [CrossRef]
- 16. Bella, V.D.; Mancini, L. Freshwater diatom and macroinvertebrate diversity of coastal permanent ponds along a gradient of human impact in a Mediterranean eco-region. *Hydrobiologia* **2009**, *634*, 25–41. [CrossRef]
- 17. Berthon, V.; Bouchez, A.; Rimet, F. Using diatom life-forms and ecological guilds to assess organic pollution and trophic level in rivers: A case study of rivers in south-eastern France. *Hydrobiologia* **2011**, 673, 259–271. [CrossRef]
- 18. Sayer, C.D. Problems with the application of diatom-total phosphorus transfer functions: Examples from a shal-low English lake. *Freshw. Biol.* **2001**, *46*, 743–757. [CrossRef]
- 19. Wu, N.; Faber, C.; Sun, X.; Qu, Y.; Wang, C.; Ivetic, S.; Riis, T.; Ulrich, U.; Fohrer, N. Importance of sampling fre-quency when collecting diatoms. *Sci. Rep.* **2016**, *6*, 36950. [CrossRef] [PubMed]
- 20. Vis, C.; Hudon, C.; Cattaneo, A.; Pinel-Alloul, B. Periphyton as an indicator of water quality in the St Lawrence River (Québec, Canada). *Environ. Pollut.* **1998**, *101*, 13–24. [CrossRef]
- 21. Revsbech, N.P.; Nielsen, J.; Hansen, P.K. Benthic Primary Production and Oxygen Profiles. In *Nitrogen Cycling in Coastal Marine Environments*; Blackburn, T.H., Sørensen, J., Eds.; John Wiley & Sons Ltd.: Hoboken, NY, USA, 1988; pp. 69–81.
- Battarbee, R.W.; Jones, V.J.; Flower, R.J.; Cameron, N.G.; Bennion, H.; Carvalho, L.; Juggins, S. Diatoms. In *Tracking Environmental Change Using Lake Sediments: Terrestrial, Algal, and Siliceous Indicators, Developments in Paleoenvironmental Research*; Smol, J.P., Birks, H.J., Last, W.M., Eds.; Springer: Dordrecht, The Netherlands, 2001; pp. 155–202.
- Kröpfl, K.; Vladár, P.; Szabó, K.; Ács, É.; Borsodi, A.K.; Szikora, S.; Caroli, S.; Záray, G. Chemical and biological characterisation of biofilms formed on different substrata in Tisza river. *Environ. Pollut.* 2006, 144, 626–631. [CrossRef]
- 24. Rimet, F.; Bouchez, A. Life-forms, cell-sizes and ecological guilds of diatoms in European rivers. *Knowl. Manag. Aquat. Ecosyst.* **2012**, *406*, 01. [CrossRef]
- 25. Rimet, F.; Berthon, V.; Bouchez, A. *Formes de vie, Guildes Écologiques et Classes de Tailles des Diatomées d'eau Douce*; INRA, Station d'hydrobiologie lacustre: Thonon, France, 2010; p. 10.
- 26. Zelnik, I.; Sušin, T. Epilithic Diatom Community Shows a Higher Vulnerability of the River Sava to Pollution during the Winter. *Diversity* **2020**, *12*, 465. [CrossRef]
- De Marco, P.; Nogueira, D.S.; Correa, C.C.; Vieira, T.B.; Silva, K.D.; Pinto, N.S.; Bichsel, D.; Hirota, A.S.V.; Vieira, R.R.S.; Carneiro, F.M.; et al. Patterns in the organization of Cerrado pond biodiversity in Brazilian pasture landscapes. *Hydrobiologia* 2014, 723, 87–101. [CrossRef]
- Šumberová, K.; Vild, O.; Ducháček, M.; Fabšičová, M.; Potužák, J.; Fránková, M. Drivers of Macrophyte and Diatom Diversity in a Shallow Hypertrophic Lake. Water 2021, 13, 1569. [CrossRef]
- Cantonati, M.; Lange-Bertalot, H.; Decet, F.; Gabrieli, J. Diatoms in very-shallow pools of the site of community importance Danta di Cadore Mires (south-eastern Alps), and the potential contribution of these habitats to diatom biodiversity conservation. *Nova Hedwig.* 2011, 93, 475–507. [CrossRef]
- Bella, D.V.; Puccinelli, C.; Marcheggiani, S.; Mancini, L. Benthic diatom communities and their relationship to water chemistry in wetlands of central Italy. J. Limnol. 2007, 43, 89–99. [CrossRef]
- 31. Blanco, S.; Olenici, A.; Ortega, F.; Jiménez-Gómez, F.; Guerrero, F. Identifying environmental drivers of benthic diatom diversity: The case of Mediterranean mountain ponds. *PeerJ* **2020**, *8*, e8825. [CrossRef]
- 32. Kochoska, H.; Zaova, D.; Videska, A.; Mitic-Kopanja, D.; Naumovska, H.; Wetzel, C.E.; Ector, L.; Levkov, Z. Sellaphora pelagonica (Bacillariophyceae), a new species from dystrophic ponds in the Republic of North Macedonia. *Phytotaxa* **2021**, 496, 2. [CrossRef]
- Vidaković, D.; Levkov, Z.; Hamilton, P.B. Neidiopsis borealis sp. nov., a new diatom species from the mountain Shar Planina, Republic of North Macedonia. Phytotaxa 2019, 402, 21. [CrossRef]
- Mihevc, A.; Gabrovšek, F.; Knez, M.; Kozel, P.; Mulec, J.; Otoničar, B.; Petrič, M.; Pipan, T.; Prelovšek, M.; Slabe, T.; et al. Karst in Slovenia. *Boletín Geológico y Minero* 2016, 127, 79–97.
- 35. Ogrin, D. Podnebni tipi v Sloveniji. Acta Geogr. K 1996, 68, 39–56.
- 36. Hofmann, G.; Werum, M.; Lange-Bertalot, H. Diatomeen im Süßwasser-Benthos von Mitteleuropa: Bestimmungsflora Kieselalgen für die Ökologische Praxis; Koeltz Scientific Books: Königstein, Germany, 2013; p. 908.
- Lange-Bertalot, H.; Hofmann, G.; Werum, M.; Cantonati, M. Freshwater Benthic Diatoms of Central Europe. Over 800 Common Species Used in Ecological Assessment; English Edition with Updated Taxonomy and Added Species; Koeltz Scientific Books: Oberreifenberg, Germany, 2017; p. 942.
- Krammer, K.; Lange-Bertalot, H. Bacillariophyceae. 1. Teil: Naviculaceae. In Süβwasserflora von Mitteleuropa; Ettl, H., Gerloff, J., Heynig, H., Mollenhauer, D., Eds.; Fischer: Jena, Germany, 1986; p. 876.
- Krammer, K.; Lange-Bertalot, H. Bacillariophyceae. 2. Teil: Epithemiaceae, Surirellaceae. In Süβwasserflora von Mitteleuropa; Ettl, H., Gerloff, J., Heynig, H., Mollenhauer, D., Eds.; Fischer: Jena, Germany, 1988; p. 596.
- Krammer, K.; Lange-Bertalot, H. Bacillariophyceae—Teil 3: Centrales, Fragilariaceae, Eunotiaceae. In Süβwasserflora von Mitteleuropa; Ettl, H., Gerloff, J., Heynig, H., Mollenhauer, D., Eds.; Fischer: Jena, Germany, 1991; p. 576.

- Krammer, K.; Lange-Bertalot, H. Bacillariophyceae. 4. Teil: Achnanthaceae, Kritische Ergänzungen zu Navicula (Lineolatae) und Gomphonema Gesamtliteraturverzeichnis. In Süβwasserflora von Mitteleuropa; Ettl, H., Gerloff, J., Heynig, H., Mollenhauer, D., Eds.; Fischer: Jena, Germany, 1991; p. 437.
- 42. Hammer, Ø.; Harper, D.A.T.; Ryan, P.D. PAST: Paleontological Statistics Software Package for Education and Data Analysis. *Palaeontol. Electron.* **2001**, *4*, 1–9.
- 43. Rott, E.; Pipp, E.; Pfister, P.; Van Dahm, H.; Ortler, K.; Binder, N.; Pall, K. *Indikationslisten fur Aufwuchsalgen in Östereichen Fließgevessern, Teil 2: Trophienindikation so vie geochemische Präferenz, Taxonomische und toxicologische Anmerkungen*; Bundesministerium für Land und Forstwirtschaft: Wien, Austria, 1999; p. 248.
- 44. Braak, C.J.F.; Šmilauer, P. CANOCO Reference Manual and CanoDraw for Windows User's Guide: Software for Canonical Community Ordination; Microcomputer Power: Ithaca, NY, USA, 2002; p. 500.
- 45. Pan, Y.; Hughes, R.; Herlihy, A.; Kaufmann, P. Non-wadeable river bioassessment: Spatial variation of benthic diatom assemblages in Pacific Northwest rivers, USA. *Hydrobiologia* **2012**, *684*, 241–260. [CrossRef]
- Heine-Fuster, I.; López-Allendes, C.; Aránguiz-Acuña, A.; Véliz, D. Differentiation of Diatom Guilds in Extreme Environments in the Andean Altiplano. Front. Environ. Sci. 2021, 9, 266. [CrossRef]
- 47. Licursi, M.; Gómez, N.; Sabater, S. Effects of nutrient enrichment on epipelic diatom assemblages in a nutrient-rich lowland stream, Pampa Region, Argentina. *Hydrobiologia* **2016**, *766*, 135–150. [CrossRef]
- Passy, S.I. Diatom ecological guilds display distinct and predictable behavior along nutrient and disturbance gradients in running waters. *Aquat. Bot.* 2007, *86*, 171–178. [CrossRef]
- Gottschalk, S.; Kahlert, M. Shifts in taxonomical and guild composition of littoral diatom assemblages along environmental gradients. *Hydrobiologia* 2012, 694, 41–56. [CrossRef]
- 50. Béres, V.; Török, P.; Kókai, Z.; Krasznai, E.T.; Tóthmérész, B.; Bácsi, I. Ecological diatom guilds are useful but not sensitive enough as indicators of extremely changing water regimes. *Hydrobiologia* **2014**, *738*, 191–204. [CrossRef]
- 51. Zelnik, I.; Balanč, T.; Toman, M.J. Diversity and Structure of the Tychoplankton Diatom Community in the Limnocrene Spring Zelenci (Slovenia) in Relation to Environmental Factors. *Water* **2018**, *10*, 361. [CrossRef]
- Peszek, Ł.; Zgrundo, A.; Noga, T.; Kochman-Kędziora, N.; Poradowska, A.; Rybak, M.; Puchalski, C.; Lee, J. The influence of drought on diatom assemblages in aemperate climate zone: A case study from the Carpathian Mountains, Poland. *Ecol. Indic.* 2021, 125, 107579. [CrossRef]
- 53. Lavoie, I.; Lento, J.; Morin, A. Inadequacy of size distributions of stream benthic diatoms for environmental monitoring. J. N. Am. Benthol. Soc. 2010, 29, 586–601. [CrossRef]
- 54. Soininen, J.; Jamoneau, A.; Rosebery, J.; Passy, S.I. Global patterns and drivers of species and trait composition in diatoms: Global compositional patterns in stream diatom. *Glob. Ecol. Biogeogr.* **2016**, *25*, 940–950. [CrossRef]
- Scinto, L.J.; Reddy, K.R. Biotic and abiotic uptake of phosphorus by periphyton in a subtropical freshwater wetland. *Aquat. Bot.* 2003, 77, 203–222. [CrossRef]
- 56. Haubois, A.G.; Sylvestre, F.; Guarini, J.M.; Richard, P.; Blanchard, G.F. Spatio-temporal structure of the epipelic diatom assemblage from an intertidal mudflat in Marennes-Oléron Bay, France. *Estuar. Coast. Shelf Sci.* **2005**, *64*, 385–394. [CrossRef]
- 57. Körner, C. The use of 'altitude' in ecological research. Trends. Ecol. Evol. 2007, 22, 569–574. [CrossRef] [PubMed]
- van der Grinten, E.; Janssen, A.P.H.; de Mutsert, K.; Barranguet, C.; Admiraal, W. Temperature- and Light-Dependent Performance of the Cyanobacterium Leptolyngbya Foveolarum and the Diatom Nitzschia Perminuta in Mixed Biofilms. *Hydrobiologia* 2005, 548, 267–278. [CrossRef]
- 59. Jurczak, T.; Wojtal-Frankiewicz, A.; Kaczkowski, Z.; Oleksińska, Z.; Bednarek, A.; Zalewski, M. Restoration of a shady urban pond—The pros and cons. *J. Environ. Manag.* 2018, 217, 919–928. [CrossRef] [PubMed]
- 60. DeNicola, D.M. A review of diatoms found in highly acidic environments. Hydrobiologia 2000, 433, 111–122. [CrossRef]
- Krivograd-Klemenčič, A.; Smolar-Žvanut, N.; Istenič, D.; Griessler-Bulc, T. Algal community patterns in Slovenian bogs along environmental gradients. *Biologia* 2010, 65, 422–437. [CrossRef]
- 62. Schmutzer, A.C.; Gray, M.J.; Burton, E.C.; Miller, D.L. Impacts of cattle on amphibian larvae and the aquatic environment. *Freshw. Biol.* **2008**, *53*, 2613–2625. [CrossRef]
- 63. Connell, J.H. Diversity in tropical rainforests and coral reefs. Science 1978, 199, 1302–1310. [CrossRef]
- 64. Gascón, S.; Boix, D.; Sala, J. Are different biodiversity metrics related to the same factors? A case study from Mediter-ranean wetlands. *Biol. Conserv.* 2009, *11*, 2602–2612. [CrossRef]
- 65. Feio, M.; Almeida, S.; Craveiro, S.; Calado, A. Diatoms and macroinvertebrates provide consistent and complementary information on environmental quality. *Fundam. Appl. Limnol.* **2007**, *169*, 247–258. [CrossRef]
- Lange, K.; Liess, A.; Piggott, J.J.; Townsend, C.R.; Matthaei, C.D. Light, nutrients and grazing interact to determine stream diatom community composition and functional group structure: Diatom responses to light, nutrients and grazing. *Freshw. Biol.* 2011, 56, 264–278. [CrossRef]