

Short Note

9,19-Cyclolanost-24-en-3-one,21,23-epoxy-21,22-dihydroxy (21*R*, 22*S*, 23*S*) from the Leaves of *Lansium domesticum* Corr cv Kokossan

Tri Mayanti ¹, Julinton Sianturi ¹, Desi Harneti ¹, Darwati ¹, Unang Supratman ^{1,*}, Mohamad Mustaqim Rosli ² and Hoong-Kun Fun ²

- ¹ Department of Chemistry, Faculty of Mathematics and Natural Sciences, Padjadjaran University, Jalan Raya Bandung-Sumedang Km 21, Jatinangor 45363, Sumedang, Indonesia;
 E-Mails: t.mayanti@yahoo.co.id (T.M.); julintons@gmail.com (J.S.);
 d_harneti@yahoo.com (D.H.); darwatititi@yahoo.co.id (D.)
- ² X-ray Crystallography Unit, School of Physics, Universiti Sains Malaysia, 11800 USM Penang, Malaysia; E-Mails: mustaqim@usm.my (M.M.R.); hkfun@usm.my (H.-K.F.)
- * Author to whom correspondence should be addressed; E-Mail: u_supratman@unpad.ac.id; Tel./Fax: +62-22-7794391.

Academic Editor: Norbert Haider

Received: 12 October 2015 / Accepted: 1 December 2015 / Published: 4 December 2015

Abstract: A new cycloartan-type triterpenoid, 9,19-cyclolanost-24-en-3-one,21,23-epoxy-21,22-dihydroxy (21*R*, 22*S*, 23*S*), was isolated from the leaves of *Lansium domesticum* Corr cv kokossan. The chemical structure of **1** was elucidated on the basis of spectroscopic data, X-ray diffraction and comparison with those related compounds previously reported.

Keywords: cycloartan-type triterpenoid; Lansium domesticum; Meliaceae

1. Introduction

Lansium domesticum Corr cv kokossan is a species of small tree from the Melliaceae family and widely distributed in Thailand and surrounding countries in Southern Asia [1,2]. Previous phytochemical studies on *L. domesticum* have resulted in the isolation of several types of triterpenoids [3–8], which possess interesting biological activities, such as anticancer [9], antibacterial [10], insecticides [11], antimalarial [12], cosmetic [13] and antifeedant activities [14].

As a part of our studies on triterpenoids compounds from Indonesian Meliaceae plants, we isolated and described two tetranortriterpenoids, kokosanolide A and C and three onoceranoid-type triterpenoids including, kokosanolide B, 8,14-secogammacera-7,14-diene-3,21-dione and 8,14-secogammacera-7,14(27)-diene-3,21-dione from the seeds and the bark of the species kokossan [12]. In further screening for triterpenoid compounds, we found that the ethyl acetate extract from the leaves of *L. domesticum* cv kokossan exhibited the presence of triterpenoid compounds, based on a phytochemical test using the Liebermann-Burchard reagent. We report herein the isolation and structure elucidation of the new cycloartan-type triterpenoid, 9,19-cyclolanost-24-en-3-one,21,23-epoxy-21,22-dihydroxy (21*R*, 22*S*, 23*S*) (1) from the leaves of the *L. domesticum* cv kokossan. The chemical structure of compound 1 was established by NMR data and X-ray diffraction, as well as by comparison with those related compounds previously reported.

2. Result and Discussion

The ethyl acetate extract was separated and purified over a chromatography column packed with silica gel 60 by gradient elution. The chromatography column fractions were repeatedly subjected to normal-phase column chromatography and recrystallized in acetone to afford compound 1 (Figure 1). 9,19-cyclolanost-24-en-3-one,21,23-epoxy-21,22-dihydroxy (21R, 22S, 23S) was obtained as a white needle-like crystals from acetone; m.p. 145–147 °C, $[\alpha]_{D}^{20}$ –5.2° (c, 0.5 CHCl₃). The molecular formula of 1 was established to be C₃₀H₄₆O₄ by HR-ESI-TOFMS spectrum $[M + H]^+ m/z$ 471.3381 (calcd. 470.3396), together with NMR data (Table 1), thus requiring eight degrees of unsaturation. Its UV spectrum exhibited no conjugated group based on maximum absorption at 200 nm. The IR spectrum of 1 exhibited the presences of O-H stretch (3401 cm⁻¹), CH sp^3 stretch (2928 cm⁻¹), C=O ketone stretch (1699 cm⁻¹), C=C (1453 cm⁻¹), and ether group (1078 cm⁻¹). The ¹H-NMR spectrum exhibited the presences of six methyl groups were resonated at $\delta_{\rm H}$ 0.97, 1.73, 1.66, 1.09, 1.16, and 0.96 for Me-18, Me-26, Me-27, Me-28, Me-29, and Me-30, respectively. The characteristic of C-9 and C-10 cyclopropyl methylene group from a cycloartanone-type triterpenoid resonating at $\delta_{\rm H}$ 0.66 and $\delta_{\rm H}$ 0.70 (J = 3.9 Hz), respectively. An olefinic proton (H-24) was observed at δ_{H} 5,48 (1H, d, J = 8.4 Hz) together with two geminal vinvl methyls proton were observed at $\delta_{\rm H}$ 1.73 (3H, s) and $\delta_{\rm H}$ 1.66 (3H, s) which assigned for H-26 and H-27 respectively, thus indicating a $\Delta^{24,25}$ double bond in the side chain. Additional functionalities included the three protons bonded to carbon bearing oxygen were resonated at $\delta_{\rm H}$ 5.17 (1H, d, J = 5.8 Hz, H-21) as a doublet as well as $\delta_{\rm H}$ 3.96 (1H, dd, J = 4.5, 6.5 Hz, H-22) and $\delta_{\rm H}$ 4.68 (1H, dd, J = 6.5, 8.4 Hz, H-23) as double doublet. A total of thirty carbon signals were observed in the ¹³C-NMR spectrum. These were assigned by DEPT and HMQC experiments to two sp^2 carbon signals at $\delta_{\rm C}$ 123.8 and 135.8, one singlet carbon for ketone group at $\delta_{\rm C}$ 215.1, six methyls sp³, eight methylenes, three sp^3 oxygenated carbons at $\delta_{\rm C}$ 101.6, 76.3 and 97.2, and five sp^3 quaternary carbons. A downfield signal of methine was observed at $\delta_{\rm C}$ 101.6 ($\delta_{\rm H}$ 5.17, 1H, d, J = 1.2 Hz) corresponding to a hemiketalic carbon. These functionalities accounted for two out of the total eight degrees of unsaturation. The remaining six degrees of unsaturation were consistent to a cycloartan-type structure with one additional ring [15].



Figure 1. 9,19-Cyclolanost-24-en-3-one,21,23-epoxy-21,22-dihydroxy (21R, 22S, 23S) (1).

The gross structure of 1 was deduced from the ¹H-¹H COSY and HMBC spectra (Figure 2). The ¹H-¹H- COSY spectrum showed coupling between H-17/H-20, H-20/H-21, H-20/H-22 and H-22/H-23, supporting the presence of a cycloartane triterepenoid structure. A long-range correlation was exhibited in the HMBC spectrum, between methine sp^2 proton signal at $\delta_{\rm H}$ 5.48 (H-24) and oxygenated carbon signal at δ_C 97.2 (C-23), as well as a correlation between δ_H 2.19 (H-20) and δ_H 4.82 (H-22) to a downfield signal of methine at $\delta_{\rm C}$ 101.6 (C-21) which suggests that the furan ring was built at C-21, C-20, C-22 and C-23, respectively. The position of olefinic carbon signals δ_C 123.8 (C-24) and δ_C 35.8 (C-25) were confirmed by long-range correlation of gem-dimethyl vinyl group (Me-26 and Me-27), whereas the position of the C-3 carbonyl was confirmed by showing correlations of H-2, Me-28 and Me-29 to the C-3 carbonyl group ($\delta_{\rm C}$ 215.1). The NMR data of 1 was similar to those of argenteanone [16], the main difference was stereochemistry at C-21, C-22 and C-23. The structure of compound 1 was further confirmed by X-ray diffraction analysis (Figure 3). The asymmetric unit of compound 1 contain one unit of 3-(2,4-dihydroxy-5-(2-methylprop-1-en-1-yl)tetrahydrofuran-3-yl)-2a,5a,8,8tetramethyltetradecahydrocyclo-penta[a]-cyclopropa[e]-phenanthren-9(1H)-one plus half unit of water molecule. It was crystallized in P21 space group with a = 13.4825(2) Å, b = 6.2433(1) Å, c =17.0838(3) Å, $\beta = 109.3068(8)^{\circ}$ and Z = 4. The molecular structure contains three six-membered rings of cyclohexane, two five membered rings, a cyclopentane and oxolane, and one cyclopropane. Three six membered rings, cyclopentane and cyclopropane are *trans* fused when the cyclopropane is in axial position relative to the mean plane cyclohexane and cyclopentane, therefore confirming the absolute configuration of 1. Therefore, compound 1 was determined as a new cycloartan-type triterpenoid and was named 24(E)-cyclolanost-24-en-3-one, 21,23-epoxy-21,22-dihydroxy (21R, 22S, 23S) (1).

Position	¹³ C-NMR	¹ H-NMR
	δ_{C} ppm (mult., ppm)	$\delta_{\rm H}$ ppm (Int, mult., $J = {\rm Hz}$)
1	33.9 (t)	1.86 (1H, <i>m</i>)
		1.60 (1H, <i>m</i>)
2	37.8 (t)	2.21 (1H, ddd, 3.9, 6.5, 10.4)
		2.73 (1H, ddd, 6.0, 6.0, 10.4)
3	215.1 (s)	-
4	50.7 (s)	-
5	46.7 (d)	1.93 (1H, <i>m</i>)
6	22.1 (t)	1.59 (1H, <i>m</i>)
		1.50 (1H, <i>m</i>)
7	28.1 (t)	1.20 (1H, <i>m</i>)
		1.86 (1H, <i>m</i>)
8	48.8 (d)	1.64 (1H, <i>m</i>)
9	21.7 (s)	-
10	26.9 (s)	-
11	26.7 (t)	1.93 (1H, <i>m</i>)
		1.07 (1H, <i>m</i>)
12	32.8 (t)	1.57 (1H, <i>m</i>)
		1.64 (1H, <i>m</i>)
13	48.7 (s)	-
14	46.5 (s)	-
15	36.3 (t)	1.05 (1H, <i>m</i>)
		1.34 (1H, <i>m</i>)
16	26.9 (t)	1.86 (1H, <i>m</i>)
		2.00 (1H, <i>m</i>)
17	49.2 (d)	1.,57 (1H, <i>m</i>)
18	19.4 (q)	0.97 (1H, <i>s</i>)
19	27.1 (t)	0.70 (1H, <i>d</i> , 3,9)
		0.66 (1H, <i>d</i> , 3,9)
20	59.4 (d)	2.19 (1H, <i>m</i>)
21	101.6 (d)	5.17 (1H, <i>d</i> , 5.8)
22	78.2 (d)	3.96 (1H, <i>dd</i> , 4.5, 6.5)
23	79.0 (d)	4.68 (1H, <i>dd</i> , 6.5, 8.4)
24	123.8 (d)	5.48 (1H, <i>d</i> , 8,4)
25	135.8 (s)	-
26	26.1 (q)	1.73 (3H, <i>s</i>)
27	18.4 (q)	1.66 (3H, <i>s</i>)
28	21.1 (q)	1.09 (3H, <i>s</i>)
29	19.7 (q)	1.16 (3H, <i>s</i>)
30	19.4 (q)	0.96 (3H, <i>s</i>)

Table 1. NMR data (500 MHz for 1 H and 125 MHz for 13 C, in acetone-*d*₆) for **1**.



Figure 2. Selected HMBC and ¹H-¹H-COSY correlations for 1.



Figure 3. ORTEP drawing for 1.

3. Experimental Section

3.1. General

Melting points were measured on an electrothermal melting point apparatus and are uncorrected. Optical rotations were recorded on an ATAGO AP-300 automatic polarimeter. The UV spectrum spectra were recorded on Shimazu series 1800 spectrophotometer (Kyoto, Japan). The IR spectra were recorded on a Perkin-Elmer spectrum-100 FT-IR (Waltham, MA, USA) in KBr. HR-ESI-TOFMS spectra were obtained with a Waters LCT Premier XE mass spectrometer instruments (Santa Clara, CA, USA). ¹H and ¹³C-NMR spectra were obtained with a JEOL JNM A-500 spectrometer using TMS as internal standard (Tokyo, Japan). Chromatographic separations were carried out on silica gel 60 (Merck, Darmstadt, Germany). TLC plates were precoated with silica gel GF₂₅₄ (Merck, 0.25 mm) and detection was achieved by spraying with 10% H₂SO₄ in ethanol, followed by heating. Crystallographic data were collected on a Bruker SMART APEXII CCD area-detector diffractometer with a graphite monochromated Mo-K α radiation ($\lambda = 0.71073$ Å) at 100.0 K with the Oxford Cryosystem Cobra low-temperature attachment (Billerica, MA, USA).

3.2. Plant Material

The leaves of *L. domesticum* cv kokossan were collected in Cililin District, Bandung, West Java Province, Indonesia in July 2013. The plant was identified by the staff of the Laboratory of Plant Taxonomy, Department of Biology, Padjadjaran University, Indonesia. A voucher specimen (No. 10188) was deposited at the herbarium of the Padjadjaran University.

3.3. Extraction and Isolation

The dried leaves (2.0 kg) were extracted with methanol exhaustively (5 L) at room temperature for 3 days. After removal of the solvent under reduced pressure, the viscous concentrate of MeOH extract (49.7 g) was first suspended in H₂O and then partitioned successively with *n*-hexane and EtOAc. A portion of ethyl acetate (16.7 g) was subjected to column chromatography over silica gel using a gradient of *n*-hexane/EtOAc (10:0–0:10) to afford 22 fractions (A01–A22). Fraction A06 (1.9 g) eluted by *n*-hexane/EtOAc = 6:4 was subjected to silica gel column chromatography, eluted with the mixtures of *n*-hexane/CHCl₃ (10:0–8:2) as eluting solvents to afford 17 fractions (B01–B17). The Fraction of B06 to B07 were combined and to give the white crystal (30.0 mg) after crystallized using acetone solvent.

NMR, IR and HRMS spectra for the title compound are available in the Supplementary Information.

Acknowledgments

This research was supported by grants from Directorate Generalof Higher Education, Ministry of Education and Culture, Indonesia (2013–2014 by TM).

Author Contributions

Unang Supratman designed the whole experiment and contributed to the manuscript. Tri Mayanti and Julinton Sianturi researched data and wrote the manuscript, Desi Harneti and Darwati analyzed the NMR and LCMS/MS spectra, Mohamad Mustaqim Rosli and Hoong-Kun Fun analyzed the X-ray diffraction. All authors read and approved the final manuscript.

Conflicts of Interest

The authors declare no conflict of interest.

References

- Omar, S.; Marcotte, M.; Fields, P.; Sanchez, P.E.; Poveda, L.; Matta, R.; Jimenez, A.; Durst, T.; Zhang, J.; Kinnon, M.; *et al.* Antifeedant activities of triterpenoids isolated from tropical Rutales. *J. Stored Prod. Res.* 2007, *43*, 92–96.
- Leaman, D.J.; Arnason, J.T.; Yusuf, R.; Sangat-Roemantyo, H.; Soedjito, H.; Angerhofer, C.K.; Pezzuto, J.M. Malaria remedies of the Kenyah of the Apo Kayan, East Kalimantan, Indonesian Borneo: A quantitative assessment of local consensus as an indicator of biological efficacy. *J. Ethnopharmacol.* 1995, 49, 1–16.

- 3. Nishizawa, M.; Nishide, H.; Hayashi, Y.; Kosela, S. The structure of lansioside A: A novel triterpene glycoside with amino-sugar from *Lansium domesticum*. *Tetrahedron Lett.* **1982**, *23*, 1349–1350.
- 4. Nishizawa, M.; Nishide, H.; Kosela, S.; Hayashi, Y. Structure of lansiosides: Biologically active new triterpene glycosides from *Lansium domesticum*. J. Org. Chem. **1983**, 48, 4462–4466.
- Tanaka, T.; Ishibashi, M.; Fujimoto, H.; Okuyama, E.; Koyano, T.; Kowiyhayakorn, T.; Hayashi, M.; Komiyama, K. New Onoceranoid Constituents from *Lansium domesticum*. J. Nat. Prod. 2002, 65, 1709–1711.
- Habaguchi, K.; Watanabe, M.; Nakadaira, Y.; Nakanishi, K.; Kaing, A.K.; Lim, F.L. The full structures of lansic acid and its minor congener, an unsymmetric onoceradienedione. *Tetrahedron Lett.* 1986, *34*, 3731–3734.
- 7. Mayanti, T.; Supratman, U.; Mukhtar, M.R.; Awang, K.; Ng, S.W. Kokosanolide from the seed of *Lansium domesticum* Corr. *Acta Crystallogr.* **2009**, *E65*, o750.
- Supratman U.; Mayanti, T.; Awang, K.; Mukhtar, M.R.; Ng, S.W.; 14-Hydroxy-8,14secogammacera-7-ene-3,21-dione from the bark of *Lansium domesticum* Corr. *Acta Crystallogr*. 2010, *E66*, o1621.
- 9. Manosroi, A.; Jantrawut, P.; Sainakham, M.; Manosroi, W.; Manosroi, J. Anticaner activities oh the extract from longkong (*Lansium domesticum*) young fruits. *Pharm Biol.* **2012**, *50*, 1397–1407.
- 10. Ragasa, C.Y.; Labrador, P.; Rideout, J.A. Antimicrobial terpenoid from *Lansium domesticum*. *Philipp. Agric. Sci.* **2006**, *89*, 101–105.
- Leatemia, J.A.; Isman, M.B. Insecticidal Activity of Crude Seed Extracts of Annona. spp., Lansium domesticum and Sandoricum koetjape Against Lepidopteran Larvae. Phytopatasitica 2004, 32, 30–37.
- 12. Saewan, N.; Sutherland, J.D.; Chantrapromma, K. Antimalarial tetranortriterpenoids from the seed of *Lansium domesticum* Corr. *Phytrochemistry* **2006**, *67*, 2288–2293.
- 13. Tilaar, M.; Wong, W.; Ranti, S.; Wasitaatmadja, M.; Junardy, D. Review of *Lansium domesticum* Corrêa and its use in cosmetics. *Bol. Latinoam. Caribe Plant. Med. Aromat.* **2008**, *7*, 183–189.
- Mayanti, T.; Tjokronegoro, R.; Supratman, U.; Mukhtar, M.R.; Awang, K.; Hadi, A.H.A. Antifeedant Triterpenoids from the Seeds and Bark of *Lansium domesticum* cv Kokossan (Meliaceae). *Molecules* 2011, *16*, 2785–2795.
- Awang, K.; Loong, X.; Leong, M.F.K.; Supratman, U.; Litaudon, U.; Mukhtar, M.M.; Mohamad, R.K. Triterpenes and steroids from the leaves of *Aglaia exima* (Meliaceae). *Fitoterapia* 2012, *83*, 1391–1395.
- 16. Ombuwajo, O.R.; Martin, M.T.; Perromat, G.; Sevenet, T.; Awang, K.; Pais, M. Cytotoxic cycloartantes from *Aglaia argentea*. *Phytochemistry* **1996**, *41*, 1325–1328.

© 2015 by the authors; licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution license (http://creativecommons.org/licenses/by/4.0/).