

Figure S1. *Mcph1* DEGs were not associated with transcriptional regulation of the SWI/SNF complex and E2F1. (A) The Venn diagrams represent the number of intersecting genes between *Mcph1*-KO DEGs and the target genes of the core proteins of the SWI/SNF complex. The core proteins in this complex are BAF155, BAF170, BRG1, and BRM. $p > 0.05$ indicates no statistical significance. Statistical analysis was performed using Fisher's exact test. (B) *Mcph1*-KO DEGs transcription factor prediction analysis.

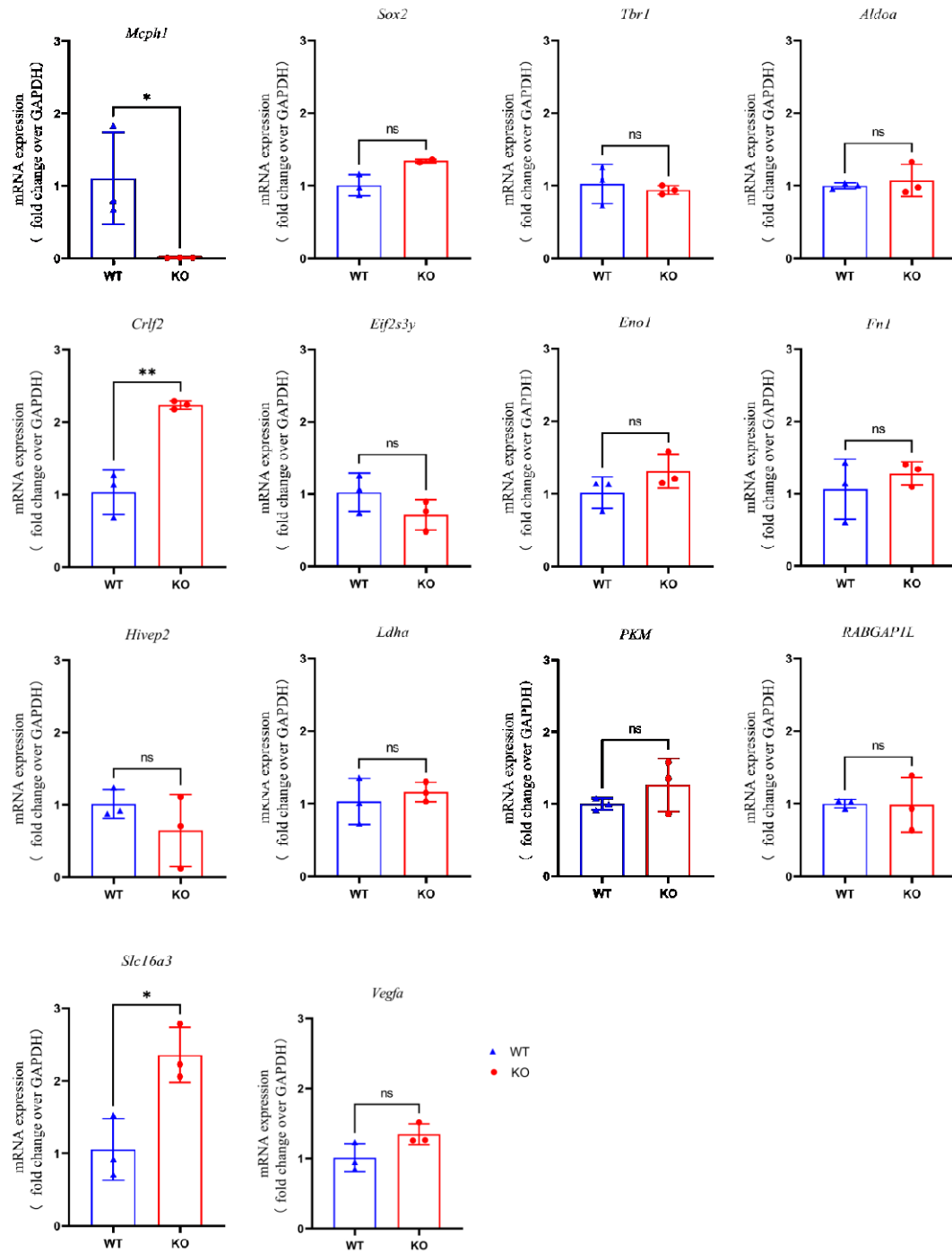


Figure S2. Expression verification of intersecting genes related to neurodevelopment. WT is the control group, and KO is the knockout of the *Mcph1* group. The expression of DEGs in the control and *Mcph1*-KO groups was detected by RT-qPCR.

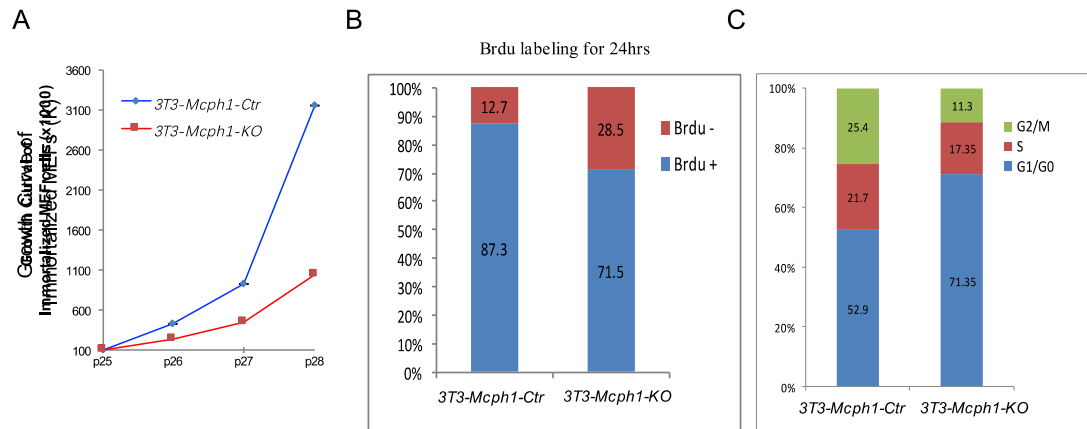


Figure S3. *Mcph1* affects cell proliferation in immortalized MEFs. (A) Growth curves of *Mcph1*-Ctr and *Mcph1*-KO immortalized MEFs. The immortalized cells have a good proliferation capability. Cells were harvested by trypsinization and counted once per passage. P25, cells at passage 25; P26, cells at passage 26; P27, cells at passage 27, and P28, cells at passage 28. (B) Immobilized MEFs, stained with BrdU antibody, and quantitative flow cytometric analysis showed that BrdU-labeled *Mcph1*-KO cells decreased at 24 h. (C) The distribution of the cell cycle was detected by flow cytometry with PI staining. The percentages of the G1, S, and G2/M phases were calculated.

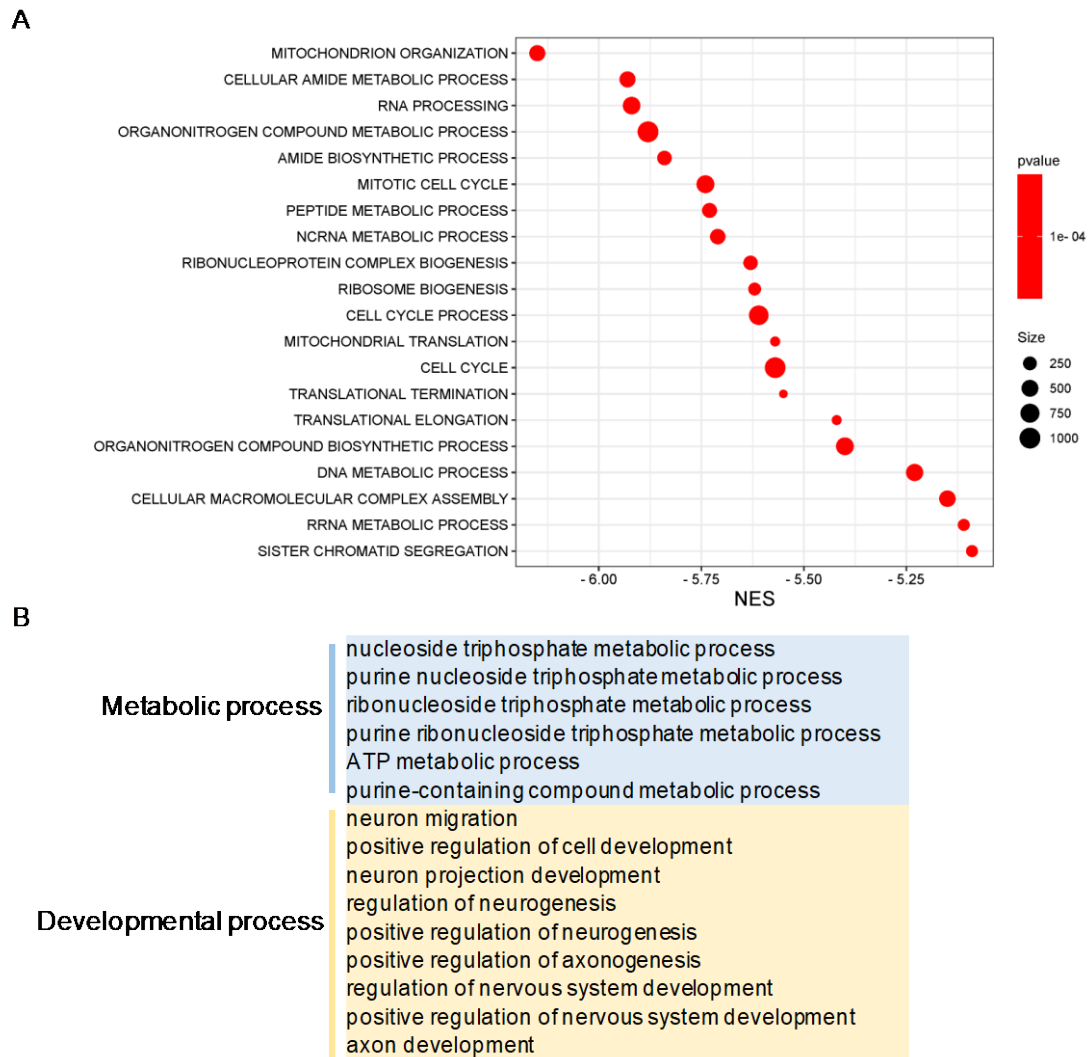


Figure S4. *Mcph1* may cause cell cycle-dependent metabolic changes (A) The biological process of the DEGs in the results of Nathalie Journiac et al[1]. The figure shows the top 20 biological processes, including metabolism-related and cell cycle related (B) We used metascape to perform the same analysis on our DEGs and their DEGs[1]. The intersection of the two different genes focuses on metabolism and development.

Table S1. Summary of intersecting genes information

NO.	Intersecting Gene	log2FoldChange	P adj	Site Count*	Remark
1	<i>Ldha</i>	0.53258291	1.40E-06	4	<i>Ldha</i> can affect neuronal excitability[2].
2	<i>Tcf7l1</i>	0.534401708	0.047987897	4	-
3	<i>Vegfa</i>	0.584031025	0.00033305	3	<i>Vegfa</i> affects embryonic angiogenesis[3].
4	<i>Hey1</i>	0.460385188	0.029574475	3	-
5	<i>Aldoa</i>	0.310349563	0.047987897	3	<i>Aldoa</i> is related to cytoskeletal development and function[4].
6	<i>Slc16a3</i>	1.274481753	9.16E-06	2	Deletion of <i>Slc16a3</i> is likely to result in decreased lactate in the embryo's blood and a decrease in the embryo's growth[5, 6].
7	<i>Tpi1</i>	0.444301423	8.25E-05	2	-
8	<i>Eno1</i>	0.436598496	0.000176167	2	<i>Eno1</i> inactivation causes delayed brain development[7, 8].
9	<i>Crlf2</i>	0.884397108	0.000936195	2	<i>Crlf2</i> is involved in neuronal signaling, and patients have symptoms of microcephaly[9].
10	<i>Fn1</i>	0.378091826	0.001353569	2	Mouse embryos lacking <i>Fn1</i> have defects in mesoderm and neural tube development[10].
11	<i>Lcp1</i>	0.521461793	0.002515204	2	-
12	<i>Pkm</i>	0.338713642	0.003225653	2	<i>Pkm</i> is related to synaptic plasticity[2, 11].
13	<i>Rabgap1l</i>	-0.342916124	0.00422707	2	<i>Rabgap1l</i> is differentially methylated in Zika-induced microcephaly[12].
14	<i>Spp1</i>	1.198484624	0.011059793	2	-
15	<i>Hivep2</i>	-0.348226286	0.016053953	2	Mutations in the <i>Hivep2</i> gene cause developmental delay/intellectual disability[13].
16	<i>Nxn</i>	0.40763114	0.01743748	2	-
17	<i>Eif2s3y</i>	-0.42710408	0.019913806	2	<i>Eif2s3y</i> can inhibit the pluripotency state of embryonic stem cells in mice[8].
18	<i>Gm37844</i>	-0.88957664	0.021387194	2	-
19	<i>Inhba</i>	-0.8904405	0.046788412	2	-
20	<i>Ddit4</i>	0.469127086	0.047987897	2	-
21	<i>Satb2</i>	-0.72840142	2.19E-06	1	Mutations in the <i>Satb2</i> gene cause developmental delay/intellectual disability[6].
22	<i>Sla</i>	-0.603553268	0.000122916	1	-
23	<i>9130024F11</i> <i>Rik</i>	-0.710580795	0.00033305	1	-

Table S1. (Continued)

NO.	Intersecting Gene	log2FoldChange	P adj	Site Count*	Remark
24	<i>Mef2c</i>	-0.414368576	0.001570872	1	-
25	<i>Fam49a</i>	-0.342475004	0.002471334	1	-
26	<i>Alas2</i>	-1.196964327	0.002515204	1	-
27	<i>Crb2</i>	0.509924247	0.002920479	1	-
28	<i>Pfkl</i>	0.373654061	0.003110287	1	-
29	<i>Dlk1</i>	0.556029226	0.003225653	1	-
30	2610318N02 <i>Rik</i>	0.754241912	0.003260079	1	-
31	<i>Dok6</i>	-0.398228958	0.004321018	1	-
32	<i>Pgk1</i>	0.352225734	0.006154432	1	-
33	<i>Gnai1</i>	-0.359240218	0.007113982	1	-
34	<i>Dll3</i>	0.612639822	0.009379012	1	-
35	<i>Cdkn1c</i>	0.335603094	0.011059793	1	Mutations in the <i>Cdkn1c</i> gene cause developmental delay/intellectual disability[14].
36	<i>Fut10</i>	-0.926136173	0.011932979	1	-
37	<i>Neurog2</i>	0.401683379	0.011932979	1	-
38	<i>Ccnd3</i>	0.44717396	0.019519963	1	-
39	1700048O20 <i>Rik</i>	0.804420254	0.019519963	1	-
40	<i>Slc2a1</i>	0.399721042	0.030723065	1	-
41	<i>Nr4a3</i>	-0.425268553	0.030892622	1	-
42	<i>Kcnn1</i>	0.55633904	0.031913054	1	-
43	<i>Gria2</i>	-0.279563023	0.032771223	1	-
44	<i>Tmem132b</i>	-0.320917809	0.032952499	1	-
45	<i>Mctp1</i>	-0.698451514	0.03545619	1	-
46	<i>Neurog1</i>	0.770946401	0.040967524	1	-
47	<i>Rhbdl3</i>	0.390346143	0.043599623	1	-
48	<i>Necab1</i>	-0.486480137	0.046488149	1	-
49	<i>Mpped1</i>	-0.309551635	0.047922476	1	-
50	<i>Ntrk3</i>	-0.277612814	0.047987897	1	-
51	<i>Zic3</i>	0.368405971	0.049918514	1	-

* Count of E2F1 binding sequence

Table S2. Primers used in this study

Gene name	Primer	Sequence (5'to 3')
<i>Satb2</i>	Forward Primer	GAGATGAGTTGAAGAGGGCTAGTG
	Reverse Primer	CCCTGTGTGCGGTTGAAT
<i>Ldha</i>	Forward Primer	AGCGTACCCGTGATGCTAAC
	Reverse Primer	CAGGGTTGGCAGATCGACAT
<i>Aldoa</i>	Forward Primer	AACGGTCACACACTTCGTCG
	Reverse Primer	TACTTTCCTTGACAAGCGAGGC
<i>Vegfa</i>	Forward Primer	GCAGGCTGCTGTAACGATGAA
	Reverse Primer	TGCTTTCTCCGCTCTGAACAA
<i>Eif2s3y</i>	Forward Primer	ATCTTGTCTCAACCTCAGACT
	Reverse Primer	TTCTTTAGCCTGGCTTTCTTTCA
<i>Hivep2</i>	Forward Primer	CTCCTTTCTCCTCCCGAGCG
	Reverse Primer	GATCCCGAGGCTACTGGCTG
<i>Rabgap1l</i>	Forward Primer	ACTGGGAATCTTCATGAGAAGCTGA
	Reverse Primer	TCACATTACTGTGCTTGATACACCA
<i>Pkm</i>	Forward Primer	GGAGGAGGAATGCAGGACTGG
	Reverse Primer	GGAGTGCACAAGAAGTGGGGA
<i>Fn1</i>	Forward Primer	AACAAGAGACCACTGGCACC
	Reverse Primer	AGAGGATTGCTTTCCCTGCC
<i>Eno1</i>	Forward Primer	CGACTGTATGGAATCCAAGGCA
	Reverse Primer	CCAGCTTTGCAGACAGCCA
<i>Crlf2</i>	Forward Primer	GCAGGTGATGTCACAGTCGT
	Reverse Primer	GCGCTGCCTAGCCTTAAACA
<i>Slc16a3</i>	Forward Primer	GGCTGTTTTATCATCACGGGT
	Reverse Primer	GTGTCGCTGTAGCCAATCCC
<i>Cdkn1c</i>	Forward Primer	AGCTGAAGGACCAGCCTCTCTC
	Reverse Primer	ACGTCGTTCGACGCCTTGTTCT
<i>mGapdh</i>	Forward Primer	GCACAGTCAAGGCCGAGAAT
	Reverse Primer	GCCTTCTCCATGGTGGTGAA
1000-Cdkn1c-pGL3	Forward Primer	TTACGCGTGCTAGCCCCGGGCTCGAGGTTG
		GAGGGCTAGATGGGGAACCTT
	Reverse Primer	ACCTTAGTTGGCTGGAAGTAGTTATGCTA
2000-Cdkn1c-pGL3		GAAAAG
	Forward Primer	TTACGCGTGCTAGCCCCGGGCTCGAGGGGG
	Reverse Primer	GTCGAATATGGCCTGA
		atgcagatcgagatctcgagCTGCACCAACTGATT
		AGGGCTT

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