

A) The primer sequences used for genotyping and expected size of PCR products are as follows:

1. **OGT**: Mutant: 487 bp; WT: 338 bp  
 F- CAT CTC TCC AGC CCC ACA AAC TG  
 R- GAC GAA GCA GGA GGG GAG AGC AC
2. **ApoE**: Mutant: 245 bp; WT: 155 bp  
 Common GCC TAG CCG AGG GAG AGC CG  
 Wild type Reverse TGT GAC TTG GGA GCT CTG CAG C
3. **Myh11-Cre<sup>ERT2</sup>**: Transgene: 287 bp; Internal positive control: 180 bp  
 Tran F- TGA CCC CAT CTC TTC ACT CC  
 Tran R- AGT CCC TCA CAT CCT CAG GTT  
 Positive control F- CAG CCA ACT TTA CGC CTA GC  
 Positive control R- TCT CAA GAT GGA CCT AAT ACG G

B) PCR Cycling conditions

1. For Cre<sup>tg</sup> and OGT<sup>F</sup>:

Temperature	Time	Repetitions
94°C	5 min	
94°C	20s	10X (lower 1C each cycle)
65-55°C	15s	
68°C	10s	
94°C	15s	
60°C	15s	30X
72°C	10s	
72°C	2 min	
15°C	hold	

2. For ApoE:

Temperature	Time	Repetition
94°C	4 min	
94°C	30s	35X
69°C	1 min	
72°C	1 min	
72°C	2 min	
15°C	hold	

**Supplementary Figure S1.** Primer sequences and PCR conditions for mice genotyping. A) Shown are the primer sequences used for OGT, ApoE and Myh11-Cre<sup>tg</sup> genotyping, B) PCR cycling conditions used for mice genotyping.

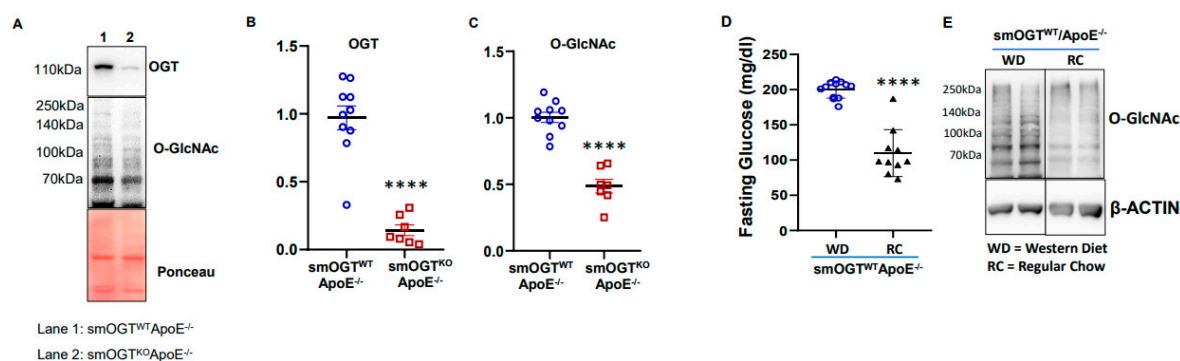
### A. Primer sequences utilized for qPCR reactions:

	Gene	Forward	Reverse
1	<i>Acta2</i>	5'- CATCTTTCATTGGGATGGAG- 3'	5'- TTAGCATAGAGATCCTTCCTG- 3'
3	<i>Myocd</i>	5'- GATGGGCTCTCTCCAGATCAG- 3'	5'- GGCTGCATCATTCTTGTCACTT- 3'
4	<i>Myh11</i>	5' TATCTTCTGGAAAAGTCTAGGG- 3'	5'- AAAGCAGGTCACCTTTTCATC- 3'
6	<i>36b4</i>	5'- AGATTCTGGGATATGCTGTTGG – 3'	5'- TCGGGTCCTAGACCAGTGTTC – 3'
7	<i>18s</i>	5'- TTGACGGAAGGGCACCACCAG- 3'	5'- GCACCACCACCCACGGAATCG- 3'
8	<i>OGT</i>	5'- GACGCAACCAAACCTTTGCAGT-3'	5'-TCAAGGGTGACAGCCTTTTCA- 3'
9	<i>Cnn1</i>	5'- TGCTGAAGTAAAGAACAAGC- 3'	5'- CATTGACCTTCTTCACAGAAC- 3'
10	<i>Il6</i>	5'- TAGTCCTTCCTACCCCAATTTC- 3'	5'- TTGGTCCTTAGCCACTCCTTC- 3'
11	<i>Il1β</i>	5'- TGTAATGAAAGACGGCACACC- 3'	5'- TCTTCTTTGGGTATTGCTTGG- 3'

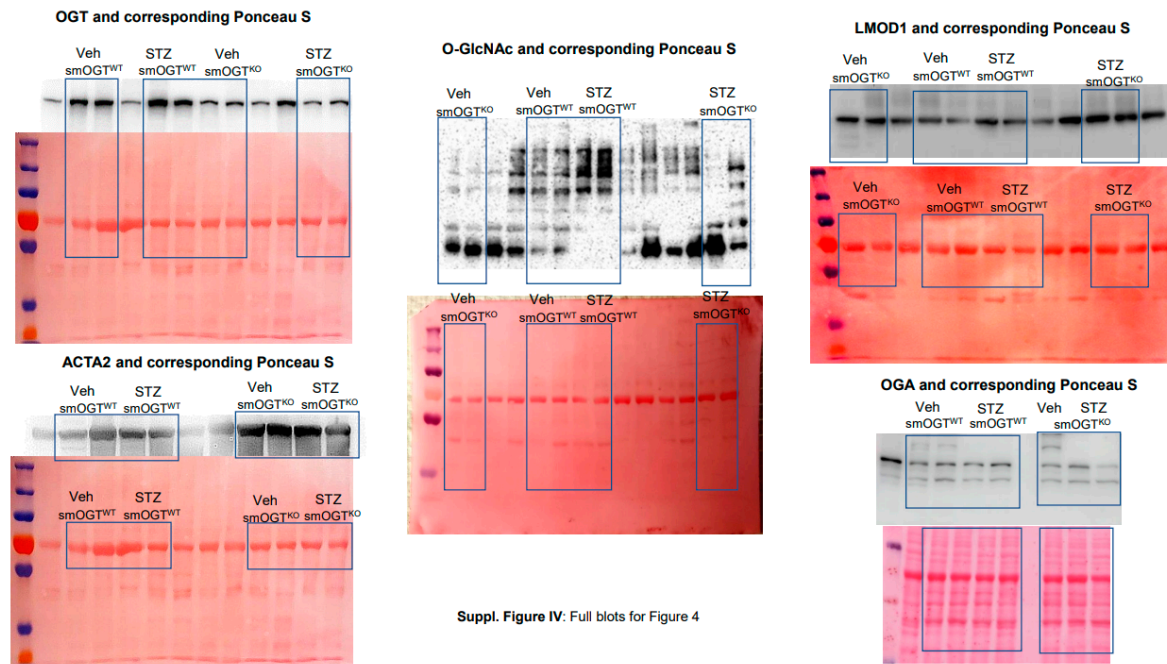
### B. qPCR cycling conditions

Thermal Cycler Profile			
Stage	Repetitions	Temperature	Time (mins)
1	1	50.0 °C	2:00
2	1	95.0 °C	10:00
3	40	95.0 °C	0:15
		60.0 °C	1:00
4 (Dissociation)	1	95.0 °C	0:15
		60.0 °C	0:20
		95.0 °C	0:15
		60.0 °C	0:15

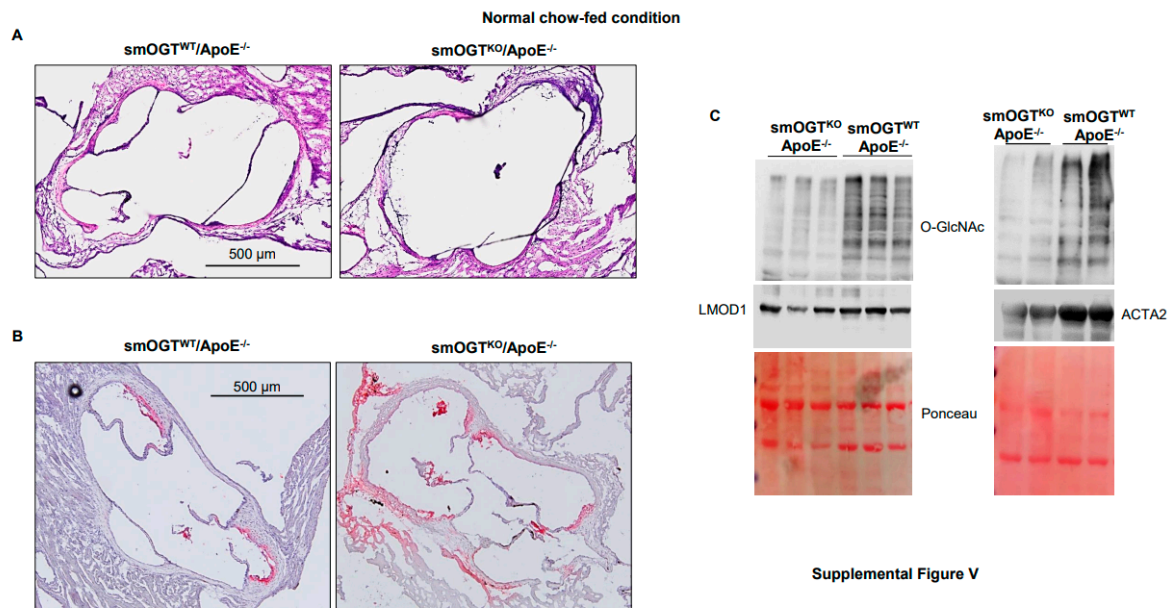
**Supplementary Figure S2.** Real-time PCR conditions. A) qPCR primer sequences and B) qPCR cycling conditions.



**Supplementary Figure S3.** (A-C) Validation of SMC-specific OGT deletion in ApoE<sup>-/-</sup> mice. Aortic lysates derived from smOGT<sup>WT</sup>ApoE<sup>-/-</sup> and smOGT<sup>KO</sup>ApoE<sup>-/-</sup> mice were subjected to immunoblotting. Shown are representative immunoblots and corresponding summary graphs depicting fold-change in protein expression normalized to Ponceau S for total protein staining. D-E) Western diet feeding increases fasting blood glucose levels accompanied with increased O-GlcNAc protein expression in the aortic vessels of smOGT<sup>WT</sup>ApoE<sup>-/-</sup> mice. n=7-11 mice per group, \*\*\*\*p<0.0001.



**Supplementary Figure S4.** Full blots for Figure 4 images. Blue box outlines the bands used in the corresponding images.



**Supplementary Figure S5.** Plaque area, lipid burden and SM contractile marker expression in normal chow-fed smOGT<sup>WT</sup>ApoE<sup>-/-</sup> and smOGT<sup>KO</sup>ApoE<sup>-/-</sup> mice. Shown are representative A) hematoxylin & eosin (H&E)- and B) Oil red O-stained images of aortic root sections derived from 14-wks old normal chow-fed smOGT<sup>WT</sup>ApoE<sup>-/-</sup> and smOGT<sup>KO</sup>ApoE<sup>-/-</sup> mice. Representative immunoblots of aortic lysates from normal chow-fed smOGT<sup>WT</sup>ApoE<sup>-/-</sup> and smOGT<sup>KO</sup>ApoE<sup>-/-</sup> mice depicting O-GlcNAc, LMOD1 and ACTA2 expression are shown in panel C); Ponceau S shows total protein staining.

**Supplementary material:** The following Supplemental data are included in this manuscript: Supplementary Figures S1-S5.