



Communication

Novel Risk Variants in the Oxytocin Receptor Gene (*OXTR*) Possibly Linked to and Associated with Familial Type 2 Diabetes

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Abstract: The oxytocin system is well-known for its role in social bonding and reproduction. Recently, the oxytocin system was found to play other metabolic roles such as regulation of food intake, peripheral glucose uptake, and insulin sensitivity. Variants in *OXTR* gene have been associated with overeating, increased cardiovascular risk, and type 2 diabetes (T2D). We tested 20 microarray-derived single nucleotide polymorphisms in the *OXTR* gene in 212 Italian families with rich family history for T2D and found four novel and one previously reported variant suggestively significant for linkage and association with the risk of T2D. Our study has shed some light into the genetics of susceptibility to T2D at least in Italian families.

Keywords: oxytocin; OXT; oxytocin receptor; *OXTR*; type 2 diabetes; T2D; single nucleotide polymorphisms; SNPs; linkage; linkage disequilibrium; association



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1. Background

Oxytocin (OXT) is a neuropeptide hormone produced by the hypothalamus and secreted by the posterior pituitary gland [1]. Oxytocin notably plays important roles in social bonding and reproduction [2,3]. Recent evidence suggests that oxytocin also plays other metabolic roles such as regulation of food intake [4], peripheral glucose uptake [5], insulin sensitivity [6], β -cell function and survival, and insulin secretion [7], which prompted investigating the implication of oxytocin pathway in type 2 diabetes (T2D). Of note, administration of oxytocin in humans improved insulin sensitivity and responsiveness of pancreatic β -cells [8]. The oxytocin receptor (*OXTR*) is a G-protein coupled receptor that mediates the central and peripheral actions of oxytocin [9]. It is encoded by the oxytocin receptor gene (*OXTR*) and is expressed in a variety of central and peripheral tissues such as the brain, heart, kidneys and pancreas [10]. *OXTR*^{-/-} knockout mice show insulin insensitivity and impaired glucose tolerance and lack protection from cell death induced by cytotoxic and metabolic stress [11], and variants in *OXTR* gene have been associated in humans with overeating [12], increased cardiovascular risk [13] and T2D [14], and they possibly act through regulation of glucose tolerance and insulin sensitivity [15,16]. The association of *OXTR*-risk variants with T2D has been found in population-based case control studies [14], but familial studies which are powerful in detecting linkage signals, not common and rare variants, and inheritance models underlying the conferred risk, are still lacking. In this study, we aimed at analyzing *OXTR* gene variants for familial linkage with T2D.

2. Results and Discussion

Five variants (rs237887, rs60345038, rs77943865, rs115356575 and rs4686302) were suggestively significant for linkage to and linkage disequilibrium (LD) with T2D ($p < 0.05$) (Figure 1) across different inheritance models (Table 1). All variants were independent (i.e., no LD blocks were found). With the exception of (rs237887), all variants are novel and have not been reported before with T2D or other related phenotype. The non-risk allele (G) of the variant (rs237887) was part of a haplotype reported to be weakly associated with the protection against T2D in a case-control study [14]. We performed in silico functional predictions for the suggestively significant single nucleotide polymorphisms (SNPs) in our study (pathogenicity [SIFT] [17], splicing [SpliceAI] [18], transcription-factor binding [SNPnexus] [19] and SNP2TFBS [20], regulatory potential [RegulomeDB] [21], and miRNA binding [mirSNP] [22]) and predicted that all T2D risk variants were associated with repressed chromatin state in the endocrine pancreas which potentially leads to negative expression of the *OXTR* gene [21]. We found no functional predictions related to affected binding of transcription factors, miRNAs or impaired splicing. These findings however, are consistent with the T2D-related phenotypes (decreased insulin sensitivity and impaired glucose tolerance) observed in *OXTR*^{-/-} knockout mice [11]. The results of our study indicate that *OXTR*-variants may confer risk not only to sporadic cases [14] but also to familial T2D, at least in Italian families. The *OXTR* gene is expressed in both the brain and the endocrine pancreas [10]; its implication in T2D could be mediated by its role in appetite [12] and/or insulin sensitivity [15,16]. However, the variants in our study are only suggestively significant since correction for multiple comparison testing has not been applied. Therefore, functional and replication studies in diverse ethnic groups are still needed to validate these results.

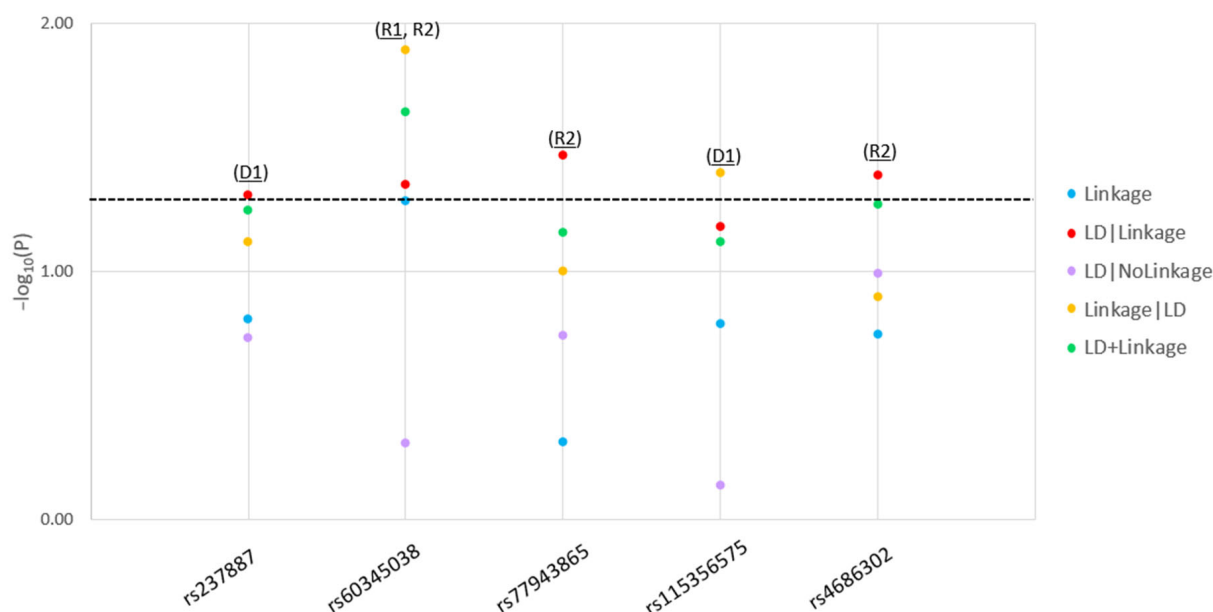


Figure 1. T2D *OXTR* risk single nucleotide polymorphisms (SNPs) linkage and linkage-disequilibrium (LD) analysis results. **Legend:** For each significant risk single nucleotide polymorphism (SNP) in the *OXTR* gene, we present the $-\log_{10}(P)$ as a function of each test statistic (Linkage, linkage disequilibrium [LD] | Linkage, LD | NoLinkage, Linkage | LD, and LD+Linkage) and label the inheritance model: D1: dominant, complete penetrance, R1: recessive, complete penetrance, R2: recessive, incomplete penetrance. For each SNP we present the most significant test statistics across the significant models.

Table 1. *OXTR* risk single nucleotide polymorphisms (SNPs) linked/in linkage-disequilibrium (LD) to/with type 2 diabetes (T2D).

Model ¹	SNP	Position	Ref	Alt	Risk Allele	Consequence	RAF ²	P ³	Reported in T2D?
D1	rs237887	8755356	G	A	A	Intronic	0.54	0.04	Yes [14]
<u>R1</u> , R2	rs60345038	8760830	C	T	C	Intronic	0.62	0.01	Novel
R2	rs77943865	8762293	G	A	G	Intronic	0.95	0.03	Novel
D1	rs115356575	8764840	G	A	G	Intronic	0.96	0.04	Novel
R2	rs4686302	8767536	C	T	C	Missense	0.89	0.04	Novel

Legend. ¹ Models: D1: dominant, complete penetrance, R1: recessive, complete penetrance, R2: recessive, incomplete penetrance. ² RAF: risk allele frequency. ³ P: *p*-value of the most significant test statistics. The most significant model is underlined.

3. Methods

3.1. Patients

We recruited 1156 subjects from 212 Italian families with rich family history of T2D diagnosed following the National Diabetes Data Group Criteria (22): “Presence of the classical signs and/or symptoms of diabetes plus elevated hyperglycemia, or by elevated fasting plasma glucose ≥ 140 mg/dL”. This was followed by reviewing the American Diabetes Association criteria: fasting glycemia at 126 mg/dL or higher in at least two measurements and/or random glycemia of at least 200 mg/dL or higher with symptoms, and/or at least 200 mg/dL or higher 2 h after an oral glucose tolerance test of 75 mg of glucose, after excluding secondary causes of diabetes (e.g., pancreatectomy). According to these criteria, 650 patients were diagnosed with T2D. The average male:female ratio was 1.04. The age of onset of T2D ranged from 7 to 81 years with a mean age of 47.8.

3.2. Genetic Testing and Analysis

We tested 20 microarray-derived SNPs in the *OXTR* gene (Supplementary Table S1). The SNPs were amplified using Affymetrix SNP array. PLINK tool was used to exclude Mendelian and genotyping errors [23]. Using Pseudomarker [24], we performed 2-point parametric linkage and linkage-disequilibrium (LD) analyses with T2D, initially testing the recessive model with incomplete penetrance (R2). Subsequently, we ran a secondary analysis with the models of recessive complete penetrance (R1), dominant complete penetrance (D1) and dominant incomplete penetrance (D2). Each SNP was tested for 1. Linkage, 2. LD|Linkage, 3. LD|NoLinkage, 4. Linkage|LD and 5. LD+Linkage. We report in Supplementary Table S1 the *p* value of the following analyses: LD/linkage, the most important, as it tests for LD given the presence of linkage; linkage/LD, which tests for linkage given the presence of LD; and LD + linkage tests, which is a combinatorial test. Presence or absence of LD blocks was imputed according to LD correlations from SNPs available in the Tuscany Italian population from the 1000 Genomes Project (<https://www.internationalgenome.org/data-portal/population/TSI> [accessed on 15 September 2022]). SNPs with $r^2 \geq 0.9$ were considered “in LD” (Supplementary Table S2). The study was approved by the Bios Ethical Committee.

Supplementary Materials: The following supporting information can be downloaded at: <https://www.mdpi.com/article/10.3390/ijms24076282/s1>.

Author Contributions: C.G. conceived and supervised the project, including statistical analysis and manuscript drafting. M.A. helped with the bioinformatic analysis, literature search, and manuscript drafting. R.W. critically helped in data interpretation and critical revision of the manuscript. All authors have approved the final manuscript. All authors have read and agreed to the published version of the manuscript.

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Institutional Review Board Statement: The study was conducted in accordance with the Declaration of Helsinki guidelines, and approved by the Bios Ethical Committee, which defined the research “non-human subjects research” as the data were fully deidentified (#311708, 15 February 2017).

Data Availability Statement: The study data are available on reasonable request; due to lacking specific patients’ consent and privacy restrictions, they are not publicly available.

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Conflicts of Interest: The authors have declared that they have no conflict of interest.

References

1. Baribeau, D.A.; Anagnostou, E. Oxytocin and vasopressin: Linking pituitary neuropeptides and their receptors to social neurocircuits. *Front. Neurosci.* **2015**, *9*, 335. [[CrossRef](#)] [[PubMed](#)]
2. Jones, C.; Barrera, I.; Brothers, S.; Ring, R.; Wahlestedt, C. Oxytocin and social functioning. *Dialogues Clin. Neurosci.* **2017**, *19*, 193–201. [[CrossRef](#)] [[PubMed](#)]
3. Veening, J.G.; de Jong, T.R.; Waldinger, M.D.; Korte, S.M.; Olivier, B. The role of oxytocin in male and female reproductive behavior. *Eur. J. Pharmacol.* **2015**, *753*, 209–228. [[CrossRef](#)] [[PubMed](#)]
4. Onaka, T.; Takayanagi, Y. Role of oxytocin in the control of stress and food intake. *J. Neuroendocrinol.* **2019**, *31*, e12700. [[CrossRef](#)]
5. Lee, E.S.; Uhm, K.O.; Lee, Y.M.; Kwon, J.; Park, S.H.; Soo, K.H. Oxytocin stimulates glucose uptake in skeletal muscle cells through the calcium-CaMKK-AMPK pathway. *Regul. Pept.* **2008**, *151*, 71–74. [[CrossRef](#)]
6. Ding, C.; Leow, M.K.; Magkos, F. Oxytocin in metabolic homeostasis: Implications for obesity and diabetes management. *Obes. Rev.* **2019**, *20*, 22–40. [[CrossRef](#)]
7. Mohan, S.; Khan, D.; Moffett, R.C.; Irwin, N.; Flatt, P.R. Oxytocin is present in islets and plays a role in beta-cell function and survival. *Peptides* **2018**, *100*, 260–268. [[CrossRef](#)]
8. Zhang, H.; Wu, C.; Chen, Q.; Chen, X.; Xu, Z.; Wu, J.; Cai, D. Treatment of obesity and diabetes using oxytocin or analogs in patients and mouse models. *PLoS ONE* **2013**, *8*, e61477. [[CrossRef](#)]
9. Chatterjee, O.; Patil, K.; Sahu, A.; Gopalakrishnan, L.; Mol, P.; Advani, J.; Mukherjee, S.; Christopher, R.; Prasad, T.S. An overview of the oxytocin-oxytocin receptor signaling network. *J. Cell Commun. Signal.* **2016**, *10*, 355–360. [[CrossRef](#)]
10. Kimura, T.; Saji, F.; Nishimori, K.; Ogita, K.; Nakamura, H.; Koyama, M.; Murata, Y. Molecular regulation of the oxytocin receptor in peripheral organs. *J. Mol. Endocrinol.* **2003**, *30*, 109–115. [[CrossRef](#)]
11. Watanabe, S.; Wei, F.Y.; Matsunaga, T.; Matsunaga, N.; Kaitsuka, T.; Tomizawa, K. Oxytocin Protects against Stress-Induced Cell Death in Murine Pancreatic β -Cells. *Sci. Rep.* **2016**, *6*, 25185.
12. Davis, C.; Patte, K.; Zai, C.; Kennedy, J.L. Polymorphisms of the oxytocin receptor gene and overeating: The intermediary role of endophenotypic risk factors. *Nutr. Diabetes* **2017**, *7*, e279. [[CrossRef](#)]
13. Jacodino, C.B.; Borges, C.A.; Rosemberg, L.S.; da Silva, I.G.; da Luz Correa, B.; Valle Gottlieb, M.G. Association of oxytocin levels and oxytocin receptor gene polymorphism (rs2254298) with cardiovascular risk factors in Brazilian elderly from Primary Health Care. *Arch. Gerontol. Geriatr.* **2019**, *84*, 103903. [[CrossRef](#)] [[PubMed](#)]
14. Salonen, J.T.; Uimari, P.; Aalto, J.M.; Pirskanen, M.; Kaikkonen, J.; Todorova, B.; Hyppönen, J.; Korhonen, V.P.; Asikainen, J.; Devine, C.; et al. Type 2 diabetes whole-genome association study in four populations: The DiaGen consortium. *Am. J. Hum. Genet.* **2007**, *81*, 338–345.
15. Chang, H.H.; Chang, W.H.; Chi, M.H.; Peng, Y.C.; Huang, C.C.; Yang, Y.K.; Chen, P.S. The OXTR Polymorphism Stratified the Correlation of Oxytocin and Glucose Homeostasis in Non-Diabetic Subjects. *Diabetes Metab. Syndr. Obes.* **2019**, *12*, 2707–2713. [[CrossRef](#)]
16. Saravani, R.; Esmaeeli, E.; Kordi Tamendani, M.; Nejad, M.N. Oxytocin Receptor Gene Polymorphisms in Patients with Diabetes. *Gene Cell Tissue* **2015**, *2*, e60171. [[CrossRef](#)]
17. Ng, P.C.; Henikoff, S. SIFT: Predicting amino acid changes that affect protein function. *Nucleic Acids Res.* **2003**, *31*, 3812–3814. [[CrossRef](#)] [[PubMed](#)]
18. Jaganathan, K.; Kyriazopoulou Panagiotopoulou, S.; McRae, J.F.; Darbandi, S.F.; Knowles, D.; Li, Y.I.; Kosmicki, J.A.; Arbelaez, J.; Cui, W.; Schwartz, G.B.; et al. Predicting Splicing from Primary Sequence with Deep Learning. *Cell* **2019**, *176*, 535–548.e24. [[CrossRef](#)] [[PubMed](#)]
19. Dayem Ullah, A.Z.; Oscanoa, J.; Wang, J.; Nagano, A.; Lemoine, N.R.; Chelala, C. SNPnexus: Assessing the functional relevance of genetic variation to facilitate the promise of precision medicine. *Nucleic Acids Res.* **2018**, *46*, W109–W113. [[CrossRef](#)] [[PubMed](#)]

20. Kumar, S.; Ambrosini, G.; Bucher, P. SNP2TFBS—A database of regulatory SNPs affecting predicted transcription factor binding site affinity. *Nucleic Acids Res.* **2017**, *45*, D139–D144. [[PubMed](#)]
21. Boyle, A.P.; Hong, E.L.; Hariharan, M.; Cheng, Y.; Schaub, M.A.; Kasowski, M.; Karczewski, K.J.; Park, J.; Hitz, B.C.; Weng, S.; et al. Annotation of functional variation in personal genomes using RegulomeDB. *Genome Res.* **2012**, *22*, 1790–1797. [[CrossRef](#)] [[PubMed](#)]
22. Liu, C.; Zhang, F.; Li, T.; Lu, M.; Wang, L.; Yue, W.; Zhang, D. MirSNP, a database of polymorphisms altering miRNA target sites, identifies miRNA-related SNPs in GWAS SNPs and eQTLs. *BMC Genom.* **2012**, *13*, 661. [[CrossRef](#)] [[PubMed](#)]
23. Purcell, S.; Neale, B.; Todd-Brown, K.; Thomas, L.; Ferreira, M.A.; Bender, D.; Maller, J.; Sklar, P.; de Bakker, P.I.; Daly, M.J.; et al. PLINK: A tool set for whole-genome association and population-based linkage analyses. *Am. J. Hum. Genet.* **2007**, *81*, 559–575. [[CrossRef](#)] [[PubMed](#)]
24. Hiekkalinna, T.; Schaffer, A.A.; Lambert, B.; Norrgrann, P.; Goring, H.H.; Terwilliger, J.D. PSEUDOMARKER: A powerful program for joint linkage and/or linkage disequilibrium analysis on mixtures of singletons and related individuals. *Hum. Hered.* **2011**, *71*, 256–266. [[CrossRef](#)] [[PubMed](#)]

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