



Article

[4+2]-Cycloaddition to 5-Methylidene-Hydantoins and 5-Methylidene-2-Thiohydantoins in the Synthesis of Spiro-2-Chalcogenimidazolones

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Abstract: Novel hydantion and thiohydantoin-based spiro-compounds were prepared via the Diels–Alder reactions between 5-methylidene-hydantoins or 5-methylidene-2-thiohydantoins and 1,3-dienes (cyclopentadiene, cyclohexadiene, 2,3-dimethylbutadiene, isoprene). It was shown that the cycloaddition reactions proceed regioselectively and stereoselectively with the formation of exo-isomers in the reactions with cyclic dienes and the less sterically hindered products in the reactions with isoprene. Reactions of methylideneimidazolones with cyclopentadiene proceed via co-heating the reactants; reactions with cyclohexadiene, 2,3-dimethylbutadiene, and isoprene require catalysis by Lewis acids. It was demonstrated that ZnI_2 is an effective catalyst in the Diels–Alder reactions of methylidene-thiohydantoins with non-activated dienes. The possibility of alkylation and acylation of the obtained spiro-hydantoins at the N(1) nitrogen atoms with $PhCH_2Cl$ or Boc_2O and the alkylation of the spiro-thiohydantoins at the S atoms with MeI or $PhCH_2Cl$ in high yields have been demonstrated. The preparative transformation of spiro-thiohydantoins into corresponding spiro-hydantoins in mild conditions by treating with 35% aqueous H_2O_2 or nitrile oxide has been carried out. The obtained compounds show moderate cytotoxicity in the MTT test on MCF7, A549, HEK293T, and VA13 cell lines. Some of the tested compounds demonstrated some antibacterial effect against *Escherichia coli* (*E. coli*) BW25113 DTC-pDualrep2 but were almost inactive against *E. coli* BW25113 LPTD-pDualrep2.

Keywords: spiro-compounds; hydantoins; 2-thiohydantoins; Diels–Alder reaction; [4+2]-cycloaddition; cytotoxicity; antibacterial activity



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1. Introduction

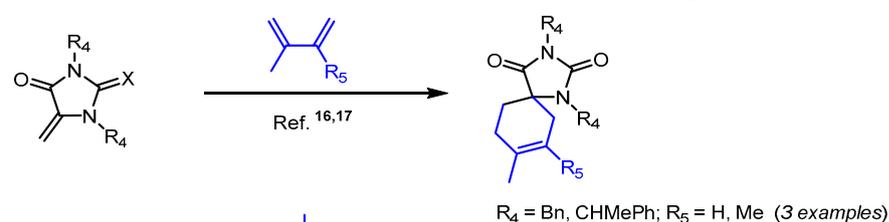
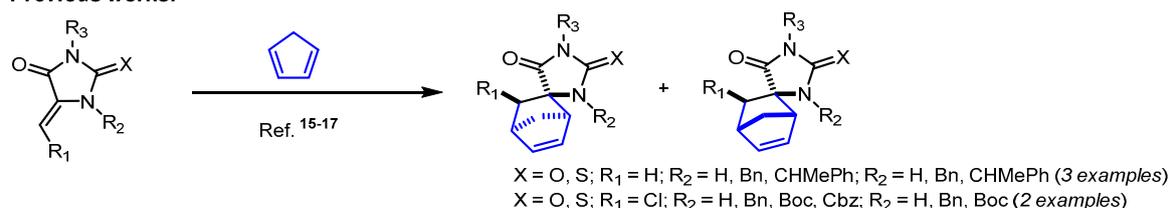
Introducing conformationally rigid lipophilic moieties into a molecule is a common approach in the medicinal chemistry [1–4] that may lead to significant increases in biological activity [5–8]. One of the convenient methods for the synthesis of such conformationally rigid polycyclic derivatives is the Diels–Alder reaction of functionalized alkenes containing exocyclic C=C bonds with dienes. Dienes variation makes it possible to fine-tune the structure of the resulting lipophilic framework [9–11], and the presence of a C=C double bond in the [4+2]-cycloaddition adduct opens the way for further modifications of the resulting molecules.

We have recently shown that spiro-2-chalcogenimidazolones, obtained by 1,3-dipolar-cycloaddition reactions to 5-methylidene-substituted hydantoins, as well as their 2-thio- and 2-seleno-analogs, possess high cytotoxicity probably due to their ability to inhibit the p53-MDM2 proteins interaction [12–14]. However, there were no general methods for the synthesis of 5-methylidene-imidazol-4-one derivatives containing spiro-linked 5- and 6-membered rings described previously, and these compounds have not been previously studied for biological activity.

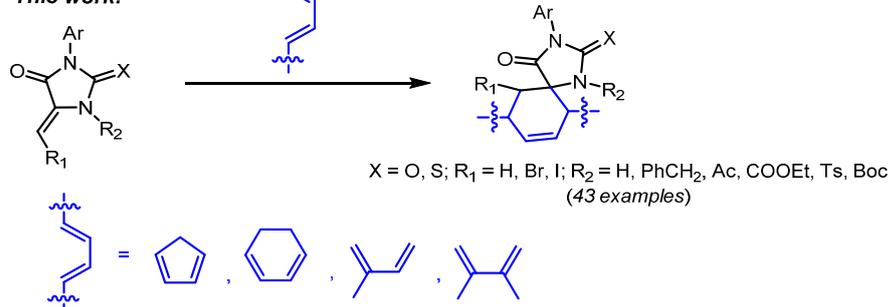
Diels–Alder reactions of 5-methylene-substituted imidazolones are described in the literature mainly as the example of highly reactive cyclopentadiene [15–17] and as single examples of the reactions with isoprene and 2,3-dimethylbutadiene [16,17] (Scheme 1). In the present article, a systematic study of [4+2]-cycloaddition reactions of cyclopentadiene, cyclohexadiene, 2,3-dimethylbutadiene, and isoprene with 5-methylene-2-chalcogenimidazolone derivatives, both N-unsubstituted and containing substituents of various natures at nitrogen atoms, is carried out. In this work, we first demonstrated the possibility of methylenethiohydantoin interaction with low active conjugated dienes, which makes it possible to obtain spirocyclic imidazolones containing various lipophilic frameworks. In addition, the possibility of post-synthetic transformations of the formed [4+2]-cycloaddition products using alkylating and acylating agents was demonstrated; these reactions can be an alternative method for the synthesis of those spiro-imidazolones that are formed in the reactions with dienes in low yields or as the inseparable diastereomers mixtures.

Various types of biological activities have been previously described for hydantoin and their spiro derivatives [18,19]. In this case, if, for example, anticonvulsants require low toxicity, then inhibition of the growth of bacteria and tumor cells is determined by the cytotoxic effect. Therefore, for the initial assessment of the biological effect on cells of various types, a number of compounds synthesized in this work were tested on cytotoxicity and antibacterial activity on several human cell lines of various etiologies and several strains of *E. coli*.

Previous works:



This work:



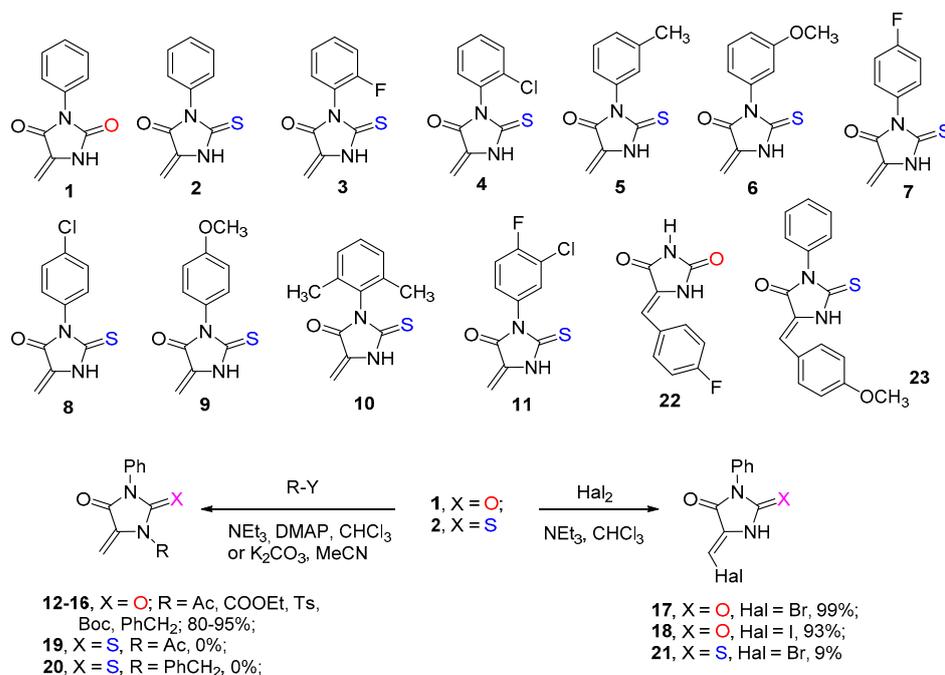
Scheme 1. Diels–Alder reactions with methylenethiohydantoin and methylenethiohydantoin [15–17].

2. Results and Discussion

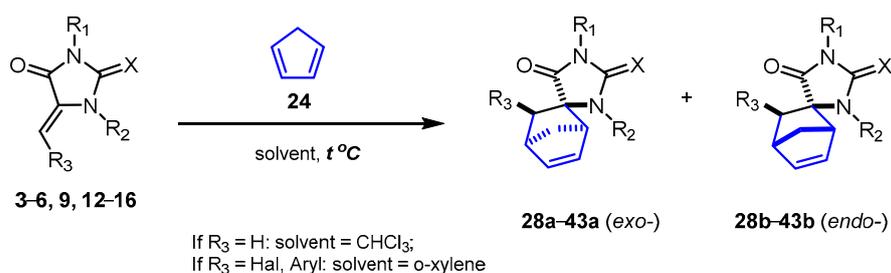
2.1. Synthesis of Dienophiles

Initial dienophiles **1–11**, **22**, and **23** were prepared according to previously described procedures [20–23]. Compounds **12–21**, containing both N(1)-substituted and N(1)-unsubstituted 2-chalcogenimidazolone moieties with exocyclic C=CH₂, C=CHHal, and C=CHAr fragments in five positions, were synthesized according to Scheme 2. The substituents with different electronic and steric effects at the C=C double bond should significantly affect

the reactivity of the dienophile, which can allow controlling the rate and selectivity of the Diels–Alder reaction with these substrates. It was found that methylenedihydroantoin **1** gives high yields of alkylation and acylation products **12–16**, and its reaction with halogens (Br_2 , I_2) in the presence of triethylamine leads to the corresponding halomethylene-substituted imidazolones **17** and **18** as single isomers (Scheme 2). The configuration of the C=C double bond in compounds **17** and **18** was confirmed by ^1H NMR spectra of the products of their reactions with cyclopentadiene (Scheme 3 and Supplementary Information).



Scheme 2. Starting dienophiles and synthesis of compounds **12–21** from methylenedihydroantoin **1,2**.



Scheme 3. Reactions of methylenedihydroantoin derivatives with cyclopentadiene.

Our attempts to introduce methylenethioantoin **2** in the reactions with acetyl chloride and benzyl bromide under the same conditions were unsuccessful. We found that thioantoin **2** rapidly degrades in the presence of bases (NEt_3 , K_2CO_3), and its alkylation or acylation reactions are not formed even in trace amounts (according to ^1H NMR spectra of reaction mixtures). When methylenethioantoin **2** reacted with bromine at 0°C , resinification of the reaction mixture occurred, and the target halogen-substituted methylenethioantoin **21** could be isolated from the reaction mixture only in 9% yield (see Scheme 2 and Supplementary Information).

Next, we started studying the Diels–Alder reactions with dienophiles **1–23**. It should be noted that for each specific diene (cyclopentadiene **24**, cyclohexadiene **25**, 2,3-dimethylbutadiene **26**, isoprene **27**), the optimal reaction conditions turned out to be different and were selected depending on its reactivity.

2.2. Reactions of 5-Methylideneimidazolones 1-6, 9, 12-18, 22, and 23 with Cyclopentadiene

In our previous communication, we demonstrated the possibility of spiro-norbornene derivatives synthesis by the [4+2]-cycloaddition of highly active dienecyclopentadiene with methylideneimidazolones **1** and **2** at an eight-fold excess of the diene in refluxing chloroform [24]. Hydantoins **3-6**, **9**, and **12-16** were now introduced into the reactions with cyclopentadiene under the same conditions (Scheme 3, Table 1). In all cases, cycloaddition reactions proceeded in high yields with the predominant formation of the *exo* products **28a-43a**.

Table 1. Diels–Alder reactions of methylideneimidazolone derivatives with cyclopentadiene.

| Products | X | R ₁ | R ₂ | R ₃ | <i>exo:endo</i> ¹ | Yield, % |
|------------------|---|------------------------------------|--------------------|------------------------------------|------------------------------|----------------------------|
| 28a + 28b | O | Ph | H | H | 91:9 | 81 + 9 ^{2,3} [24] |
| 29a + 29b | O | Ph | Ac | H | 77:23 | 73 (78:22) ^{3,4} |
| 30a + 30b | O | Ph | COOEt | H | 79:21 | 83 (83:17) ^{3,4} |
| 31a + 31b | O | Ph | Ts | H | 87:13 | 91 (89:11) ^{3,4} |
| 32a + 32b | O | Ph | Boc | H | 85:15 | 73 (85:15) ^{3,4} |
| 33a | O | Ph | CH ₂ Ph | H | 100:0 | 10 ^{3,5} |
| 34a + 34b | S | Ph | H | H | 91:9 | 79 + 8 ^{2,3} [24] |
| 35a + 35b | S | 2-FC ₆ H ₄ | H | H | 91:9 | 71 + 7 ^{2,3} |
| 36a + 36b | S | 2-ClC ₆ H ₄ | H | H | 91:9 | 68 + 0 ^{2,3} |
| 37a + 37b | S | 3-MeC ₆ H ₄ | H | H | 91:9 | 80 + 8 ^{2,3} |
| 38a + 38b | S | 3-MeOC ₆ H ₄ | H | H | 91:9 | 81 + 9 ^{2,3} |
| 39a + 39b | S | 4-MeOC ₆ H ₄ | H | H | 91:9 | 78 + 8 ^{2,3} |
| 40a | O | Ph | H | Br | 100:0 | 64 ^{6,7} |
| 41a | O | Ph | H | I | 100:0 | 37 ^{6,7} |
| 42a | O | H | H | 4-FC ₆ H ₄ | - | 0 ⁷ |
| 43a | S | Ph | H | 4-MeOC ₆ H ₄ | - | 0 ⁷ |

¹ Based on ¹H NMR spectra of reaction mixture. ² Isolated yields for major and minor isomers. ³ Conditions: cyclopentadiene (8 equivalents.), CHCl₃, reflux, 6h. ⁴ Isolated yields for the isomers mixture; isomers ratio is indicated in the parentheses. ⁵ Yield by ¹H NMR spectra of reaction mixture. ⁶ Isolated yields. ⁷ Conditions: cyclopentadiene (20 equivalents), *o*-xylene, reflux, 13 h.

It should be noted that, unlike *exo*- and *endo*-products **29-33a** and **29-33b**, which do not contain unsubstituted nitrogen atoms, N(1)-unsubstituted compounds **28a**, **34a-39a**, and **28b**, **34b-39b** can be separated by column chromatography. Probably, the difference in the chromatographic mobility of the isomers is determined by the ability of the CONH and CSNH fragments of the molecules to form hydrogen bonds with the silica gel surface.

The structures of the resulting spiro-imidazolones **28-43** were determined using ¹H and ¹³C NMR spectroscopy; for compounds **28** and **34**, their spectra correspond to those described in the literature [24]. According to ¹H NMR spectroscopy data, *exo*- and *endo*-products **a** and **b** differ in chemical shifts of HC=CH protons of the norbornene framework: the characteristic doublets of the main products are shifted to a weaker field (at 6.20–6.60 ppm) relative to the signals of minor products (at 6.15–6.45 ppm) (Figure 1a), which is consistent with the literature data [15,17,24]. In addition, compounds **28**, **34-39** are characterized by a very large difference in the NH protons shifts of *exo*- and *endo*-isomers (>1 ppm in CDCl₃), which can be explained by the influence of the anisotropy of the double C=C bond, which is quite close to the amide proton in isomer **a**.

The criterion for assigning *exo*- and *endo*-stereoisomers of compounds **28**, **34-39**, in addition to the chemical shifts of vinyl and NH protons, could be the NOESY spectra shown in the Supplementary Information for compounds **35a** and **35b**. In the NOESY spectrum of spiro-derivative **35a**, the interaction of the NH proton with the double bond proton, as well as with the proton of the nearest methylene group of the bicycle, is manifested, and there is no interaction with the proton at the C(7) atom of the norbornene framework (see Figure 1b). On the contrary, for the minor isomer **35b**, when the NH proton was irradiated,

the Overhauser effect is observed for the proton at the C(7) atom of norbornene and is absent for the proton at the C=C double bond.

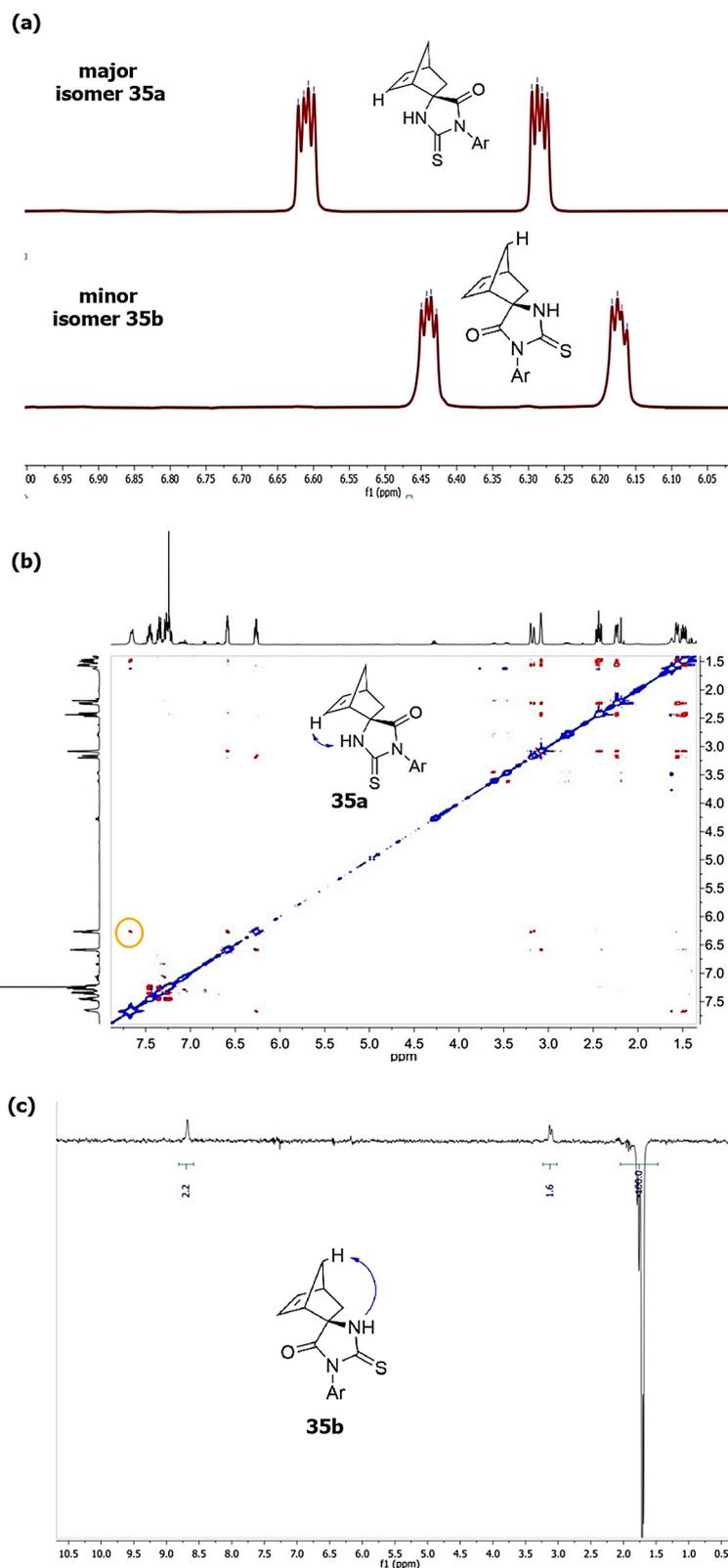


Figure 1. (a) Characteristic doublets of doublet set of HC=CH protons of compounds 35a and 35b in their ^1H NMR spectra (CDCl_3). (b) Characteristic correlations observed in ^1H - ^1H NOESY NMR spectrum of the compounds 35a. (c) ^1H NOESY NMR spectrum of compound 35b.

The diastereoselectivity of the cyclopentadiene Diels–Alder reaction with methylenehydantoin was found to be sensitive to the nature of the substituent at position 1 of the imidazolone moiety. Despite the increase in steric hindrance near the exocyclic double bond of the dienophile, the interaction of cyclopentadiene with hydantoin **12–15** containing electron-withdrawing Ac, COOEt, Ts, Boc substituents proceeded less selectively compared to N(1) unsubstituted hydantoin **1**. Such results may indicate that the electronic factors of the substituents have a stronger effect on the reactivity of the dienophile than the steric ones. The important role of the substituent in position 1 of the initial heterocycle on the reaction course is also confirmed by the fact that for the imidazolone **16** with a donor benzyl fragment, the yields of the cycloaddition product **33** are low, and only the isomers **33a** are formed (see Table 1).

Variation of the exocyclic chalcogen atom of the starting heterocycle does not affect the stereoselectivity of the Diels–Alder reaction (Table 1, compare products **28** and **34**). The same stereoselectivity of the Diels–Alder reaction is observed for thiohydantoin **2–6** and **9** with different substituents at N(3) atom. Apparently, the electronic properties of substituents in the C(2) and N(3) positions of the heterocycle have little effect on the reactivity of the dienophile.

An increase in steric hindrance at the C=C bond of methylenimidazolone should hinder the [4+2]-cycloaddition reaction. Indeed, halomethylenhydantoin **17, 18** react with cyclopentadiene only when refluxed in ortho-xylene. In the course of these reactions, only exo-isomers **40a** and **41a** were formed in moderate yields. In the ¹H NMR spectra of compounds **40a** and **41a**, in the region of about 4.7 ppm, there are doublets with *J* ~3.3 Hz, which confirms the spatial arrangement of the CHBr and CHI groups in the norbornene framework.

Imidazolones **22, 23**, containing the bulkier structure and less acceptor compared to halogens aryl substituents at the double C=C bond, did not form the reaction products with cyclopentadiene under similar conditions even in trace amounts (according to ¹H NMR spectroscopy of the reaction mixtures), probably for both electronic and steric reasons.

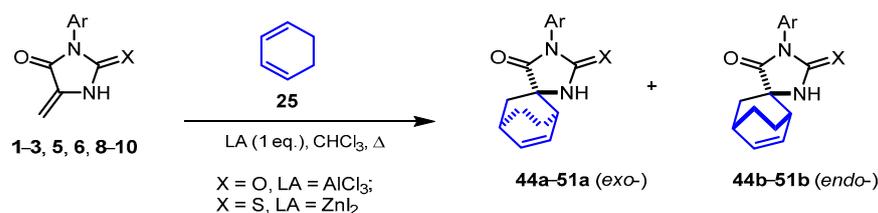
2.3. Reactions of 5-Methylenimidazolones 1-3, 5, 6, 8-10 with Cyclohexadiene

Reactions of 5-methylenimidazolones with dienes **25–27**, which are less reactive than cyclopentadiene, do not occur even when refluxing in toluene with a 20-fold excess of dienes. Under microwave irradiation, when methylenhydantoin **1** was heated in benzene to 140°C with an excess of cyclohexadiene **25**, the formation of the target products only in trace amounts was detected. The reactions of cyclohexadiene with compounds **2–11** were successfully carried out by catalysis of the reaction with Lewis acids; BF₃·Et₂O, AlCl₃, and ZnI₂ were tested in the target reactions.

It was found that cyclohexadiene **25** rapidly decomposed in the presence of BF₃·Et₂O, and no products of its Diels–Alder reaction were formed. In the presence of AlCl₃, the decomposition of the diene also occurred, which accelerated upon heating the reaction mixture; however, the target product **44a** could be isolated if diene **25** was added slowly into a refluxing solution of dienophile **1** and Lewis acid.

In the presence of 1 equivalent of ZnI₂, cyclohexadiene did not react with 5-methylenhydantoin **1** even when heated, but methylenethiohydantoin **2, 3, 5, 6, 8–10** reacted with this diene under the same conditions to form the mixtures of diastereomeric products **45a–51a** and **45b–51b** in a ratio of ~3:1 (Scheme 4, Table 2), which could be separated by column chromatography.

It should be noted that the selectivity of the reaction is practically independent of the substituents in position 3 of the heterocycles **2, 3, 5, 6, 8–10**. In the presence of an excess of Lewis acid, the yields of the target products somewhat decreased, presumably due to the acceleration of the decomposition of the diene on the catalyst, and with a submolar amount of ZnI₂, the formation of products slowed down.



Scheme 4. Reactions of methylidenethiohydantoin 1-3, 5, 6, 8-10 with cyclohexadiene.

Table 2. Diels–Alder reactions of 5-methylideimidazolones with cyclohexadiene.

| Products | X | LA | Ar | Yield a ¹ , % | Yield b ¹ , % |
|-----------|---|-------------------|---|--------------------------|--------------------------|
| 44a + 44b | O | AlCl ₃ | Ph | 16 | 0 |
| | | ZnI ₂ | | 0 | 0 |
| 45a + 45b | S | ZnI ₂ | Ph | 54 | 19 |
| 46a + 46b | S | ZnI ₂ | 2-FC ₆ H ₄ | 44 | 15 |
| 47a + 47b | S | ZnI ₂ | 3-MeC ₆ H ₄ | 60 | 25 |
| 48a + 48b | S | ZnI ₂ | 3-MeOC ₆ H ₄ | 51 | 21 |
| 49a + 49b | S | ZnI ₂ | 4-ClC ₆ H ₄ | 55 | 20 |
| 50a + 50b | S | ZnI ₂ | 4-MeOC ₆ H ₄ | 50 | 19 |
| 51a + 51b | S | ZnI ₂ | 2,6-Me ₂ C ₆ H ₃ | 44 | 15 |

¹ Isolated yields.

Different results of the reaction of diene **25** with hydantoin **1** and thiohydantoin **2**, **3**, **5**, **6**, **8-10** in the presence of ZnI₂ may be due to the fact that ZnI₂ as a soft Lewis acid is predominantly coordinated to the sulfur atom of 2-thioimidazolone, and varying the chalcogen (O or S) strongly affects the efficiency of binding the dienophile to the Lewis acid.

The structures of spiro-imidazolones **44-51** were confirmed by ¹H and ¹³C NMR spectroscopy data; the configuration of compound **46a** was determined via two-dimensional NMR techniques COSY and gHSQC (see Supplementary Information, Figures S67 and S68).

It may be noted that the ¹H, ¹³C (and ¹⁹F NMR in cases where the compounds contained fluorine) spectra of all ortho-phenyl-N(3)-substituted spiro-imidazolones (for example, compounds **35**, **46**, and **51**, see Supplementary Information, Figures S23, S63, and S87 and similar) demonstrate two sets of signals. This can be explained by the hindered rotation around the single bond C(Ar)-N(imidazolone) of the imidazolone fragment and, consequently, the existence of each ortho-phenyl-substituted spiro-compounds as two atropisomers (axially chiral heterocyclic analogs of biaryl derivatives similar to those described for others ortho-substituted 2-thiohydantoin) [25–27]. The NMR spectra of these compounds at ambient temperatures do not indicate the presence of intramolecular dynamic processes, apparently due to the high barrier to internal rotation. Thus, weak line broadening in the ¹⁹F NMR spectrum of product **35a** is observed only at temperatures above 80 °C (Figure S30); this indicates that the rotational barrier is greater than 20 kcal/mol.

As in the reactions with cyclopentadiene, the isomeric *exo*- and *endo*-Diels–Alder products of the reactions of methylidenethiohydantoin **2**, **3**, **5**, **6**, **8-10** with cyclohexadiene can be distinguished by the chemical shifts of the protons of the CH=CH group: for the main isomers **a**, a characteristic doublet of doublets observed in the region of 6.62–6.34 ppm, and for minor products **b**, such doublet of doublets is present in the region of 6.48–6.30 ppm (in CDCl₃).

The structure of compound **51a** was additionally confirmed by the X-ray diffraction data. The crystal cell shown in Figure 2 includes two spiro-thiohydantoin molecules linked by N-HS hydrogen bonds to form an eight-membered cycle. The dihedral angle between the planes of the cycles at the spiro junction is close to 90°; imidazolone five-membered rings are near the planar.

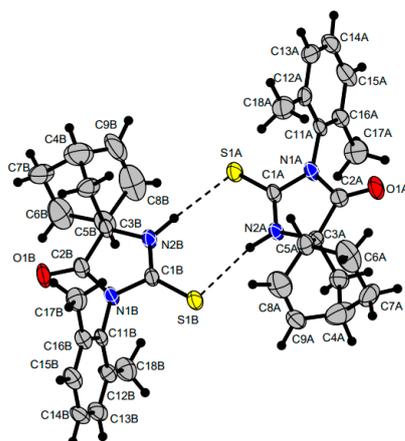
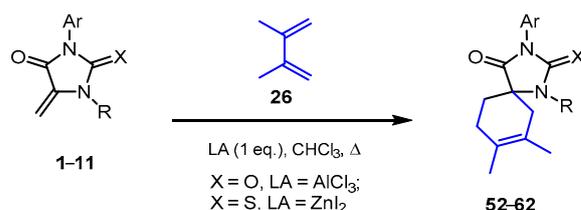


Figure 2. Molecular structure of compound **51a**. Thermal ellipsoids are given a 30% probability.

2.4. Reactions of 5-Methylideneimidazolones 1–12 with 2,3-Dimethylbutadiene

Reactions of methylideneimidazolones **1–12** with 2,3-dimethylbutadiene also require Lewis acids as the catalysis but can proceed in the presence of both AlCl_3 and ZnI_2 (Scheme 5, Table 3). The reactions of diene **26** with dienophiles **1** and **2** under the action of AlCl_3 proceeded in moderate yields giving the compounds **52** and **53**.



Scheme 5. Reactions of methylideneimidazolones **1–12** with 2,3-dimethylbutadiene.

Table 3. Diels–Alder reactions of methylideneimidazolones with 2,3-dimethylbutadiene.

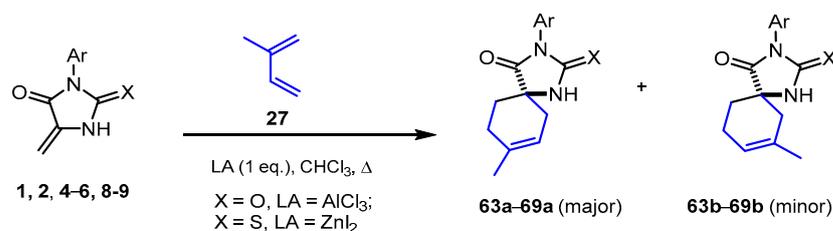
| Product | X | LA | Ar | R | Yield ¹ , % |
|-----------|---|-----------------|--|---|------------------------|
| 52 | O | AlCl_3 | Ph | H | 47 |
| | | ZnI_2 | | | 0 |
| 53 | S | AlCl_3 | Ph | H | 45 |
| | | ZnI_2 | | | 92 |
| 54 | S | ZnI_2 | 2- FC_6H_4 | H | 84 |
| 55 | S | ZnI_2 | 2- ClC_6H_4 | H | 71 |
| 56 | S | ZnI_2 | 3- MeC_6H_4 | H | 85 |
| 57 | S | ZnI_2 | 3- MeOC_6H_4 | H | 79 |
| 58 | S | ZnI_2 | 4- FC_6H_4 | H | 74 |
| 59 | S | ZnI_2 | 4- ClC_6H_4 | H | 82 |
| 60 | S | ZnI_2 | 4- MeOC_6H_4 | H | 88 |
| 61 | S | ZnI_2 | 2,6- $\text{Me}_2\text{C}_6\text{H}_3$ | H | 68 |
| 62 | S | ZnI_2 | 3- Cl ,4- FC_6H_3 | H | 81 |

¹ isolated yields.

As well as in reactions with cyclohexadiene, ZnI_2 activates thiohydantoin **2–11** in the reactions with 2,3-dimethylbutadiene with the formation of target compounds **54–62** but does not catalyze the reactions with oxygen analogs, probably due to the selective coordination of the Lewis acid to the C=S bond of the starting heterocycle. A significant increase in the yield of thiohydantoin **53** when AlCl_3 was replaced by the softer ZnI_2 may be due to a decrease in the rate of the side process of diene polymerization under the action of the Lewis acid.

2.5. Reactions of 5-Methylideneimidazolones 1, 2, 4-6, 8, and 9 with Isoprene

Using isoprene **27**, the regioselectivity of the Diels–Alder reaction of methylideneimidazolones **1**, **2**, **4-6**, **8**, and **9** were studied. Boiling of hydantoin **1** mixture with a 10-fold excess of this diene in chloroform led to the formation of cycloaddition product **63a** in 70% yield and trace amounts of the minor regioisomer **63b**. Under similar conditions, thiohydantoin **2**, **4-6**, **8**, and **9** formed an inseparable mixture of regioisomers **64a-69a** and **64b-69b** in good yields in a ratio of ~87:13 (Scheme 6, Table 4 and Supplementary Information).



Scheme 6. Reactions of methylideneimidazolones **1**, **2**, **4-6**, **8**, and **9** with isoprene.

Table 4. Diels–Alder reactions of methylideneimidazolones with isoprene.

| Product | X | LA | Ar | Yield a + b ¹ , % |
|------------------|---|-------------------|------------------------------------|------------------------------|
| 63a + 63b | O | AlCl ₃ | Ph | 70 |
| 64a + 64b | S | ZnI ₂ | Ph | 82 |
| 65a + 65b | S | ZnI ₂ | 2-ClC ₆ H ₄ | 61 |
| 66a + 66b | S | ZnI ₂ | 3-MeC ₆ H ₄ | 64 |
| 67a + 67b | S | ZnI ₂ | 3-MeOC ₆ H ₄ | 76 |
| 68a + 68b | S | ZnI ₂ | 4-ClC ₆ H ₄ | 68 |
| 69a + 69b | S | ZnI ₂ | 4-MeOC ₆ H ₄ | 71 |

¹ isolated yields.

The structures of regioisomers were confirmed by ¹H NOESY1D, ¹H-¹³C HSQC, and ¹H-¹³C HMBC NMR spectroscopy data for the compounds **67a** and **67b** (see Supplementary Information, Figures S123–S125). In particular, the position of the methine proton in the six-membered ring of compound **67a** was confirmed by the presence of a cross peak in the HMBC spectrum, which is responsible for the vicinal interaction of this proton with the quaternary spiro-carbon.

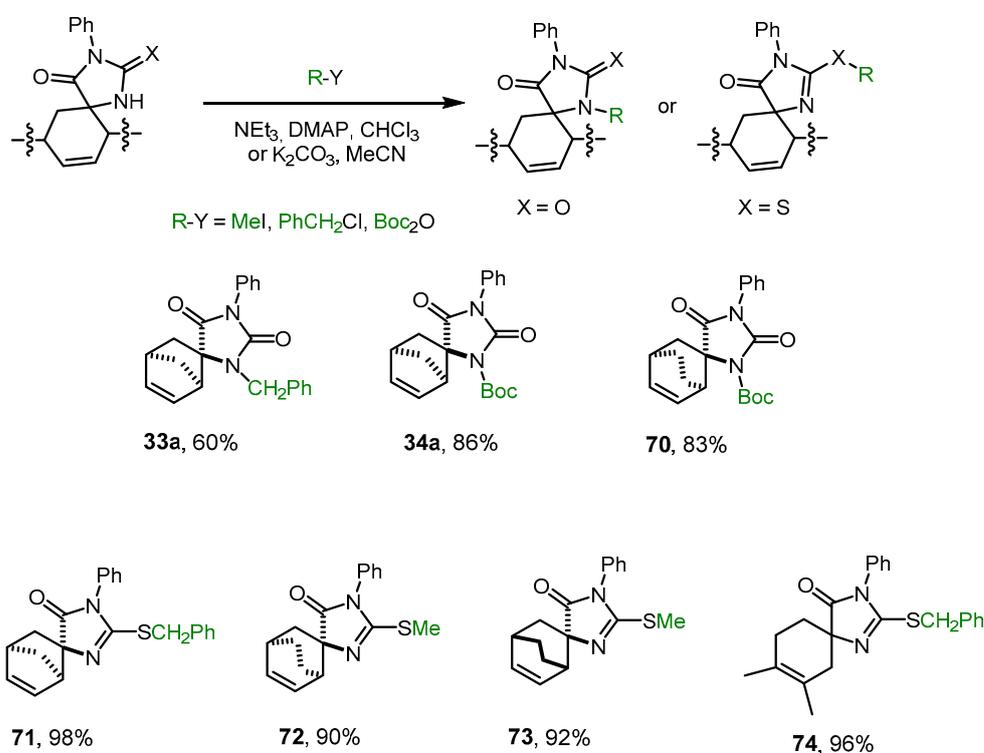
The results obtained demonstrate that AlCl₃ is an effective catalyst for the reactions of low-activity dienes with hydantoin and ZnI₂ with thiohydantoin. At the same time, AlCl₃ similarly catalyzes reactions with hydantoin and thiohydantoin (see Table 3). It may be supposed that the harder Lewis acid AlCl₃ is coordinated to the N(1) atom of the starting imidazolone and, thus, catalyzes both reactions with hydantoin and thiohydantoin, while the softer Lewis acid ZnI₂ is coordinated to the S atom and, thus, activates only thiohydantoin; however, exact proof of this assumption requires additional research.

2.6. Alkylation and Acylation of [4+2]-Cycloaddition Reaction Products

Studying the Diels–Alder reactions with methylideneimidazolones **1-18**, it was found that in the case of N(1)-unsubstituted dienophiles **1** and **2**, the resulting diastereomeric pair of spiroheterocycles can be successfully separated into individual isomers, while N(1)-substituted derivatives are formed either as an inseparable mixture of diastereomers or in extremely low yields (Table 1). Therefore, we studied the possibility of post-modification at the nitrogen atom of N(1)-unsubstituted Spiro derivatives **28a**, **34a**, **44a**, **45a**, **45b**, and **53**.

We found that hydantoin **28a** and **44a** can be alkylated or acylated with PhCH₂Cl or Boc₂O in high yields by refluxing in chloroform or acetonitrile (Scheme 7). The spectral data of the resulting products **33a** and **34a** completely coincided with the spectral data of

the main products of the Diels–Alder reaction of methylidenehydantoin **15** and **16** with cyclopentadiene (see Section 2.2).



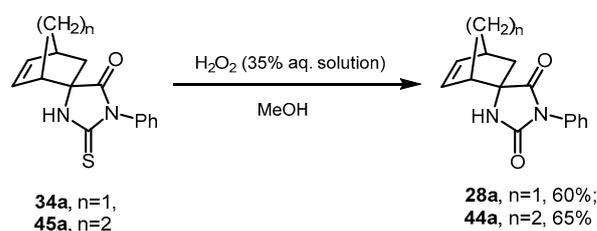
Scheme 7. Alkylation and acylation of spirocyclic imidazolones.

Thiohydantoin **34a**, **45a**, **45b**, and **54** were alkylated with MeI and PhCH₂Cl under milder conditions at room temperature in acetonitrile in the presence of K₂CO₃, exclusively at the sulfur atom. The formation of S–CH₂Ph and S–Me bonds in corresponding imidazolones **72–75** was confirmed by ¹³C NMR spectroscopy data.

2.7. Desulfurization of Spiro-Thiohydantoin

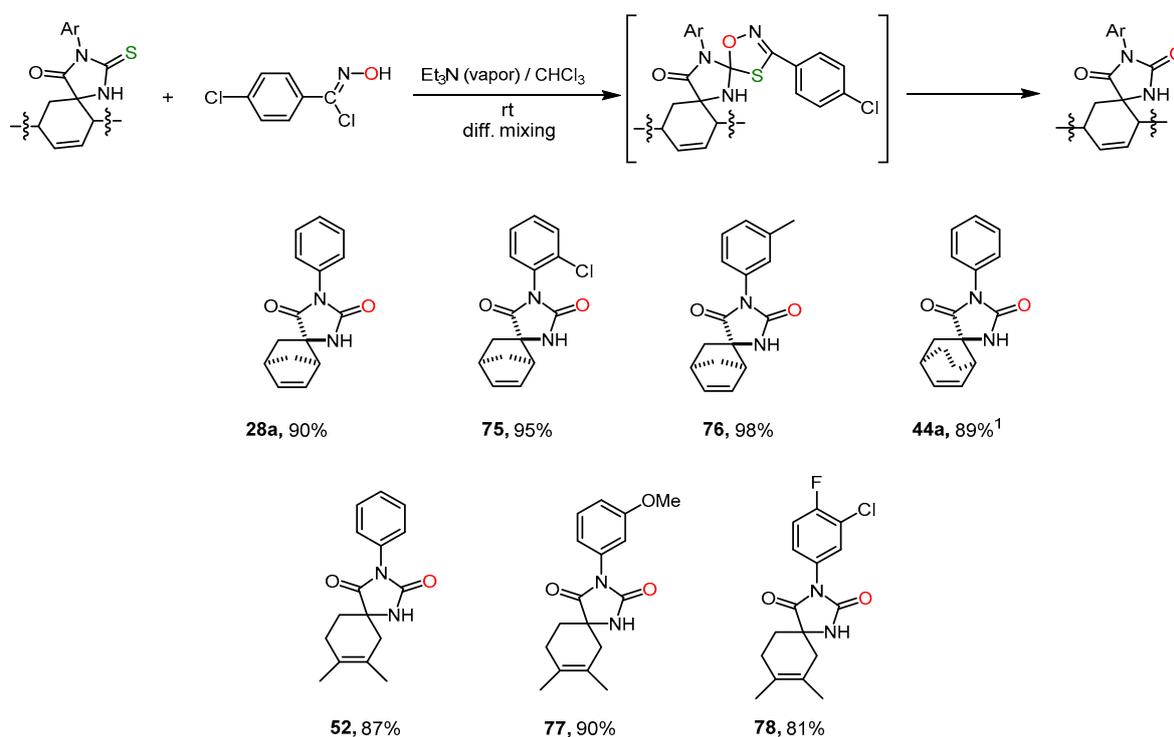
Since methylidenehydantoin **1** reacted with dienes **25** and **26** with low conversion even in the presence of Lewis acids (Sections 2.3 and 2.4), we proposed an alternative scheme for the synthesis of spirocyclic hydantoin from their thiohydantoin analogs. The procedure described in the literature for the hydrolysis of S-alkylated thiohydantoin under the action of HCl in refluxing ethanol [13,28] applied to compound **72** did not lead to the formation of the corresponding hydantoin **28a**. In the ¹H NMR spectrum of the reaction mixture, there were no characteristic CH=CH signals of the norbornene skeleton protons, which probably indicates the ongoing processes of cationic polymerization under the reaction conditions.

The desired transformation of spirothiohydantoin **34a** and **45a** into spirohydantoin **28a** and **44a** was achieved under milder conditions by treating methanolic solutions of the starting compounds with 35% aqueous H₂O₂ at room temperature in good yields (Scheme 8).



Scheme 8. Desulfurization of spiro-thiohydantoin by H₂O₂.

However, an even more effective way of thiohydantoin desulfurizing turned out to be their interaction with nitrile oxides [29] (Scheme 9). The reactive nitrile oxide was generated in situ from N-hydroxyimidoyl chloride under the action of a tertiary amine. As a result of subsequent N-hydroxyimidoyl chloride 1,3-dipolar cycloaddition at the C=S bond, an unstable oxatriazole was obtained, which then decomposed to form hydantoin, analogously to that described in [30]. For this reaction, we used a recently proposed convenient method of diffusion reagents mixing, which made it possible to suppress undesirable dimerization processes of the 1,3-dipole and introduce thiohydantoin and the dipole precursor into the reaction in a strictly equimolar ratio [31]. Under these conditions, hydantoin **28a**, **52**, **75–78** were formed in high yields without the side products of dipole addition to the C=C bond. During the synthesis of compound **44a**, a certain amount (no more than 10%) of the addition product of nitrile oxide to the C=C bond was formed.



Scheme 9. ¹ This product was isolated as an inseparable mixture with the addition products of nitrile oxide at the C=C bond. Desulfurization of spiro-thiohydantoin by the nitrile oxide action.

2.8. Cytotoxicity against Human Cell Lines

Some of the obtained spiro-derivatives were tested for cytotoxicity using the standard MTT assay [32]. Cytotoxicity was evaluated using the cell lines of various etiologies: breast cancer MCF7, human lung carcinoma A549, non-cancer human embryonic kidney cell line HEK293T, and non-cancer lung fibroblast VA13 cell line. Lung cancers and breast cancers are among the most common causes of tumor lesions and related death in the world [33], and non-cancerous cells were applied for the specificity of action evaluation. The results are shown in Table 5; the dose-response dependency graphs are available in Supplementary Information.

Generally, the tested compounds show rather low or moderate cytotoxicity to all tested cell lines; their IC_{50abs} values (IC_{50abs} is the concentration resulting in a two-fold decrease in the number of cells in comparison with untreated cells) against A549 and MCF7 cell lines mostly ranging from 20 to 100 μM. Additionally, the tested compounds show little selectivity to all cell lines tested. Despite the nominal IC_{50abs} selectivity of the compounds **55** and **61** against A549 compared to VA13, their IC₅₀ values (IC₅₀ is the concentration resulting in half of the maximal cytotoxic effect) were quite similar (Table S1).

Table 5. Cytotoxicity effects of the compounds calculated as IC_{50abs} (IC_{50abs}—concentration of compounds, which caused 50% death of cells) for 65-h-treatment of the cell lines of different etiology using the standard MTT assay.

| Compound | Cell line | | | | C _{max} | Units |
|-----------|-------------|-------------|-------------|-------------|------------------|-------|
| | HEK293T | MCF7 | VA13 | A549 | | |
| 28a | >>100 | >>100 | >>100 | >>100 | 100 | μM |
| 28b | >>100 | >>100 | >>100 | >>100 | 100 | μM |
| 34a | 83.0 ± 3.7 | >>100 | >>100 | >>100 | 100 | μM |
| 35b | 87.4 ± 9.9 | >>100 | >>100 | >>100 | 100 | μM |
| 37a | >>100 | >>100 | >>100 | >>100 | 100 | μM |
| 37b | 115 ± 17 ** | >>100 | >>100 | >>100 | 100 | μM |
| 38a | 111 ± 9 ** | >>100 | >>100 | >>100 | 100 | μM |
| 38b | 112 ± 8 ** | >>100 | >>100 | >>100 | 100 | μM |
| 39b | 120 ± 13 ** | >>100 | >>100 | >>100 | 100 | μM |
| 40a | 56.4 ± 2.5 | >>100 | >>100 | >>100 | 100 | μM |
| 41a | 69.1 ± 4.8 | >>100 | >>100 | >>100 | 100 | μM |
| 45a | 21.3 ± 2.3 | 141 ± 28 ** | >>100 | 85.2 ± 10.1 | 100 | μM |
| 48a | 98.0 ± 9.7 | >>100 | >>100 | >>100 | 100 | μM |
| 48b | 108 ± 12 ** | >>100 | >>100 | >>100 | 100 | μM |
| 49a | 6.7 ± 0.7 | 37.8 ± 6.1 | 30.9 ± 3.4 | 16.9 ± 1.9 | 100 | μM |
| 50a | 41.9 ± 3.8 | >>100 | >>100 | >>100 | 100 | μM |
| 50b | 109 ± 12 ** | >>100 | >>100 | >>100 | 100 | μM |
| 51a | >>100 | >>100 | >>100 | >>100 | 100 | μM |
| 51b | 87.5 ± 7.2 | 104 ± 11 ** | >>100 | 140 ± 21 ** | 100 | μM |
| 54 | 29.6 ± 3 | 122 ± 29 ** | >>100 | 49.5 ± 5 | 100 | μM |
| 55 * | ~5 | ~100 | >>100 | ~10 | 100 | μM |
| 56 | 22.0 ± 8.0 | 102 ± 17 ** | 98.7 ± 17.6 | >>100 | 100 | μM |
| 57 | 18.3 ± 1.7 | 50.6 ± 5.2 | 80.8 ± 13.7 | 35.4 ± 4.8 | 100 | μM |
| 58 | 13.7 ± 2.3 | 59.0 ± 13.3 | 45.0 ± 7.5 | 23.2 ± 3.5 | 100 | μM |
| 59 | 4.4 ± 0.6 | 76.6 ± 16 | 52.6 ± 7.8 | 26.2 ± 3.9 | 100 | μM |
| 60 | 10.2 ± 1.3 | 98.3 ± 19.8 | 75.3 ± 17 | 52.2 ± 5.7 | 100 | μM |
| 61 * | ~4 | ~100 | N/A | ~10 | 100 | μM |
| 62 | 6.8 ± 0.3 | 11.1 ± 0.6 | 9.8 ± 0.4 | 7.9 ± 0.4 | 100 | μM |
| 66a + 66b | 44.6 ± 4.9 | 146 ± 22 ** | 110 ± 13 ** | 144 ± 36 ** | 100 | μM |
| 72 | 60.1 ± 3.4 | >>100 | >>100 | >>100 | 100 | μM |
| 75 | 99.0 ± 11.1 | >>100 | >>100 | >>100 | 100 | μM |
| 76 | 48.6 ± 2.4 | 86.4 ± 7.3 | 132 ± 17 ** | 135 ± 14 ** | 100 | μM |

C_{max}—the highest concentration studied in the test.>>100—compound shows little or no cytotoxicity. IC_{50abs} values could not be calculated. *—approximate IC_{50abs} values are given, see IC₅₀ values in Table 6. **—IC_{50abs} value, calculated by data extrapolation, exceeds C_{max}.N/A—not available.

Table 6. Antibacterial effects shown as MIC for *E. coli* BW25113 DTC and *E. coli* K-12. MIC values are given in μM.

| Strain | 28a | 28b | 35b | 40a | 41a | 51a | 55 | 60 | 62 | 72 | 75 | 76 |
|--------------------|-------|-------|-------|-------|-------|-------|-------|-------|-------|-------|-------|-------|
| <i>E. coli</i> DTC | >1000 | >1000 | 1000 | 1000 | 1000 | >1000 | >1000 | >1000 | 1000 | 500 | 500 | 125 |
| <i>E. coli</i> K12 | >1000 | >1000 | >1000 | >1000 | >1000 | >1000 | >1000 | >1000 | >1000 | >1000 | >1000 | >1000 |

>1000—compound is inactive in this test.

It may also be noticed that the introduction into the tested spiro-imidazolones of halogenated phenyl ring instead of non-halogenated one in most cases increased cytotoxicity against all tested cell lines. It can be seen by comparing the data obtained for compound **63**, which is the most cytotoxic of all the tested compounds and has a dihalogenated phenyl ring, with the data obtained for compounds **49a**, **55**, **58**, and **59**, which have monohalogenated phenyl ring, and the non-halogenated compounds **28**, **34**. The compounds with

OMe- or Me-substituent in the phenyl rings at N(3) imidazolone atom had no significant cytotoxicity on the most tested cell lines (Table 5, compounds 37–39, 48, 50, 56, 60, 66).

2.9. Cytotoxicity against *E. coli*

All compounds investigated for cytotoxicity effects were also analyzed for antibacterial activity. Tests were carried out on *E. coli* BW25113 DTC-pDualrep2 strain that has a normal cell wall and the *E. coli* BW25113 LPTD-pDualrep2 strain that has damaged cell walls. Some of the tested compounds showed a noticeable antibacterial effect against *E. coli* BW25113 DTC-pDualrep2 but were almost inactive against *E. coli* BW25113 LPTD-pDualrep2. All the tested compounds were analyzed for the mechanism of antibacterial activity by means of the pDualrep2 [34] reporter system, which allows for the detection of translational inhibitors and SOS-response inducers, but none of these mechanisms is involved.

Then the minimal inhibitory concentration (MIC) was measured for *E. coli* BW25113 DTC and *E. coli* K-12 (WT) for the 12 most bacteriotoxic compounds from the plate test. MIC of compound 77 against *E. coli* BW25113 DTC is 125 μ M, while the other compounds either show MIC \sim 500 μ M, 1 mM, or are inactive in this test (Table 6).

All the tested compounds were completely inactive against *E. coli* K-12 at concentrations of 1 mM and below. It indicates that the studied compounds are toxic to *E. coli* but are effectively removed from the cell by efflux systems and, therefore, do not affect the wild type of *E. coli*.

3. Materials and Methods

All solvents were purified using standard procedures [35]. Starting compounds were purchased from commercial sources (Sigma–Aldrich, ABCR, AKSci, Burlington, VT, USA). Reactions were checked by TLC analysis using silica plates ^1H , and ^{13}C NMR spectra were recorded on BrukerAvance or Agilent 400-MR spectrometers (400 MHz for ^1H , 100 MHz for ^{13}C). Chemical shifts are reported in parts per million relative to TMS.

Electrospray ionization high-resolution mass spectra were recorded on a TripleTOF 5600+ quadrupole time-of-flight mass spectrometer (ABSciex, Concord, Vaughan, ON, Canada) with DuoSpray ion source in positive ion mode. The capillary voltage was 5.5 kV; nebulizing and curtain gas pressure—15 and 25 psi, respectively; ion source temperature ambient; declustering potential 20 V; m/z range 100–1200. Elemental compositions of the detected ions were determined based on accurate masses and isotopic distributions using Formula Finder software (ABSciex, Concord, ON, Canada).

The X-Ray data for compound 51a were collected via STOE diffractometer Pilatus100K detector, Cu K α (1.54086 Å) radiation, rotation method mode. STOE X-AREA software was used for cell refinement and data reduction. Data collection and image processing were performed with X-Area 1.67 (STOE & Cie GmbH, Darmstadt, Germany, 2013). Intensity data were scaled with LANA (part of X-Area) in order to minimize differences in intensities of symmetry-equivalent reflections (multi-scan method).

The structures were solved and refined with SHELX(1) program [36]. The non-hydrogen atoms were refined by using the anisotropic full matrix least-square procedure. Hydrogen atoms were placed in the calculated positions and allowed to ride on their parent atoms [C-H 0.93–0.98; Uiso 1.2 Ueq(parent atom)]. Hydrogen atoms at nitrogen atoms N2A and N2B (see Figure 2) were localized from Fourier syntheses and refined freely. Molecular geometry calculations were performed with the SHELX program, and the molecular graphics were prepared by using DIAMOND(2) version 4 software [37].

Some crystallographic data for 51a are listed in Table S2; hydrogen bonds are given in Table S3 (see Supplementary part of this article); CCDC-2232689 contain the Supplementary crystallographic data. These data can be obtained free of charge from The Cambridge Crystallographic Data Centre via www.ccdc.cam.ac.uk/data_request/cif, 1 February 2023.

Experimental procedures and characteristic data for all synthesized compounds are given in the Supplementary Information.

Cell cultures. Human embryonic kidney HEK293T cell line was kindly provided by Dr. E. Knyazhanskaya; immortalized human fibroblasts cell line VA13 was kindly provided by Dr. M. Rubtsova; human breast cancer cell line MCF7 and human lung adenocarcinoma cell line A549 were kindly provided by Dr. S. Dmitriev. MCF7, VA13, A549, and HEK293T cell lines were maintained in DMEM/F-12 (Thermo Fisher Scientific, Waltham, Massachusetts, USA) culture medium containing 10% fetal bovine serum (Thermo Fisher Scientific, Waltham, Massachusetts, USA) and 50 µg/mL penicillin and 0.05 mg/mL streptomycin at 37°C (Thermo Fisher Scientific, Waltham, Massachusetts, USA) in 5% CO₂. Cells were maintained at 37°C in a humidified incubator MCO-18AC (Sanyo, Japan) supplied with 5% CO₂. Cell cultures were tested for the absence of mycoplasma.

In Vitro Survival Assay (MTT Assay). The cytotoxicity was tested using the MTT (3-(4,5-dimethylthiazol-2-yl)2,5-diphenyl tetrazolium bromide) assay [32] with some modifications. A total of 2500 cells per well for the MCF7, HEK293T cell lines, 3000 cells for the A549 cell line, or 4000 cells per well for the VA-13 cell line were plated out in 135 µL of DMEM-F12 media (Gibco, USA) in a 96-well plate and then incubated in the 5% CO₂ incubator for first 20 h without treating. After this, 15 µL of media-DMSO solutions of tested substances were added to the cells (final DMSO concentrations in the media were 0,5% or less) and treated for 65 h with 45 nM–100 µM (eight dilutions) of our substances (triplicate each) and doxorubicin as a control substance. Then the MTT reagent (Paneco LLC, Moscow, Russia) was added to cells up to the final concentration of 0.5 g/L (10 × stock solution in PBS was used), and cells were incubated for 2 h at 37 °C under an atmosphere of 5% CO₂. Then the MTT solution was discarded, and 140 µL of DMSO (PharmaMed LLC, Russia) was added. The plates were swayed on a shaker (200 rpm) to dissolve the formazan. The absorbance was measured using a microplate reader (VICTOR ×5 Light Plate Reader, PerkinElmer, USA) at a wavelength of 555 nm (in order to measure formazan concentration). The results were used to construct dose-response graphs and to estimate IC_{50abs} values (IC_{50abs} is the concentration resulting in a two-fold decrease in the number of cells in comparison with the untreated cells) and, in some cases, IC₅₀ values (IC₅₀ is the concentration resulting in half of the maximal cytotoxic effect) with GraphPadSoftware, Inc., San Diego, CA, USA.

E. coli cytotoxicity assay with a screening of Mechanism of Action. A total of 50 µL of 20 mM solution of each compound in DMSO and DMSO as a control substance was added into wells punched in an agar plate containing a lawn of the *E. coli* BW25113 DTC-pDualrep2 or *E. coli* BW25113 LPTD-pDualrep2, which are hypersensitive to antibiotics. *E. coli* DTC [38] lacks the *tolC* gene, which encodes the outer membrane component of several multidrug transporters [39]. *E. coli* LPTD [40] has the 23-amino-acid deletion in the *lptD* gene, an inessential protein functioning in the final stages of the assembly of lipopolysaccharides into the outer membrane [41]. After overnight incubation at 37 °C, the plate was scanned with the ChemiDoc (Bio-Rad, Hercules, CA, USA) system with two channels, including “Cy3-blot” (553/574 nm, green pseudocolor) for RFP fluorescence and “Cy5-blot” (588/633 nm, red pseudocolor) for Katushka2S fluorescence. Cells without reporter construction could be detected in both channels. The induction of the expression of Katushka2S is triggered by translation inhibitors, while RFP is up-regulated by the induction of DNA damage and SOS response. In addition, 2 µL of levofloxacin (25 µg/mL) and erythromycin (5 mg/mL) were used as positive controls for DNA biosynthesis and ribosome inhibitors, respectively.

Determination of Minimal Inhibitory Concentration (MIC). MIC values were determined by monitoring growth in 96-well plates of *E. coli* cultures exposed to serial dilutions of compounds. Specifically, overnight *E. coli* BW25113 DTC and K12 cultures were diluted in 96-well plates to an OD₅₉₀ of 0.01 in LB medium. The wells were then supplemented with the solution of each tested compound at concentrations of 1 mM, 500 µM, 250 µM, 125 µM, 62.5 µM, 31.25 µM, 15.63 µM, 7.81 µM, 3.91 µM, and 1.95 µM. Then the plates were incubated with shaking (200 r.p.m.) overnight at 37°C, and cell growth was assessed by scanning OD₅₉₀ each well with a VictorX5 reader.

4. Conclusions

In the present study, a series of new spiro-derivatives containing hydantoin and thiohydantoin fragments was synthesized by [4+2]-cycloaddition of 1,3-dienes (cyclopentadiene, cyclohexadiene, 2,3-dimethylbutadiene, isoprene) to 5-methylidene-substituted imidazolones (hydantoins or thiohydantoins). It was shown that the studied cycloaddition reactions proceed regioselectively and stereoselectively. Reactions of methylideneimidazolones with cyclopentadiene proceed by co-heating the reactants; reactions with cyclohexadiene, 2,3-dimethylbutadiene, and isoprene require catalysis by Lewis acids. It was demonstrated that ZnI_2 is an effective catalyst for the interaction of methylidenehydantoins with non-activated dienes. The possibility of alkylation and acylation of the obtained spiro-hydantoins at the N(1) nitrogen atoms with $PhCH_2Cl$ or Boc_2O and the alkylation of the spiro-thiohydantoins at the S atoms with MeI or $PhCH_2Cl$ in high yields have been demonstrated. The preparative transformation of spiro-thiohydantoins into corresponding spiro-hydantoins in mild conditions by treating with 35% aqueous H_2O_2 or nitrile oxide has been carried out.

Some synthesized spiro-imidazolones in preliminary biological tests demonstrated moderate cytotoxic activity on MCF7, A549, HEK293T, and VA13 cell lines. They show only little (at concentrations more than 100 μM) antibacterial effects in the experiments with wild type of *E. coli*.

Supplementary Materials: The following supporting information can be downloaded at: <https://www.mdpi.com/article/10.3390/ijms24055037/s1>: the experimental details.

Author Contributions: Conceptualization, E.K.B. and M.E.K.; methodology, D.E.S.; validation, D.E.S., Y.K.G. and V.A.R.; formal analysis, Y.K.G.; investigation, Y.S.H., D.E.S., V.A.T. and D.A.S.; resources, Y.K.G.; data curation, M.E.K.; writing—original draft, D.E.S.; supervision, E.K.B.; project administration, M.E.K. and N.V.Z.; funding acquisition, E.K.B. and M.E.K. All authors have read and agreed to the published version of the manuscript.

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