

# Novel 1,4-Dihydropyridine Derivatives as Mineralocorticoid Receptor Antagonists

Felipe Luis Pérez-Gordillo <sup>1</sup>, Natalia Serrano-Morillas <sup>2</sup>, Luz Marina Acosta-García <sup>2</sup>,  
María Teresa Aranda <sup>1</sup>, Daniela Passeri <sup>3</sup>, Roberto Pellicciari <sup>3</sup>, María Jesús Pérez de Vega <sup>1</sup>,  
Rosario González-Muñiz <sup>1</sup>, Diego Alvarez de la Rosa <sup>2,\*</sup> and Mercedes Martín-Martínez <sup>1,\*</sup>

<sup>1</sup> Instituto de Química Médica (IQM-CSIC), Juan de la Cierva, 3, 28006 Madrid, Spain

<sup>2</sup> Departamento de Ciencias Médicas Básicas and Instituto de Tecnologías Biomédicas, Universidad de La Laguna, 38200 La Laguna, Tenerife, Spain

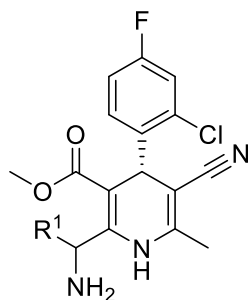
<sup>3</sup> Tes Pharma, via P. Togliatti 20, 06073 Taverne di Corciano, Italy

\* Correspondence: dalrosa@ull.edu.es (D.A.d.l.R.); mmartin@iqm.csic.es (M.M.-M.)

## Supporting information

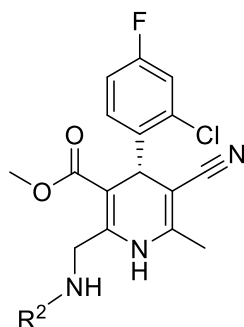
### S1. Focused libraries

**Table S1.** 1<sup>st</sup> focused library, based on proteinogenic amino acids as element of structural diversity. L and D amino acids have been studied.



DHP	Conf.	Amino acid	R <sup>1</sup>	DHP	Config	Amino acid	R <sup>1</sup>
L1.1		Gly	H	L1.10	L	Asn	
				L1.11	D		
L1.2	L	Ala	CH <sub>3</sub>	L1.12	L	Gln	
L1.3	D			L1.13	D		
L1.4	L	Val		L1.14	L	Asp	
L1.5	D			L1.15	D		
L1.6	L	Phe		L1.16	L	Glu	
L1.7	D			L1.17	D		
L1.8	L	Tyr		L1.18	L	Lys	
L1.9	D			L1.19	D		

**Table S2.** 2<sup>nd</sup> focused library, starting from DHP L1.1 (Table S1) and attaching different substituents at the NH<sub>2</sub> at position 2 of the DHP ring.



DHP	R <sup>2</sup>	DHP	R <sup>2</sup>
<b>L2.1</b>	COCH <sub>3</sub>	<b>L2.5</b>	COCH <sub>2</sub> COOH
<b>L2.2</b>	COCH <sub>2</sub> OH	<b>L2.6</b>	CO(CH <sub>2</sub> ) <sub>2</sub> COOH
<b>L2.3</b>	CO(CH <sub>2</sub> ) <sub>2</sub> OH	<b>L2.7</b>	
<b>L2.4</b>	COCOOH		

## S2. Synthetic procedures

General: All reagents and solvents were of commercial quality. The 3-aminocrotonates and the aldehydes are commercially available. The  $\beta$ -keto esters are either commercially available or their preparation has been described within publications [1-6].

Reaction monitoring: TLC silica gel plates (Merck 60 F254, Spain) and a HPLC-MS Waters system composed of a 2695 separations module, a 2996 Photodiode Array (UV, 230-700 nm), a MicroMass ZQ 2000 (electrospray positive mode, m/z: 100-1500) and a Sunfire C18 Column, 100 Å, 2.5  $\mu$ m, 2.1 x 50 mm; mobile phase (A:B:C), CH<sub>3</sub>CN(A):H<sub>2</sub>O(B):H<sub>2</sub>O(1% TFA)(C); flux, 0.2 mL/min. Chromatographic separations: flash column, silica gel Merck 60 (230–400), HPLC-MS Waters semipreparative purification system composed of a 2767 sample manager, a SFO system fluidic organizer, a 2545 binary gradient module, a 515 auxiliary pump, a 2998 photodiode array, a 3100 Mass Detector and a Sunfire C18 Column, 100 Å, 5  $\mu$ m, 19 x 150 mm; mobile phase (A:B), CH<sub>3</sub>CN (0.1% formic acid)(A): H<sub>2</sub>O(0.1% formic acid)(B); flux, 17 mL/min. <sup>1</sup>H NMR spectra: Varian INOVA-300 (300 MHz), Bruker 300 (300 MHz), Varian INOVA-400 (400 MHz) and Varian System-500 (500 MHz), with TMS as internal standard. <sup>13</sup>C NMR spectra: Varian INOVA-300 (75 MHz), Bruker 300 (75 MHz), Varian INOVA-400 (101

MHz) and Varian System-500 (126 MHz). Chemical shifts are expressed in ppm, the coupling constants are expressed in Hz. Mass spectra; Waters MicroMass ZQ 2000 (electrospray positive mode, m/z: 100-1500). Exact mass: high resolution mass spectra (ESI-HRMS) were recorded on an Agilent 6520 Q-TOF instrument.

## S2.1. Synthesis of dihydropyridines

Following the methods included in the manuscript section 4.2.1.

**Methyl 2-[1(S)-(benzyloxycarbonyl)amino]ethyl-4-(2-chloro-4-fluoro)phenyl-5-cyano-6-methyl-1,4-dihydropyridine-3-carboxylate (12ab)** From methyl (4S)-[(benzyloxycarbonyl)amino]-3-oxopentanoate, 2-chloro-4-fluorobenzaldehyde and 3-aminocrotonate. Method A: 5 min (40 °C), 15 min (100 °C) and 3 days (CHCl<sub>3</sub>). Yield 55%. Method B. Yield 80%. Syrup. Eluent: Hexane:EtOAc (3:2), HPLC-MS: Gradient from 30 to 95% ACN/H<sub>2</sub>O (0.05% TFA) in 10 min, *t<sub>R</sub>*=8.10 min (m/z: 484.08 M+H<sup>+</sup>), this purification do not allow the isolation of diastereoisomers. HRMS (ESI pos) m/z Calculated C<sub>25</sub>H<sub>23</sub>ClFN<sub>3</sub>O<sub>4</sub> 483.13611, found 483.13704 (1.92 ppm). Diastereoisomers were subsequently resolved by semi-preparative HPLC-MS. Gradient from 40 to 70% ACN/H<sub>2</sub>O (0.1% formic acid) in 70 min (diastereoisomers A:B 1:1), FP531a (*t<sub>R</sub>*=48min, m/z: 484.04 M+H<sup>+</sup>) and FP531b (*t<sub>R</sub>*=52min, m/z: 484.04 M+H<sup>+</sup>). **12a:** <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>, 50 °C): δ 7.34 (m, 6H, 1-H, Ph Cbz), 7.17 (dd, *J* = 8.7, 6.1 Hz, 1H, Ph 6-H), 7.08 (dd, *J* = 8.6, 2.6 Hz, 1H, Ph 3-H), 6.91 (ddd, *J* = 8.7, 8.2, 2.6 Hz, 1H, Ph 5-H), 5.47 (d, *J*=6.7 Hz, 1H, NHCO), 5.44 (s, 1H, Et CH), 5.20 (s, 1H, 4-H), 5.17 (d, *J* = 12.2 Hz, 1H, Cbz CH<sub>2</sub>), 5.09 (d, *J* = 12.2 Hz, 1H, Cbz CH<sub>2</sub>), 3.54 (s, 3H, OCH<sub>3</sub>), 1.96 (s, 3H, 6-CH<sub>3</sub>), 1.55 (d, *J* = 6.7 Hz, 3H, Et CH<sub>3</sub>). <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>, 50 °C): δ 166.8 (3-CO), 161.5 (d, *J* = 249.5 Hz, 4-C Ph), 157.1 (Cbz CO), 150.0 (2-C), 145.3 (6-C), 139.2 (d, *J* = 3.6 Hz, Ph 1-C), 136.1 (Cbz Ph C), 133.5 (d, *J* = 10.3 Hz, Ph 2-C), 131.6 (d, *J* = 8.8 Hz, Ph 6-C), 128.8, 128.7, 128.3 (Cbz Ph CH), 118.8 (CN), 117.1 (d, *J* = 24.5 Hz, Ph 3-C), 114.8 (d, *J* = 21.2 Hz, Ph 5-C), 100.0 (3-C), 86.2 (5-C), 67.7 (Cbz CH<sub>2</sub>), 51.4 (OCH<sub>3</sub>), 47.4 (Et CH), 38.4 (4-C), 18.9 (Et CH<sub>3</sub>), 18.4 (6-CH<sub>3</sub>). **12b:** <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>, 50 °C): δ 7.35 (m, 5H, 6-H, Cbz), 7.21 (bs, 1H, Ph 6-H), 7.07 (dd, *J* = 8.6, 2.6 Hz, 1H, Ph 3-H), 6.87 (bs, 1H, Ph 5-H), 5.50 (bs, 1H, Et CH), 5.28 (d, *J* = 5.1 Hz, 1H, Cbz NH), 5.18 (s, 1H, 4-H), 5.16 (s, 2H, Cbz CH<sub>2</sub>), 3.55 (s, 3H, OCH<sub>3</sub>), 2.04 (s, 3H, 6-CH<sub>3</sub>), 1.49 (d, *J* = 7.0 Hz, 3H, Et CH<sub>3</sub>). <sup>13</sup>C NMR (126 MHz, CDCl<sub>3</sub>, 50 °C): δ 166.7 (3-CO), 161.4 (d, *J* = 249.2 Hz, Ph 4-C), 156.8 (Cbz CO), 150.3 (2-C), 145.1 (6-C), 139.6 (d, *J* = 3.5 Hz, Ph 1-C), 136.0 (Cbz Ph C), 132.9 (d, *J* = 10.4 Hz, Ph 2-C), 131.6 (d, *J* = 8.8 Hz, Ph 6-C), 128.8, 128.7, 128.2 (Cbz Ph CH), 118.9 (CN), 116.7 (d, *J* = 24.3 Hz, Ph 3-C), 115.0 (d, *J* = 21.1 Hz, Ph 5-C), 100.0 (3-C), 86.7 (5-C), 67.7 (Cbz CH<sub>2</sub>), 51.5 (OCH<sub>3</sub>), 47.7 (Et CH), 37.5 (4-C), 19.0 (Et CH<sub>3</sub>), 18.5 (6-CH<sub>3</sub>).

**Methyl 2-[1(S)-(benzyloxycarbonyl)amino]isobutyl-4-(2-chloro-4-fluoro)phenyl-5-cyano-6-methyl-1,4-dihydropyridine-3-carboxylate (13ab).** From (4S)-methyl-4-[(benzyloxycarbonyl)amino]-5-methyl-3-oxohexanoate, 2-chloro-4-fluorobenzaldehyde and 3-aminocrotonate. Method A: 3 min (40 °C), 3 min (100 °C) and 2 days (CHCl<sub>3</sub>). Yield

59%. Syrup. Method B: Yield 74%. Syrup. Eluent: Hexane:EtOAc (3:1). It is obtained as a diastereoisomeric mixture that could not be resolved. HPLC-MS: Gradient from 30 to 95% ACN/H<sub>2</sub>O (0.05% TFA) in 10 min, *t<sub>R</sub>*: 9.06, (m/z: 512.06 M+H<sup>+</sup>). HRMS (ESI pos) m/z Calculated C<sub>27</sub>H<sub>27</sub>ClFN<sub>3</sub>O<sub>4</sub> 511.16741, found 511.16597 (-2.83 ppm). <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>) δ 7.73 (bs, 2H, 1-H) 7.39 – 7.29 (m, 10H, Cbz), 7.22 (dd, *J* = 8.7, 6.0 Hz, 2H, Ph 6-H), 7.09-7.05 (m, 2H, Ph 3-H), 6.93-6.87 (m, 2H, Ph 5-H), 5.88 (bs, 1H, NHCO), 5.69 (bs, 1H, NHCO), 5.24 (s, 1H, 4-H), 5.21 (s, 1H, 4-H), 5.19-5.06 (m, 6H, Cbz CH<sub>2</sub>, <sup>i</sup>Bu 1-H), 3.54 (s, 3H, OCH<sub>3</sub>), 3.52 (s, 3H, OCH<sub>3</sub>), 2.02 (s, 3H, 6-CH<sub>3</sub>), 1.97 (s, 3H, 6-CH<sub>3</sub>), 1.04-0.97 (m, 14H, <sup>i</sup>Bu 2-H, CH<sub>3</sub>). <sup>13</sup>C NMR (126 MHz, CDCl<sub>3</sub>): δ 167.2 (3-CO), 161.4 (d, *J* = 249.5 Hz, Ph 4-C), 161.4 (d, *J* = 249.1 Hz, Ph 4-C), 157.7, 157.6 (Cbz CO), 148.8 (2-C), 145.5, 145.27 (6-C), 139.7 (d, *J* = 3.6 Hz, Ph 1-C), 139.5 (d, *J* = 3.7 Hz, Ph 1-C), 136.2, 136.1 (Cbz Ph C), 133.2 (d, *J* = 10.3 Hz, Ph 2-C), 133.1 (d, *J* = 10.1 Hz, Ph 2-C), 131.6 (d, *J* = 8.8 Hz, Ph 6-C), 131.5 (d, *J* = 8.9 Hz, Ph 6-C), 128.8, 128.7, 128.60, 128.59, 128.1 (Cbz Ph CH), 119.0, 118.9 (CN), 116.9 (d, *J* = 24.1 Hz, Ph 3-C), 116.8 (d, *J* = 24.6 Hz, Ph 3-C), 114.9 (d, *J* = 21.0 Hz, Ph 5-C), 101.7, 101.6 (3-C), 86.3, 86.1 (5-C), 67.6, 67.5 (Cbz CH<sub>2</sub>), 58.1, 57.6 (<sup>i</sup>Bu 1-C) 51.5, 51.4 (OCH<sub>3</sub>), 38.0, 37.7 (4-C), 29.8, 29.5 (<sup>i</sup>Bu 2-C), 26.0, 24.9, 20.0, 19.8, 19.6, 19.2 (<sup>i</sup>Bu CH<sub>3</sub>), 18.4, 18.3 (6-CH<sub>3</sub>).

***tert*-Butyl-2-[(benzyloxycarbonyl)amino]methyl-4-(2-chloro-4-fluoro)phenyl-5-cyano-6-methyl-1,4-dihydropyridine-3-carboxylate (15).** From *tert*-butyl 4-[(benzyloxycarbonyl)amino]-3-oxo-butanoate, 2-chloro-4-fluorobenzaldehyde and 3-aminocrotonate. Method A: 5 min (40 °C), 10 min (100 °C) and 2 days (CHCl<sub>3</sub>). Yield 14%. Syrup. Purified by isocratic semi-preparative HPLC-MS, 55% CH<sub>3</sub>CN (0.1% formic acid): 45% H<sub>2</sub>O (0.1% formic acid) (*t<sub>R</sub>*=39 min). HPLC-MS: Gradient from 30 to 95% ACN/H<sub>2</sub>O (0.05% TFA) in 10 min, *t<sub>R</sub>*: 9.80, (m/z: 512.06 M+H<sup>+</sup>). HRMS (ESI pos) m/z Calculated C<sub>27</sub>H<sub>27</sub>ClFN<sub>3</sub>O<sub>4</sub> 511.16741, found 511.16622 ( -2.33 ppm). <sup>1</sup>H RMN (500 MHz, CDCl<sub>3</sub>): δ 7.45 – 7.32 (m, 5H, Cbz), 7.20-7.17 (m, 2H, 1-H, Ph 6-H), 7.10 (ddd, *J* = 8.6, 2.6, 0.7 Hz, 1H, Ph 3-H), 6.93 (ddd, *J* = 8.6, 8.3, 2.6 Hz, 1H, Ph 5-H), 5.75 (t, *J* = 6.5 Hz, 1H, NHCO), 5.15 (s, 2H, Cbz CH<sub>2</sub>), 5.12 (s, 1H, 4-C), 4.29 (m, 2H, 2-CH<sub>2</sub>), 2.01 (s, 3H, 6-CH<sub>3</sub>), 1.23 (s, 9H, <sup>i</sup>Bu CH<sub>3</sub>). <sup>13</sup>C RMN (126 MHz, CDCl<sub>3</sub>): δ 166.1 (3-CO), 161.3 (d, *J* = 249.4 Hz, Ph 4-C), 158.6 (Cbz CO), 145.5, 145.4 (2-C, 6-C), 139.3 (Ph 1-C), 136.1 (Cbz Ph C), 133.2 (d, *J* = 10.3 Hz, Ph 2-C), 131.4 (d, *J* = 8.8 Hz, Ph 6-C), 128.8, 128.6, 128.2 (Cbz Ph CH), 119.1 (CN), 116.9 (d, *J* = 24.5 Hz, Ph 3-C), 114.7 (d, *J* = 21.1 Hz, Ph 5-C), 103.6 (3-C), 85.4 (5-C), 81.4 (<sup>i</sup>Bu C), 67.7 (Cbz CH<sub>2</sub>), 41.2 (2-CH<sub>3</sub>), 37.8 (4-C), 28.1 (<sup>i</sup>Bu CH<sub>3</sub>), 18.5 (6-CH<sub>3</sub>).

**Methyl 2-[(benzyloxycarbonyl)amino]methyl-5-carbamoyl-4-(2-chloro-4-fluoro)phenyl-6-methyl-1,4-dihydropyridine-3-carboxylate (17).** From methyl-4-[(benzyloxycarbonyl)amino]-3-oxo-butanoate, 2-chloro-4-fluorobenzaldehyde and 3-amino-2-butenamida. Method A: 5 min (40 °C), 10 min (100 °C) and 2 days (CHCl<sub>3</sub>). Yield 29%. Syrup. Eluent: Hexane:EtOAc (1:9). HPLC-MS: Gradient from 30 to 95% ACN/H<sub>2</sub>O (0.05% TFA) in 10 min, *t<sub>R</sub>* = 5.86, (m/z: 488.06 M+H<sup>+</sup>). HRMS (ESI pos) m/z Calculated C<sub>24</sub>H<sub>23</sub>ClFN<sub>3</sub>O<sub>5</sub> 487.13103, found 487.13160 (1.18 ppm). <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>): δ 7.40 – 7.31 (m, 6H, Cbz Ph and Ph 6-H), 7.08 (s, 1H, 1-H), 6.99 (dd, *J* = 8.6, 2.6 Hz, 1H, Ph 3-H), 6.89 (ddd, *J* = 8.7, 8.3, 2.6 Hz, 1H, Ph 5-H), 5.74 (t, *J* = 6.7 Hz, 1H, NHCO), 5.51 (bs,

2H, 5-CONH<sub>2</sub>), 5.13 (s, 1H, 4-H), 5.13 (s, 2H, Cbz CH<sub>2</sub>), 4.37 (dd, *J* = 14.9, 6.4 Hz, 1H, 2-CH<sub>2</sub>), 4.23 (dd, *J* = 15.0, 6.9 Hz, 1H, 2-CH<sub>2</sub>), 3.61 (s, 3H, 3-COOCH<sub>3</sub>), 2.24 (s, 3H, 6-CH<sub>3</sub>). <sup>13</sup>C NMR (126 MHz, CDCl<sub>3</sub>): δ 169.9 (5-CO), 167.6 (3-CO), 161.2 (d, *J* = 249.6 Hz, Ph 1-C), 158.3 (Cbz CO), 146.9 (3-C), 141.1 (d, *J* = 2.7 Hz, Ph 1-C), 140.9 (5-C), 136.1 (Cbz Ph C), 132.3 (d, *J* = 8.7 Hz, Ph 6-C), 131.3 (d, *J* = 10.1 Hz, Ph 2-C), 128.8, 128.5, 128.2 (Cbz Ph CH), 116.3 (d, *J* = 24.7 Hz, Ph 3-C), 115.6 (d, *J* = 21.0 Hz, Ph 5-C), 106.2 (6-C), 102.4 (2-C), 67.5 (Cbz CH<sub>2</sub>), 51.3 (3-COOCH<sub>3</sub>), 41.4 (2-CH<sub>2</sub>), 37.2 (4-C), 19.2 (6-CH<sub>3</sub>).

## S2.2. Catalytic hydrogenation

General method: To a solution of the benzyloxycarbonyl derivative (0.1 mmol) in MeOH (10 mL) at 0°C is added Pd/C (10%). The suspension is hydrogenated at room temperature and 25 psi during 2h 30 min. After filtration, the solvent is evaporated to dryness.

**Methyl 2-aminomethyl-4-(2-chloro-4-fluoro)phenyl-5-cyano-6-methyl-1,4-dihydropyridine-3-carboxylate (18).** From **11**. Yield: 93%. Amorphous solid. HPLC-MS: Gradient from 15 to 95% ACN/H<sub>2</sub>O (0.05% TFA) in 10 min, *t<sub>R</sub>* = 4.40, (*m/z*: 336.35 *M*+H<sup>+</sup>). This compound decomposed over time.

**Methyl 2-[(S)-2-Amino-5-*tert*-butoxi-5-oxopentanamido]methyl-5-cyano-4-(2-chloro-4-fluoro)phenyl-6-methyl-1,4-dihydropyridine-3-carboxylate (23ab).** From **22ab**. Eluent: DCM:MeOH (20:1). Yield 93% (diastereoisomers A:B 1:1). Syrup. Diastereoisomer A: [*α*]<sub>D</sub><sup>20</sup> = -86.8° (*c* 0.90, CHCl<sub>3</sub>). HPLC-MS: Gradient from 15 to 95% ACN/H<sub>2</sub>O (0.05% TFA) in 15 min, *t<sub>R</sub>*: 5.17 (*m/z*: 521.69 *M*+H<sup>+</sup>). HRMS (ESI pos) *m/z* Calculated C<sub>25</sub>H<sub>30</sub>ClFN<sub>4</sub>O<sub>5</sub> 520.18888, found 520.19028 (2.7 ppm). **23a**: <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>): δ 8.14 (t, *J* = 6.6 Hz, 1H, CONH), 7.98 (s, 1H, 1-H), 7.21 (dd, *J* = 8.7, 6.1 Hz, 1H, Ph 6-H), 7.07 (dd, *J* = 8.6, 2.6 Hz, 1H, Ph 3-H), 6.95 (ddd, *J* = 8.7, 7.8, 2.6 Hz, 1H, Ph 5-H), 5.15 (s, 1H, 4-H), 4.55 (dd, *J* = 14.3, 6.6 Hz, 1H, 2-CH<sub>2</sub>), 4.43 (dd, *J* = 14.3, 6.6 Hz, 1H, 2-CH<sub>2</sub>), 3.56 (s, 3H, OCH<sub>3</sub>), 3.49 (dd, *J* = 7.6, 4.9 Hz, 1H, Pen 2-H), 2.37 (m, 1H, Pen 4-H), 2.34 (m, 1H, Pen 4-H), 2.12 (dtd, *J* = 14.9, 7.5, 4.9 Hz, 1H, Pen 3-H), 2.05 (s, 3H, 6-CH<sub>3</sub>), 1.92 (br s, 2, Pen 2-NH<sub>2</sub>), 1.84 (dt, *J* = 21.9, 7.6 Hz, 1H, Pen 3-H), 1.45 (s, 9H, <sup>t</sup>Bu CH<sub>3</sub>). <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>): δ 177.4, 172.6, 167.2 (CO), 161.4 (d, *J* = 249.3 Hz, Ph 4-C), 146.6, 145.7 (2-C, 6-C), 139.3 (d, *J* = 3.6 Hz, Ph 1-C), 133.0 (d, *J* = 10.4 Hz, Ph 2-C), 131.6 (d, *J* = 8.8 Hz, Ph 6-C), 119.1 (CN), 116.9 (d, *J* = 24.4 Hz, Ph 3-C), 114.9 (d, *J* = 21.0 Hz, Ph 5-C), 101.6 (3-C), 85.7 (5-C), 81.1 (<sup>t</sup>Bu C), 54.5 (Pen 2-C), 51.6 (OCH<sub>3</sub>), 39.1 (2-CH<sub>2</sub>), 37.5 (4-C), 31.9 (Pen 4-C), 30.1 (Pen 3-C), 28.2 (<sup>t</sup>Bu CH<sub>3</sub>), 18.5 (6-CH<sub>3</sub>). **23b**: [*α*]<sub>D</sub><sup>20</sup> = +89.7° (*c* 0.92, CHCl<sub>3</sub>). HPLC-MS: Gradient from 15 to 95% ACN/H<sub>2</sub>O (0.05% TFA) in 15 min, *t<sub>R</sub>*: 5.17, (*m/z*: 521.69 *M*+H<sup>+</sup>). HRMS (ESI pos) *m/z* Calculated C<sub>25</sub>H<sub>30</sub>ClFN<sub>4</sub>O<sub>5</sub> 520.18888, found 520.18863 (-0.47 ppm). <sup>1</sup>H RMN (400 MHz, CDCl<sub>3</sub>): δ 8.07 (br t, 1H, CONH), 7.99 (s, 1H, 1-H), 7.21 (dd, *J* = 8.7, 6.0 Hz, 1H, Ph 6-H), 7.07 (dd, *J* = 8.6, 2.6 Hz, 1H, Ph 3-H), 6.93 (ddd, *J* = 8.7, 7.9, 2.6 Hz, 1H, Ph 5-H), 5.16 (s, 1H, 4-H), 4.59 (dd, *J* = 14.5, 6.4 Hz, 1H, 2-CH<sub>2</sub>), 4.46 (dd, *J* = 14.5, 6.7 Hz, 1H, 2-CH<sub>2</sub>), 3.56 (s, 3H, OCH<sub>3</sub>), 3.49 (dd, *J* = 7.6, 5.2 Hz, 1H, Pen 2-H), 2.43 – 2.26 (m, 2H, Pen 4-H), 2.12 (ddd, *J* = 14.2, 7.4, 5.2 Hz, 1H, Pen 3-H), 2.05 (s, 3H, 6-CH<sub>3</sub>), 1.93 (br s,

2, Pen 2-NH<sub>2</sub>), 1.85 (td, *J* = 14.2, 7.6 Hz, 1H, Pen 3-H), 1.45 (s, 9H, <sup>t</sup>Bu CH<sub>3</sub>). <sup>13</sup>C RMN (101 MHz, CDCl<sub>3</sub>): δ 177.6, 172.6, 167.2 (CO), 161.4 (d, *J* = 249.4 Hz, Ph 4-C), 146.6, 145.7 (2-C, 6-C), 139.3 (d, *J* = 3.5 Hz, Ph 1-C), 133.1 (d, *J* = 10.4 Hz, Ph 2-C), 131.6 (d, *J* = 8.8 Hz, Ph 6-C), 119.1 (5-CN), 116.9 (d, *J* = 24.4 Hz, Ph 3-C), 114.9 (d, *J* = 21.2 Hz, Ph 5-C), 101.6 (3-C), 85.7 (5-C), 81.1 (<sup>t</sup>Bu C), 54.5 (Pen 2-C), 51.6 (3-COOCH<sub>3</sub>), 39.0 (2-CH<sub>2</sub>), 37.6 (4-C), 31.9 (Pen 4-C), 30.2 (Pen 3-C), 28.2 (<sup>t</sup>Bu CH<sub>3</sub>), 18.4 (6-CH<sub>3</sub>).

### S2.3. Acylation of primary amides

Following the methods included in the manuscript section 4.2.2.

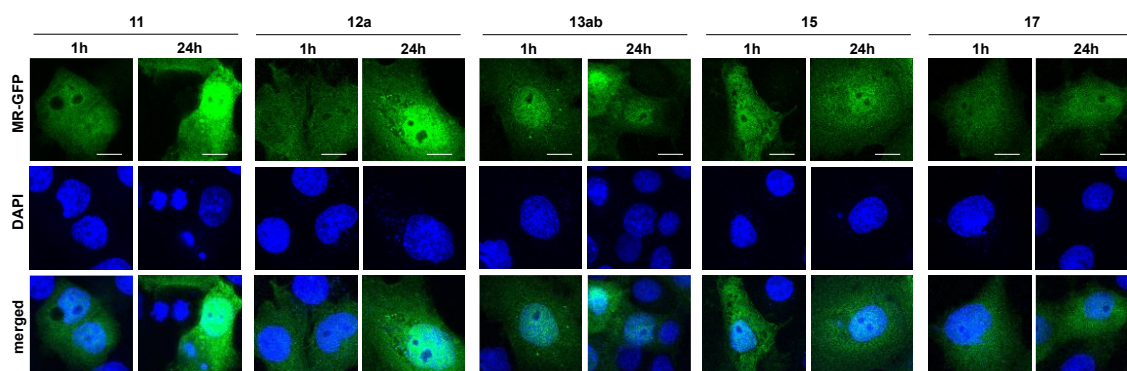
**Methyl (4S)-2-[[2-(benzyloxycarbonyl)amino-5-tert-butoxy-5-oxopentanamide]methyl]-4-(2-chloro-4-fluoro)phenyl-5-cyano-6-methyl-1,4-dihydropyridine-3-carboxylate (22ab).** From **18** and Z-Glu(O<sup>t</sup>Bu)OH. Method B: Yield 57%, it is obtained as diastereoisomeric mixture that could not be isolated. Eluent: Hexane:EtOAc (1:1). HPLC-MS: Gradient from 15 to 95% ACN/H<sub>2</sub>O (0.05% TFA) in 10 min, *t<sub>R</sub>* = 9.16, (*m/z*: 655.57 M+H<sup>+</sup>). HRMS (ESI pos) *m/z* Calculated C<sub>33</sub>H<sub>36</sub>ClFN<sub>4</sub>O<sub>7</sub> 654.22566, found 654.22655 (1.36 ppm). <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>): δ 7.67 (t, *J* = 11.9 Hz, 2H, Pen CONH), 7.32 (m, 14H, 1-H, Cbz Ph), 7.31 (m, 1H, H-1), 7.24 - 7.17 (m, 2H, Ph 6-H), 7.09 - 7.03 (m, 2H, Ph 3-H), 6.99 - 6.90 (m, 2H, Ph 5-H), 5.99 - 5.86 (m, 2H, Cbz-NH), 5.23 - 5.00 (m, 8H, 4-H, Cbz CH<sub>2</sub>, NHCO), 4.60 - 4.33 (m, 4H, 2-CH<sub>2</sub>), 4.18 (m, 2H, Pen 2-H), 3.53 (m, 6H, OCH<sub>3</sub>), 2.46 - 1.83 (m, 10H, 2-CH<sub>3</sub>, Pen 3-H, 4-H), 1.51 - 1.32 (m, 18H, <sup>t</sup>Bu CH<sub>3</sub>). <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>): δ 174.5, 173.7, 167.3(CO), 161.3 (d, *J* = 249.2 Hz, Ph 4-C), 156.7, 156.6 (Cbz CO), 146.1, 145.73, 145.69 (2-C, 6-C), 139.4 (d, *J* = 3.1 Hz, Ph 1-C), 136.0 (Cbz Ph C), 132.9 (d, *J* = 10.1 Hz, Ph 2-C), 131.6 (d, *J* = 8.8 Hz, Ph 6-C), 128.72, 128.70, 128.6, 128.5, 128.4, 128.3, 128.2, 128.1 (Cbz Ph CH), 119.0 (CN), 116.8 (d, *J* = 24.3 Hz, Ph 3-C), 115.0 (d, *J* = 20.9 Hz, Ph 5-C), 101.2, 101.1 (3-C), 86.1 (5-C), 81.6, 81.2 (<sup>t</sup>Bu C), 67.4, 67.2 (Cbz CH<sub>2</sub>), 55.3, 55.2 (Pen 2-C), 51.61, 51.60 (OCH<sub>3</sub>), 39.9, 39.8 (2-CH<sub>2</sub>), 37.3 (4-C), 31.8 (Pen 4-C), 28.1 (<sup>t</sup>Bu CH<sub>3</sub>), 27.5, 27.3 (Pen 3-C) 18.24, 18.21 (6-CH<sub>3</sub>).

### S2.4. Hydrolysis of *tert*-butyl esters

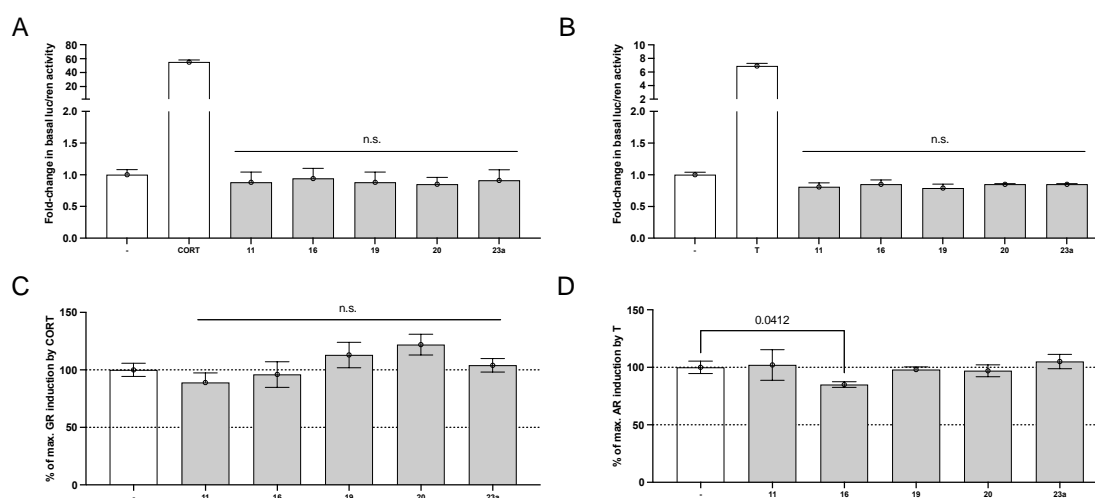
**Methyl 4-(2-chloro-4-fluoro)phenyl-5-cyano-2-[(malonamide)methyl]-6-methyl-1,4-dihydropyridine-3-carboxylate (21).** The corresponding *tert*-butyl esters DHP **20** (50 mg, 0.1 mmol) was dissolved in a mixture 1:1 of CH<sub>2</sub>Cl<sub>2</sub>/TFA (1 mL). After 2 h of stirring at room temperature, the solvent was evaporated to dryness, with coevaporations with CH<sub>2</sub>Cl<sub>2</sub>. Then, the resulting residue was dissolved in a ACN/H<sub>2</sub>O and it was lyophilized. In this conditions **21** was obtained with quantitative yield as a syrup. HPLC-MS: Gradient from 15 to 95% ACN/H<sub>2</sub>O (0.05% TFA) in 10 min, *t<sub>R</sub>* = 4.83, (*m/z*: 422.56 M+H<sup>+</sup>). HRMS (ESI pos) *m/z* Calculated C<sub>19</sub>H<sub>17</sub>ClFN<sub>3</sub>O<sub>5</sub> 421.08408, found 421.08286 (-2.88 ppm). <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>): δ 8.98 (s, 1H, 1-H), 8.40 (br t, 1H, NHCO), 7.37 (dd, *J* = 8.9, 2.6 Hz, 1H, Ph 3-H), 7.33 (dd, *J* = 8.6, 6.2 Hz, 1H, Ph 6-H), 7.20 (ddd, *J* = 8.6, 8.4, 2.6 Hz, 1H, Ph 5-H), 5.06 (s, 1H, 4-H), 4.39 (dd, *J* = 15.5, 5.5, 1H, 2-CH<sub>2</sub>), 4.34 (dd, *J* = 15.5, 5.7, 1H, 2-CH<sub>2</sub>), 3.47 (s, 3H, 3-COOCH<sub>3</sub>), 3.27 (d, *J* = 3.3 Hz, 2H, Pr CH<sub>2</sub>), 3.17 (s, 1H, COOH) 2.02 (s, 3H, 6-CH<sub>3</sub>). <sup>13</sup>C NMR (101 MHz, DMSO-*d*<sub>6</sub>): δ 170.3, 166.8, 166.2 (CO), 160.5 (d, *J* = 247.0 Hz, Ph 4-C), 146.8, 146.6 (2-C, 6-C), 140.2 (d, *J* = 3.5 Hz, Ph 1-C), 131.8 (d, *J* = 9.4

Hz, Ph 6-C), 131.4 (d,  $J = 10.2$  Hz, Ph 2-C), 119.1 (5-CN), 116.0 (d,  $J = 24.4$  Hz, Ph 3-C), 115.4 (d,  $J = 20.9$  Hz, Ph 5-C), 99.3 5-C, 83.9 (3-C), 51.2 (OCH<sub>3</sub>), 42.5 (CH<sub>2</sub>), 39.1 (2-CH<sub>2</sub>), 36.6 (4-C), 17.5 (6-CH<sub>3</sub>).

### S3. Biological evaluation



**Supplementary Figure S1.** Effect of the indicated 1,4-DHP derivatives on MR subcellular localization. See Fig.6 legend for experimental details. White bars, 10  $\mu$ m.

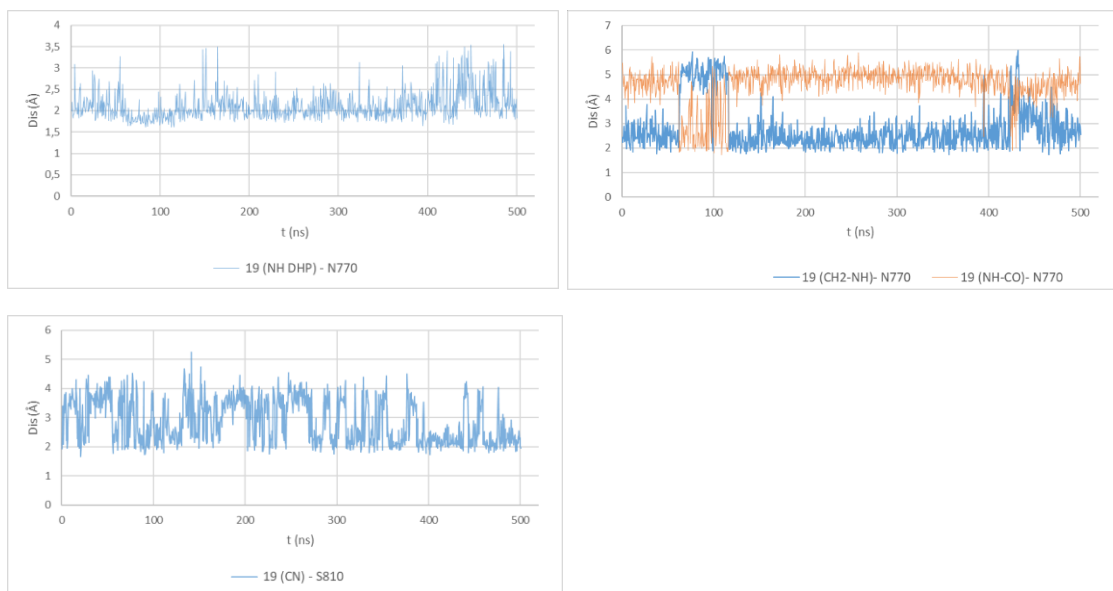


**Supplementary Figure S2.** Functional effects of 1,4-DHP derivatives on GR and AR transactivation properties. Bars represent the average value obtained from three technical replicates in one assay  $\pm$  SE. Statistical analysis was performed with one-way ANOVA followed by Dunnett multiple comparison test where the average of each group was compared to the average of control (unstimulated) condition; n.s., not significant. A. Effects of 5  $\mu$ M ligand treatment on basal GR activity. Induction of GR activity by 500 nM cortisol (CORT) is included to facilitate comparison. Averages were normalized to basal luciferase activity in the absence of any ligand. B. Effects of 5  $\mu$ M ligand treatment on basal AR activity. Induction of AR activity by 10 nM testosterone (T) is included to facilitate comparison. Averages were normalized to basal luciferase activity in the absence of any ligand. C. Effect of 1,4-DHP derivatives (5  $\mu$ M) on GR stimulation by 500 nM cortisol. Averages were normalized to maximal luciferase activity induced by 16h of treatment with the agonist. D. Effect of 1,4-DHP derivatives (5  $\mu$ M) on AR

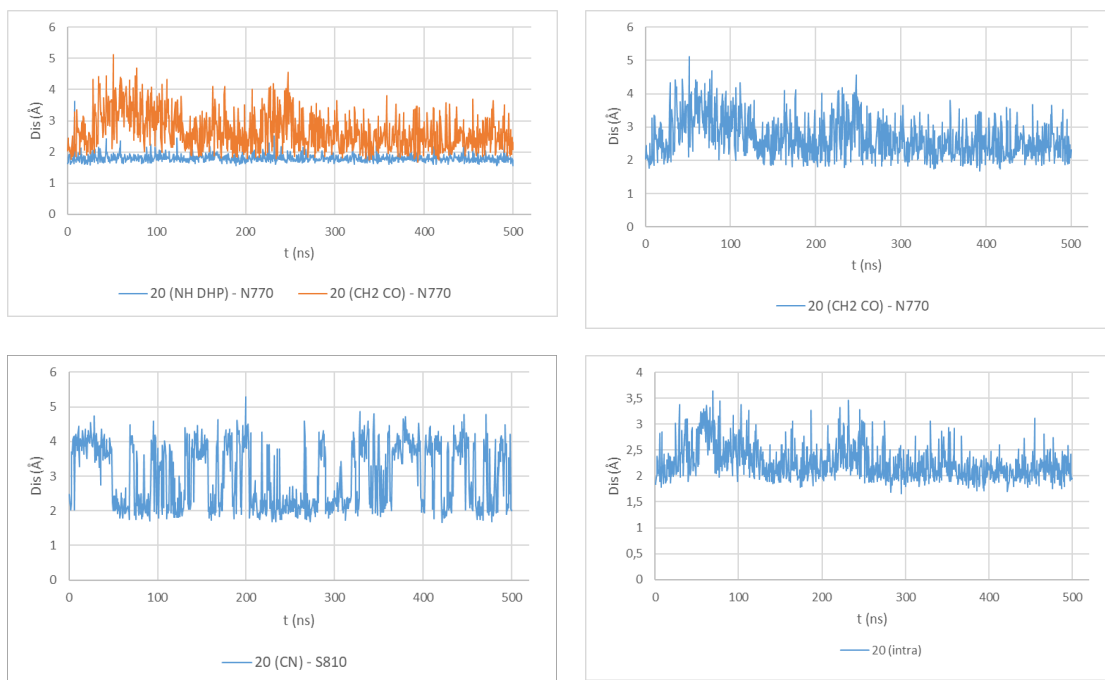
stimulation by 10 nM testosterone. Averages were normalized to maximal luciferase activity induced by 16h of treatment with the agonist.

#### S4. Molecular dynamics studies

Representative distances and angles along the 500 ns simulation for derivatives **11**, **19** and **20** are included in Figures S3-S6.

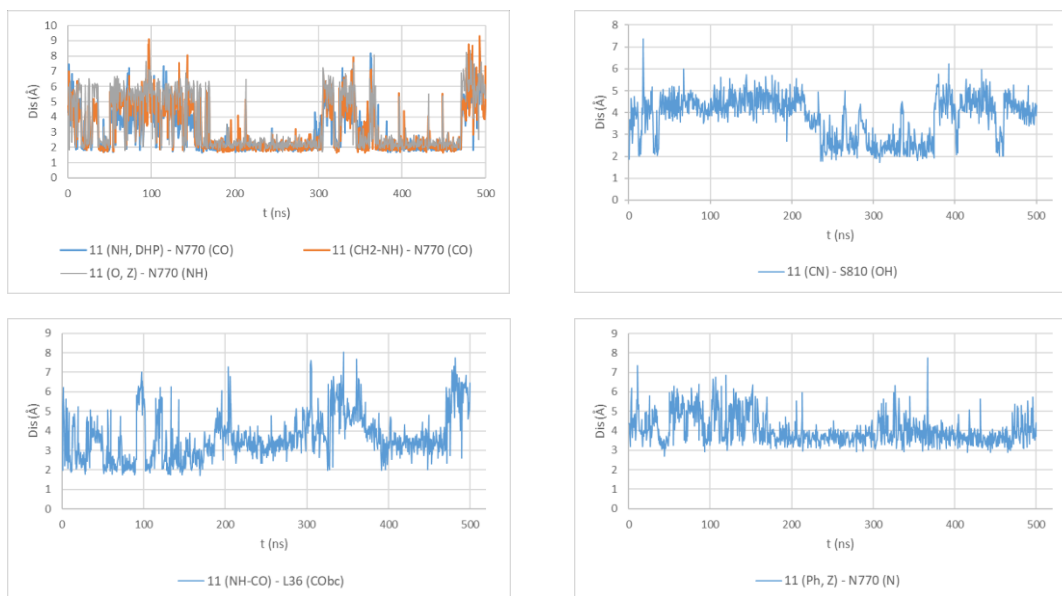


**Supplementary Figure S3.** Representative distances along the 500 ns MD between selected 1,4-DHP **19** and MR atoms.

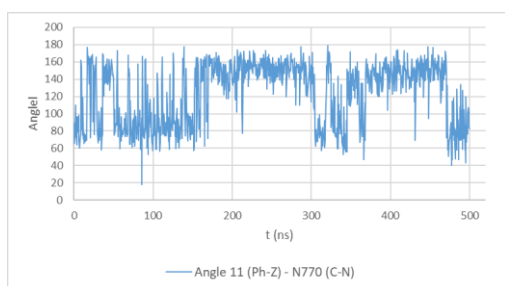


**Supplementary Figure S4.** Representative distances along the 500 ns MD between selected 1,4-DHP **20** and MR atoms.





**Supplementary Figure S5.** Representative distances along the 500 ns MD between selected 1,4-DHP **11** and MR atoms. bc indicates backbone, otherwise it refers to atoms in amino acid side chains.



**Supplementary Figure S6.** Representative angle along the 500 ns MD between the centroid of the benziloxycarbonyl phenyl group of 1,4-DHP **11**, and the N-C atoms of N770 sidechain.

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