



Early-Onset Ovarian Cancer <30 Years: What Do We Know about Its Genetic Predisposition?

Klara Horackova ¹, Marketa Janatova ¹, Petra Kleiblova ^{1,2}, Zdenek Kleibl ^{1,3}, and Jana Soukupova ^{1,*}

- ¹ Institute of Medical Biochemistry and Laboratory Diagnostics, First Faculty of Medicine, Charles University and General University Hospital in Prague, 128 00 Prague, Czech Republic; klara.horackova@lf1.cuni.cz (K.H.); mjana@lf1.cuni.cz (M.J.); pekleje@lf1.cuni.cz (P.K.); zdekleje@lf1.cuni.cz (Z.K.)
- ² Institute of Biology and Medical Genetics, First Faculty of Medicine, Charles University and General University Hospital in Prague, 128 00 Prague, Czech Republic
- ³ Institute of Pathological Physiology, First Faculty of Medicine, Charles University, 128 00 Prague, Czech Republic
- * Correspondence: jana.soukupova@lf1.cuni.cz; Tel.: +420-2249-64266

Abstract: Ovarian cancer (OC) is one of the leading causes of cancer-related deaths in women. Most patients are diagnosed with advanced epithelial OC in their late 60s, and early-onset adult OC diagnosed \leq 30 years is rare, accounting for less than 5% of all OC cases. The most significant risk factor for OC development are germline pathogenic/likely pathogenic variants (GPVs) in OC predisposition genes (including *BRCA1*, *BRCA2*, *BRIP1*, *RAD51C*, *RAD51D*, Lynch syndrome genes, or *BRIP1*), which contribute to the development of over 20% of all OC cases. GPVs in *BRCA1/BRCA2* are the most prevalent. The presence of a GPV directs tailored cancer risk-reducing strategies for OC patients and their relatives. Identification of OC patients with GPVs can also have therapeutic consequences. Despite the general assumption that early cancer onset indicates higher involvement of hereditary cancer predisposition, the presence of GPVs in early-onset OC is rare (<10% of patients), and their heritability is uncertain. This review summarizes the current knowledge on the genetic predisposition to early-onset OC, with a special focus on epithelial OC, and suggests other alternative genetic factors (digenic, oligogenic, polygenic heritability, genetic mosaicism, imprinting, etc.) that may influence the development of early-onset OC in adult women lacking GPVs in known OC predisposition genes.

Keywords: ovarian cancer; early-onset; genetic predisposition; germline pathogenic variant

1. Introduction

Ovarian cancer (OC) accounts for 4.7% of cancer-related deaths in women worldwide [1]. Early detection of OC remains challenging due to the predominance of nonspecific symptoms that occur mainly at advanced clinical stages [2]. The majority of patients are diagnosed with advanced disease with an unfavorable prognosis (5-year survival rate of approximately 50%), which is even worse in cases with metastatic disease (5-year survival rate of approximately 30%) [3]. Thus, it is important to identify women at increased OC risk early, especially when we know that the proportion of hereditary OC is high, reaching even over 20% [4–6]. Surprisingly, despite all assumptions, the abundance of germline pathogenic/likely pathogenic variant (GPV) carriers among early-onset OC patients (<30 years) falls below 10% [7–9], and the genetic predisposition to early-onset OC remains uncertain.

This review focuses on the characterization of early-onset adult OC patients (diagnosed between 18 and 30 years old) from the perspective of cancer predisposition, with a special focus on epithelial OC.



Citation: Horackova, K.; Janatova, M.; Kleiblova, P.; Kleibl, Z.; Soukupova, J. Early-Onset Ovarian Cancer <30 Years: What Do We Know about Its Genetic Predisposition? *Int. J. Mol. Sci.* 2023, 24, 17020. https://doi.org/ 10.3390/ijms242317020

Academic Editor: Marcus Vetter

Received: 25 October 2023 Revised: 27 November 2023 Accepted: 29 November 2023 Published: 30 November 2023

Correction Statement: This article has been republished with a minor change. The change does not affect the scientific content of the article and further details are available within the backmatter of the website version of this article.



Copyright: © 2023 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (https:// creativecommons.org/licenses/by/ 4.0/).

2. Characteristics of Early-Onset and Late-Onset OC

Ovarian cancer is a heterogeneous group, including tumors in the ovaries, fallopian tubes, and peritoneum. Approximately 90% of OC cases are of epithelial origin. The remaining 10% of OC are non-epithelial tumors consisting of germ cell, sex cord-stromal, and other rare non-epithelial ovarian tumors (including sarcoma or small cell carcinoma).

Epithelial OC can be stratified into five major histologic subtypes, including highgrade serous carcinoma (HGSC; accounting for up to 70% of all epithelial OC cases), endometrioid (\sim 10%), clear cell (\sim 10%), mucinous (\sim 3%), and low-grade serous carcinoma (LGSC; <5%) [10]. The histopathologic classification of epithelial OC largely determines the clinical course of the disease. From this perspective, epithelial OC is divided into tumors with a good (type I) or poor (type II) prognosis. Type II OC is more prevalent as it comprises the most common HGSC and several less common OC subtypes, including carcinosarcoma and other mixed or undefined epithelial OC. Type II is characterized by frequent abnormalities in p53-related and homologous recombination (HR) DNA repair pathways, resulting in genome instability, and includes high-grade tumors with high proliferative potential and rapid progression, contributing to the late diagnosis at advanced stages [11,12]. In contrast, less common type I ovarian tumors include LGSC, clear cell carcinoma, endometrioid carcinoma, mucinous carcinoma, seromucinous carcinoma, and transitional cell carcinoma. Compared to the known genetic instability of type II tumors, type I tumors are relatively genetically stable. Type I OC are typically low-grade tumors with low proliferative activity and slow progression developing from benign lesions, particularly borderline tumors of the ovary, and, therefore, these tumors are more often diagnosed at an earlier stage [13,14].

Typically, OC develops in late adulthood, with a median age at diagnosis of 63 years [3]. Extremely early-onset ovarian tumors diagnosed between 18 and 30 years of age account for less than 5% of all OC cases (Figure 1) [3,15], and owing to their rarity, only a few studies have been conducted up-to-date. However, all the reports pointed out some striking differences between late- and early-onset adult OC, including genetic background, clinicopathologic features, or clinical outcomes compared with late-onset tumors (Table 1). Notably, while type II OC and particularly HGSC predominate in late-onset OC, approximately 50% of early-onset ovarian tumors are of germ-cell origin (representing a juvenile form of ovarian tumors diagnosed most frequently between the ages of 15 and 20 years), and only approximately 40% of the tumors belong to epithelial OC, particularly LGSC [16]. Correspondingly, late-onset OC is typically diagnosed at advanced stages, frequently with metastatic spread [3,17,18], whereas early-onset OC patients are typically diagnosed with localized disease. This would imply a better prognosis for early-onset OC patients [3,17,19], as also indicated by the significant survival advantage shown in a population-based study of OC patients [17]. However, the age at disease onset has not been confirmed as an independent stratifying factor concerning the prognosis for early-onset OC patients [19–22]. Particularly, in the long-term follow-up study, Gershenson et al. [22] observed a significantly worsened outcome, including both progression-free survival and overall survival in early-onset (\leq 35 years) compared with late-onset OC patients. Although LGSC generally have a more favorable prognosis in general [3,16,18], LGSC in early-onset OC patients have a worse prognosis and lower 5-year survival [22], providing another reason for separating this cohort from the majority of late-onset OC.

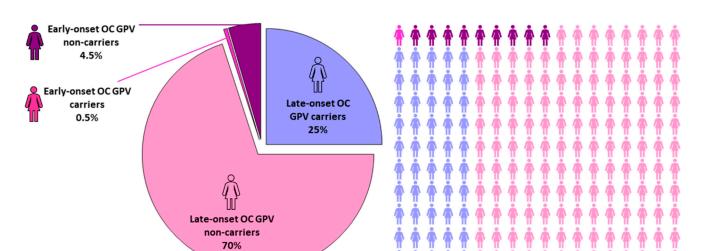


Figure 1. Frequency of germline pathogenic/likely pathogenic variants in established cancer predisposition genes in early-onset vs. late-onset OC patients.

Patients	Early-Onset OC	Late-Onset OC	
Incidence in females	1.6/100,000 [3,15]	22.0/100,000 [3,15]	
5-year relative survival rate	58–87% [17,19,20,23] -lower in LGSC [22]	Approx. 50% [3,17]	
Clinicopathology			
	~40% epithelial—LGSC prevails [16,17]	~90% epithelial—HGSC prevails (70%) [13,26]	
Histology	~50% germ-cell [16,24]	~6% sex cord-stromal [6]	
	~10% sex cord-stromal [24,25]	~3% germ-cell [6]	
Dominant tumor stage	Localized disease [3]	Distant disease [3]	
Genetic predispositions			
GPV	Low <10% [7–9]	High >20% [4,5,9]	
	GPV, germline pathogenic/likely pathogenic varia	nt; HGSC, high-grade serous carcinoma; LGSC, low-grade	

Table 1. Characteristics of early-onset and late-onset OC.

GPV, germline pathogenic/likely pathogenic variant; HGSC, high-grade serous carcinoma; LGSC, low-grade serous carcinoma; OC, ovarian cancer.

3. OC Risk Factors and Predisposition

The lifetime OC risk is about 1.1% in developed countries [3] and positively associated with lifetime ovulatory years (except for rare mucinous tumors) [27]. Moreover, the relative risk (RR) of OC development can increase approximately three times in cases of positive family epithelial OC history [28]. This is related to the high proportion of hereditary forms of OC, as >20% of OC patients are carriers of a GPV in an established cancer predisposition gene (Figure 1) [4,5,9]. Nevertheless, the frequency of GPV carriers differs substantially among patients with various histological subtypes. The highest proportion of GPVs are present in epithelial type II OC and, particularly in HGSC, the most prevalent late-onset OC, while the proportion of GPVs decreases in type I OC, including LGSC, and is the lowest in clear cell and mucinous type I carcinomas [29]. The predisposition to non-epithelial ovarian tumors is much less understood.

3.1. Established OC Predisposition Genes

The genetic predisposition to epithelial OC is well established, with major contributions from GPVs in genes coding for HR and mismatch repair (MMR) proteins (Table 2). The *BRCA1* and *BRCA2* HR genes are the most commonly germline-altered cancer predisposition genes responsible for the development of hereditary breast and ovarian cancer (HBOC) syndrome. GPVs in both genes could be found in nearly 20% of all OC patients [30]. The OC risk for *BRCA1* and *BRCA2* carriers rises significantly from the age of 35 and 45 years and reaches 58% and 29%, respectively [31,32]. An order of magnitude lower frequency but still significant risk is associated with carriers of GPVs in other HR genes, RAD51D, RAD51C, and BRIP1, having a life-time risk of epithelial OC of 20%, 15%, and 15%, respectively [33,34]. The median age of OC onset in BRCA1 GPV carriers is significantly lower (53 years) compared to BRCA2, RAD51D, RAD51C, and BRIP1 GPV carriers (59, 57, 62, and 65 years, respectively) or the general population; however, tumors before the age of 30 are rare, which is also reflected by the clinical management guidelines [31,33–36]. An increased risk of epithelial OC has also been documented for carriers of GPVs in MMR genes associated with Lynch syndrome (LS) (Table 2). The life-time risk varies in a range from a few percent (MSH6 and PMS2) up to 20% (MLH1) and 38% (MSH2/EPCAM) [37]. However, while MLH1, MSH2, and MSH6 are strongly associated with OC, the role of PMS2 in OC predisposition is limited. Lynch syndrome OC patients are typically younger, with a median age at diagnosis of 43 and 46 years for MSH2 and MLH1, respectively [38,39], and have also been described in early-onset OC patients [40]. In addition to the high-penetrance epithelial OC predisposition genes, GPVs in other genes involved in double strand break repair, including ATM and PALB2, have been associated with moderate risk to late-onset epithelial OC [41–43]. However, the clinical utility of the moderate-penetrance genes is low and largely depends on family OC history.

In addition, other rarely mutated genes are associated with non-epithelial ovarian tumors (Table 2). The GPVs in *STK11*, causing rare Peutz–Jeghers syndrome, are associated with a non-epithelial ovarian tumor risk exceeding 10% and the development of early-onset tumors in patients <30 years [44–46]. Only a few episodical reports document the development of early-onset epithelial OC in carriers of GPVs in *STK11* (Table 2) [4,41]. GPVs in *DICER1* have been linked to early-onset, non-epithelial sex cord-stromal ovarian neoplasms [47,48] accounting for nearly half of the stromal ovarian tumors [49]. However, *DICER1* appears to be exclusively characteristic of non-epithelial, early-onset ovarian tumors. Furthermore, GPVs in *SMARCA4* were associated with small cell carcinoma of the ovary hypercalcemic type (SCCOHT), a rare, aggressive OC similar to malignant rhabdoid tumors that primarily affects women under 40 years of age [50]. GPVs in *SMARCA4* have been reported recently in two early-onset OC patients; nevertheless, selectively in SCCOHT [51].

Generally, early cancer onset indicates the involvement of hereditary cancer predispositions [52,53]. Thus, one might anticipate an enrichment of GPV carriers in high-penetrance genes in early-onset OC patients. However, this assumption does not apply for early-onset epithelial OC patients, who were found to rarely carry GPVs, with the frequency not exceeding 10% (Figure 1) [4,7–9,38]. Nevertheless, only a limited number of studies focusing on the genetic predispositions of early-onset OC have been performed so far (Table 3).

Table 2. Established OC predisposition genes.

	Heterozygote	Homozygote/				
Gene	Associated OC Histotype	Absolute Risk for OC [35]	GPV Identified in Early-Onset OC	Other Associated Cancer Types [35]	Compound Heterozygote [54]	
High penetrance						
BRCA1	Epithelial [41]	39–58%	Yes [4,9]	BC, PaC, PrC	FA-S	
BRCA2	Epithelial [41]	13-29%	Yes [4]	BC, PaC, PrC, MM	FA-D1	
BRIP1	Epithelial [41]	5-15%	Yes [4,9]	BC, CrC, EC	FA-J	
DICER1	Sex cord-stromal [47]	NA	Yes [47,48]	DICER1 sy	-	
MLH1	Epithelial [37,41]	4–20%	No	Lynch sy—CrC, EC, PaC	CMMRD	
MSH2	Epithelial [37]	8–38%	Yes [40,52]	Lynch sy—CrC, EC, PaC	CMMRD	

	Heterozygote	Heterozygote						
Gene	Associated OC Histotype	Absolute Risk for OC [35]	GPV Identified in Early-Onset OC	Other Associated Cancer Types [35]	Compound Heterozygote [54]			
High penetranc	ce							
RAD51C	Epithelial [41]	10-15%	Yes [4]	BC	FA-O			
RAD51D	Epithelial [41]	10-20%	No	BC	-			
SMARCA4	SCCOHT [50]	NA	Yes [50,51]	Rhabdoid tumor predisposition sy	-			
STK11	Non-epithelial [45]	>10%	Yes [46]	Peutz–Jeghers sy, BC, PaC, CrC	-			
Moderate penet	trance/Insufficient evidenc	e						
ATM	Epithelial [41]	2–3%	Yes [4,9]	PaC	AT			
MSH6	Epithelial [37]	1–13%	No	Lynch sy—CrC, EC, PaC	CMMRD			
PMS2	Epithelial [37]	1–3%	Yes [40,52]	Lynch sy—CrC, EC	CMMRD			
PALB2	Epithelial [42]	3–5%	No	BC, PaC	FA-N			

Table 2. Cont.

AT, ataxia-telangiectasia; BC, breast cancer; CMMRD, constitutional mismatch repair deficiency syndrome; CrC, colorectal cancer; EC, endometrial cancer; FA, Fanconi anemia; GPV, germline pathogenic/likely pathogenic variant; MM, malignant melanoma; NA, not available; OC, ovarian cancer; PaC, pancreatic cancer; PrC, prostate cancer; SCCOHT, small cell carcinoma of the ovary hypercalcemic type; sy, syndrome.

Table 3. Studies involving early-onset OC patients	tients.
----------------------------------------------------	---------

		Study Details	No. of Tested Genes *	No. of All OC Patients	No. of Early-Onset OC Patients	Range of Early-Onset Patients' Age at Dg.	Early-Onset OC Patients		
Study Population	Population						No. of High- Penetrance GPV Carriers	GPVs in Established High- Penetrance OC Predis- position Genes	GPVs in Candidate OC Predis- position Genes
Stratton (1999) [8]	UK	Early-onset epithelial OC	4 **	169	169	13–30	0 ***	0	0
Carter (2018) [9]	US	OC	15	4439	147	6–30	2 (1.4%)	1×BRCA1; 1×BRIP1	3×ATM; 1×BARD1; 5×CHEK2
Lhotova (2020) [4]	CZ	OC	219	1333	84	15–30	6 (7.1%)	2×BRCA1; 1×BRCA2; 2×RAD51C; 1×STK11	1×ATM; 1×BARD1; 4×CHEK2; 1×NBN;
Herold (2023) [51]	GER	OC	25	206	83	13–30	3 (3.6%)	1× BRIP1; 2×SMARCA4	$\begin{array}{c} 1 \times FANCM; \\ 1 \times MUTYH \\ het; \\ 1 \times PMS2; \\ 1 \times TP53 \end{array}$
Flaum (2023) [40]	UK	Early-onset OC	15	77	77	15–30	4 (5.2%)	$4 \times MSH2$	$1 \times PMS2$
Bernards (2015) [38]	US	Early-onset OC	18	47	5	27–30	0	0	0
Felicio (2020) [55]	BRA	BRCA neg., TP53 neg. HBOC patients	WES	11	3	20–21	0	0	1×CHEK2
Da Costa (2020) [56]	BRA	HBOC	21	6	2	22–30	0	0	0
Boyd (2000) [57]	Jew	OC	2	189	1	25	0	0	0
Hajkova (2019) [58]	CZ	Synchronous EC and OC	73	22	1	29	0	0	1×BARD1
Jarhelle (2019) [59]	NOR	HBOC BRCA1/BRCA2 neg.	94	20	1	27	0	0	1×CHEK2

Cont.

							Earl	y-Onset OC Pati	ents
Study	Population	Study Details	No. of Tested Genes *	No. of All OC Patients	No. of Early-Onset OC Patients	Range of Early-Onset Patients' Age at Dg.	No. of High- Penetrance GPV Carriers	GPVs in Established High- Penetrance OC Predis- position Genes	GPVs in Candidate OC Predis- position Genes
Risch (2001) [7]	US	OC	2	649	NA (96 <40 yo)	20–30	0	0	0
Koczkowska (2018) [60]	PL	OC	25	333	NA	NA	0	0	1×CHEK2
Pal (2005) [61]	US	epithelial OC	2	209	NA (11 <40 yo)	18–30	0	0	0
Ryan (2017) [52]	UK	LS assoc. preselected OC positive for LS-GPV	4	53	NA	24–30	2	2×MSH2	1× <i>PMS2</i> biallelic
Hajkova (2019) [58]	CZ	Synchronous EC and OC	73	22	1	29	0	0	1×BARD1

Dg, diagnosis; GPV, germline pathogenic variants; HBOC, hereditary breast and ovarian cancer; LS, Lynch syndrome; OC, ovarian cancer; NA, not available; No.; number; WES, whole exome sequencing; yo, years old; * The list of tested genes in each study is available in Supplementary Table S1; ** in the case of *BRCA2*, only the OC cluster region was analyzed; *** identified variants; *MLH1*: c.1853A > C/p.Lys618Thr and *MLH1*: c.1000G > C/p.Asp304His are not currently considered GPVs.

This controversy was first acknowledged by Stratton et al. in 1999 [8], who tested only a few genetic loci (BRCA1, MLH1, MSH2, and a part of BRCA2) in the largest cohort of early-onset OC patients up to date. Since then, few studies have been conducted, but all of them confirmed a low frequency of GPVs among early-onset OC patients diagnosed <30 years [4,7,9,38,40,51,56,57]. Four recent larger studies, Carter et al. (2018) [9], Lhotova et al. (2020) [4], Flaum et al. (2023) [40], and Herold et al. (2023) [51], identified 2/147 (1.4%), 6/84 (7.1%), 4/77 (5.2%), and 3/83 (3.6%) GPV carriers in established high-penetrance OC predisposition genes (including 0.7%, 3.6%, 0%, and 0% mutations in BRCA1/BRCA2), respectively (Table 3). All four studies used the panel NGS approach, but with a different range of analyzed genes. While Carter [9] and Flaum [40] analyzed 15 and Herold [51] included 25 established cancer predisposition genes in their panel (Supplementary Table S1), Lhotova [4] used a much wider panel, targeting 219 established and candidate cancer predisposition genes. Despite the different design of the analysis, the similar results pointing to a very low frequency of GPV carriers in early-onset OC patients were strikingly similar. Interestingly, Flaum [40] associated GPVs in MSH2 with early-onset OC, as 5.2% (4/77) of the patients carried the same MSH2 GPV. This association was in coherence with the findings of another study focusing on OC in LS patients that included also three early-onset OC patients (Table 3) [52]; however, no other study further supported MSH2-association with early-onset OC. Due to the rarity of early-onset OC, further studies evaluating genetic predisposition to OC have included only a few OC cases with diagnoses at such a young age [52,55,58–60], leading to a limited understanding of the genetic factors underlying early-onset OC development. The overall lack of GPV carriers among earlyonset OC patients is, however, evident and implies the need for the separation of this cohort from OC. Based on the frequency of GPVs in established OC predisposition genes, the cut-off age for distinguishing between early- and late-onset epithelial OC is around 30, as the frequency of BRCA1 GPVs (a major genetic contributor to OC) starts to increase from age 35 [4].

3.2. Candidate OC Predisposition Genes

Candidate OC predisposition genes can be proposed based on their predisposition to other cancer types or associated diseases, which, however, have not yet been associated with OC (Table 4). The high-penetrance cancer predisposition genes *APC*, *BMPR1A*, *BAP1*, *FH*, *MEN1*, *PTEN*, *VHL*, *WT1*, and *TP53* (on top of their strong association with gastrointestinal tumors, melanoma, leiomyomatosis, multiple endocrine neoplasia, hamartomas, kidney

tumors, breast cancer (BC), central nervous system tumors, and sarcomas, respectively) were also identified in OC patients and/or families [5,62–70], suggesting a potential wider cancer manifestation in these syndromes. Moreover, *BARD1* and *CHEK2* [5,69–75] were associated with a high to moderate risk of other cancer types, but their contribution to OC risk remains to be confirmed. Nevertheless, GPVs in both of these genes have been reported in several early-onset OC patients [4,9,55,58–60].

Furthermore, as the majority of established OC predisposition genes (Table 2) code for proteins involved in DNA repair and/or DNA damage response, the new candidate OC predisposition genes are often sought among genes involved in these pathways. GPVs in genes of the Fanconi anemia complex (including *FANCA*, *FANCC*, *FANCL*, *FANCM*, and *SLX4*), MRN complex genes (*MRE11-RAD50-NBN*), and other genes associated with DNA repair pathways have been reported in OC patients, suggesting their possible role in OC predisposition [5,41,51,55,68,75–85]. Moreover, new potential candidate genes emerged from complex sequencing studies of OC patients without being previously clearly established with any cancer, namely *ABRAXAS1* (also known as *FAM175A*), *CNKSR1*, and *PIK3C2G* [56,86,87]. However, it cannot be ruled out that all these more or less isolated findings may be coincidental and do not significantly affect the OC risk. Summarizing, the role of all the above-mentioned, candidate genes in early-onset OC is unclear, calling for further investigation in this field.

Gene	GPVs Identified in Early-Onset OC	Associated Disease (Inheritance Mode) [54]	OC-Association Reported in
ABRAXAS1 (FAM175A)	No	-	[56,86]
ATR	No	Cutaneous telangiectasia and cancer sy (AD)	[78]
APC	Yes (early 30s) [63]	Familial adenomatous polyposis (AD)	[62,63]
BAP1	No	Melanoma (AD)	[68]
BARD1	Yes [6,25,55,59,60]	BC (AD)	[71]
BLM	No	Bloom sy (AR)	[5,76]
BMPR1A	Yes [64]	Juvenile polyposis sy, primary ovarian insufficiency (AD)	[64]
BRAT	Yes (in their 30s) [85]	Neurodevelopmental disorder (AR)	[85]
CNKSR1	No	-	[87]
CDKN2A	No	MM, MM-PaC sy (AD)	[5,72]
CHEK2	Yes [6,25,55,59,60]	BC (AD)	[5,73]
ERCC3	No	Trichothiodystrophy, xeroderma pigmentosum (AR)	[88,89]
FANCA	No	FA (AR)	[77,78]
FANCC	No	FA (AR)	[77]
FANCL	No	FA (AR)	[77]
FANCM	Yes [51]	-	[77,79]
		Leiomyomatosis and renal cell	
FH	No	cancer (AD), fumarase	[68]
		deficiency (AR)	
MEN1	No	Multiple endocrine neoplasia (AD)	[65]
MRE11	No	AT-like disorder (AR)	[81,84]
NBN	Yes [25]	Nijmegen breakage sy (AR)	[41]
NF1	No	Neurofribromatosis (AD)	[5]
PIK3C2G	No	-	[87]
POLD1	No	CrC, EC (AD)	[74]

Table 4. Candidate OC predisposition genes.

Gene	GPVs Identified in Early-Onset OC	Associated Disease (Inheritance Mode) [54]	OC-Association Reported in
POLE	No	CrC, EC (AD), IMAGE-I sy (AR)	[73]
POLK	No	-	[75]
PTEN	Yes [66]	Cowden sy (AD)	[66,67]
RAD50	No	Nijmegen breakage syndrome-like disorder (AR)	[84]
RAD51B	No	-	[5,82]
RAD52	No	-	[83]
RAD54B	No	-	[83]
RAD54L	No	-	[55]
RB1	No	Retinoblastoma (AD)	[5]
RTEL1	No	Dyskeratosis congenita (AD/AR), telomere-related pulmonary fibrosis, and/or bone marrow failure sy (AD)	[68]
SLX4	No	FA (AR)	[75,81]
TP53	Yes [51,69]	Li–Fraumeni sy (AD)	[5,69,70]
TSC2	No	Tuberous sclerosis (AD)	[68]
VHL	No	von Hippel–Lindau sy, pheochromocytoma (AD)	[68]
WT1	No	Wilms tumor (AD)	[68]
XRCC3	No	-	[82]

Table 4. Cont.

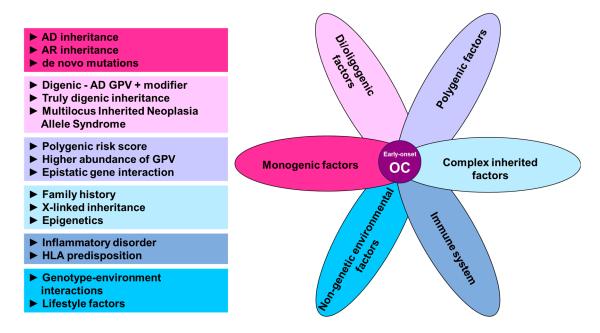
AD, autosomal dominant; AR, autosomal recessive; AT, ataxia-telangiectasia; BC, breast cancer; CrC, colorectal cancer; FA, Fanconi anemia; IMAGE, intrauterine growth restriction, metaphyseal dysplasia, adrenal hypoplasia congenita, and genitourinary abnormalities; MM, malignant melanoma; PaC, pancreatic cancer; sy, syndrome.

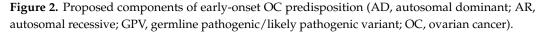
4. Alternative Approaches to Germline Genetic Testing in Early-Onset OC

The lack of identified GPVs in established and candidate cancer predisposition genes in early-onset OC patients could indicate a different disease-causing genetic basis compared to late-onset OC. Applying wider-scope germline whole exome sequencing (WES) or even whole genome sequencing (WGS) may provide new insight into early-onset OC genetics. WES has been used for the genetic analysis of OC (including some early-onset cases) [5,55,84]; however, the results have not shed light on an apparent, clinically relevant, new predisposition gene. Promisingly, WGS could identify yet missed variants (including deep intronic, untranslated regions, and copy number variants) in established and candidate or completely newly associated genes involved in OC predisposition [90–92]. Considering complex WES/WGS data analysis and interpretation, complementation with RNA-NGS could help to better understand the cancer-predisposition molecular mechanisms [93]. Nonetheless, these methods are currently not commonly used in clinical practice, as the limitations are not only the substantial cost of the testing and the massive abundance of the NGS data, but most importantly, the unclear clinical significance of the identified variants [94,95].

4.1. Alternative Ways of Cancer Predisposition Inheritance

Apart from applying new testing and analytical methods, various genetic and nongenetic factors (Figure 2) need to be considered in a complex analysis of early-onset OC predisposition. The association of proposed genetic components, as well as other factors predisposing to OC mentioned below has not been established for early-onset OC yet, but there are some pieces of evidence suggesting it could cause or contribute to early-onset cancer or to OC development in general.





In addition to autosomal dominant Mendelian inheritance, recessive inheritance may rarely contribute to the early-onset of OC, as exemplified by isolated cases of biallelic GPV carriers in *BRCA1* [96] or *PMS2* [52]. These sporadic cases suggest that recessive inheritance represents an uncommon cause of early-onset OC. Similarly, the development of de novo GPVs in *BRCA1* has been reported in rare cases of early-onset HBOC patients [97,98]. Also, a de novo GPV in *SMARCA4* has been observed in a patient with early-onset OC and other types of childhood/early-onset rhabdoid tumors [99]. The low frequency of de novo mutations among OC patients may have been biased by the HBOC genetic testing criteria, which used to prioritize patients with strongly positive family histories [100]. As early-onset OC patients lack a strong family cancer history [8], the de novo or compound heterozygous OC-predisposition mutations may be under-reported, and their identification, e.g., using a trio WES (analysis of the patient + her parents) similar to other rare childhood/early-onset diseases [101] may help to uncover the underlying genetic causes.

In addition, constitutional mosaicism of GPVs (with lower variant allele frequency in peripheral blood) in established/candidate genes with Mendelian inheritance may be a specific issue. Constitutional mosaicism of GPVs in *BRCA1* and *BRCA2* [98,102,103] has been previously reported in HBOC patients, suggesting that even the low-level mosaic GPVs in peripheral blood may be significant for the phenotype.

Finally, epigenetic inactivation can contribute to hereditary OC, as shown by promoter methylation of *BRCA1* [104], or *MLH1* in LS patients recently [105]. However, the prevalence of this phenomenon is largely unknown due to the limited data.

4.2. Family History and X-Linked Inheritance

Despite the low proportion of strong cancer family history in early-onset OC patients compared to patients with OC diagnosed > 30 years, the familiar form of early-onset OC still raises some important questions. Stratton et al. [8] described a slightly elevated OC risk, but also significantly enriched non-Hodgkin lymphoma and myeloma among first-degree relatives of early-onset OC patients (in the majority of cases without known germline genetic predisposition). Similarly, Rantala et al. [106] associated an increased incidence of early-onset (<40 years) OC and testicular cancer in patients whose aunts suffered from early-onset BC. On the other hand, Imbert-Bouteille et al. [107] found no relation between the occurrence of early-onset OC in families with *BRCA1/BRCA2* mutations and the age at

diagnosis of BC or OC in their relatives. When focusing on the sex of the affected relatives, Stratton et al. [108] noticed that an affected woman's sisters are at higher risk of disease than their mothers. In sync, Eng et al. described an X-linked association between prostate cancer in men and OC in their mothers and daughters [109]. These somewhat contradictory findings about the importance of family history in early-onset OC predisposition may point in the direction of an X-linked inheritance.

Apart from single nucleotide variants, X-linked OC predisposition can include complex genetic effects like X chromosome inactivation (XCI). Buller et al. [110] described a higher frequency of skewed XCI among OC patients, suggesting X-linked tumor suppressor genes or X-linked low penetrance susceptibility alleles affected by the inactivation pattern [111]. On the other hand, skewed XCI has been described at an increased frequency in BRCA1 mutation carriers compared with controls, and is associated with a statistically significant increase in age at diagnosis of breast and ovarian cancer in *BRCA1/BRCA2* GPV carriers [112]. In addition to the effect on the age of onset, molecular signatures of XCI were associated with clinical outcomes in epithelial OC, as patients with dysregulated XCI had shorter progression and overall survival than those with regulated XCI [113], suggesting a complex involvement of the X chromosome in OC.

4.3. Polygenic Inheritance

A complex polygenic inheritance stemming from an additive effect of multiple genetic variants may also be involved in early-onset OC predisposition (Figure 2). Polygenic inheritance could also explain the observed lack of positive family history in close relatives of early-onset OC patients [8]. A polygenic model of inherited predisposition to cancer was proposed by Qing et al. [114], who identified a higher burden of germline variants in protein-coding cancer hallmark genes predicted to alter the structure, expression, or function in early-onset patients compared to late-onset ones. They hypothesized that the early-onset carriers of more germline low-risk variants needed to harbor fewer somatic mutations for malignant transformation. Their hypothesis was supported by a significant association with several cancer types, including the OC.

Currently, hundreds of cancer-risk single nucleotide polymorphisms (SNPs) have been identified by genome-wide association studies, improving their understanding but not fully uncovering their polygenic heritability. These SNPs are considered causal or linked to causal variants. Using a polygenic risk score (PRS), some studies have recently shown a cumulative impact of SNPs in patients with several cancer diagnoses, including OC [115]. Especially serous OC showed association with PRS [116,117], and, interestingly, association was also found between PRS and early diagnosis of BC [118], together suggesting PRS might be associated with early-onset OC. In addition to PRS, another OC risk score evaluating epistatic gene interactions via chromosomal-scale length variation was proposed [119].

4.4. Di/Oligenic Inheritance

Since both monogenic and polygenic inheritance in OC have been demonstrated, the involvement of digenic to oligogenic inheritance might presumably also be a part of the OC genetic predisposition (Figure 2). In general, several di/oligogenic mechanisms have been suggested.

Firstly, a higher incidence and earlier onset of cancer have been proposed in carriers of two or more GPVs in high-penetrance cancer predisposition genes called multilocus inherited neoplasia allele syndrome (MINAS). However, *BRCA1/BRCA2*-MINAS patients did not develop OC significantly earlier than single GPV carriers [120], further disputing the *BRCA1/BRCA2* involvement in early-onset OC predisposition.

Secondly, a combination of a monogenic GPVs and a phenotype-modifying (e.g., age of onset) variant were reported. Carriers of GPVs in established OC-predisposition genes *BRCA1/BRCA2*, together with the newly associated gene *PPARGC1A*, were diagnosed at significantly earlier ages, suggesting *PPARGC1A* is a modifier of OC onset in *BRCA1/BRCA2* carriers [121]. Moreover, ethnically-specific modifiers were proposed to

influence the phenotype in *BRCA1* GPV carriers [122], suggesting missing clues even in *BRCA1/BRCA2* otherwise well-established OC predisposition. On the other hand, focusing on *BRCA1/BRCA2* negative OC patients, Eng et al. [109] proposed one X-linked SNP in the *MAGEC3* gene to advance the age of OC onset by almost seven years. The OC risk or age at onset might also be modified by genetic variants in regulatory elements such as miRNA. Dysregulation of *BRCA1/BRCA2* functions in OC was reported in the absence of *BRCA1/BRCA2* GPVs stemming from miRNA dysregulation [123], and, especially, miR-146a polymorphism was associated with an earlier age of onset in *BRCA1/BRCA2*-negative HBOC patients [124].

Thirdly, a truly digenic disorder might be considered. Despite several clearly reported oligogenic disease associations, such as familial hemophagocytic lymphohistiocytosis, primary immunodeficiency, or familial hypercholesterolemia [125–128], the majority of proposed di/oligogenic allele combinations found in affected patients remain of uncertain significance [129]. Currently, we can only hypothesize about combinations of genetic variants that are separately nonpathogenic (e.g., missense) but together pathogenic when present in mutually interacting domains of proteins involved in OC predisposition, asking for a battery of functional tests and clinical investigation. Despite the currently unavailable clear association with early-onset OC, some evidence about the di/oligogenic inheritance involvement in cancer and, particularly, OC predisposition was proposed. Mouse model-based results suggest that oligogenic inheritance is also a part of cancer predisposition [130], supported by the first few reported clinical cases of digenic colorectal and gastric cancer in the OLIDA (OLIgogenic diseases DAtabase) [129].

4.5. Immune-Related Modifiers of OC

To further expand the complexity of OC predisposition, the immune system is also involved in cancer development (Figure 2). It has already been established that local chronic inflammation and autoimmune disease, including genetic immune dysregulations, predispose to certain types of cancer, such as autoimmune hepatitis-induced cirrhosis to hepatocellular carcinoma [131], Helicobacter pylori infection to gastric cancer [132], or inflammatory bowel disease to colorectal cancer [133]. Therefore, one could speculate that a systemic or ovarian-localized inflammatory condition may also, with time, lead to OC, possibly on an immunogenetic inherited basis. Currently, the knowledge of inherited immunity aspects associated with OC is limited, but a few examples have been described in the literature. Namely, systemic lupus erythematosus was reported to be associated with a higher incidence of cancer, including ovarian cancer [134]. Moreover, a significantly increased incidence of specific HLA-class II haplotypes (namely, DRB1*0301-DQA1*0501-DQB1*0201 and DRB1*1001-DQA1*0101-DQB1*0501) has been observed in OC patients, suggesting their role in OC pathogenesis [135].

4.6. Non-Genetic Factors

Similarly, genotype-environment interactions are a great unknown in the field of OC predisposition. The fact that the interaction of exposure to environmental carcinogens and constitutional genetics modulates cancer risk was first suggested in autosomal recessive Bloom and Werner syndromes caused by biallelic mutations in *BLM* and *WRN*, respectively [136]. Since then, several genotype-environment interactions have been recognized, including the combined presence of GPVs in *BAP1* and exposure to asbestos fibers, which together increase the risk of disease more than either component alone [137]. Moreover, the genotype-environment interaction may possibly modulate phenotypic heterogeneity, as shown in Birt–Hogg–Dubé syndrome caused by GPVs in *FLCN* or suggested for *DICER1* [138,139], which were also both reported in families with OC cases [47,48].

Moreover, lifestyle can also modify pre-existing OC predisposition. Namely, alcohol intake elevates the risk for OC [140], as well as exposure to smoking in childhood. Interestingly, smoking exposure was more likely associated with LGSC and non-serous OC [141], which are more frequent OC types among early-onset OC patients [16,17]. On the other

hand, smoking has also been identified as a protective factor in endometroid and clear-cell OC [142]; however, smokers among early-onset OC patients have a short smoking history, limiting the effect of this risk factor.

Finally, we can also raise the question of whether survival differs in extremely young OC patients compared to histology-matched late-onset patients, as shown in early-onset LGSC OC patients with a less favorable prognosis and decreased 5-year survival [22]. Isolated studies point to some predictive markers associated with better survival in ovarian cancer patients, such as *PRDM1* variant rs2185379, which is suggested to be positively associated with the long-term recurrence-free survival of advanced OC [143]. On the other hand, the *EXO1* variant rs851797 was negatively associated with progression-free and overall survival in OC patients [144]. However, these isolated association studies typically deal with HGSC with late-onset and focus on patients from a specific population and should be interpreted with caution.

5. Conclusions

Despite being rare, early-onset OC represents a significant healthcare and socioeconomic problem, and its development is currently poorly understood. The risk of the disease can be modulated not only by genetics but also by environmental/lifestyle factors and/or their interactions. Supported by the rarity of the disease, it cannot be ruled out that early-onset OC (<30 years) is an unpredictable stochastic event caused just due to chance by random variables of different values.

Knowledge of the genetic factors could help identify women at risk of early-onset OC and might also be of predictive and prognostic value. Nevertheless, this is currently impossible, as we do not fully understand the factors associated with OC risk at such a young age. Since early-onset OC shows a significantly distinct germline mutation status compared to late-onset OC and the number of studies analyzing the GPVs in early-onset OC is limited, it is necessary to investigate early-onset OC patients in detail and separately from late-onset OC. Finding new approaches to germline genetic testing of early-onset OC patients is crucial for the identification of the expected yet unknown genetic causes, if present. These new insights could not only help to better understand the specifics of early-onset OC and provide new potential diagnostic and preventive targets for patient care and counseling, but they could also help to tailor the treatment modalities, thus improving the quality of life and survival of the early-onset OC patients.

Supplementary Materials: The following supporting information can be downloaded at: https://www.mdpi.com/article/10.3390/ijms242317020/s1.

Author Contributions: K.H.: writing—original draft. M.J.: funding acquisition, writing—review and editing. P.K.: funding acquisition, writing—review and editing. Z.K.: funding acquisition, writing—review and editing. J.S.: Funding acquisition, writing—original draft. All authors have read and agreed to the published version of the manuscript.

Funding: This work has been supported by the Ministry of Health of the Czech Republic: NU20-03-00016, NU20-09-00355, NU23-03-00150, RVO-VFN 00064165; Charles University: COOPERATIO, SVV260516; and the Ministry of Education Youth and Sports of the Czech Republic: Programme EXCELES, ID Project No. LX22NPO5102—Funded by the European Union—Next Generation EU.

Institutional Review Board Statement: Not applicable.

Informed Consent Statement: Not applicable.

Data Availability Statement: The data presented in this study are available on request from the corresponding author.

Conflicts of Interest: No potential conflicts of interest were reported. We declare that the results summarized in this manuscript have not been published previously and have not been submitted for consideration to any other journal.

Abbreviations

AD	Autosomal Dominant
AR	Autosomal Recessive
AT	Ataxia-Telangiectasia
BC	Breast Cancer
CMMRD	Constitutional mismatch repair deficiency syndrome
CrC	Colorectal Cancer
Dg	Diagnosis
EC	Endometrial Cancer
FA	Fanconi Anemia
GPV	Germline Pathogenic/Likely Pathogenic Variant
HBOC	Hereditary Breast and Ovarian Cancer
HGSC	High-Grade Serous Carcinoma
HR	Homologous Recombination
1 IK	
IMAGE	Intrauterine growth restriction, Metaphyseal dysplasia,
LGSC	Adrenal hypoplasia congenita, and Genitourinary abnormalities Low-Grade Serous Carcinoma
LGSC	
	Lynch Syndrome
MINAS	Multilocus Inherited Neoplasia Allele Syndrome
MM	Malignant Melanoma
MMR	Mismatch Repair
NA	Not Available
NGS	Next-Generation Sequencing
No.	Number
OC	Ovarian cancer
OLIDA	Oligogenic Diseases Database
PaC	Pancreatic Cancer
PrC	Prostate Cancer
PRS	Polygenic Risk Score
SCCOHT	Small Cell Carcinoma of the Ovary Hypercalcemic Type
SNP	Single Nucleotide Polymorphism
Sy	Syndrome
WES	Whole Exome Sequencing
WGS	Whole Genome Sequencing
XCI	X Chromosome Inactivation
Yo	Years Old

References

- Sung, H.; Ferlay, J.; Siegel, R.L.; Laversanne, M.; Soerjomataram, I.; Jemal, A.; Bray, F.; Bsc, M.F.B.; Me, J.F.; Soerjomataram, M.I.; et al. Global Cancer Statistics 2020: GLOBOCAN Estimates of Incidence and Mortality Worldwide for 36 Cancers in 185 Countries. *CA Cancer J. Clin.* 2021, 71, 209–249. [CrossRef] [PubMed]
- Kim, M.-K.; Kim, K.; Kim, S.M.; Kim, J.W.; Park, N.-H.; Song, Y.-S.; Kang, S.-B. A hospital-based case-control study of identifying ovarian cancer using symptom index. J. Gynecol. Oncol. 2009, 20, 238–242. [CrossRef] [PubMed]
- 3. National Cancer Institute: Surveillance, Epidemiology, and End Results Program. Available online: www.seer.cancer.gov/ (accessed on 1 September 2023).
- Ray-Coquard, I.; Morice, P.; Lorusso, D.; Prat, J.; Oaknin, A.; Pautier, P.; Colombo, N. Non-epithelial ovarian cancer: ESMO Clinical Practice Guidelines for diagnosis, treatment and follow-up. *Ann. Oncol.* 2018, 29 (Suppl. S4), iv1–iv18. [CrossRef] [PubMed]
- Lhotova, K.; Stolarova, L.; Zemankova, P.; Vocka, M.; Janatova, M.; Borecka, M.; Cerna, M.; Jelinkova, S.; Kral, J.; Volkova, Z.; et al. Multigene Panel Germline Testing of 1333 Czech Patients with Ovarian Cancer. *Cancers* 2020, 12, 956. [CrossRef] [PubMed]
- Kanchi, K.L.; Johnson, K.J.; Lu, C.; McLellan, M.D.; Leiserson, M.D.M.; Wendl, M.C.; Zhang, Q.; Koboldt, D.C.; Xie, M.; Kandoth, C.; et al. Integrated analysis of germline and somatic variants in ovarian cancer. *Nat. Commun.* 2014, *5*, 3156. [CrossRef] [PubMed]
- Risch, H.A.; McLaughlin, J.R.; Cole, D.E.; Rosen, B.; Bradley, L.; Kwan, E.; Jack, E.; Vesprini, D.J.; Kuperstein, G.; Abrahamson, J.L.; et al. Prevalence and penetrance of germline BRCA1 and BRCA2 mutations in a population series of 649 women with ovarian cancer. *Am. J. Hum. Genet.* 2001, *68*, 700–710. [CrossRef] [PubMed]

- Stratton, J.F.; Thompson, D.; Bobrow, L.; Dalal, N.; Gore, M.; Bishop, D.; Scott, I.; Evans, G.; Daly, P.; Easton, D.F.; et al. The genetic epidemiology of early-onset epithelial ovarian cancer: A population-based study. *Am. J. Hum. Genet.* 1999, 65, 1725–1732. [CrossRef]
- 9. Carter, N.J.; Marshall, M.L.; Susswein, L.R.; Zorn, K.K.; Hiraki, S.; Arvai, K.J.; Torene, R.I.; McGill, A.K.; Yackowski, L.; Murphy, P.D.; et al. Germline pathogenic variants identified in women with ovarian tumors. *Gynecol. Oncol.* **2018**, *151*, 481–488. [CrossRef]
- 10. Prat, J. New insights into ovarian cancer pathology. Ann. Oncol. 2012, 23 (Suppl. S10), x111-x117. [CrossRef]
- 11. Cancer Genome Atlas Research Network. Integrated genomic analyses of ovarian carcinoma. Nature 2011, 474, 609–615. [CrossRef]
- Morden, C.R.; Farrell, A.C.; Sliwowski, M.; Lichtensztejn, Z.; Altman, A.D.; Nachtigal, M.W.; McManus, K.J. Chromosome instability is prevalent and dynamic in high-grade serous ovarian cancer patient samples. *Gynecol. Oncol.* 2021, 161, 769–778. [CrossRef] [PubMed]
- Matz, M.; Coleman, M.P.; Sant, M.; Chirlaque, M.D.; Visser, O.; Gore, M.; Allemani, C.; Bouzbid, S.; Hamdi-Chérif, M.; Zaidi, Z.; et al. The histology of ovarian cancer: Worldwide distribution and implications for international survival comparisons (CONCORD-2). *Gynecol. Oncol.* 2017, 144, 405–413. [CrossRef] [PubMed]
- 14. Kurman, R.J.; Shih, I.M. The Dualistic Model of Ovarian Carcinogenesis: Revisited, Revised, and Expanded. *Am. J. Pathol.* **2016**, *186*, 733–747. [CrossRef] [PubMed]
- 15. GLOBOCAN 2020. Available online: https://gco.iarc.fr/ (accessed on 1 September 2023).
- 16. Lockley, M.; Stoneham, S.J.; Olson, T.A. Ovarian cancer in adolescents and young adults. *Pediatr. Blood Cancer* 2019, 66, e27512. [CrossRef]
- 17. Chan, J.K.; Urban, R.; Cheung, M.K.; Osann, K.; Husain, A.; Teng, N.N.; Kapp, D.S.; Berek, J.S.; Leiserowitz, G.S. Ovarian cancer in younger vs older women: A population-based analysis. *Br. J. Cancer.* **2006**, *95*, 1314–1320. [CrossRef] [PubMed]
- Grimley, P.M.; Matsuno, R.K.; Rosenberg, P.S.; Henson, D.E.; Schwartz, A.M.; Anderson, W.F. Qualitative Age Interactions between Low-grade and High-grade Serous Ovarian Carcinomas. *Cancer Epidemiol. Biomark. Prev.* 2009, 18, 2256–2261. [CrossRef] [PubMed]
- 19. Massi, D.; Susini, T.; Savino, L.; Boddi, V.; Amunni, G.; Colafranceschi, M. Epithelial ovarian tumors in the reproductive age group: Age is not an independent prognostic factor. *Cancer Interdiscip. Int. J. Am. Cancer Soc.* **1996**, 77, 1131–1136. [CrossRef]
- Yoshikawa, K.; Fukuda, T.; Uemura, R.; Matsubara, H.; Wada, T.; Kawanishi, M.; Tasaka, R.; Kasai, M.; Hashiguchi, Y.; Ichimura, T.; et al. Age-related differences in prognosis and prognostic factors among patients with epithelial ovarian cancer. *Mol. Clin. Oncol.* 2018, *9*, 329–334. [CrossRef]
- Schildkraut, J.M.; Halabi, S.; Bastos, E.; Marchbanks, P.A.; McDonald, J.A.; Berchuck, A. Prognostic factors in early-onset epithelial ovarian cancer: A population-based study. *Obstet. Gynecol.* 2000, 95, 119–127. [CrossRef]
- Gershenson, D.M.; Bodurka, D.C.; Lu, K.H.; Nathan, L.C.; Milojevic, L.; Wong, K.K.; Malpica, A.; Sun, C.C. Impact of Age and Primary Disease Site on Outcome in Women With Low-Grade Serous Carcinoma of the Ovary or Peritoneum: Results of a Large Single-Institution Registry of a Rare Tumor. J. Clin. Oncol. 2015, 33, 2675–2682. [CrossRef]
- Lalrinpuii, E.; Bhageerathy, P.S.; Sebastian, A.; Jeyaseelan, L.; Thomas, V.; Thomas, A.; Chandy, R.; Peedicayil, A. Ovarian Cancer in Young Women. *Indian J. Surg. Oncol.* 2017, *8*, 540–547. [CrossRef]
- Zhang, J.; Ugnat, A.-M.; Clarke, K.; Mao, Y. Ovarian cancer histology-specific incidence trends in Canada 1969–1993: Age-periodcohort analyses. Br. J. Cancer 1999, 81, 152–158. [CrossRef] [PubMed]
- Schneider, D.T.; Terenziani, M.; Cecchetto, G.; Olson, T.A.; Schneider, D.T.; Terenziani, M.; Olson, T.A.; Schneider, D.T.; Cecchetto, G.; Olson, T.A.; et al. Gonadal and Extragonadal Germ Cell Tumors, Sex Cord Stromal and Rare Gonadal Tumors. In *Rare Tumors in Children and Adolescents*; Springer International Publishing: Cham, Switzerland, 2012; pp. 327–402.
- Huang, Y.; Ming, X.; Li, B.; Li, Z. Histological Characteristics and Early-Stage Diagnosis Are Associated With Better Survival in Young Patients With Epithelial Ovarian Cancer: A Retrospective Analysis Based on Surveillance Epidemiology and End Results Database. *Front. Oncol.* 2020, 10, 595789. [CrossRef]
- Fu, Z.; Brooks, M.M.; Irvin, S.; Jordan, S.; Aben, K.K.H.; Anton-Culver, H.; Bandera, E.V.; Beckmann, M.W.; Berchuck, A.; Brooks-Wilson, A.; et al. Lifetime ovulatory years and risk of epithelial ovarian cancer: A multinational pooled analysis. *JNCI J. Natl. Cancer Inst.* 2023, *115*, 539–551. [CrossRef] [PubMed]
- Pavanello, M.; Chan, I.H.; Ariff, A.; Pharoah, P.D.; Gayther, S.A.; Ramus, S.J. Rare Germline Genetic Variants and the Risks of Epithelial Ovarian Cancer. *Cancers* 2020, 12, 3046. [CrossRef] [PubMed]
- Witjes, V.M.; van Bommel, M.H.; Ligtenberg, M.J.; Vos, J.R.; Mourits, M.J.; Ausems, M.G.; de Hullu, J.A.; Bosse, T.; Hoogerbrugge, N. Probability of detecting germline BRCA1/2 pathogenic variants in histological subtypes of ovarian carcinoma. A meta-analysis. *Gynecol. Oncol.* 2022, 164, 221–230. [CrossRef]
- Toss, A.; Tomasello, C.; Razzaboni, E.; Contu, G.; Grandi, G.; Cagnacci, A.; Schilder, R.J.; Cortesi, L. Hereditary ovarian cancer: Not only BRCA 1 and 2 genes. *BioMed Res. Int.* 2015, 2015, 341723. [CrossRef]
- Chen, J.; Bae, E.; Zhang, L.; Hughes, K.; Parmigiani, G.; Braun, D.; Rebbeck, T.R. Penetrance of Breast and Ovarian Cancer in Women Who Carry a BRCA1/2 Mutation and Do Not Use Risk-Reducing Salpingo-Oophorectomy: An Updated Meta-Analysis. *JNCI Cancer Spectr.* 2020, 4, pkaa029. [CrossRef]
- Kuchenbaecker, K.B.; Hopper, J.L.; Barnes, D.R.; Phillips, K.A.; Mooij, T.M.; Roos-Blom, M.J.; Jervis, S.; Van Leeuwen, F.E.; Milne, R.L.; Andrieu, N.; et al. Risks of Breast, Ovarian, and Contralateral Breast Cancer for BRCA1 and BRCA2 Mutation Carriers. JAMA 2017, 317, 2402–2416. [CrossRef] [PubMed]

- 33. Weber-Lassalle, N.; Hauke, J.; Ramser, J.; Richters, L.; Groß, E.; Blümcke, B.; Gehrig, A.; Kahlert, A.-K.; Müller, C.R.; Hackmann, K.; et al. BRIP1 loss-of-function mutations confer high risk for familial ovarian cancer, but not familial breast cancer. *Breast Cancer Res.* 2018, 20, 7. [CrossRef]
- Lilyquist, J.; LaDuca, H.; Polley, E.; Davis, B.T.; Shimelis, H.; Hu, C.; Hart, S.N.; Dolinsky, J.S.; Couch, F.J.; Goldgar, D.E. Frequency of mutations in a large series of clinically ascertained ovarian cancer cases tested on multi-gene panels compared to reference controls. *Gynecol. Oncol.* 2017, 147, 375–380. [CrossRef]
- National Comprehensive Cancer Network. Genetic/Familial High-Risk Assessment: Breast, Ovarian, and Pancreatic (Version 1.2023). 2022. Available online: https://www.nccn.org/professionals/physician_gls/pdf/genetics_bop.pdf (accessed on 1 September 2023).
- Cummings, S.; Roman, S.S.; Saam, J.; Bernhisel, R.; Brown, K.; Lancaster, J.M.; Usha, L. Age of ovarian cancer diagnosis among BRIP1, RAD51C, and RAD51D mutation carriers identified through multi-gene panel testing. *J. Ovarian Res.* 2021, 14, 61. [CrossRef] [PubMed]
- Dominguez-Valentin, M.; Sampson, J.R.; Seppälä, T.T.; ten Broeke, S.W.; Plazzer, J.-P.; Nakken, S.; Engel, C.; Aretz, S.; Jenkins, M.A.; Sunde, L.; et al. Cancer risks by gene, age, and gender in 6350 carriers of pathogenic mismatch repair variants: Findings from the Prospective Lynch Syndrome Database. *Genet. Med.* 2020, 22, 15–25. [CrossRef] [PubMed]
- Bernards, S.S.; Norquist, B.M.; Harrell, M.I.; Agnew, K.J.; Lee, M.K.; Walsh, T.; Swisher, E.M. Genetic characterization of early onset ovarian carcinoma. *Gynecol. Oncol.* 2016, 140, 221–225. [CrossRef] [PubMed]
- Curtius, K.; Gupta, S.; Boland, C.R. Review article: Lynch Syndrome-a mechanistic and clinical management update. *Aliment. Pharmacol. Ther.* 2022, 55, 960–977. [CrossRef]
- 40. Flaum, N.; Crosbie, E.J.; Woodward, E.R.; Lalloo, F.; Morgan, R.; Ryan, N.; Evans, D.G. MSH2 is the very young onset ovarian cancer predisposition gene, not BRCA1. *J. Med. Genet.* **2023**, *60*, 576–577. [CrossRef] [PubMed]
- 41. Kurian, A.W.; Hughes, E.; Handorf, E.A.; Gutin, A.; Allen, B.; Hartman, A.-R.; Hall, M.J. Breast and Ovarian Cancer Penetrance Estimates Derived From Germline Multiple-Gene Sequencing Results in Women. *JCO Precis. Oncol.* **2017**, *1*, 1–12. [CrossRef]
- Yang, X.; Leslie, G.; Doroszuk, A.; Schneider, S.; Allen, J.; Decker, B.; Dunning, A.M.; Redman, J.; Scarth, J.; Plaskocinska, I.; et al. Cancer Risks Associated With Germline PALB2 Pathogenic Variants: An International Study of 524 Families. *J. Clin. Oncol.* 2020, 38, 674–685. [CrossRef]
- Narayan, P.; Ahsan, M.D.; Webster, E.M.; Perez, L.; Levi, S.R.; Harvey, B.; Wolfe, I.; Beaumont, S.; Brewer, J.T.; Siegel, D.; et al. Partner and localizer of BRCA2 (PALB2) pathogenic variants and ovarian cancer: A systematic review and meta-analysis. *Gynecol.* Oncol. 2023, 177, 72–85. [CrossRef]
- 44. Hearle, N.; Schumacher, V.; Menko, F.H.; Olschwang, S.; Boardman, L.A.; Gille, J.J.; Keller, J.J.; Westerman, A.M.; Scott, R.J.; Lim, W.; et al. Frequency and spectrum of cancers in the Peutz-Jeghers syndrome. *Clin. Cancer Res.* **2006**, *12*, 3209–3215. [CrossRef]
- Klimkowski, S.; Ibrahim, M.; Ibarra Rovira, J.J.; Elshikh, M.; Javadi, S.; Klekers, A.R.; Abusaif, A.A.; Moawad, A.W.; Ali, K.; Elsayes, K.M. Peutz-Jeghers Syndrome and the Role of Imaging: Pathophysiology, Diagnosis, and Associated Cancers. *Cancers* 2021, 13, 5121. [CrossRef] [PubMed]
- 46. Giardiello, F.M.; Brensinger, J.D.; Tersmette, A.C.; Goodman, S.N.; Petersen, G.M.; Booker, S.V.; Cruz–Correa, M.; Offerhaus, J.A. Very high risk of cancer in familial Peutz-Jeghers syndrome. *Gastroenterology* **2000**, *119*, 1447–1453. [CrossRef] [PubMed]
- 47. De Paolis, E.; Paragliola, R.M.; Concolino, P. Spectrum of DICER1 Germline Pathogenic Variants in Ovarian Sertoli-Leydig Cell Tumor. *J. Clin. Med.* **2021**, *10*, 1845. [CrossRef]
- Frio, T.R.; Bahubeshi, A.; Kanellopoulou, C.; Hamel, N.; Niedziela, M.; Sabbaghian, N.; Pouchet, C.; Gilbert, L.; O'Brien, P.K.; Serfas, K.; et al. DICER1 mutations in familial multinodular goiter with and without ovarian Sertoli-Leydig cell tumors. *JAMA* 2011, 305, 68–77. [CrossRef] [PubMed]
- Schultz, K.A.P.; Harris, A.; Doros, L.A.; Young, R.H.; Dehner, L.P.; Frazier, A.L.; Hill, D.A.; Messinger, Y.H. Clinical and genetic aspects of ovarian stromal tumors: A report from the International Ovarian and Testicular Stromal Tumor Registry. *J. Clin. Oncol.* 2014, 32 (Suppl. S15), 5520. [CrossRef]
- Witkowski, L.; Carrot-Zhang, J.; Albrecht, S.; Fahiminiya, S.; Hamel, N.; Tomiak, E.; Grynspan, D.; Saloustros, E.; Nadaf, J.; Rivera, B.; et al. Germline and somatic SMARCA4 mutations characterize small cell carcinoma of the ovary, hypercalcemic type. *Nat. Genet.* 2014, 46, 438–443. [CrossRef] [PubMed]
- Herold, N.; Schmolling, J.; Ernst, C.; Ataseven, B.; Blümcke, B.; Schömig-Markiefka, B.; Heikaus, S.; Göhring, U.; Engel, C.; Lampe, B.; et al. Pathogenic germline variants in SMARCA4 and further cancer predisposition genes in early onset ovarian cancer. *Cancer Med.* 2023, 12, 15256–15260. [CrossRef]
- 52. Ryan, N.; Evans, D.; Green, K.; Crosbie, E. Pathological features and clinical behavior of Lynch syndrome-associated ovarian cancer. *Gynecol. Oncol.* 2017, 144, 491–495. [CrossRef]
- 53. Huang, K.L.; Mashl, R.J.; Wu, Y.; Ritter, D.I.; Wang, J.; Oh, C.; Paczkowska, M.; Reynolds, S.; Wyczalkowski, M.A.; Oak, N.; et al. Pathogenic Germline Variants in 10,389 Adult Cancers. *Cell* **2018**, *173*, 355–370.e14. [CrossRef]

- 54. McKusick-Nathans Institute of Genetic Medicine Johns Hopkins University; Baltimore, M. Online Mendelian Inheritance in Man, OMIM[®]. 2022. Available online: https://omim.org/ (accessed on 1 September 2023).
- 55. Felicio, P.S.; Grasel, R.S.; Campacci, N.; de Paula, A.E.; Galvão, H.C.; Torrezan, G.T.; Sabato, C.S.; Fernandes, G.C.; Souza, C.P.; Michelli, R.D.; et al. Whole-exome sequencing of non-BRCA1/BRCA2 mutation carrier cases at high-risk for hereditary breast/ovarian cancer. *Hum. Mutat.* 2021, *42*, 290–299. [CrossRef]
- 56. da Costa e Silva Carvalho, S.; Cury, N.M.; Brotto, D.B.; De Araujo, L.F.; Rosa, R.C.A.; Texeira, L.A.; Plaça, J.R.; Marques, A.A.; Peronni, K.C.; Ruy, P.D.C.; et al. Germline variants in DNA repair genes associated with hereditary breast and ovarian cancer syndrome: Analysis of a 21 gene panel in the Brazilian population. *BMC Med. Genom.* 2020, *13*, 21. [CrossRef] [PubMed]
- 57. Boyd, J.; Sonoda, Y.; Federici, M.G.; Bogomolniy, F.; Rhei, E.; Maresco, D.L.; Saigo, P.E.; Almadrones, L.A.; Barakat, R.R.; Brown, C.L.; et al. Clinicopathologic features of BRCA-linked and sporadic ovarian cancer. *JAMA* **2000**, *283*, 2260–2265. [CrossRef] [PubMed]
- 58. Cibula, D.; Laco, J.; Dundr, P.; Hájková, N.; Tichá, I.; Hojný, J.; Němejcová, K.; Bártů, M.; Michálková, R.; Zikán, M.; et al. Synchronous endometrioid endometrial and ovarian carcinomas are biologically related: A clinico-pathological and molecular (next generation sequencing) study of 22 cases. Oncol. Lett. 2019, 17, 2207–2214.
- Jarhelle, E.; Stensland, H.M.F.R.; Hansen, G.M.; Skarsfjord, S.; Jonsrud, C.; Ingebrigtsen, M.; Strømsvik, N.; Van Ghelue, M. Identifying sequence variants contributing to hereditary breast and ovarian cancer in BRCA1 and BRCA2 negative breast and ovarian cancer patients. *Sci. Rep.* 2019, *9*, 19986. [CrossRef] [PubMed]
- Koczkowska, M.; Krawczynska, N.; Stukan, M.; Kuzniacka, A.; Brozek, I.; Sniadecki, M.; Debniak, J.; Wydra, D.; Biernat, W.; Kozlowski, P.; et al. Spectrum and Prevalence of Pathogenic Variants in Ovarian Cancer Susceptibility Genes in a Group of 333 Patients. *Cancers* 2018, *10*, 442. [CrossRef] [PubMed]
- Pal, T.; Permuth-Wey, J.; Betts, J.A.; Krischer, J.P.; Fiorica, J.; Arango, H.; LaPolla, J.; Hoffman, M.; Martino, M.A.; Wakeley, K.; et al. BRCA1 and BRCA2 mutations account for a large proportion of ovarian carcinoma cases. *Cancer* 2005, 104, 2807–2816. [CrossRef] [PubMed]
- 62. Anand, L.; Padmavathi, V.; Dhivya, V.; Mahalaxmi, I.; Balachandar, V. De novo germ-line mutation of APC gene in periampullary carcinoma with familial adenomatous polyps—A novel familial case report in South India. *Karbala Int. J. Mod. Sci.* **2016**, *2*, 266–270. [CrossRef]
- Vibert, R.; Le Gall, J.; Buecher, B.; Mouret-Fourme, E.; Bataillon, G.; Becette, V.; Trabelsi-Grati, O.; Moncoutier, V.; Dehainault, C.; Carriere, J.; et al. APC germline pathogenic variants and epithelial ovarian cancer: Causal or coincidental findings? *J. Med. Genet.* 2023, 60, 460–463. [CrossRef]
- 64. Babovic, N.; Simmons, P.S.; Moir, C.; Thorland, E.C.; Scheithauer, B.; Gliem, T.J.; Babovic-Vuksanovic, D. Mucinous cystadenoma of ovary in a patient with juvenile polyposis due to 10q23 microdeletion: Expansion of phenotype. *Am. J. Med. Genet. A* **2010**, 152A, 2623–2627. [CrossRef]
- 65. Lou, L.; Zhou, L.; Wang, W.; Li, H.; Li, Y. Atypical ovarian carcinoid tumor with widespread skeletal metastases: A case report of multiple endocrine neoplasia type 1 in a young woman. *BMC Cancer* **2019**, *19*, 1107. [CrossRef]
- 66. Yauy, K.; Imbert-Bouteille, M.; Bubien, V.; Lindet-Bourgeois, C.; Rathat, G.; Perrochia, H.; MacGrogan, G.; Longy, M.; Bessis, D.; Tinat, J.; et al. Ovarian Clear Cell Carcinoma in Cowden Syndrome. *J. Natl. Compr. Cancer Netw.* **2019**, *17*, 7–11. [CrossRef]
- Cho, M.-Y.; Kim, H.S.; Eng, C.; Kim, D.S.; Kang, S.J.; Eom, M.; Yi, S.Y.; Bronner, M.P. First report of ovarian dysgerminoma in Cowden syndrome with germline PTEN mutation and PTEN-related 10q loss of tumor heterozygosity. *Am. J. Surg. Pathol.* 2008, 32, 1258–1264. [CrossRef] [PubMed]
- Sia, T.Y.; Maio, A.; Kemel, Y.M.; Arora, K.S.; Gordhandas, S.B.; Kahn, R.M.; Salo-Mullen, E.E.; Sheehan, M.A.; Tejada, P.R.; Bandlamudi, C.; et al. Germline Pathogenic Variants and Genetic Counseling by Ancestry in Patients With Epithelial Ovarian Cancer. JCO Precis. Oncol. 2023, 7, e2300137. [CrossRef] [PubMed]
- 69. Janavičius, R.; Andrėkutė, K.; Mickys, U.; Rudaitis, V.; Brasiūnienė, B.; Griškevičius, L. Apparently "BRCA-related" breast and ovarian cancer patient with germline TP53 mutation. *Breast J.* 2011, *17*, 409–415. [CrossRef] [PubMed]
- Blanco, A.; Grana, B.; Fachal, L.; Santamarina, M.; Cameselle-Teijeiro, J.; Ruíz-Ponte, C.; Carracedo, A.; Vega, A. Beyond BRCA1 and BRCA2 wild-type breast and/or ovarian cancer families: Germline mutations in TP53 and PTEN. *Clin. Genet.* 2010, 77, 193–196. [CrossRef] [PubMed]
- 71. Norquist, B.M.; Harrell, M.I.; Brady, M.F.; Walsh, T.; Lee, M.K.; Gulsuner, S.; Bernards, S.S.; Casadei, S.; Yi, Q.; Burger, R.A.; et al. Inherited Mutations in Women With Ovarian Carcinoma. *JAMA Oncol.* **2016**, *2*, 482–490. [CrossRef] [PubMed]
- 72. Dace, P.; Olita, H.; Ludmila, E.; Ingrida, D. Tumour suppressor gene CDKN2A/p16 germline mutations in melanoma patients with additional cancer and cancer in their family history. *Acta Univ. Latv.* **2003**, *662*, 25–32.
- 73. Eoh, K.J.; Kim, J.E.; Park, H.S.; Lee, S.T.; Park, J.S.; Han, J.W.; Lee, J.Y.; Kim, S.; Kim, S.W.; Kim, J.H.; et al. Detection of Germline Mutations in Patients with Epithelial Ovarian Cancer Using Multi-gene Panels: Beyond BRCA1/2. *Cancer Res. Treat.* 2018, 50, 917–925. [CrossRef]
- 74. Mur, P.; García-Mulero, S.; del Valle, J.; Magraner-Pardo, L.; Vidal, A.; Pineda, M.; Cinnirella, G.; Martín-Ramos, E.; Pons, T.; López-Doriga, A.; et al. Role of POLE and POLD1 in familial cancer. *Genet. Med.* **2020**, *22*, 2089–2100. [CrossRef]
- 75. Song, H.; Dicks, E.M.; Tyrer, J.; Intermaggio, M.; Chenevix-Trench, G.; Bowtell, D.D.; Traficante, N.; AOCS Group; Brenton, J.; Goranova, T.; et al. Population-based targeted sequencing of 54 candidate genes identifies PALB2 as a susceptibility gene for high-grade serous ovarian cancer. J. Med Genet. 2020, 58, 305–313. [CrossRef]

- 76. Schubert, S.; van Luttikhuizen, J.L.; Auber, B.; Schmidt, G.; Hofmann, W.; Penkert, J.; Davenport, C.F.; Hille-Betz, U.; Wendeburg, L.; Bublitz, J.; et al. The identification of pathogenic variants in BRCA1/2 negative, high risk, hereditary breast and/or ovarian cancer patients: High frequency of FANCM pathogenic variants. *Int. J. Cancer.* 2019, 144, 2683–2694. [CrossRef]
- del Valle, J.; Rofes, P.; Moreno-Cabrera, J.M.; López-Dóriga, A.; Belhadj, S.; Vargas-Parra, G.; Teulé, A.; Cuesta, R.; Muñoz, X.; Campos, O.; et al. Exploring the Role of Mutations in Fanconi Anemia Genes in Hereditary Cancer Patients. *Cancers* 2020, 12, 829. [CrossRef] [PubMed]
- Bertelsen, B.; Tuxen, I.V.; Yde, C.W.; Gabrielaite, M.; Torp, M.H.; Kinalis, S.; Oestrup, O.; Rohrberg, K.; Spangaard, I.; Santoni-Rugiu, E.; et al. High frequency of pathogenic germline variants within homologous recombination repair in patients with advanced cancer. NPJ Genom. Med. 2019, 4, 13. [CrossRef] [PubMed]
- Cavaillé, M.; Uhrhammer, N.; Privat, M.; Ponelle-Chachuat, F.; Gay-Bellile, M.; Lepage, M.; Molnar, I.; Viala, S.; Bidet, Y.; Bignon, Y. Analysis of 11 candidate genes in 849 adult patients with suspected hereditary cancer predisposition. *Genes Chromosomes Cancer* 2021, 60, 73–78. [CrossRef] [PubMed]
- Fostira, F.; Kostantopoulou, I.; Apostolou, P.; Papamentzelopoulou, M.S.; Papadimitriou, C.; Faliakou, E.; Christodoulou, C.; Boukovinas, I.; Razis, E.; Tryfonopoulos, D.; et al. One in three highly selected Greek patients with breast cancer carries a loss-of-function variant in a cancer susceptibility gene. *J. Med Genet.* 2019, *57*, 53–61. [CrossRef] [PubMed]
- Norquist, B.; Harrell, M.; Walsh, T.; Lee, M.; King, M.; Davidson, S.; Mannel, R.; DiSilvestro, P.; Swisher, E.; Birrer, M. Germline mutations in DNA repair genes in women with ovarian, peritoneal, or fallopian tube cancer treated on GOG protocols 218 and 262. *Gynecol. Oncol.* 2014, 133, 6. [CrossRef]
- Golmard, L.; Castéra, L.; Krieger, S.; Moncoutier, V.; Abidallah, K.; Tenreiro, H.; Laugé, A.; Tarabeux, J.; Millot, G.A.; Nicolas, A.; et al. Contribution of germline deleterious variants in the RAD51 paralogs to breast and ovarian cancers. *Eur. J. Hum. Genet.* 2017, 25, 1345–1353. [CrossRef] [PubMed]
- 83. Zhao, Q.; Yang, J.; Li, L.; Cao, D.; Yu, M.; Shen, K. Germline and somatic mutations in homologous recombination genes among Chinese ovarian cancer patients detected using next-generation sequencing. *J. Gynecol. Oncol.* **2017**, *28*, e39. [CrossRef]
- Subramanian, D.N.; Zethoven, M.; McInerny, S.; Morgan, J.A.; Rowley, S.M.; Lee, J.E.A.; Li, N.; Gorringe, K.L.; James, P.A.; Campbell, I.G. Exome sequencing of familial high-grade serous ovarian carcinoma reveals heterogeneity for rare candidate susceptibility genes. *Nat. Commun.* 2020, *11*, 1640. [CrossRef]
- 85. Srivastava, S.; Olson, H.E.; Cohen, J.S.; Gubbels, C.S.; Lincoln, S.; Davis, B.T.; Shahmirzadi, L.; Gupta, S.; Picker, J.; Yu, T.W.; et al. BRAT1 mutations present with a spectrum of clinical severity. *Am. J. Med. Genet. A* **2016**, *170*, 2265–2273. [CrossRef]
- Pennington, K.P.; Walsh, T.; Harrell, M.I.; Lee, M.K.; Pennil, C.C.; Rendi, M.H.; Thornton, A.; Norquist, B.M.; Casadei, S.; Nord, A.S.; et al. Germline and somatic mutations in homologous recombination genes predict platinum response and survival in ovarian, fallopian tube, and peritoneal carcinomas. *Clin. Cancer Res.* 2014, 20, 764–775. [CrossRef]
- Lu, C.; Xie, M.; Wendl, M.C.; Wang, J.; McLellan, M.D.; Leiserson, M.D.M.; Huang, K.-L.; Wyczalkowski, M.A.; Jayasinghe, R.; Banerjee, T.; et al. Patterns and functional implications of rare germline variants across 12 cancer types. *Nat. Commun.* 2015, 6, 10086. [CrossRef]
- Stradella, A.; Del Valle, J.; Rofes, P.; Vargas-Parra, G.; Salinas, M.; González, S.; Montes, E.; López-Doriga, A.; Gómez, C.; de Cid, R.; et al. ERCC3, a new ovarian cancer susceptibility gene? *Eur. J. Cancer* 2020, 141, 1–8. [CrossRef] [PubMed]
- Soukupova, J.; Zemankova, P.; Nehasil, P.; Kleibl, Z.; Kleibl, Z.; Soukupová, J.; Janatová, M.; Zemánková, P.; Černá, M.; Jelínková, S.; et al. Re: ERCC3, a new ovarian cancer susceptibility gene? *Eur. J. Cancer* 2021, 150, 278–280. [CrossRef]
- 90. Meienberg, J.; Bruggmann, R.; Oexle, K.; Matyas, G. Clinical sequencing: Is WGS the better WES? *Hum. Genet.* 2016, 135, 359–362. [CrossRef] [PubMed]
- 91. Cavalieri, S.; Pozzi, E.; Gatti, R.A.; Brusco, A. Deep-intronic ATM mutation detected by genomic resequencing and corrected in vitro by antisense morpholino oligonucleotide (AMO). *Eur. J. Hum. Genet.* **2013**, *21*, 774–778. [CrossRef] [PubMed]
- 92. Evans, D.R.; van Veen, E.M.; Byers, H.J.; Wallace, A.J.; Ellingford, J.M.; Beaman, G.; Santoyo-Lopez, J.; Aitman, T.J.; Eccles, D.M.; Lalloo, F.I.; et al. A Dominantly Inherited 5' UTR Variant Causing Methylation-Associated Silencing of BRCA1 as a Cause of Breast and Ovarian Cancer. Am. J. Hum. Genet. 2018, 103, 213–220. [CrossRef] [PubMed]
- Rusch, M.; Nakitandwe, J.; Shurtleff, S.; Newman, S.; Zhang, Z.; Edmonson, M.N.; Parker, M.; Jiao, Y.; Ma, X.; Liu, Y.; et al. Clinical cancer genomic profiling by three-platform sequencing of whole genome, whole exome and transcriptome. *Nat. Commun.* 2018, *9*, 3962. [CrossRef] [PubMed]
- 94. Funingana, I.; Trotman, J.; Ambrose, J.; Roberts, T.; Watkins, J.; Ridley, M.; Gilson, B.; Freeman, S.; Jimenez-Linan, M.; Sosinsky, A.; et al. 7P Integration of whole genome sequencing (WGS) into NHS pathways for high-grade ovarian cancer (HGOC): A single-centre prospective experience. *ESMO Open* 2023, *8*, 100861. [CrossRef]
- 95. Guan, Z.; Begg, C.B.; Shen, R. Predicting Cancer Risk from Germline Whole-exome Sequencing Data Using a Novel Context-based Variant Aggregation Approach. *Cancer Res. Commun.* **2023**, *3*, 483–488. [CrossRef]
- Domchek, S.M.; Tang, J.; Stopfer, J.; Lilli, D.R.; Hamel, N.; Tischkowitz, M.; Monteiro, A.N.A.; Messick, T.E.; Powers, J.; Yonker, A.; et al. Biallelic deleterious BRCA1 mutations in a woman with early-onset ovarian cancer. *Cancer Discov.* 2013, *3*, 399–405. [CrossRef] [PubMed]

- 97. Scherz, A.; Stoll, S.; Rothlisberger, B.; Rabaglio, M. A New de novo BRCA1 Mutation in a Young Breast Cancer Patient: A Case Report. *Appl. Clin. Genet.* 2023, *16*, 83–87. [CrossRef] [PubMed]
- 98. Tenedini, E.; Piana, S.; Toss, A.; Marino, M.; Barbieri, E.; Artuso, L.; Venturelli, M.; Gasparini, E.; Mandato, V.D.; Marchi, I.; et al. Constitutional Mosaicism: A Critical Issue in the Definition of BRCA-Inherited Cancer Risk. *JCO Precis. Oncol.* 2022, 6, e2200138. [CrossRef] [PubMed]
- Witkowski, L.; Lalonde, E.; Zhang, J.; Albrecht, S.; Hamel, N.; Cavallone, L.; May, S.T.; Nicholson, J.C.; Coleman, N.; Murray, M.J.; et al. Familial rhabdoid tumour 'avant la lettre'—From pathology review to exome sequencing and back again. *J. Pathol.* 2013, 231, 35–43. [CrossRef] [PubMed]
- Golmard, L.; Delnatte, C.; Laugé, A.; Moncoutier, V.; Lefol, C.; Abidallah, K.; Tenreiro, H.; Copigny, F.; Giraudeau, M.; Guy, C.; et al. Breast and ovarian cancer predisposition due to de novo BRCA1 and BRCA2 mutations. *Oncogene* 2016, 35, 1324–1327. [CrossRef] [PubMed]
- von Hardenberg, S.; Wallaschek, H.; Du, C.; Schmidt, G.; Auber, B. A holistic approach to maximise diagnostic output in trio exome sequencing. *Front. Pediatr.* 2023, 11, 1183891. [CrossRef] [PubMed]
- Speight, B.; Colvin, E.; Epurescu, E.D.; Drummond, J.; Verhoef, S.; Pereira, M.; Evans, D.G.; Tischkowitz, M. Low-level constitutional mosaicism of BRCA1 in two women with young onset ovarian cancer. *Hered. Cancer Clin. Pract.* 2022, 20, 32. [CrossRef]
- 103. Alhopuro, P.; Vainionpää, R.; Anttonen, A.-K.; Aittomäki, K.; Nevanlinna, H.; Pöyhönen, M. Constitutional mosaicism for a BRCA2 mutation as a cause of early-onset breast cancer. *Fam. Cancer* 2020, *19*, 307–310. [CrossRef]
- 104. Schwartz, M.; Ibadioune, S.; Chansavang, A.; Vacher, S.; Caputo, S.M.; Delhomelle, H.; Wong, J.; Abidallah, K.; Moncoutier, V.; Becette, V.; et al. Mosaic *BRCA1* promoter methylation contribution in hereditary breast/ovarian cancer pedigrees. *J. Med. Genet.* 2023. [CrossRef]
- 105. Pinto, D.; Pinto, C.; Guerra, J.; Pinheiro, M.; Santos, R.; Vedeld, H.M.; Yohannes, Z.; Peixoto, A.; Santos, C.; Pinto, P.; et al. Contribution of MLH1 constitutional methylation for Lynch syndrome diagnosis in patients with tumor MLH1 downregulation. *Cancer Med.* 2018, 7, 433–444. [CrossRef]
- 106. Rantala, J.N.J.; Heikkinen, S.M.M.; Hirvonen, E.M.; Tanskanen, T.; Malila, N.K.; Pitkäniemi, J.M. Familial aggregation of early-onset cancers in early-onset breast cancer families. *Int. J. Cancer* 2023, *153*, 331–340. [CrossRef] [PubMed]
- 107. Imbert-Bouteille, M.; Corsini, C.; Picot, M.-C.; Mizrahy, L.; Akouete, S.; Huguet, H.; Thomas, F.; Geneviève, D.; Taourel, P.; Ychou, M.; et al. No Association of Early-Onset Breast or Ovarian Cancer with Early-Onset Cancer in Relatives in BRCA1 or BRCA2 Mutation Families. *Genes* 2021, *12*, 1100. [CrossRef] [PubMed]
- Stratton, J.F.; Pharoah, P.; Smith, S.K.; Easton, D.; Ponder, B.A.J. A systematic review and meta-analysis of family history and risk of ovarian cancer. *BJOG Int. J. Obstet. Gynaecol.* 1998, 105, 493–499. [CrossRef] [PubMed]
- 109. Eng, K.H.; Szender, J.B.; Etter, J.L.; Kaur, J.; Poblete, S.; Huang, R.Y.; Zhu, Q.; Grzesik, K.A.; Battaglia, S.; Cannioto, R.; et al. Paternal lineage early onset hereditary ovarian cancers: A Familial Ovarian Cancer Registry study. *PLoS Genet.* 2018, 14, e1007194. [CrossRef] [PubMed]
- 110. Buller, R.E.; Sood, A.K.; Lallas, T.; Buekers, T.; Skilling, J.S. Association between nonrandom X-chromosome inactivation and BRCA1 mutation in germline DNA of patients with ovarian cancer. *J. Natl. Cancer Inst.* **1999**, *91*, 339–346. [CrossRef] [PubMed]
- 111. Liu, R.; Kain, M.; Wang, L. Inactivation of X-linked tumor suppressor genes in human cancer. *Future Oncol.* **2012**, *8*, 463–481. [CrossRef] [PubMed]
- 112. Lose, F.; Duffy, D.L.; Kay, G.F.; Kedda, M.A.; Spurdle, A.B. Skewed X chromosome inactivation and breast and ovarian cancer status: Evidence for X-linked modifiers of BRCA1. *J. Natl. Cancer Inst.* **2008**, *100*, 1519–1529. [CrossRef]
- Winham, S.J.; Larson, N.B.; Armasu, S.M.; Fogarty, Z.C.; Larson, M.C.; McCauley, B.M.; Wang, C.; Lawrenson, K.; Gayther, S.; Cunningham, J.M.; et al. Molecular signatures of X chromosome inactivation and associations with clinical outcomes in epithelial ovarian cancer. *Hum. Mol. Genet.* 2019, 28, 1331–1342. [CrossRef]
- 114. Qing, T.; Mohsen, H.; Marczyk, M.; Ye, Y.; O'meara, T.; Zhao, H.; Townsend, J.P.; Gerstein, M.; Hatzis, C.; Kluger, Y.; et al. Germline variant burden in cancer genes correlates with age at diagnosis and somatic mutation burden. *Nat. Commun.* 2020, 11, 2438. [CrossRef]
- 115. Jia, G.; Lu, Y.; Wen, W.; Long, J.; Liu, Y.; Tao, R.; Li, B.; Denny, J.C.; Shu, X.-O.; Zheng, W. Evaluating the Utility of Polygenic Risk Scores in Identifying High-Risk Individuals for Eight Common Cancers. *JNCI Cancer Spectr.* **2020**, *4*, pkaa021. [CrossRef]
- 116. Yang, X.; Leslie, G.; Gentry-Maharaj, A.; Ryan, A.; Intermaggio, M.; Lee, A.; Kalsi, J.K.; Tyrer, J.; Gaba, F.; Manchanda, R.; et al. Evaluation of polygenic risk scores for ovarian cancer risk prediction in a prospective cohort study. J. Med Genet. 2018, 55, 546–554. [CrossRef] [PubMed]
- 117. Dareng, E.O.; Tyrer, J.P.; Barnes, D.R.; Jones, M.R.; Yang, X.; Aben, K.K.H.; Adank, M.A.; Agata, S.; Andrulis, I.L.; Anton-Culver, H.; et al. Polygenic risk modeling for prediction of epithelial ovarian cancer risk. *Eur. J. Hum. Genet.* 2022, 30, 349–362. [CrossRef] [PubMed]
- 118. Borde, J.; Laitman, Y.; Blümcke, B.; Niederacher, D.; Weber-Lassalle, K.; Sutter, C.; Rump, A.; Arnold, N.; Wang-Gohrke, S.; Horváth, J.; et al. Polygenic risk scores indicate extreme ages at onset of breast cancer in female BRCA1/2 pathogenic variant carriers. *BMC Cancer* **2022**, 22, 706. [CrossRef] [PubMed]

- 119. Fatapour, Y.; Brody, J.P. Genetic Risk Scores and Missing Heritability in Ovarian Cancer. Genes 2023, 14, 762. [CrossRef] [PubMed]
- 120. Rebbeck, T.R.; Friebel, T.M.; Mitra, N.; Wan, F.; Chen, S.; Andrulis, I.L.; Apostolou, P.; Arnold, N.; Arun, B.K.; Barrowdale, D.; et al. Inheritance of deleterious mutations at both BRCA1 and BRCA2 in an international sample of 32,295 women. *Breast Cancer Res.* **2016**, *18*, 112. [CrossRef] [PubMed]
- 121. Zhu, Q.; Wang, J.; Yu, H.; Hu, Q.; Bateman, N.W.; Long, M.; Rosario, S.; Schultz, E.; Dalgard, C.L.; Wilkerson, M.D.; et al. Whole-Genome Sequencing Identifies PPARGC1A as a Putative Modifier of Cancer Risk in BRCA1/2 Mutation Carriers. *Cancers* 2022, 14, 2350. [CrossRef] [PubMed]
- 122. Laitman, Y.; Michaelson-Cohen, R.; Chen-Shtoyerman, R.; Goldberg, Y.; Reish, O.; Bernstein-Molho, R.; Levy-Lahad, E.; Ben Baruch, N.E.; Kedar, I.; Evans, D.G.; et al. Age at diagnosis of cancer in 185delAG BRCA1 mutation carriers of diverse ethnicities: Tentative evidence for modifier factors. *Fam. Cancer* 2021, 20, 189–194. [CrossRef]
- 123. Lee, C.H.; Subramanian, S.; Beck, A.H.; Espinosa, I.; Senz, J.; Zhu, S.X.; Huntsman, D.; van de Rijn, M.; Gilks, C.B. MicroRNA profiling of BRCA1/2 mutation-carrying and non-mutation-carrying high-grade serous carcinomas of ovary. *PLoS ONE* **2009**, *4*, e7314. [CrossRef]
- 124. Pastrello, C.; Polesel, J.; Della Puppa, L.; Viel, A.; Maestro, R. Association between hsa-mir-146a genotype and tumor age-of-onset in BRCA1/BRCA2-negative familial breast and ovarian cancer patients. *Carcinogenesis* **2010**, *31*, 2124–2126. [CrossRef]
- Zhang, K.; Chandrakasan, S.; Chapman, H.; Valencia, C.A.; Husami, A.; Kissell, D.; Johnson, J.A.; Filipovich, A.H. Synergistic defects of different molecules in the cytotoxic pathway lead to clinical familial hemophagocytic lymphohistiocytosis. *Blood* 2014, 124, 1331–1334. [CrossRef]
- 126. Moreno-Ruiz, N.; Ambrose, J.C.; Arumugam, P.; Baple, E.L.; Bleda, M.; Boardman-Pretty, F.; Boissiere, J.M.; Boustred, C.R.; Brittain, H.; Caulfield, M.J.; et al. Assessing the digenic model in rare disorders using population sequencing data. *Eur. J. Hum. Genet.* 2022, *30*, 1439–1443. [CrossRef]
- 127. Kamar, A.; Khalil, A.; Nemer, G. The Digenic Causality in Familial Hypercholesterolemia: Revising the Genotype-Phenotype Correlations of the Disease. *Front. Genet.* **2020**, *11*, 572045. [CrossRef] [PubMed]
- 128. Ameratunga, R.; Woon, S.-T.; Bryant, V.L.; Steele, R.; Slade, C.; Leung, E.Y.; Lehnert, K. Clinical Implications of Digenic Inheritance and Epistasis in Primary Immunodeficiency Disorders. *Front. Immunol.* **2017**, *8*, 1965. [CrossRef]
- 129. Nachtegael, C.; Gravel, B.; Dillen, A.; Smits, G.; Nowé, A.; Papadimitriou, S.; Lenaerts, T. Scaling up oligogenic diseases research with OLIDA: The Oligogenic Diseases Database. *Database* **2022**, 2022, baac023. [CrossRef] [PubMed]
- 130. Fijneman, R.J.; de Vries, S.S.; Jansen, R.C.; Demant, P. Complex interactions of new quantitative trait loci, Sluc1, Sluc2, Sluc3, and Sluc4, that influence the susceptibility to lung cancer in the mouse. *Nat. Genet.* **1996**, *14*, 465–467. [CrossRef] [PubMed]
- Tansel, A.; Katz, L.H.; El-Serag, H.B.; Thrift, A.P.; Parepally, M.; Shakhatreh, M.H.; Kanwal, F. Incidence and Determinants of Hepatocellular Carcinoma in Autoimmune Hepatitis: A Systematic Review and Meta-analysis. *Clin. Gastroenterol. Hepatol.* 2017, 15, 1207–1217.e4. [CrossRef]
- 132. Helicobacter and Cancer Collaborative Group. Gastric cancer and Helicobacter pylori: A combined analysis of 12 case control studies nested within prospective cohorts. *Gut* 2001, *49*, 347. [CrossRef]
- Gausman, V.; Dornblaser, D.; Anand, S.; Hayes, R.B.; O'Connell, K.; Du, M.; Liang, P.S. Risk Factors Associated With Early-Onset Colorectal Cancer. *Clin. Gastroenterol. Hepatol.* 2020, 18, 2752–2759.e2. [CrossRef]
- 134. Bae, E.; Lim, S.; Han, K.-D.; Jung, J.-H.; Choi, H.; Kim, C.; Ma, S.; Kim, S. Systemic lupus erythematosus is a risk factor for cancer: A nationwide population-based study in Korea. *Lupus* **2019**, *28*, 317–323. [CrossRef]
- 135. Kübler, K.; Arndt, P.F.; Wardelmann, E.; Krebs, D.; Kuhn, W.; van der Ven, K. HLA-class II haplotype associations with ovarian cancer. *Int. J. Cancer.* 2006, 119, 2980–2985. [CrossRef]
- 136. Carbone, M.; Arron, S.T.; Beutler, B.; Bononi, A.; Cavenee, W.; Cleaver, J.E.; Croce, C.M.; D'andrea, A.; Foulkes, W.D.; Gaudino, G.; et al. Tumour predisposition and cancer syndromes as models to study gene-environment interactions. *Nat. Rev. Cancer* 2020, 20, 533–549. [CrossRef] [PubMed]
- 137. Novelli, F.; Bononi, A.; Wang, Q.; Bai, F.; Patergnani, S.; Kricek, F.; Haglund, E.; Suarez, J.S.; Tanji, M.; Xu, R.; et al. BAP1 forms a trimer with HMGB1 and HDAC1 that modulates gene x environment interaction with asbestos. *Proc. Natl. Acad. Sci. USA* 2021, 118, e2111946118. [CrossRef] [PubMed]
- Kurzynska-Kokorniak, A.; Koralewska, N.; Pokornowska, M.; Urbanowicz, A.; Tworak, A.; Mickiewicz, A.; Figlerowicz, M. The many faces of Dicer: The complexity of the mechanisms regulating Dicer gene expression and enzyme activities. *Nucleic Acids Res.* 2015, 43, 4365–4380. [CrossRef] [PubMed]
- 139. Nickerson, M.L.; Warren, M.B.; Toro, J.R.; Matrosova, V.; Glenn, G.; Turner, M.L.; Duray, P.; Merino, M.; Choyke, P.; Pavlovich, C.P.; et al. Mutations in a novel gene lead to kidney tumors, lung wall defects, and benign tumors of the hair follicle in patients with the Birt-Hogg-Dubé syndrome. *Cancer Cell* 2002, 2, 157–164. [CrossRef] [PubMed]
- 140. L'espérance, K.; Grundy, A.; Abrahamowicz, M.; Arseneau, J.; Gilbert, L.; Gotlieb, W.H.; Provencher, D.; Koushik, A. Alcohol intake and the risk of epithelial ovarian cancer. *Cancer Causes Control* **2023**, *34*, 533–541. [CrossRef] [PubMed]
- 141. Wang, T.; Townsend, M.K.; Vinci, C.; Jake-Schoffman, D.E.; Tworoger, S.S. Early life exposure to tobacco smoke and ovarian cancer risk in adulthood. *Int. J. Epidemiol.* **2021**, *50*, 965–974. [CrossRef] [PubMed]
- 142. Collaborative Group on Epidemiological Studies of Ovarian Cancer. Ovarian cancer and smoking: Individual participant meta-analysis including 28,114 women with ovarian cancer from 51 epidemiological studies. *Lancet Oncol.* 2012, 13, 946–956. [CrossRef] [PubMed]

- 143. Mitamura, T.; Zhai, T.; Hatanaka, K.C.; Hatanaka, Y.; Amano, T.; Wang, L.; Tanaka, S.; Watari, H. Germline PRDM1 Variant rs2185379 in Long-Term Recurrence-Free Survivors of Advanced Ovarian Cancer. *Pharmacogenomics Pers. Med.* 2022, 15, 977–984. [CrossRef] [PubMed]
- 144. Shi, T.; Jiang, R.; Wang, P.; Xu, Y.; Yin, S.; Cheng, X.; Zang, R. Significant association of the EXO1 rs851797 polymorphism with clinical outcome of ovarian cancer. *Onco Targets Ther.* **2017**, *10*, 4841–4851. [CrossRef]

Disclaimer/Publisher's Note: The statements, opinions and data contained in all publications are solely those of the individual author(s) and contributor(s) and not of MDPI and/or the editor(s). MDPI and/or the editor(s) disclaim responsibility for any injury to people or property resulting from any ideas, methods, instructions or products referred to in the content.