



Supplementary Materials

Amphiphilic Polypeptides Obtained by the Post-Polymerization Modification of Poly(Glutamic Acid) and Their Evaluation as Delivery Systems for Hydrophobic Drugs

Apollinariia Yu. Dzhuzha ^{1,2}, Irina I. Tarasenko ², Leonard Ionut Atanase ³, Antonina Lavrentieva ⁴ and Evgenia G. Korzhikova-Vlakh ^{2,*}

¹ Institute of Chemistry, Saint-Petersburg State University, 198504 St. Petersburg, Russia;

² Institute of Macromolecular Compounds, Russian Academy of Sciences, 199004 St. Petersburg, Russia;

³ Faculty of Dental Medicine, "Apollonia" University, 700399 Iasi, Romania;

⁴ Institute of Technical Chemistry, Gottfried-Wilhelm-Leibniz University, 30167 Hannover, Germany;

Correspondence: vlakh@hq.macro.ru (E.K.-V.)

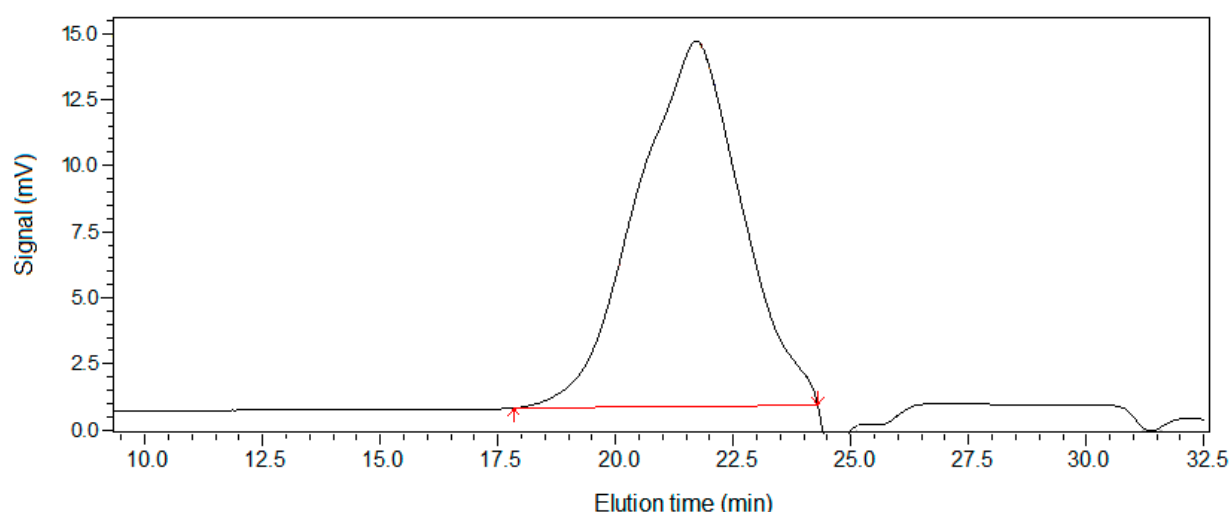
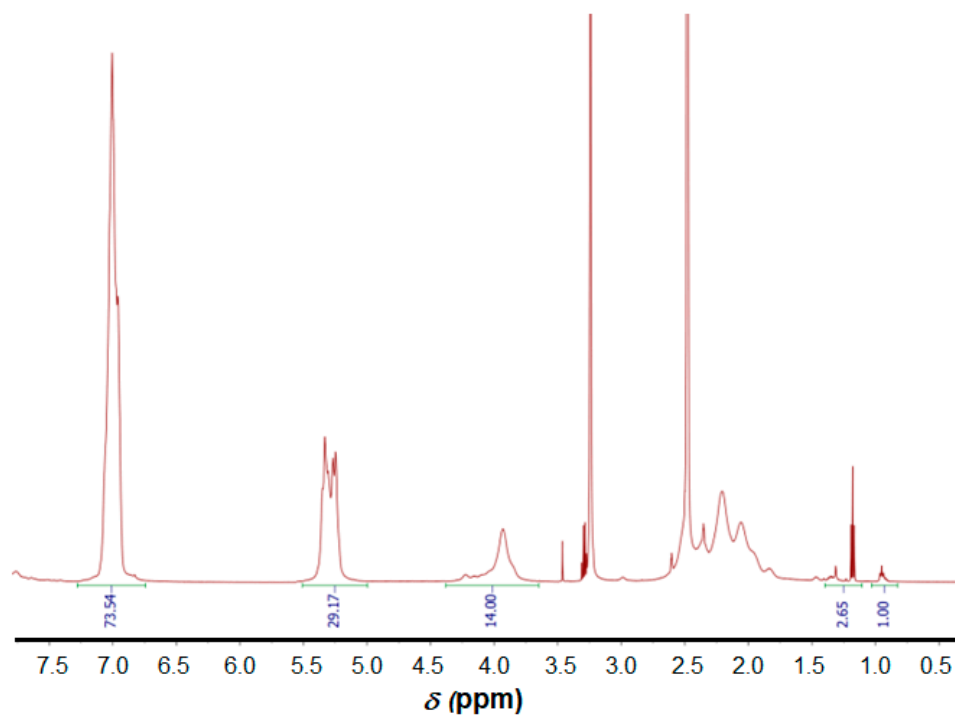
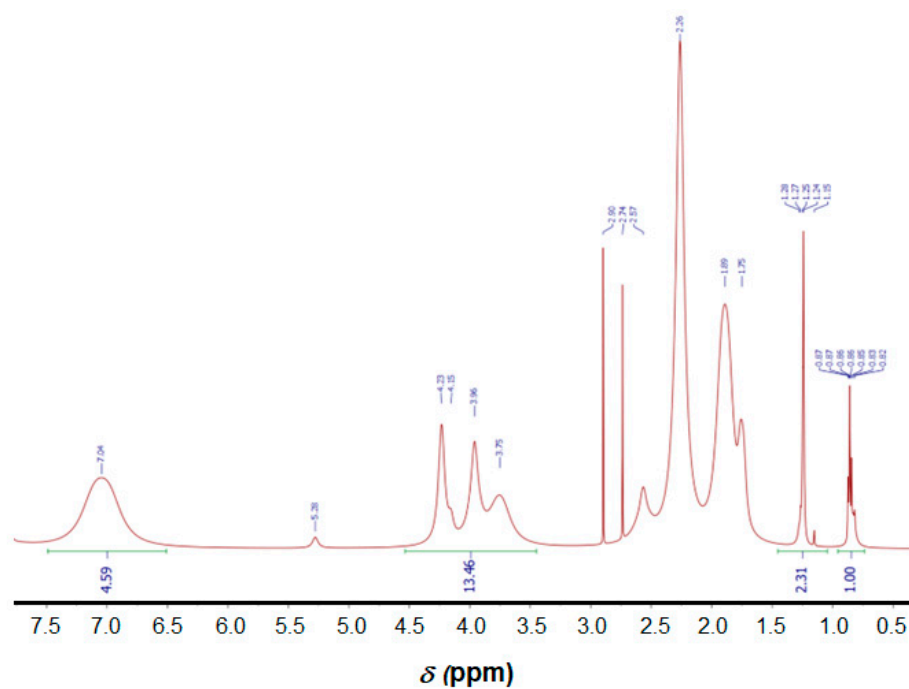


Figure S1. SEC trace of poly(α ,L-glutamic acid γ -benzyl ester). Conditions: Styragel Column, HMW6E, Waters (7.8 mm \times 300 mm, 15–20 μ m bead size), DMF containing 0.1M LiBr, 40 $^{\circ}$ C, elution rate 0.3 mL/min, refractometric detection.

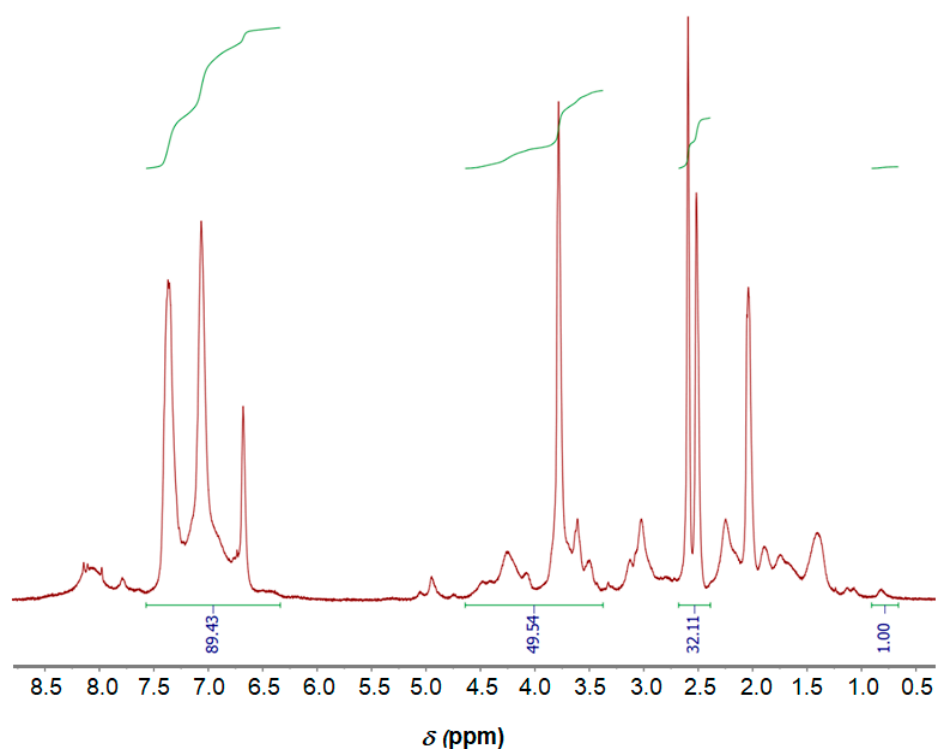


(a)

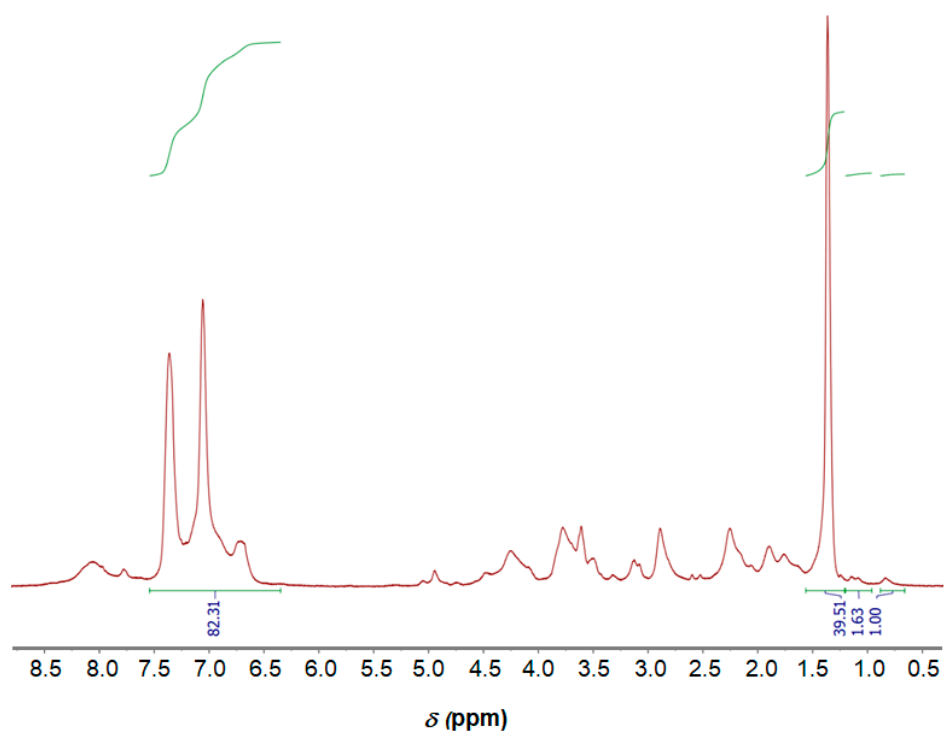


(b)

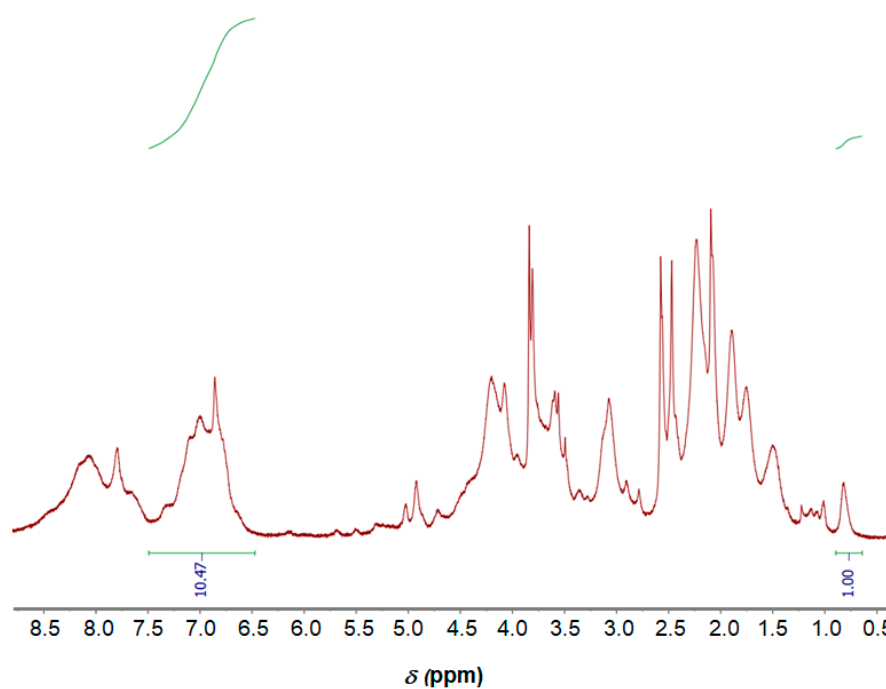
Figure S2. ^1H NMR spectra of Bzl-protected (a) and deprotected (b) poly(α ,L-glutamic acid) (DMSO- d_6 , 25 $^\circ\text{C}$): Signals (ppm): 0.80-0.85 (CH_3 , hexylamine), 1.00-1.33 (CH_2 , hexylamine), 3.5-4.5 (CH , Glu), 5.0-5.5 (O- CH_2 - C_6H_5 , Glu(OBzl)), 6.7-7.4 (O- CH_2 - C_6H_5 , Glu(OBzl)).



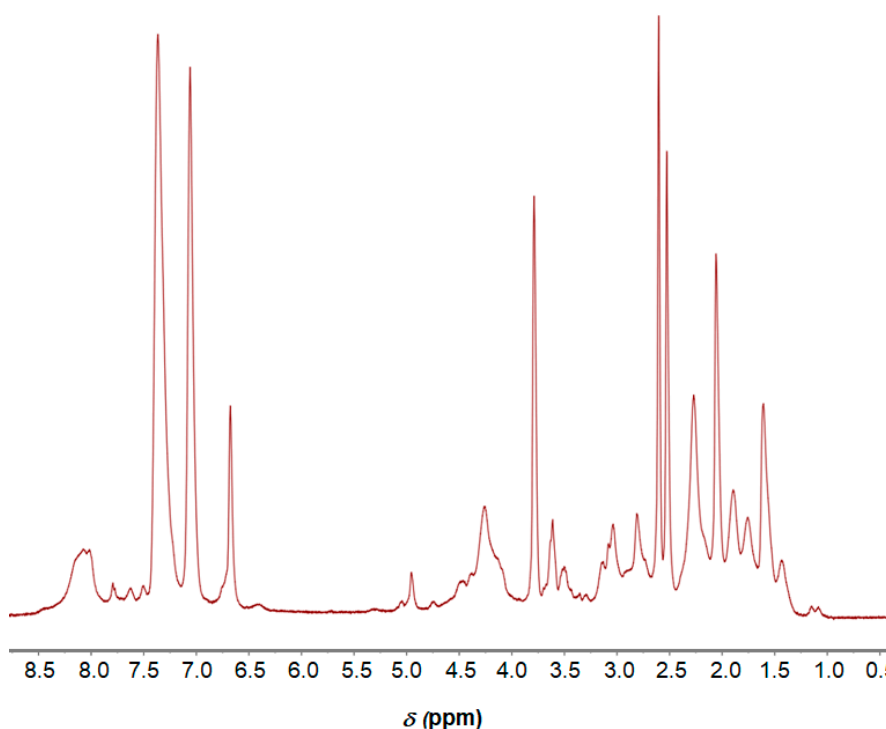
(a)



(b)

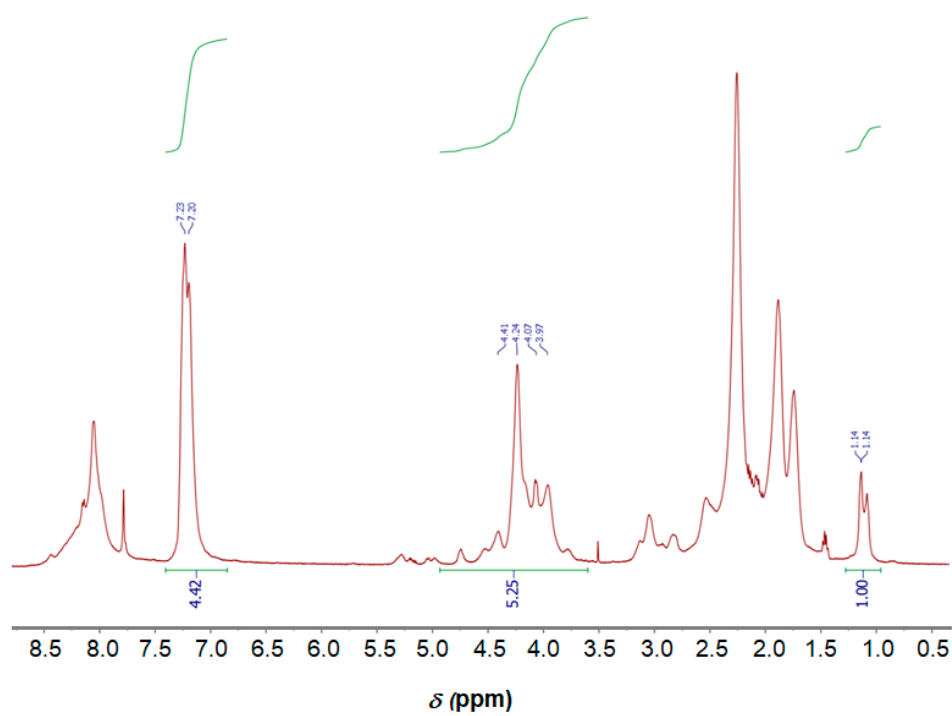


(c)

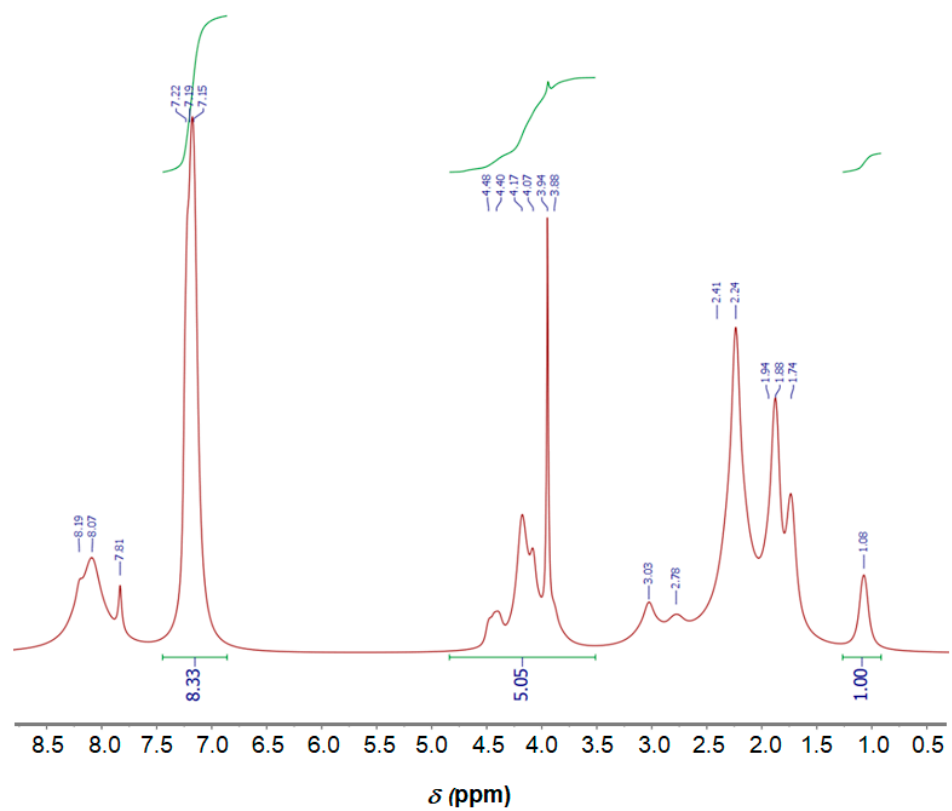


(d)

Figure S3. ^1H NMR spectra of P[EE(R(Mtr))E(H(Trt))E(I)E(Glc)] (a), P[EE(O(Boc))E(H(Trt))E(I)E(Glc)] (b), P[EE(R)E(H)E(I)E(Glc)] (with diffusion filter) (c), P[EE(R(Mtr))E(H(Trt))E(W(Boc))E(Glc)] (with diffusion filter) (d). Signals (ppm): 0.80-0.85 (CH_3 , hexylamine), 1.00-1.33 (CH_2 , hexylamine), 1.2-1.5 (2CH_2 , Arg; CH_3 , Boc of Orn and Trp), 1.5-2.4 (2CH_2 , Glu, Arg and Orn), 2.4-2.7 ($\text{CH}_3\text{-C}_6\text{H}_5$, Mtr of Arg), 2.7-3.5 (6CH_2 , glucose; CH_2 , Orn), 3.5-4.5 (CH , Glu; $\text{CH}_3\text{-O-C}_6\text{H}_5$, Arg; CH , Arg, His, Orn, Trp; CH_2 , His), 6.3-7.5 (C_6H_5 , Trt of His; 2H , His; C_6H_5 , Mtr of Arg; C_6H_5 , Trp), 1.4 (CH_3 , BOC of Orn), 7.8-8.5 (NH-CO , all polypeptides; $\text{NH}_2\text{-C=NH}$, Arg).



(a)



(b)

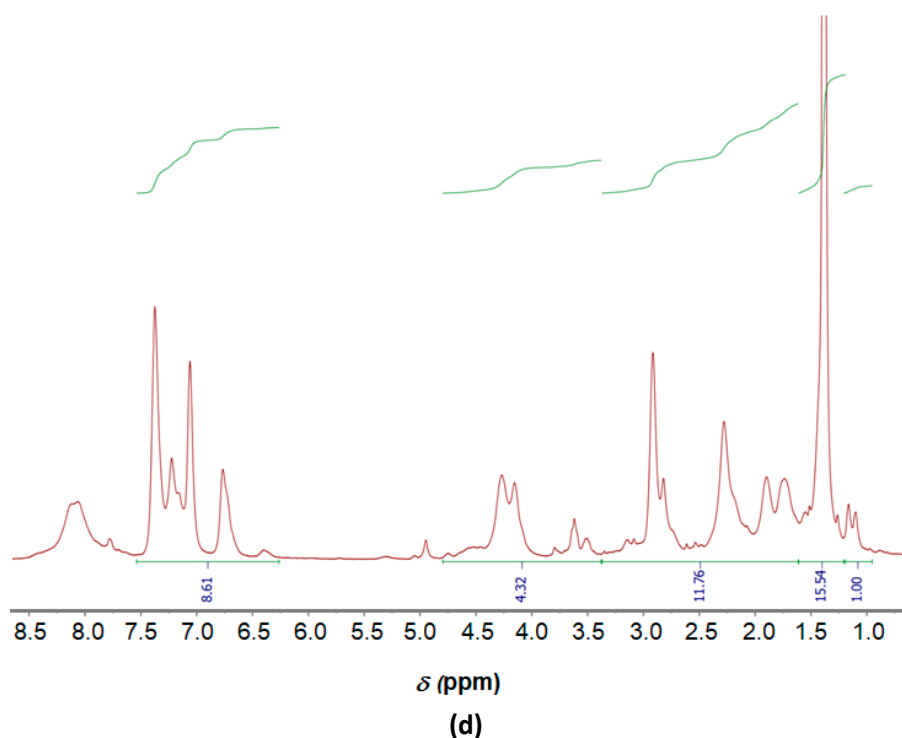
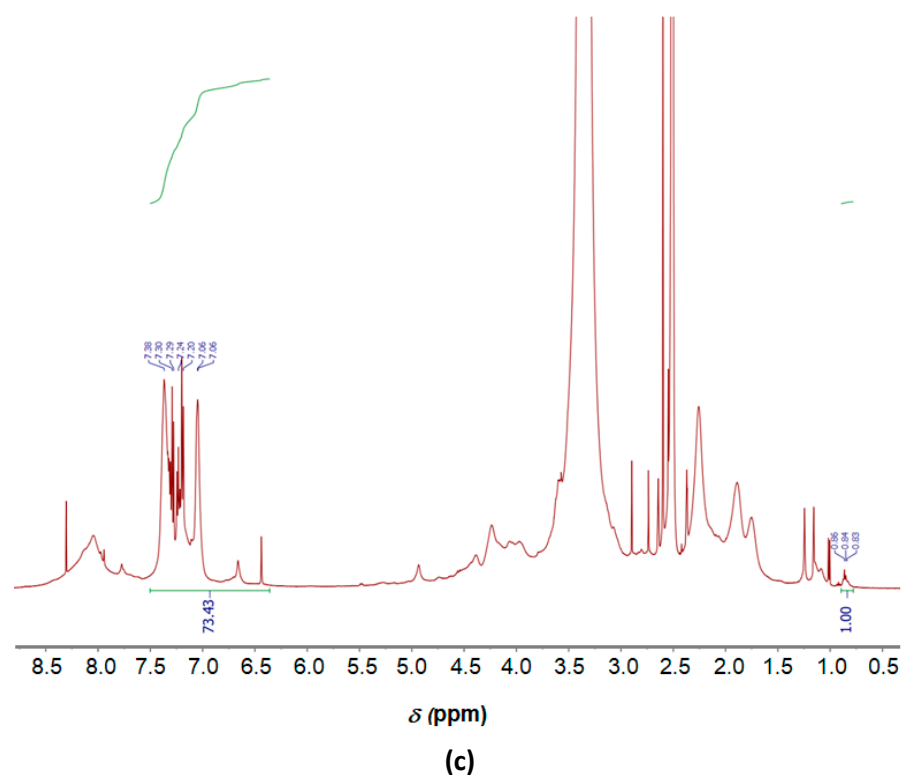
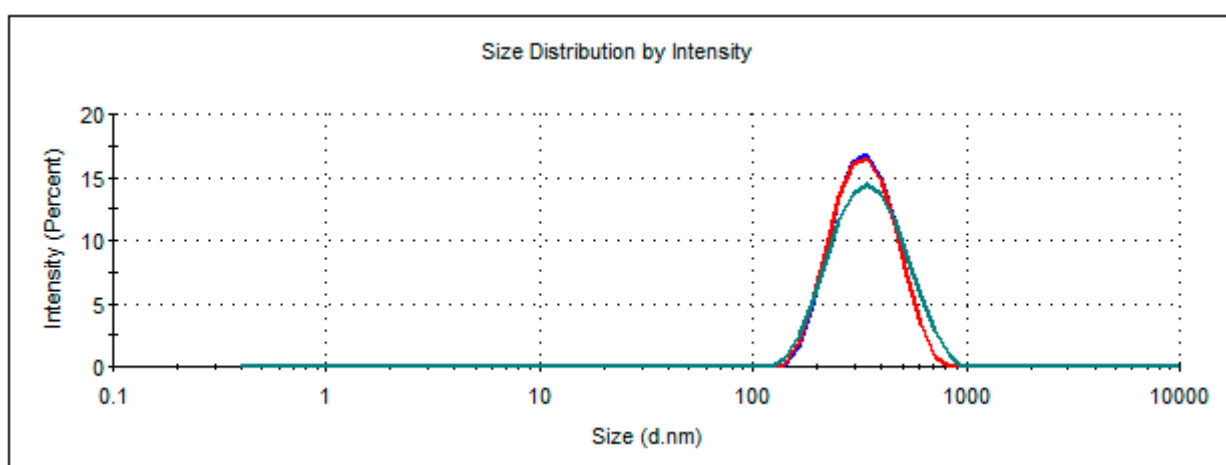


Figure S4. ^1H NMR spectra (DMSO- d_6 , 25 $^\circ\text{C}$) of poly(α ,L-glutamic acid) modified with: **(a, b)** Phe (a – precursor polymer for P[EE(O)E(H)E(F)E(Glc)]; b – precursor polymer for P[EE(O)E(F)E(H₆-pept)]); **(c)** Phe, His(Trt) and Glc; **(d)** Phe, Orn(Boc), His(Trt) and Glc (with diffusion filter). *Signals (ppm):* 0.80-0.85 (CH_3 , hexylamine), 1.00-1.3 (CH_2 , hexylamine), 1.3-1.5 (CH , Boc of Orn), 1.5-2.4 (2CH_2 , Glu, and Orn), 2.7-3.5 (6CH_2 , glucose; CH_2 , Orn), 3.5-4.5 (CH , Glu; CH , His and Orn; CH_2 , His), 6.3-7.5 (C_6H_5 , Trt of His; 2H , His), 7.8-8.5 (NH-CO , all polypeptides; $\text{NH}_2\text{-C=NH}$, Arg).

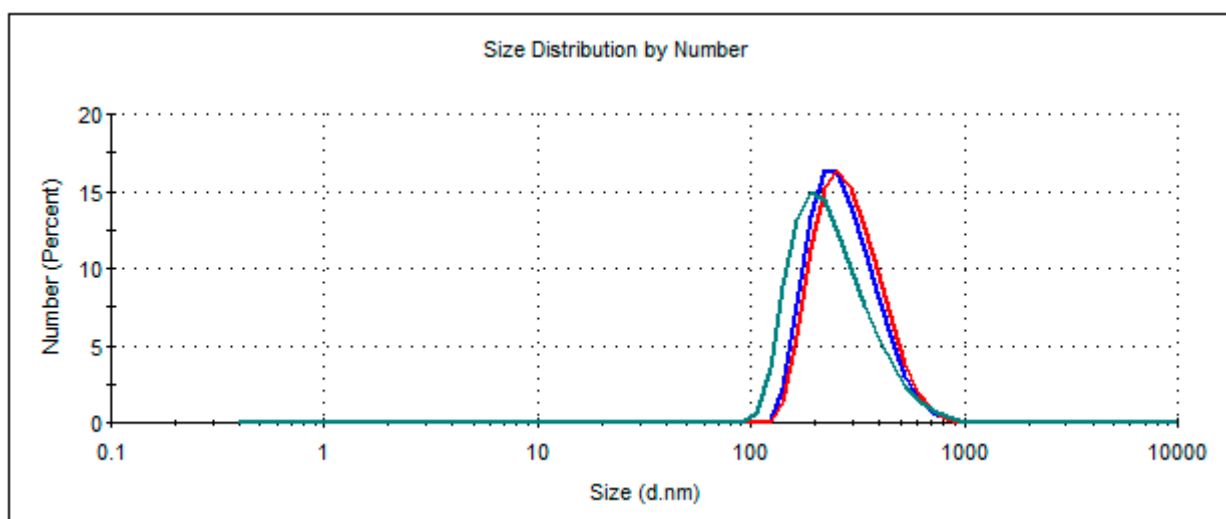
Table S1. Composition of the amphiphilic copolymers determined by ^1H NMR spectroscopy.

Sample	Modifier				
	Determined composition (mol%) *				
	<i>R</i> or <i>O</i>	<i>H</i>	<i>Glc</i>	<i>F</i> or <i>I</i>	<i>H</i> ₆ - <i>pept.</i>
P[EE(O)E(H)E(F)E(Glc)]	33	26	17	10	–
P[EE(R)E(H)E(I)E(Glc)]	25	24	21	N/D	–
P[EE(O)E(H)E(I)E(Glc)]	31	24	19	N/D	–
P[EE(O)E(F)E(H ₆ -pept)]	42	–	–	26	N/D

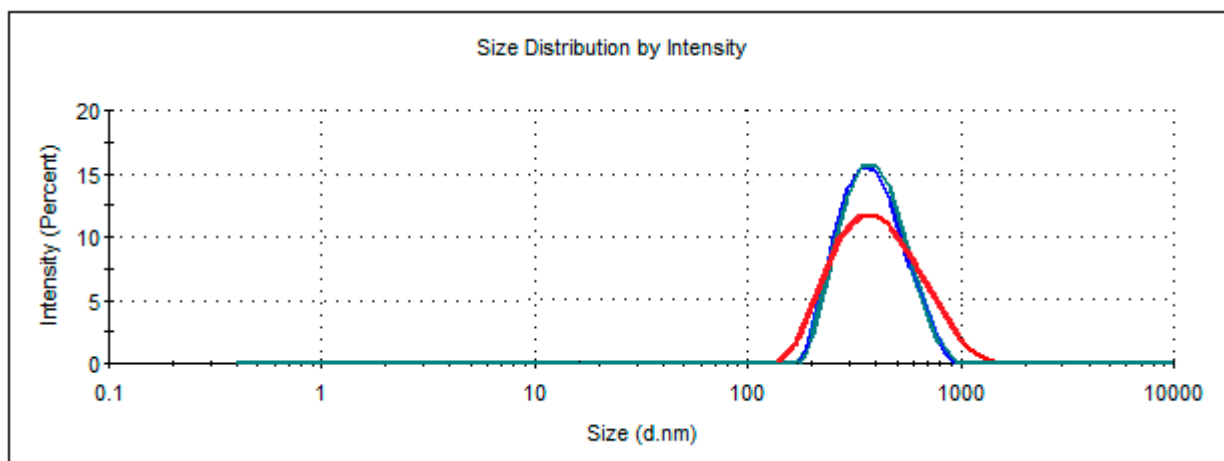
*Side-chain amino acids and glucosamine were calculated relative to glutamic acid, taken as 100%.



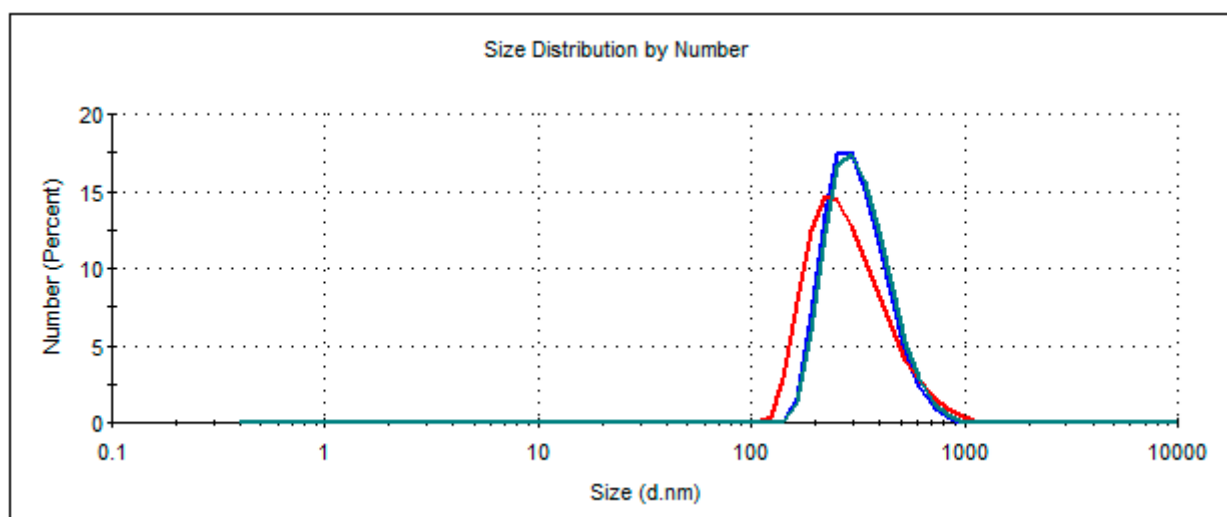
(a)



(b)

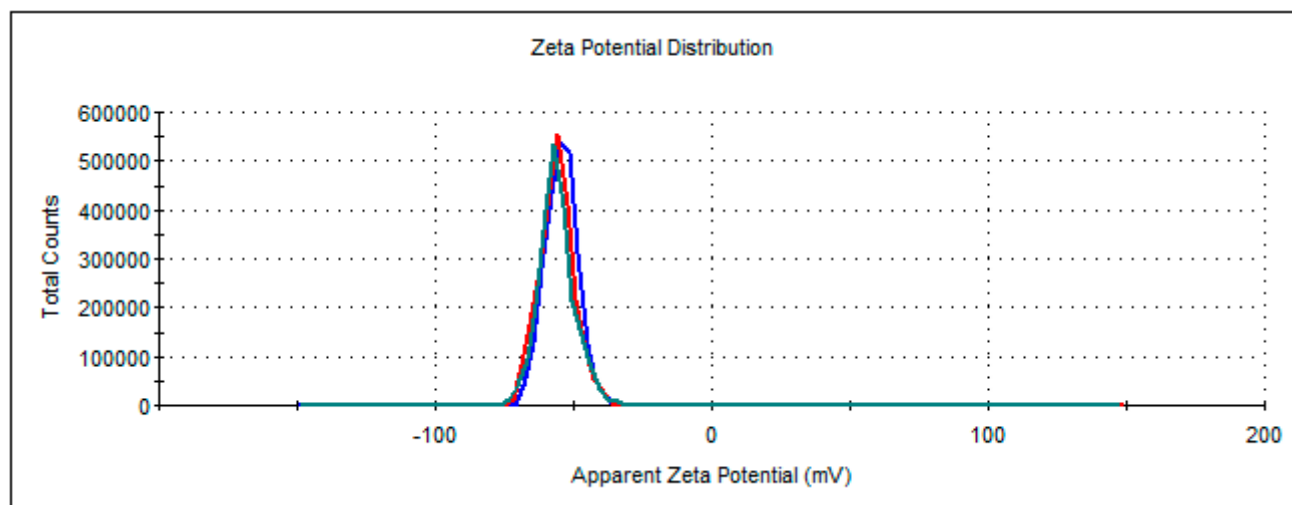


(c)

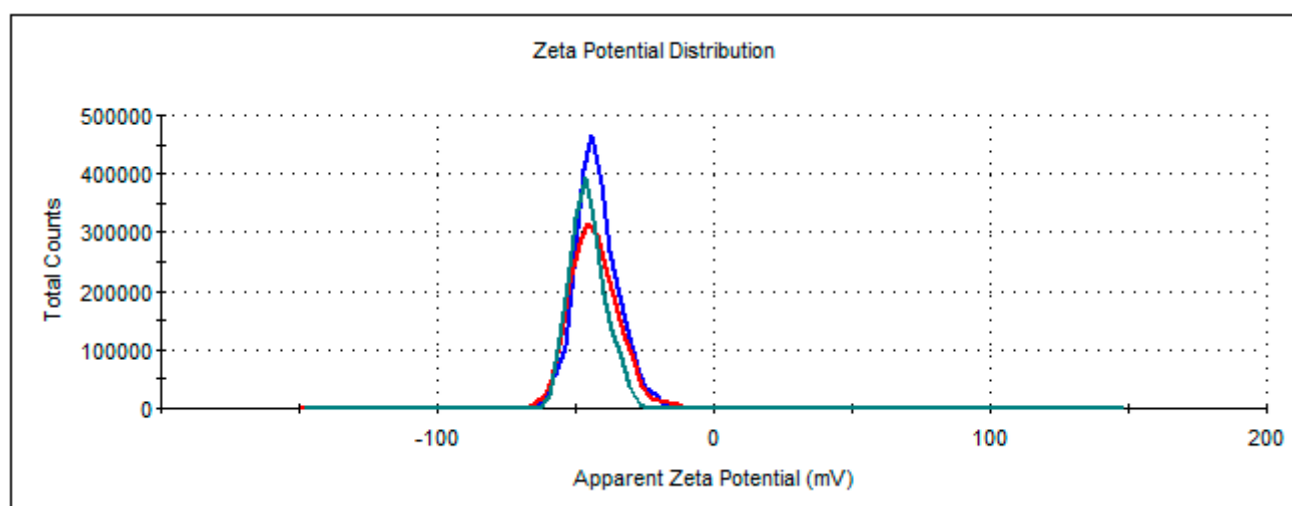


(d)

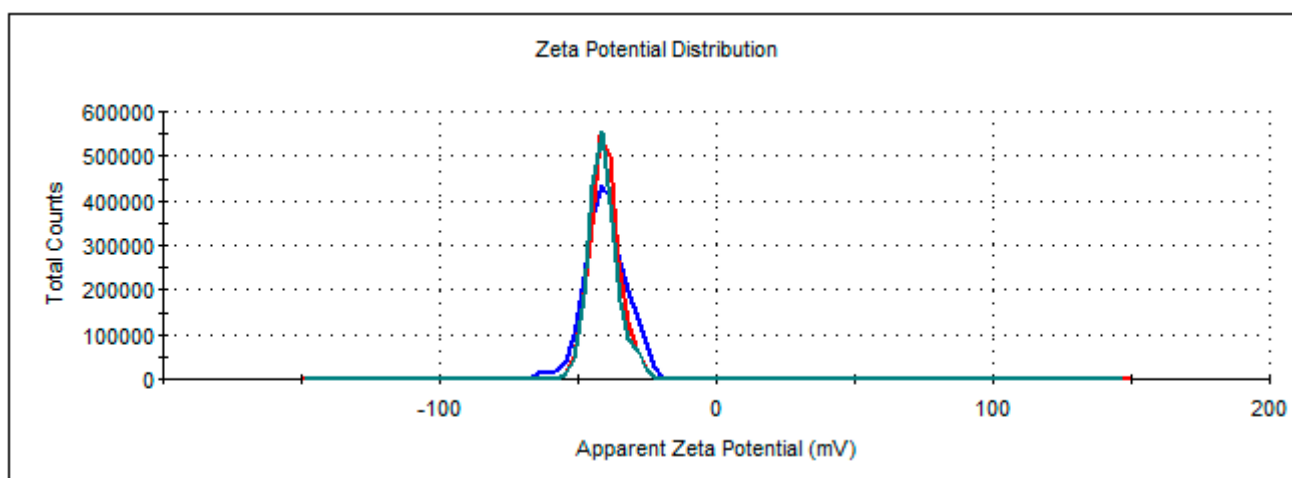
Figure S5. Hydrodynamic diameter distribution measured by intensity (a,c) and by number (b,d) (DLS) for P[EE(O)E(H)E(I)E(Glc)] (a,b) and P[EE(O)E(F)E(H₆-pept)] (c, d) nanoparticles.



(a)

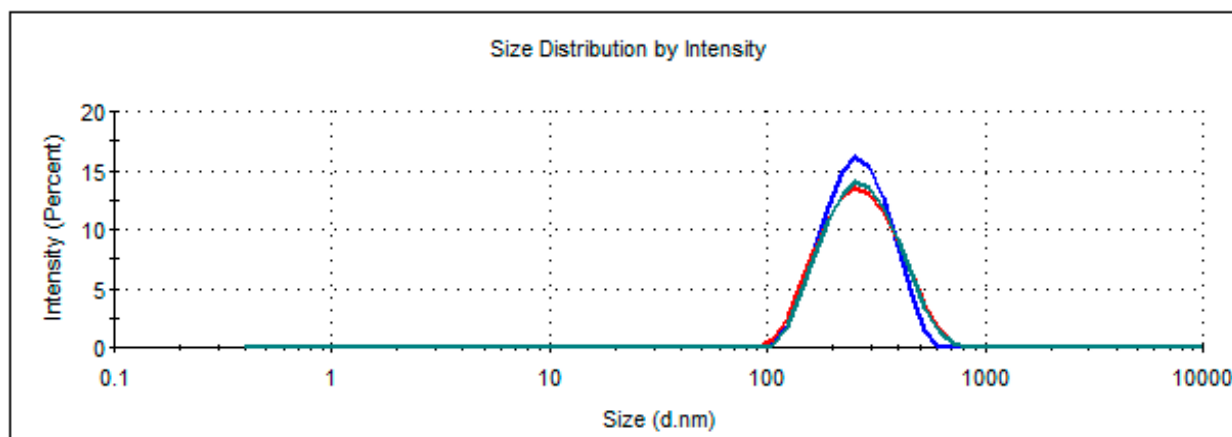


(b)

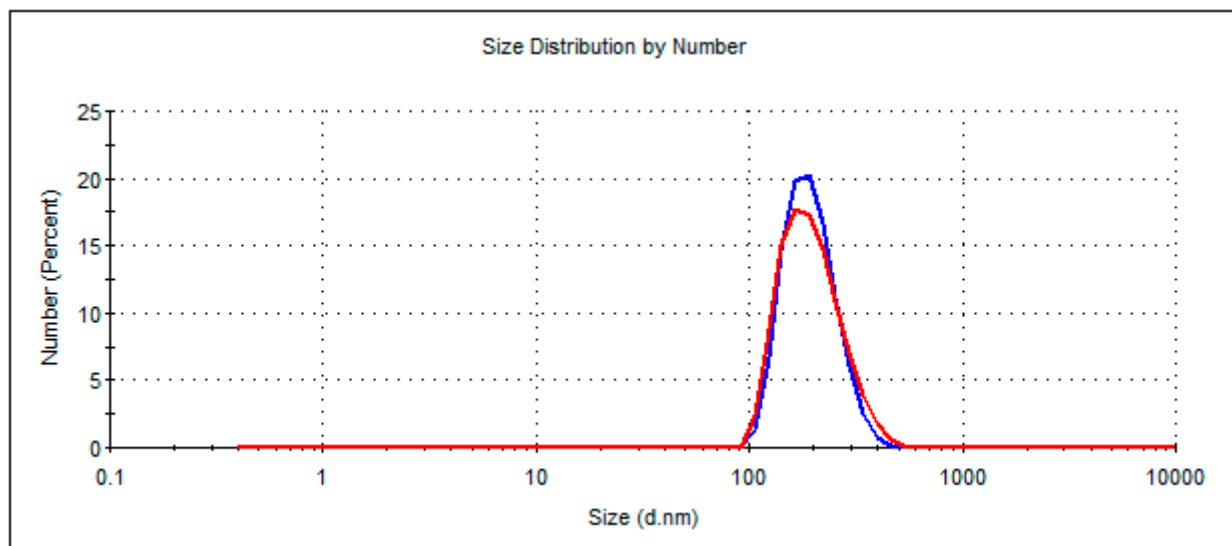


(c)

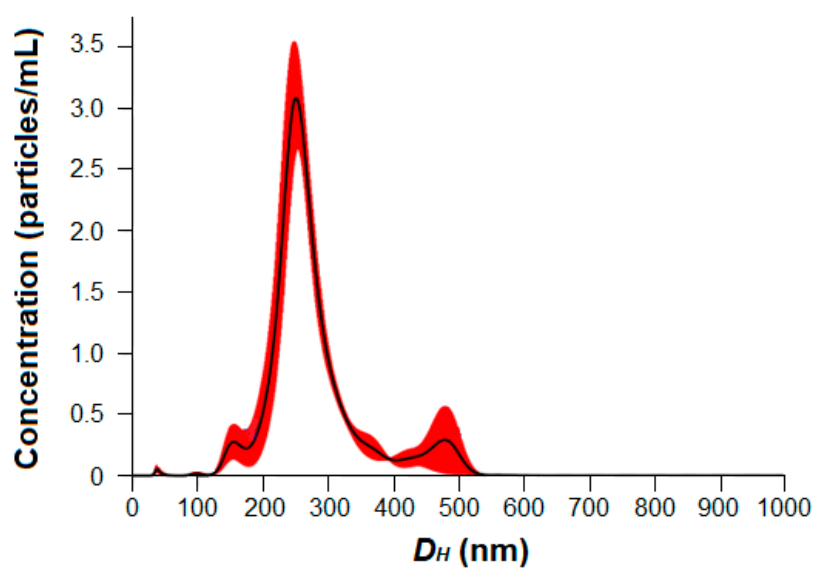
Figure S6. Zeta-potential distribution obtained by ELS for P[EE(R)E(H)E(I)E(Glc)] (a), P[EE(O)E(H)E(I)E(Glc)] (b) and P[EE(O)E(F)E(H₆-pept)] (c) nanoparticles.



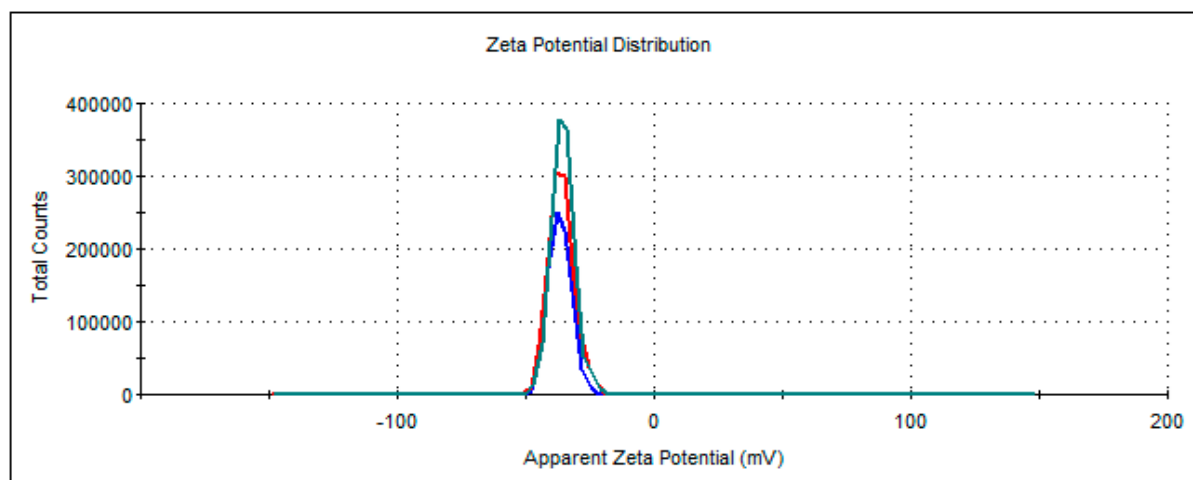
(a)



(b)

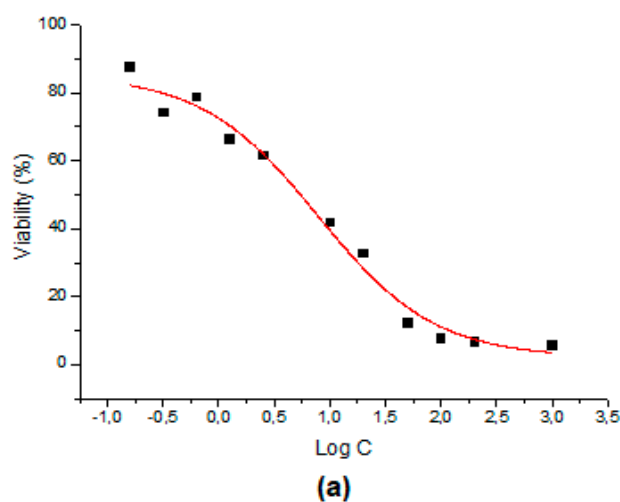


(c)

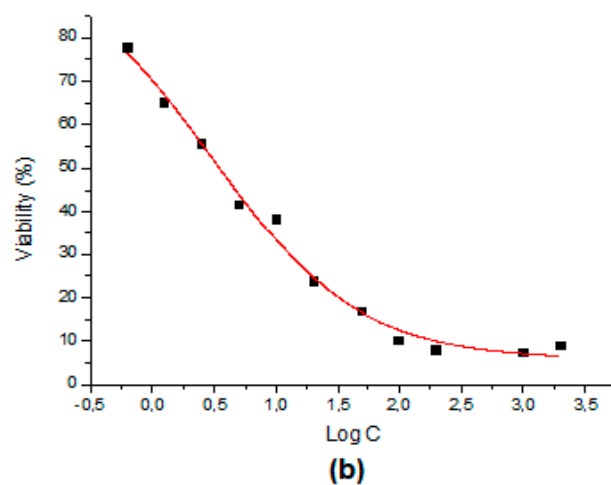


(d)

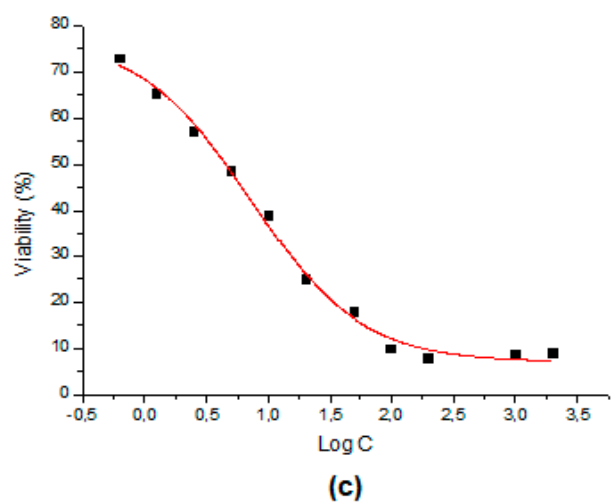
Figure S7. Hydrodynamic diameter distribution (a-c) and zeta-potential distribution (d) measured for PTX-loaded nanoparticles based on P[EE(R)E(H)E(I)E(Glc)] using: DLS by intensity (a), DLS by number (b), NTA (c), ELS (d).



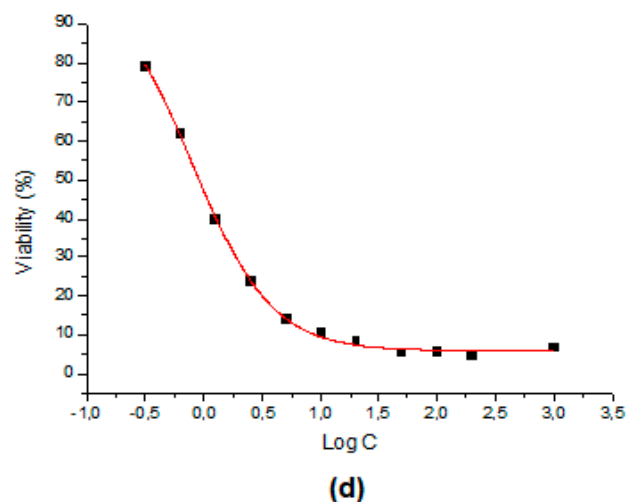
(a)



(b)

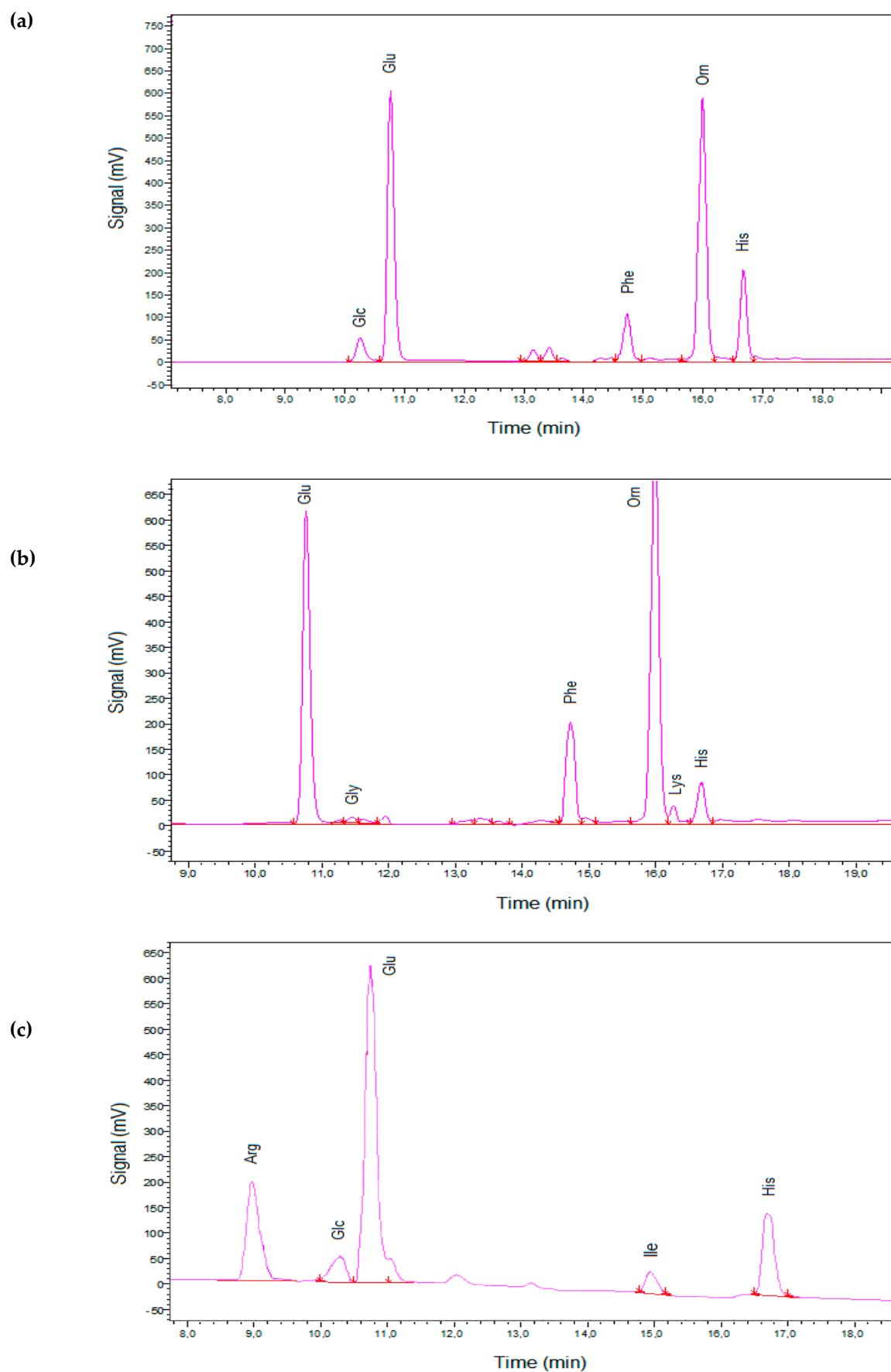


(c)



(d)

Figure S8. Dose response curves illustrating the inhibition of A549 by PTX nanoformulations (a-c) and free PTX (d): PTX/ P[EE(R)E(H)E(F)E(Glc)] (a), PTX/ P[EE(O)E(H)E(I)E(Glc)] (b), PTX/ P[EE(O)E(F)E(H₆-pept)] (c).



(d)

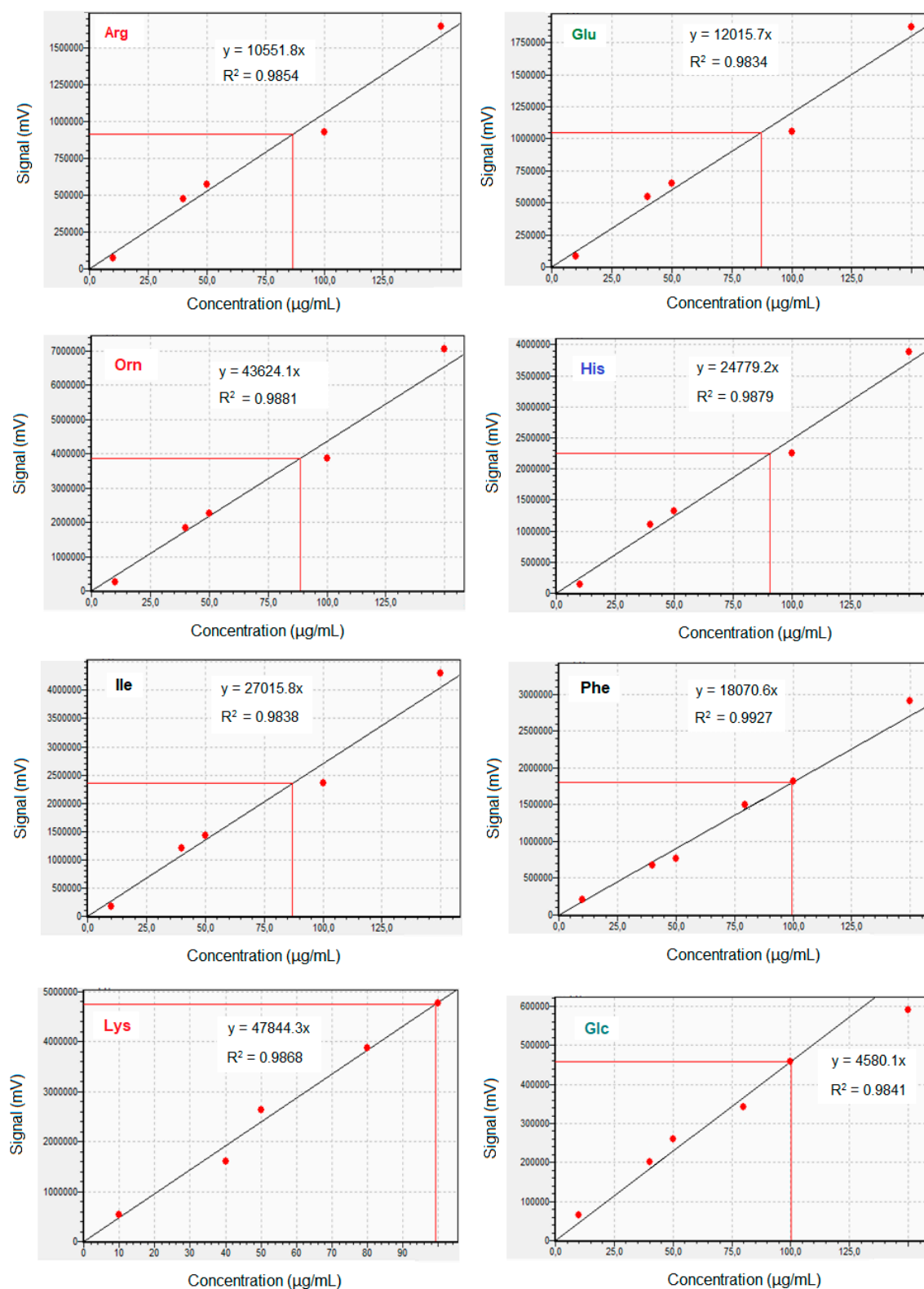


Figure S9. Chromatograms of HPLC amino acid and glucosamine analysis (a–c), obtained after total acidic hydrolysis of (glycol)polypeptides: P[EE(O)E(H)E(F)E(Glc)] (a), P[EE(O)E(F)E(H₆-pept)] (b), P[EE(R)E(H)E(F)E(Glc)] (c) and calibration curves built for individual components (d).