

## **Supplementary Materials**

Development and characterization of indole-responsive whole-cell biosensor based on the inducible gene expression system from *Pseudomonas putida* KT2440

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## Supplementary Tables

**Supplementary Table S1.** Bacterial strains used in the study.

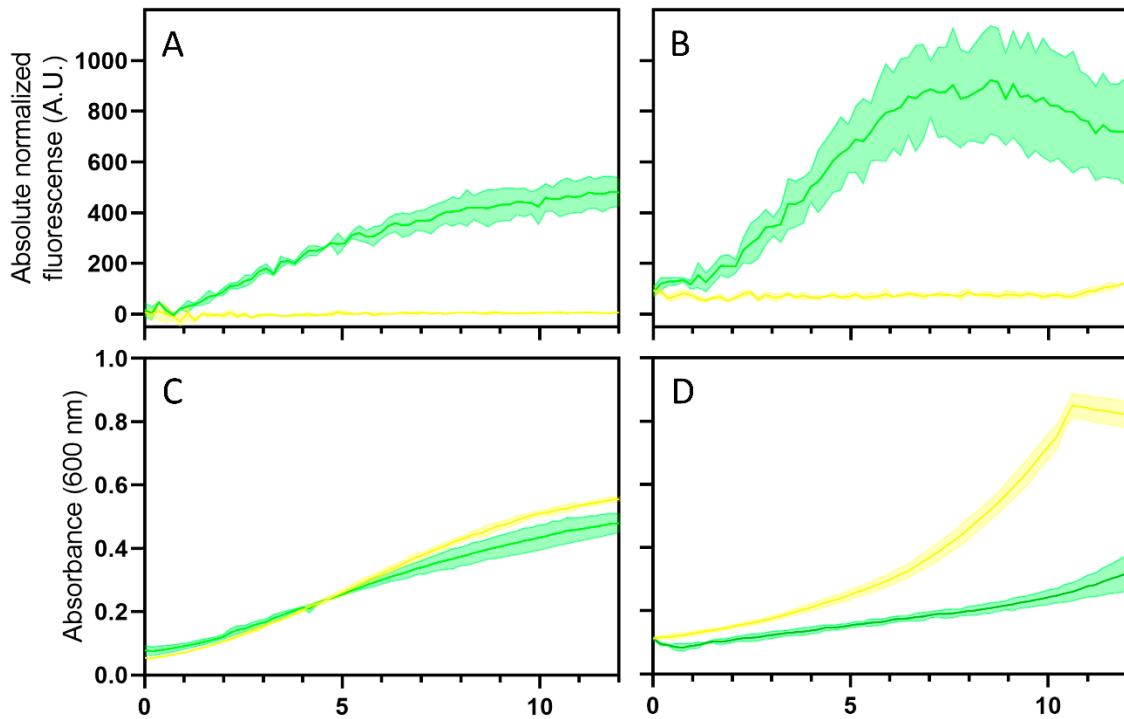
Strain	Characteristics	Reference or source
<i>Escherichia coli</i> TOP10	F <sup>-</sup> <i>mcrA</i> Δ( <i>mrr-hsdRMS-mcrBC</i> ) Φ80lacZΔM15 Δ <i>lacX74 recA1</i> <i>araD139</i> Δ( <i>araIeu7697 galU</i> <i>galK rpsL</i> (StrR) <i>endA1 nupG</i>	Thermo Fisher Scientific, Waltham, MA, USA
<i>Cupriavidus necator</i> H16	Wild-type strain	DSM 428 (DSMZ, Braunschweig, Germany)

**Supplementary Table S2.** Oligonucleotide primers used in the study.

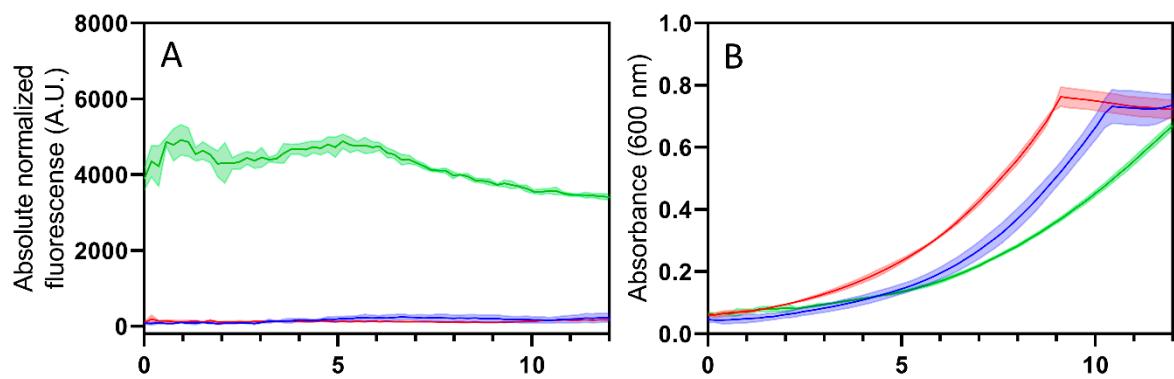
Name	Oligonucleotide primer sequence (5' → 3')
P015	gggccttcgttt <u>atgacgtctgatatgc</u> aaggaaactaagcgctctgc
P016	cgtcttcgctactcgcc <u>atatggcgctcaaggcgtag</u>
P015TgSr	gggccttcgttt <u>atgacgtcttacctgtgagtttctgaca</u> agtttac
P065	tca <u>ctccaccggtgcttaagcaaaagatctttaagaaggagatatacatatggaaaactttaaacatctccctgaaccg</u>
P066	tc <u>cttactcgagttggatccctaaactctttaagttgcggtaagtgac</u>

Restriction endonuclease recognition sites are underlined.

## Supplementary Figures



**Supplementary Figure S1.** Induction dynamics of *PpTrpI/P<sub>PP\_RS00425</sub>*-based *E. coli* and *C. necator* biosensors. Absolute normalized fluorescence (A, B) and absorbance (C, D) obtained using *E. coli* (A, C) and *C. necator* (B, D) harbouring inducible system *PpTrpI/P<sub>PP\_RS00425</sub>* (pPM0081). Measurements were performed over the period of 12 hours in MM supplemented with 1 mM (green) and without indole (yellow). Data are mean  $\pm$  SD,  $n = 3$ .



**Supplementary Figure S2.** Screening and characterization of *tnaA*-positive strains using *C. necator*/P<sub>p</sub>TrpI/P<sub>PP\_RS00425</sub> biosensor. Absolute normalized fluorescence (A) and absorbance (B) of *C. necator* harbouring inducible system P<sub>p</sub>TrpI/P<sub>PP\_RS00425</sub> without (pPM0081; control, blue) or with *tnaA* gene (pPM0083; red and green). Data are mean ± SD, n=3.

A

AatII

gacgtctgtatgcaaggaaactaagcgctctgcgcgaagtccgcctaaccactgcggcaactgtcgccgcgcgtctgcggcgccgtggcacccacaatgc  
end of *trpI* (*PP\_RS00425*)

start of *trpI*

NdeI

catcactcattcaactacgcctggagcgccatatg

start of *rfp*

B

AatII

ggcgttttacgttgatgtttctgacaagttacgcatacttgcgtttcagccgactgcccgtggtagagtaagcccatactcattcaactacgcctggagcc

NdeI

catatg

start of *rfp*

C

BgIII

agatcttaagaaggagatatacatatggaaaactttaaacatctccctgaaccgttccgcattcgttgtattgagccagtaaaacgttaccactcgcgcttatcgtgaagaggc  
start of *tnaA*

BamHI

agagaacgcggcgaatattaaaggattaaccttacgtacgaaccgaaaagtattgcgtcaactcaccgaaaaacttaagaagttaaggatcc

end of *tnaA*

**Supplementary Figure S3.** Nucleotide sequences of genetic elements of the indole-inducible system. (A) Nucleotide sequence containing *trpI* gene, encoding LysR family transcriptional regulator (locus tag *PP\_RS00430*), and intergenic region *PP\_RS00430/PP\_RS00425*, assembled in pPM0081 and corresponding to the schematic representation in Figure 1B. (B) Sequence of intergenic region *PP\_RS00430/PP\_RS00425* (in pPM0082 and Figure 1C). (C) Nucleotide sequence of *tnaA* gene encoding tryptophanase (in pPM0083 and Figure 1D). The start and end of open reading frames are displayed in bold. AatII, NdeI, BglII, and BamHI restriction sites are underlined. The nucleotides of intergenic region *PP\_RS00430/PP\_RS00425* are highlighted in green.