

Supplementary Materials

MISF2 encodes an essential mitochondrial splicing cofactor required for *nad2* mRNA processing and embryo development in *Arabidopsis thaliana*

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Supplementary data files:

Figure S1: The topology and structure of MISF2 protein.

Figure S2: MISF2 gene expression pattern during *Arabidopsis* development.

Figure S3: Nucleotide sequence of the *MISF2* gene and precise location of T-DNA insertion sites.

Figure S4: Steady-state level analysis of various mitochondrial proteins in wild type, *misf2.2* and functionally complemented *misf2.2* plants.

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Table S1: List of oligonucleotides used in this study.

Table S2: List of antibodies used in this study.

a

MPP/ICP55 cleavage site

MLTKLRISKLVSYTLPRIFQRFLVTNNNTAEEPIVAAESPELPSWIKDFLSNKPS
 SSVSKDDED**F**VIPSLANWESQKFSRQQVSEGNNVKPVEDIDKVCDFLNKKDTSHEDVV
 KELSKCDVVVTESLVLQVLRRFSNGWNQAYGFFIWANSQTGYVHS | **GHTYNA**MVDVLGKC
 RNF**DLM**WELVNEMNKNEESKL | VTI | **LDTMSK**VMRRLAKSGKYNKAVDAFLEMEKSYGVK
 TD | **TIA**MNSLMDALVKENSIEHAHEVFLKFDTIKPD | **ARTF**NILIHGFC**K**ARKFDDARA
 MMDLMKVTEFTPD | **VVTYTS**FVEAYCKEGDFRRVNEMLEEMRENGCNPN | **VVTY**TIVMHS
 LGKSKQVAEALGVYEKM**K**EDGCVPD | **AKFYSS**LIHILSKTGRFKDAAE**I**FEDMTNQGVRR
 D | **VLVY**NTMISAALHHSRDEM**AL**RLLKRM**E**DEEGESCSPN | **VETY**APLLKM**C**CHKKM**KL**
 LGILLHHMV**KNDV**SID | **VSTY**ILLIRGLCMSGKVEACLFEEAVRKGM**VPR** | DSTCKML
 VDELEKKNMAEAKLKIQS**L**VQS**K**TM**I**DSH**S**PLS**V**S

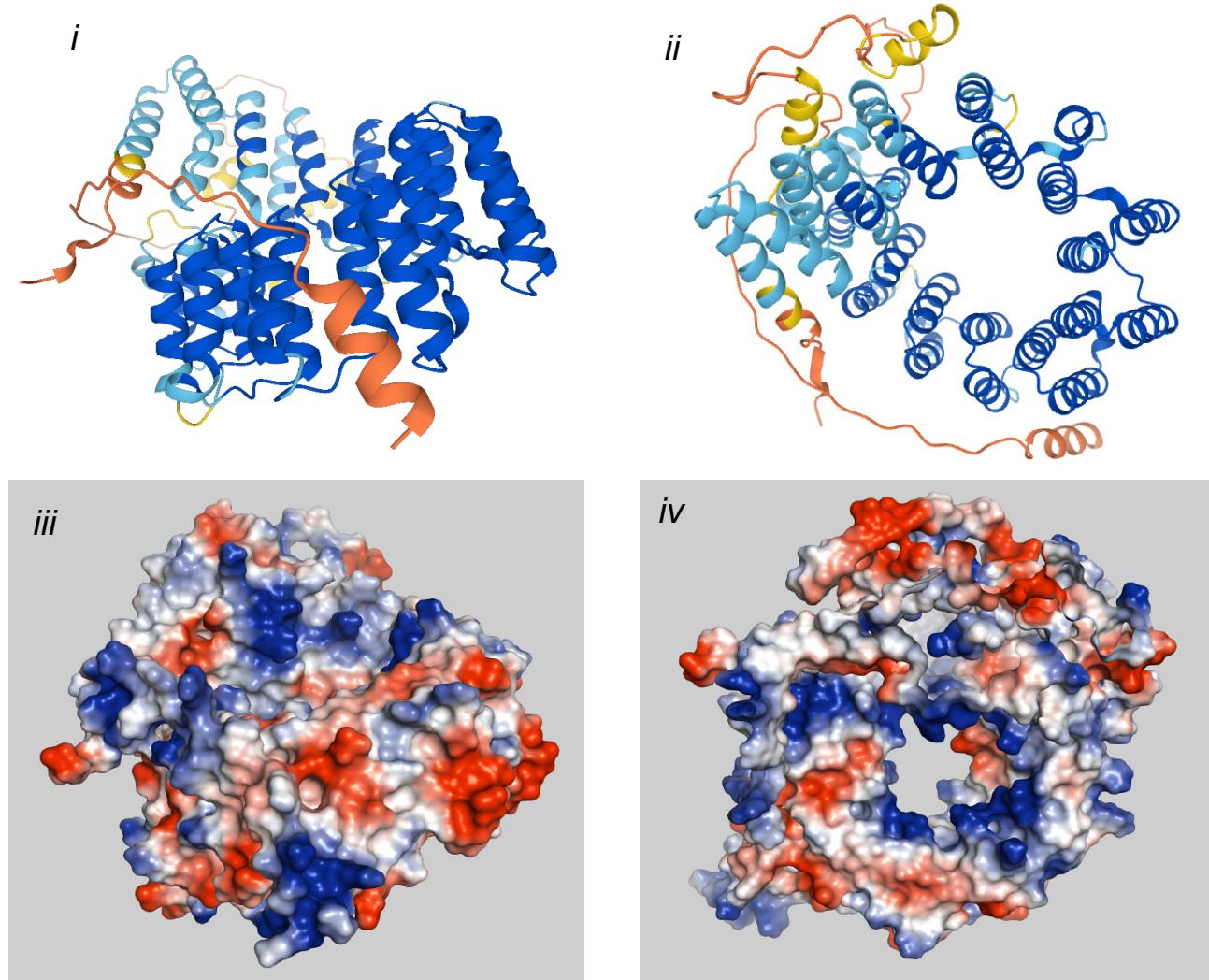
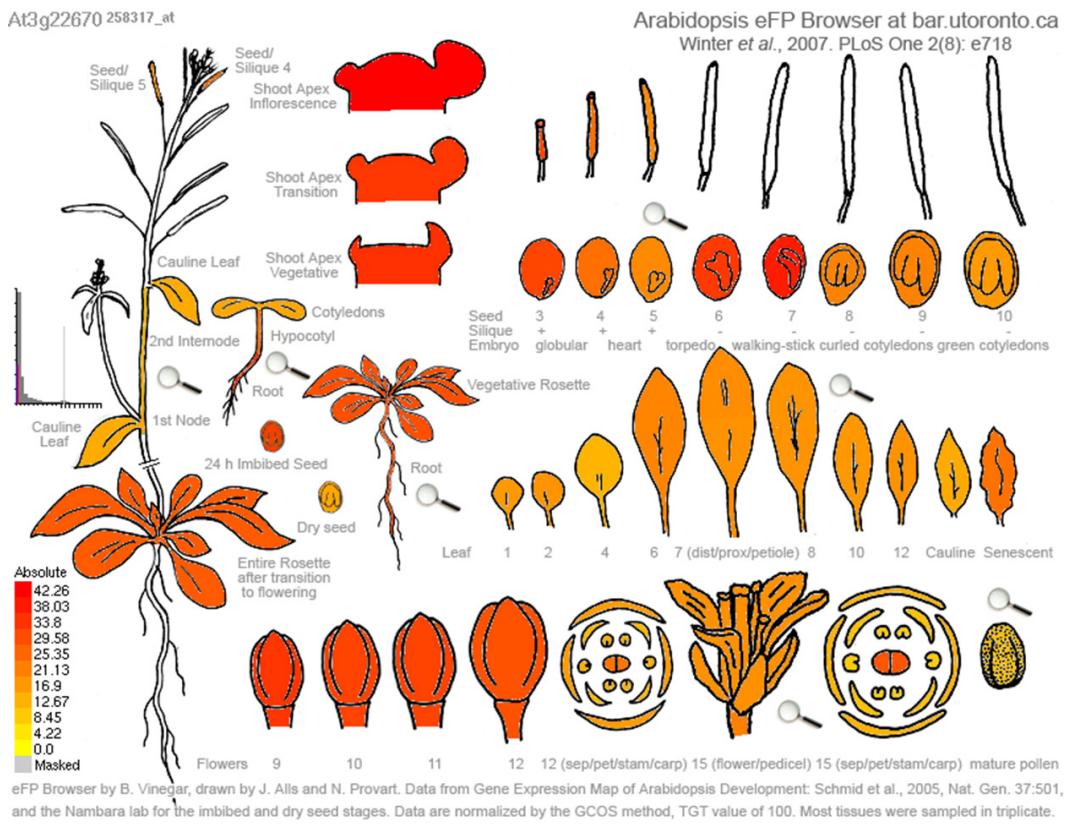
b

Figure S1. The topology and structure of MISF2 protein. (a) Amino acid sequence of the MISF2 protein (At3g22670). The region corresponding to the predicted mitochondrial targeting sequence is underlined and highlighted in red. The predicted cleavage sites of the Mitochondrial Processing Peptidase (MPP) or Intermediate Cleavage Peptidase 55 (ICP55) is indicated with a red arrow. The 10 PPR motifs (highlighted in blue letters) were predicted with the PPRfinder (Gutmann et al. 2020), PPRCODE (Shen et al. 2019), SMART (Letunic et al. 2012) and CDD (Marchler-Bauer et al. 2003) web-servers. Underlined letters indicate the 5 and 35 positions within each PPR repeat. (b) Atomic structural model of MISF2 predicted by the AlphaFold server (Jumper et al. 2021). Ribbon structures (i and ii) were generated by the PyMol software suite. Panels "iii" and "iv" represent charges on the surface of MISF2 from two different angles, with red, white, and blue for negatively, neutral, and positively charged residues, respectively.

a



b

Dataset: 10 developmental stages from data selection: AT_AFFY_ATH1-0

Showing 1 measure(s) of 1 gene(s) on selection: MSP2

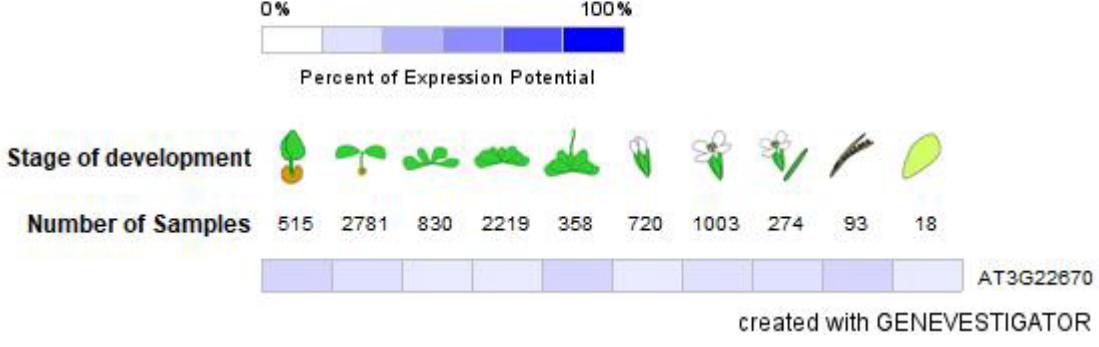
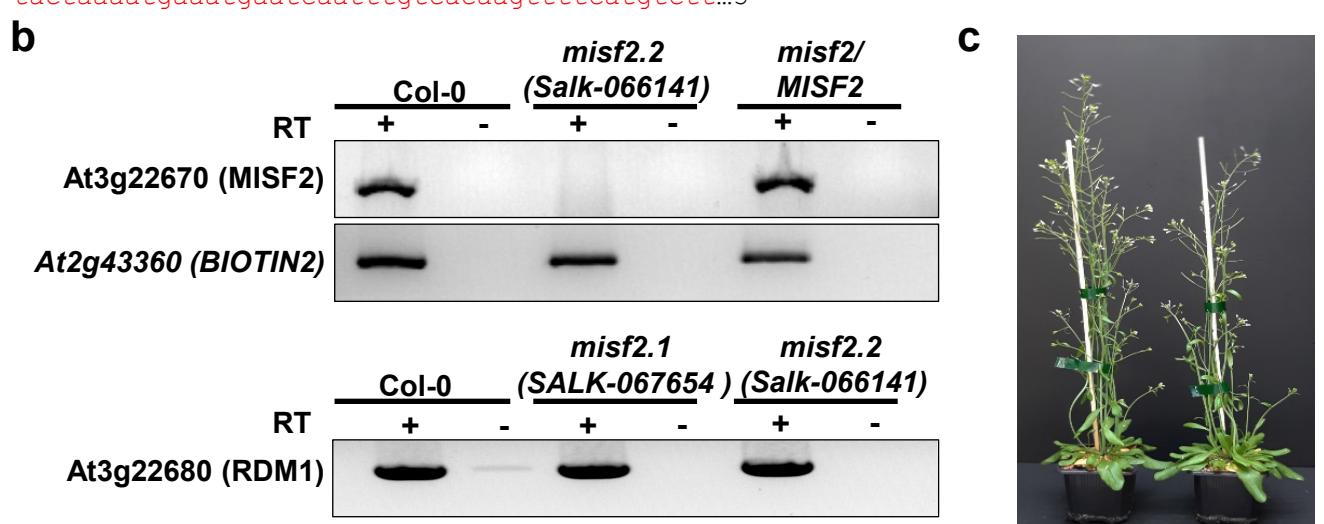


Figure S2. MISF2 gene expression pattern during *Arabidopsis* development. The expression pattern of MISF2 was analyzed using publicly available microarray and high-throughput RNAseq databases, including (a) 'The *Arabidopsis* Information Resource' (TAIR; <http://www.arabidopsis.org>), and (b) 'Genevestigator analysis toolbox' database (Hruz et al. 2008, Zimmermann et al. 2004).

a 5' ...agatggaaaggaaagattcacattttattgaataagaccctgcaattatacatataagcttaatgcttgaacaaggttctaa
aagcaaaaagttagataaaaaaggatggttcatcacaattcaagaacttgt**TCAT**TTCTCAGGAAAGATTGGGTCAATGAAAGAGT
GATACATAGGATCAGATCTCCAAAAGAAGAGCCATGTGAAGATGAGAAGGAGTGAGTCGATGAATCTCTCAACAAGCCAGATAGTA
GCTTCAGCTTAGGATGAATGATAGAATCCAACCTCTGTTCTCAGAATCAGTACCGAATCGTATTGATCCCATCGTTGAA
GAGAACATTGAGGTAATGTAGAGGCTGACCATAAAGCTGTTTCAATTGAAATGCTTAATCCAACCCAGCTGTGAACGGAATCA
ACGAGGCCACGATTGTCGGGATTGGTACTTGCTCATGTTGATACATCTGCTCTCAGCAACGAAC**Ctgcactcaaaa**
aatttcagagcattttgtaaaaatcaaaactttgaatcaaaatggtagaaaaggaaagaaaacaaggaccattgaaatcgta
cCTTCATCAGCGACGTCTGTGAGAAGTATCGAGCGGTGAAAAGCCATCGCTGATCTCAGCATCGACGTAGATGAACCGGAATC
ACCTGATGGTCGTAGTCCATTGAGCTTG**CAT**ggta**cactgtcgagcagttagatttgacggcgtagaaagcagaaaagg**
tacgaagacgatgaccacacgagttcctaaaatttagggtttagaggaagaagacgaagagaatgactgtacacgtgtcagt
aaaaatcgtaacttgtcagatgtggggtaaacaaaaatctaaacgctaaattaaatcgaaaccggggtgttaccaattaaac
gaaccgaccgcaccggcacaaggcagcaagcttaggtctaaaagattcgaaacttgcggaaaaacaggagagaagtcgtcgt
ttagatcaaacgaaacaaa**ATG**CTCACTAACGTTAGGATTAGTAAGCTTGTTCGATCTTACCTCGGAGATCTTCAAACGG
AGGTTCTGGTAACGAATAACACTGCAGAGGAGTCGCCATTGTCGCCAGAGTCACCGGAGCTACCGTCTGGATTAAAGACTT
TCTATCAAACAAGCCTCTTCTCATCGAGTTGCCAAAGATGATGAAGATTGTTATCCCTCTTGGCGAATTGGTCG
AGAGCCAGAAGTTAGTCGTAACAAGTCTCAGAGGGAAATGAGTGAAGAACCACTGGAGATATTGATAAAAGTTGCACTTT
▼ TTGAATAAGAAGGACACGTCATGA ▼ GGATGTTGTTAAAGAGTTGAGTAAATGCGATGTTGGTACAGAGAGTTGGTTT
GCAAGTCTAAGAAGGTTAGTAATGGTGAATCAAGCTACGGGTTTCATTGGCAAAATTCAAAACAGGTTATGTCATT
CAGGTCAACTTATAATGCAATGGTGTGTTGGGAAATGTAGGAATTGATGTTGAGTGGAAATTAGTCATGAGATGAA
AAGAATGAAGAATCTAACGTTACGCTTGATACGATGAGTAAGGTTATGAGGAGATTGCCAGTCAGGGAAATAATAAAAGC
TGTTGATGCTTTTAGAGATGGAGAAGAGTTAGGTGTGAAAACAGATACTATTGCTATGAAACAGTTGATGGATGCACTGTGA
AAGAGAATAGTATAGAGCATGCTCATGAAGTTTCTGAAGCTTCTGATACAATCAAGCCGGATGCGGGACATTCAACATTG
ATTCAACGTTTGAAAGCTGGAAATTGATGATGCTAGAGCGATGATGGATTGATGAAGGTTACTGAGTTCACTCCTGATGT
TGTTACTTACACTAGCTTGTGAAAGCTTGTGACTTATACTATTGATGCTATTGCAAGGAAGGGATTAGGAGGTTAATGAA
ATGTTGAGTGGAAATCTAAACAGTAGCTGAAGCTTGTGAGGAGATGCTGAGGAGATGCGAGAGA
ATGGGGTGAATCTAATGTTGACTTATACTATTGATGCTATTGCAAGGAAGGGATTAGGAGGTTAATGAAATGTTGAGAG
TATGAGAAGATGAAAGAAGATGGTGTGTTCTGATGCTAAGTTTATAGCTATTGATGAAAGACTGGTAGGTT
TAAGGATGCACTGAGATATTGAGGACATGACGAATCAAGGAGTCGCTGATGTTAGTCTACAATACCATGATTCCGCTG
CGCTTCACTCACGAGATGAGATGGCTTACGTTGCTAAAGGATGGAGGATGAGGAGGGGAATCATGTTAGTCAACACGTT
GAGACTTATGCTCCATTGTTGAAGATGTTGCCATAAGAAGAAAATGAAGCTACTTGAATATTGATCACATGGTAAAAAA
CGATGTTAGTATCGATGTTGACATATTCTCTCATTAGAGGTTATGATGAGCAGGGAAAGTTGAAAGAAGACTGGCTTCT
TTGAGGAAAGCAGTACGAAAAGGAATGGTCTAGAGATAGCAGTGCATAAGGAAATGTTGAGTGGAGCTTGTGAGAAG
GAAGCAAAACTCAAGATTGAGTTGTTCAATCGAAAACAATGATTGATTCTCACAGTCCTTATCTGTTCTG**TAG**agattaga
gttttgagcctggctcaaagtttttttcccttctgttgaatgatactcttatactatgttagatctcaaattatgtgaccc
tactaaaatgaaatgaaatcaatttgcatacaagtttcatatctt 3'



misf2/MISF2 Col-0

Figure S3. Nucleotide sequence of the MISF2 gene and precise location of T-DNA insertion sites. (a) Nucleotide sequence of MISF2 and RDM1, encoded by the At3g22670 and At3g22680 gene-loci, respectively. Lowercase letters colored in red indicate 5' and 3' untranslated regions (UTRs), as revealed by 5' and 3' RACE analysis, while uppercase letters correspond to the MISF2 open reading frame. Positions of T-DNA insertions in *misf2.1* (SALK-067654, 324 nts upstream to the AUG) and *misf2.2* (SALK-066141, 350 nts upstream to the AUG) lines are indicated. (b) RT-PCR analysis monitoring the expression of MISF2 in wild type (Col-0), homozygous *misf2.2* mutants (Salk_066141) and functionally complemented *misf2.2* plants (*misf2/MISF2*). (c) Photo of 5-week-old plants displaying the phenotypic recovery of *misf2.2* mutant by the expression of a complementary gene.

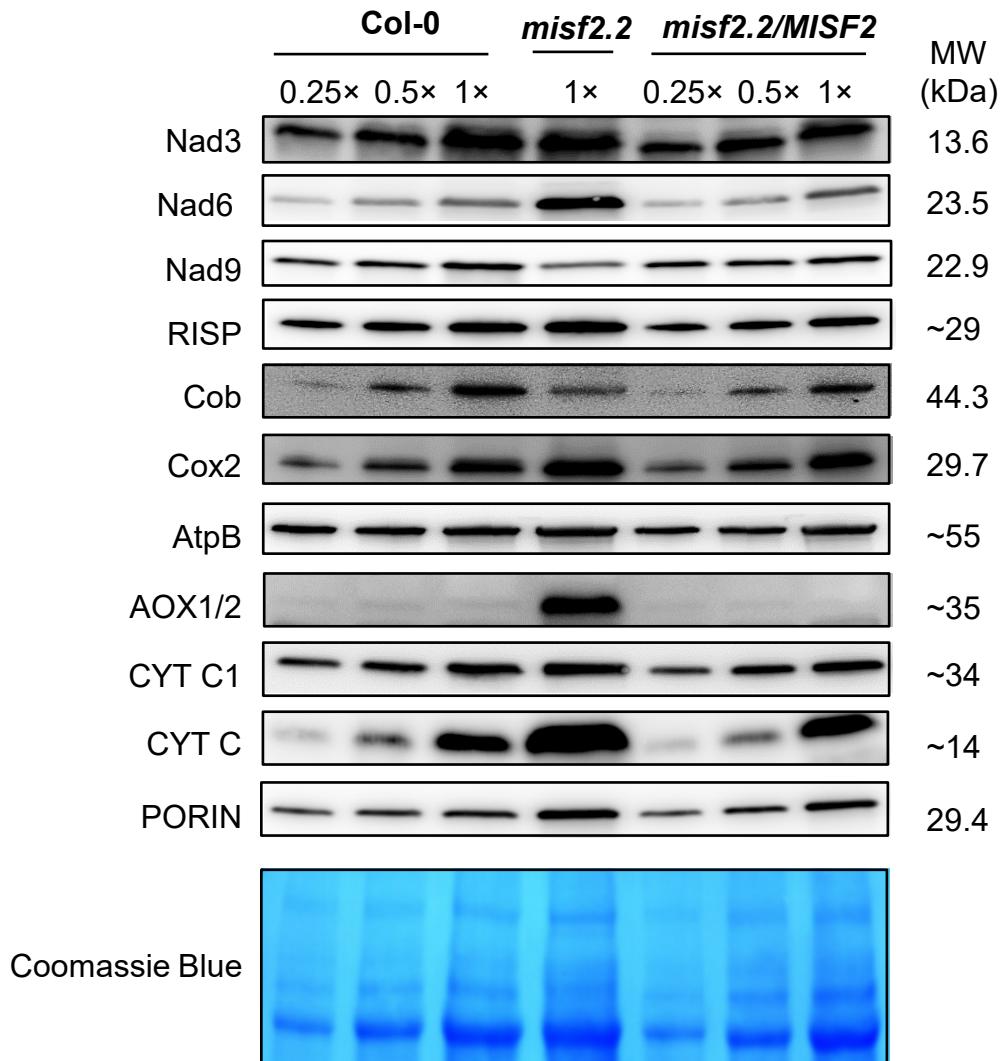


Figure S4. Steady-state level analysis of various mitochondrial proteins in wild type, *misf2.2* and functionally complemented *misf2.2* plants. Immunoblots performed with total proteins extracted from 3-week-old MS-grown wild type plants, rescued *misf2.1* plantlets, and complemented *misf2.2:35S-MISF2* plants. The blots were probed with polyclonal antibodies raised against the indicated mitochondrial proteins. Dilution series of proteins extracted from wild type and complemented plants were loaded for signal comparison with *misf2.1* extracts. Detection was carried out by chemiluminescence assays after incubation with HRP-conjugated secondary antibody. The original SDS-PAGE, following Coomassie-blue staining, is indicated below the immunoblot panels.

MISF2 homologous sequences in different angiosperm species

b Phylogenetic tree of MISF2/EMP10 sequences in plants

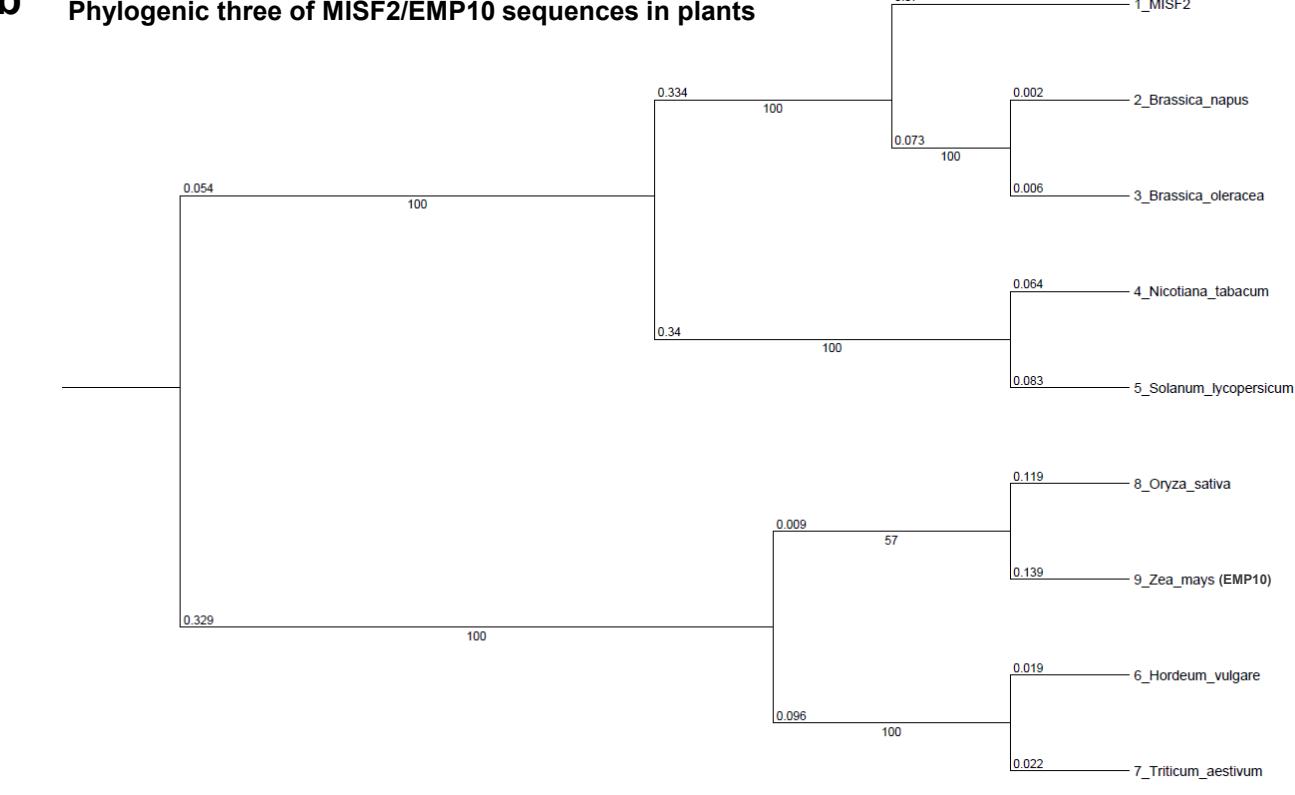


Figure S5. Phylogenetic analysis of homologous MISF2 in plants. Homologues sequences to the MISF2 protein were obtained by the Basic Local Alignment Search Tool (BLAST), against the plant genome sequence databases. (a) Alignment of homologous MISF2 proteins from *A. thaliana* (NP_188906.1), *B. napus* (XP_013736563.2), *B. oleracea* (XP_013623013.1), *N. tabacum* (XP_016433260.1), *S. lycopersicum* (XP_004247584.1), *H. vulgare* (XP_044948743.1), *T. aestivum* (XP_044382896.1), *O. sativa* (XP_015628161.1) and *Z. mays* (EMP10, NP_001183846.1), was conducted with the MAFFT multiple sequence alignment server [83] and displayed using GeneDoc [82] with the conserved residue shading mode. A phylogeny tree (b) was constructed, using the MAFFT and Archaeopteryx tree

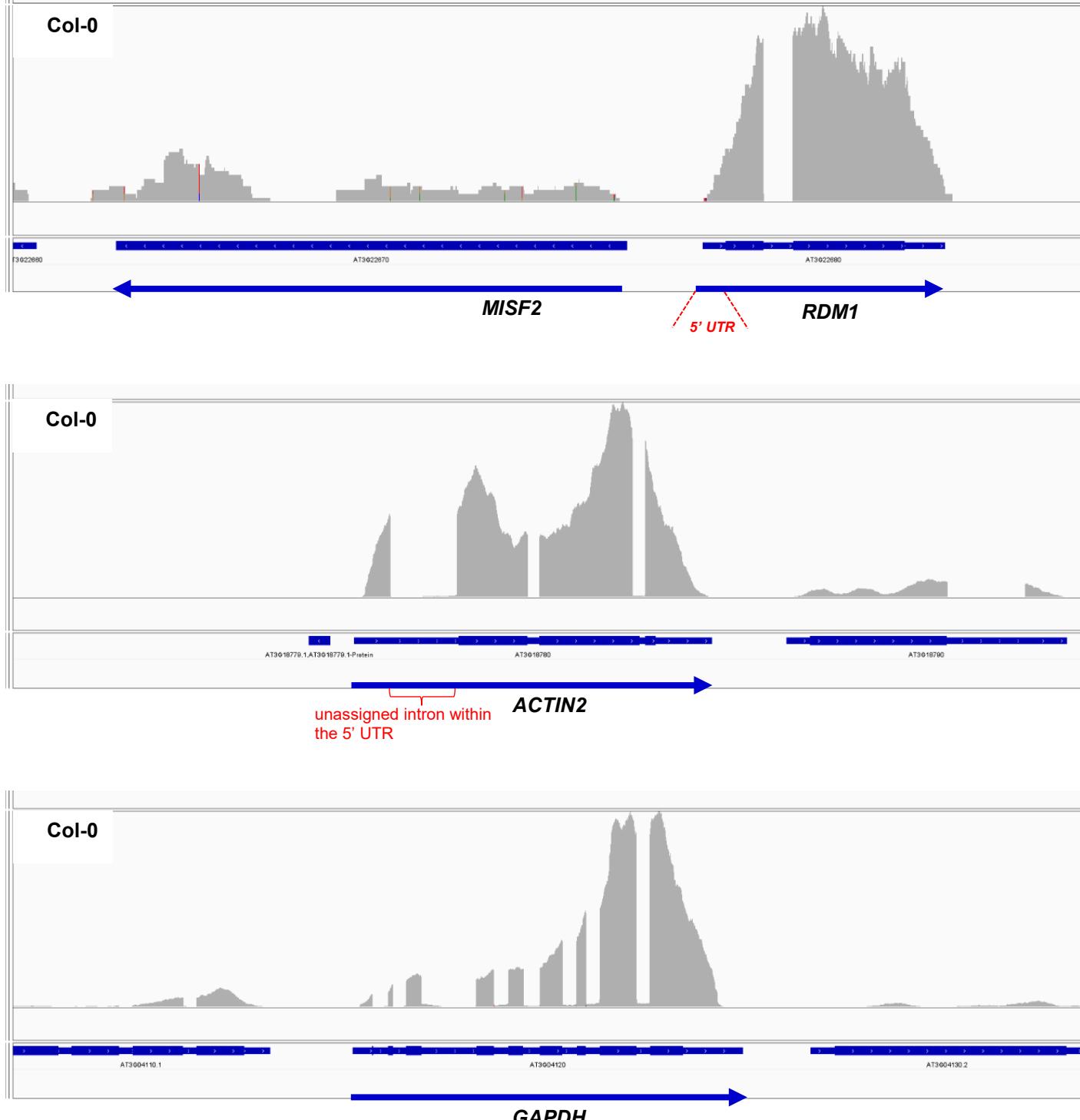


Figure S6. Transcription profiles of *RDM1* gene by RNA-seq analyses. Total RNA extracted from Col-0 plantlets was used to generate Illumina-sequencing (NovaSeq 6000, 150 bp paired-end) libraries (BioProject: PRJNA472433) [84]. Based on these datasets we analyzed the read coverage of *RDM1* (At3g22680) and *MISF2* (At3g22670), as well as *ACTIN2* (At3g18780) and *GAPDH* (At3g04120) genes, in *Arabidopsis thaliana* (Col-0) plants. Thicker boxes indicate coding sequences (exons), while thinner regions indicate UTR and intron regions in Col-0 plants. The RNA-seq data indicated that *RDM1* harbors a 5' UTR of about 50 to 80 nucleotides long. The data also revealed to unannotated introns within the 5' UTR of *ACTIN2* gene.

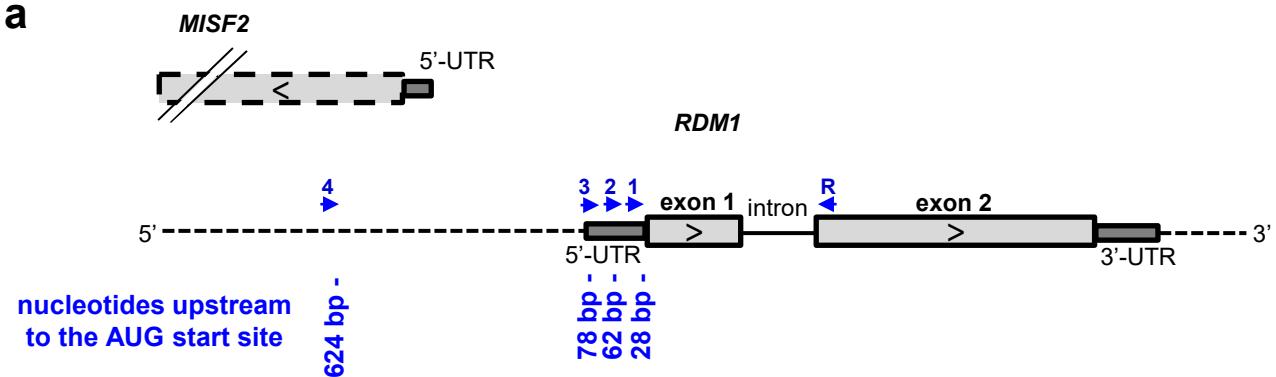
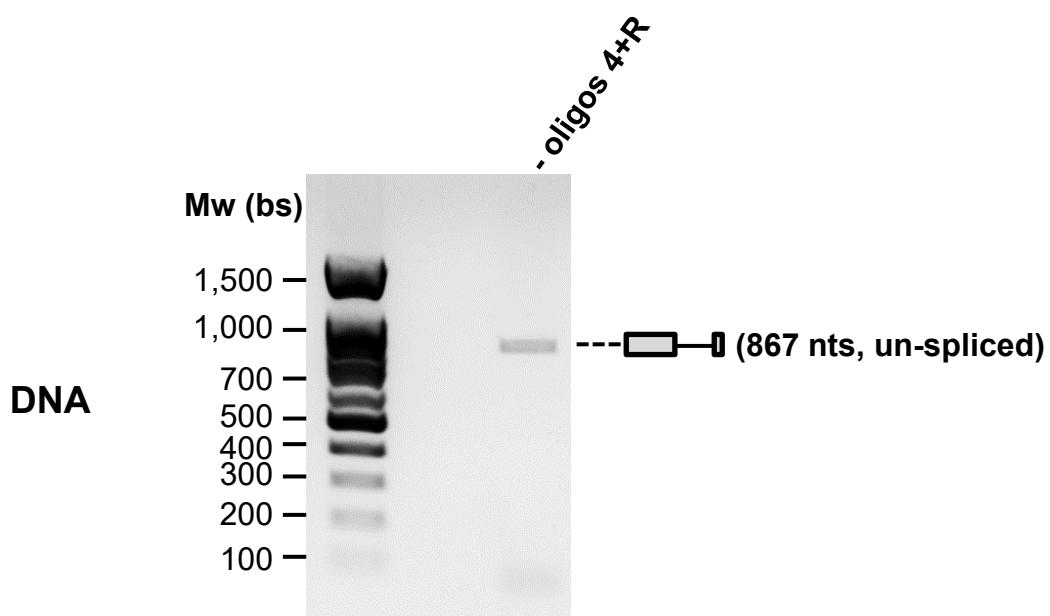
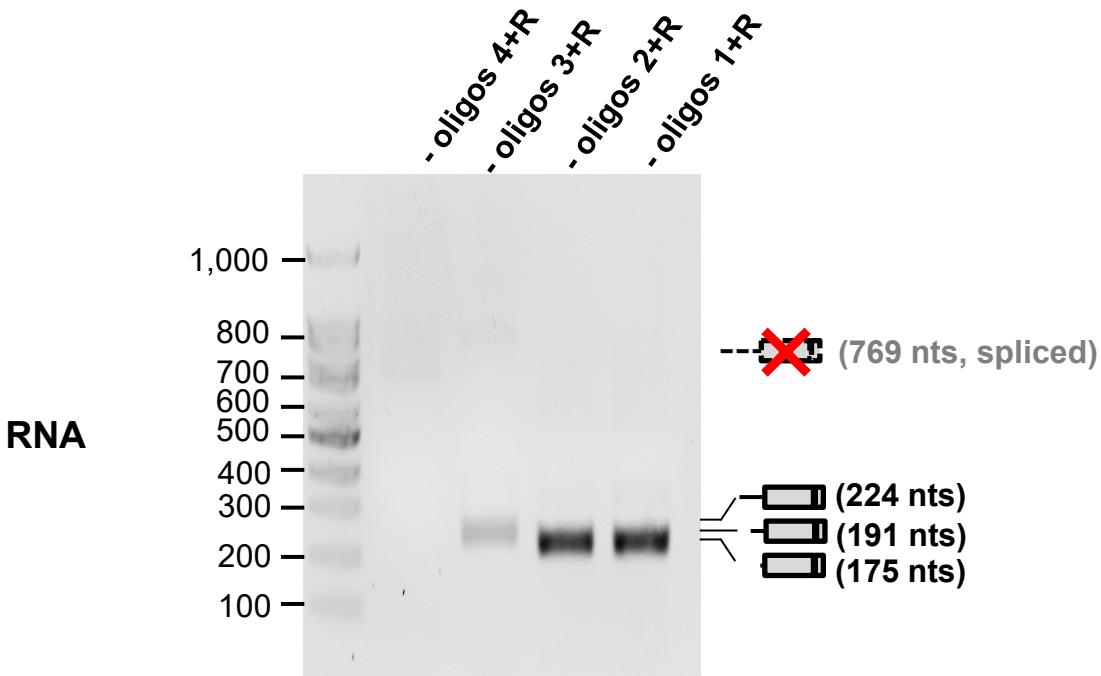
a**b****c**

Figure S7. Analysis of the 5' UTR of *RDM1* transcript by RT-PCR. (a) Scheme of the At3g22670 (*MISF2*) and At3g22680 (*RDM1*) gene-loci and gene structure. The two exons (1 and 2) consist of 124 and 368 nts, respectively, are separated by a 98 bp intron region. The position of the oligonucleotides used in PCR (panel 'b', DNA) and RT-PCR (RNA, panel 'c') are indicated. Oligonucleotides 1, 2 (28 and 62 nts upstream to the initiation codon, respectively) and the reverse primer (R, found at the very 5' of exon 2) are located inside the coding region of *RDM1* gene. Oligonucleotide 3 is found 78 nts upstream to the AUG start codon, while oligonucleotide 4 is found 624 nts upstream to the AUG site (inside the *MISF2* coding region).

Table S1. List of oligonucleotides used in this study.

List of oligonucleotides used for the analysis of the mRNA profiles of wild-type and mutant plants by RT-qPCR experiments.

gene target	oligo name	sequence (5'-to-3')
<i>AOXIA</i>	<i>aox1aF</i>	AGCATCATGTTCCAACGACGTTTC
	<i>aox1aR</i>	GCTCGACATCCATATCTCCTCTGG
<i>AOXIB</i>	<i>aox1bF</i>	GGACCGTGAATCTCTCGATGGC
	<i>aox1bR</i>	TCTAGCATCATTGCTCTGCATCCG
<i>AOXIC</i>	<i>aox1cF</i>	TCTCCAGAGGAGGTATGGTTGCC
	<i>aox1cR</i>	AGTCATAAGCATCCCTCCAACC
<i>AOXID</i>	<i>aox1dF</i>	TTTGCTGAAGAGGCTGAGAACG
	<i>aox1dR</i>	CTCGTTCGTACCAATTGGGTTGTG
<i>AOX2</i>	<i>aox2F</i>	ACGGTGATTCTGTGCTGATGAAGC
	<i>aox2R</i>	TCCTTGATTGCGAATGTCAGAACG
<i>NDA1</i>	<i>nda1F</i>	GTATCCAACCGGCGATTTCACG
	<i>nda1R</i>	AGTTACAGTCTCACATGCACCTC
<i>NDA2</i>	<i>nda2F</i>	TGGTGTGGCTCTCTCCCTTCG
	<i>nda2R</i>	TCCATT CGTCAATGCCATCCTTC
<i>NDB1</i>	<i>ndb1F</i>	TAACACATTGGCACTCCTGGTG
	<i>ndb1R</i>	CTCTGTGCATCCTCTACTCCTTG
<i>atp1</i>	<i>atp1F</i>	TCACTTCGACACGTCTTGC
	<i>atp1R</i>	GGAATGGCCTTGAATCTTGA
<i>atp6</i>	<i>atp6-1F</i>	TCTTTGCGAGTCATGCAC
	<i>atp6-1R</i>	TCTCGCGTATCTCACATTGC
<i>atp8</i>	<i>atp8F</i>	CCGTCGACTTATTGGGAAAA
	<i>atp8R</i>	TTCCCTGGCCATGTACAACA
<i>atp9</i>	<i>atp9F</i>	CATCCCTCTGACGTGAAT
	<i>atp9R</i>	TCGTCGATTCTTACCCCTCGT
<i>atp4</i>	<i>atp4F</i>	GGATCAGCTTGCAGATTGT
	<i>atp4R</i>	GCAAATTGCTCCCCACTAA
<i>ccmb</i>	<i>ccmBF</i>	TCTTCCAATCACATCCAGCA
	<i>ccmBR</i>	CGAGACCGAAATTGGAAAAAA
<i>ccmc</i>	<i>ccmCF</i>	AGCTACGGCAAATTCTCAT
	<i>ccmCR</i>	GCCGTGGCGATATAAACAAAT
<i>ccmfc</i>	<i>ccmFcF</i>	CACATGGAGGAGTGTGCATC
	<i>ccmFcR</i>	GTGGGTCCATGTAAATGATCG
<i>ccmfn-1</i>	<i>ccmFN1F</i>	AGCTCTTGGCATTGCTTGT
	<i>ccmFN1R</i>	AGTGCCACAATCCCATTCTCAT
<i>ccmfn-2</i>	<i>ccmFN2F</i>	CGTGTGTTCGTAATGGAAA
	<i>ccmFN2R</i>	TGATAAGCCCACCAACTTCC
<i>cob</i>	<i>cobF</i>	TGCCGGAATGGTATTCTTA
	<i>cobR</i>	GCCAAAAGCAACCAAAACAT
<i>cox1</i>	<i>cox1F</i>	GTAGCTGGGTGAAGTAGGC
	<i>cox1R</i>	CTGCCTGGATTGGTATCAT
<i>cox2</i>	<i>cox2F</i>	TGATGCTGTACCTGGTCGTT

	<i>cox2R</i>	TGGGGGATTAATTGATTGGA
<i>cox3</i>	<i>cox3F</i>	CCGTAACCTGGGCTCATCAT
	<i>cox3R</i>	AAACCATGAAAGCCTGTTGC
<i>mttb</i>	<i>mttBF</i>	GGGGTCTTCTTGGAAACC
	<i>mttBR</i>	TCTCCCTCATTCCACTCGTC
<i>nad1 exons a-b</i>	<i>nad1 1-2F</i>	GACCAATAGATACTTCATAAGAGACCA
	<i>nad1 1-2R</i>	TTGCCATATCTCGCTAGGTG
<i>nad1 exons b-c</i>	<i>nad1 2-3F</i>	ATTCAGCTCCGCTCTGG
	<i>nad1 2-3R</i>	TCTGCAGCTCAAATGGTCTC
<i>nad1 exons c-d</i>	<i>nad1 3-4F</i>	AAAAGAGCAGACCCCATTGA
	<i>nad1 3-4R</i>	TCCGTTGATCTCCCAGAAG
<i>nad1 exons d-e</i>	<i>nad1 4-5F</i>	AGCCCAGGATCTTCTTGA
	<i>nad1 4-5R</i>	TCTTCAATGGGGTCTGCTC
<i>nad2 exons a-b</i>	<i>nad2 exons a-bF</i>	GCGAGCAGAAGCAAGGTTAT
	<i>nad2 exons a-bR</i>	GGATCCTCCCACACATGTTC
<i>nad2 exons b-c</i>	<i>nad2 exons b-cF</i>	AAAGGAAC TGCAAGTGATCTTGA
	<i>nad2 exons b-cR</i>	AATATTGATCTTAGGTGCATTITC
<i>nad2 exons c-d</i>	<i>nad2 exons c-dF</i>	GCGCAATAGAAAGGAATGCT
	<i>nad2 exons c-dR</i>	CTATGGGTCTACTGGAGCTACCC
<i>nad2 exons d-e</i>	<i>nad2 exons d-eF</i>	CAAAGGAGAGGGGTATAGCAA
	<i>nad2 exons d-eR</i>	TATTGTTCTCGCCGCTTT
<i>nad3</i>	<i>nad3F</i>	CGAATGTGGTTTCGATCCTT
	<i>nad3R</i>	GCACCCCTTTCCATTCTATA
<i>nad4 exons a-b</i>	<i>nad4 exons a-bF</i>	ATTCTATGTTTCCGAAAGC
	<i>nad4 exons a-bR</i>	GAAAAAACTGATATGCTGCCCTG
<i>nad4 exons b-c</i>	<i>nad4 exons b-cF</i>	AATAACCCATGTTCCCGAAG
	<i>nad4 exons b-cR</i>	TGCTACCTCCAATTCCCTGT
<i>nad4 exons c-d</i>	<i>nad4 exons c-dF</i>	TTCCTCCATAAATTCTCCGATT
	<i>nad4 exons c-dR</i>	TGAAATTGCCATGTTGCAC
<i>nad4L</i>	<i>nad4L-F</i>	GGGAATCCTCCTTAATAGACG
	<i>nad4L-R</i>	AACGAAAATGGCTAACCAATA
<i>nad5 exons a-b</i>	<i>nad5 exons a-bF</i>	TGGACCAAGCTACTTATGGATG
	<i>nad5 exons a-bR</i>	CCATGGATCTCATCGGAAAT
<i>nad5 exons b-c</i>	<i>nad5 exons b-cF</i>	TACCTAAACCAATCATCATATC
	<i>nad5 exons b-cR</i>	CTGGCTCTCGGGAGTCTCTT
<i>nad5 exons c-d</i>	<i>nad5 exons c-dF</i>	AACTCGGATT CGCAAGAA
	<i>nad5 exons c-dR</i>	GATATGATGATTGGTTAGGTA
<i>nad5 exons d-e</i>	<i>nad5 exons d-eF</i>	AACATTGCAAAGGCATAATGA
	<i>nad5 exons d-eR</i>	GTTCCTGCGTTTCGGATATG
<i>nad6</i>	<i>nad6F</i>	TATGCCGAAAGGTACGAAG
	<i>nad6R</i>	GTGAGTGGGTCA GTCGTCCT
<i>nad7 exons a-b</i>	<i>nad7 exons a-bF</i>	ACCTCAACATCCTGCTGCTC
	<i>nad7 exons a-bR</i>	AAGGTAAAGCTGAAGATAAGTTTGT
<i>nad7 exons b-c</i>	<i>nad7 exons b-cF</i>	GAGGGACTGAGAAATTAAATAGAGTACA
	<i>nad7 exons b-cR</i>	TGGTACCTCGCAATTCAAAA
<i>nad7 exons c-d</i>	<i>nad7 exons c-dF</i>	ACTGTCACTGCACAGCAAGC
	<i>nad7 exons c-dR</i>	CATTGCACAATGATCCGAAG

<i>nad7</i> exons d-e	<i>nad7</i> exons d-eF	GATCAAAGCCGATGATCGTAA
	<i>nad7</i> exons d-eR	AGGTGCTTCACACTGCAGGTAT
<i>nad9</i>	<i>nad9F</i>	GGATGACCCTCGAAACCATA
	<i>nad9R</i>	CACGCATTCTGTACAAACC
<i>rpl2</i>	<i>rpl2F</i>	CCGAAGACGGATCAAGGTAA
	<i>rpl2R</i>	CGCAATTCATCACCATTG
<i>rpl5</i>	<i>rpl5F</i>	AAGGGGTTCGACAGGAAAGT
	<i>rpl5R</i>	CGTATTCGACCGGAAAATC
<i>rpl16</i>	<i>rpl16F</i>	GAGCATTGCCAAACTCACA
	<i>rpl16R</i>	CGGACACTTCATCGTGCTA
<i>rps3</i>	<i>rps3F</i>	CCGATTTCGGTAAGACTTGG
	<i>rps3R</i>	AGCCGAAGGTGAGTCTCGTA
<i>rps4</i>	<i>rps4F</i>	ACCCATCACAGAGATGCACA
	<i>rps4R</i>	TCACACAAACCTTCGATGA
<i>rps7</i>	<i>rps7F</i>	CTCGAACTGAACCGCAGTGA
	<i>rps7R</i>	AAGCTGCTCAAGGATCCAA
<i>rps12</i>	<i>rps12F</i>	AGCCAAAGTACGGTTGAGCA
	<i>rps12R</i>	TTTGGGTTTCTGCACCAT
<i>matR</i>	<i>matR-F</i>	AATTTTGCAGAGCTGGAA
	<i>matR-R</i>	TTGAACCCGTCCTGTAGAC
<i>rrn18</i>	<i>rrn18F</i>	CGTCACCTGGTCAAAACT
	<i>rrn18R</i>	GCTTGAAAACCGAAGTGAGC
<i>rrn26</i>	<i>rrn26F</i>	GACGAGACTTCGCCTTTG
	<i>rrn26R</i>	CTTGGAGCGAATTGGATGAT
<i>rrn5</i>	<i>rrn5F</i>	CCGACCTCGATATGTGGAATCGTC
	<i>rrn5R</i>	TGGACCATGTCCTCCGAACAATC
<i>18S rRNA</i> (nuclear)	<i>18S nucl-F</i>	AAACGGCTACCACATCCAAG
	<i>18S nucl-R</i>	ACTCGAAAGAGCCCGGTATT
<i>ACTIN2</i> (At3g18780, nuclear)	<i>actin2-F</i>	GGTAACATTGTGCTCAGGGTGG
	<i>actin2-R</i>	AACGACCTTAATCTCATGCTGC
<i>GAPDH</i> (At3g04120)	<i>GAPDH-F</i>	TCTCGATCTCAATTTCGAAAA
	<i>GAPDH-R</i>	CGAAACCGTTGATTCCGATT

List of oligonucleotides used for the analysis of the splicing profiles of wild-type and mutant plants by RT-qPCR experiments.

Gene	Forward primer	Reverse primer
<i>rpl2</i>	CCGAAGACGGATCAAGGTAA	CGCAATTCATCACCATTG
<i>rpl2</i> intron exon2	TTAGGAAGAGCCGTACGAGG	CGCAATTCATCACCATTG
<i>rps3</i>	AGCCGAAGGTGAGTCTCGTA	CCGATTTCGGTAAGACTTGG
<i>rps3</i> intron1 exon2	AGCCGAAGGTGAGTCTCGTA	TCTACGGCGGGGTCACTAT
<i>cox2</i>	TGGGGGATTAATTGATTGGA	TGATGCTGTACCTGGTCGTT
<i>cox2</i> intron1 exon2	TGGGGGATTAATTGATTGGA	AGCAGTACGAGCTGAAAGGC
<i>ccmFc</i>	GTGGGTCCATGTAAATGATCG	CACATGGAGGAGTGTGCATC
<i>ccmFc</i> intron1 exon1	CCCGGATCGAACATCAGAGTT	CACATGGAGGAGTGTGCATC
<i>nad1</i> exon1-2	GACCAATAGATACTTCATAAGAGACCA	TTGCCATATCTCGCTAGGTG
<i>nad1</i> intron1 exon2	GACCAATAGATACTTCATAAGAGACCA	CGTGCTCGTACGGTTCATAG
<i>nad1</i> exon2-3	ATTCAAGCTTCCGCTTCTGG	TCTGCAGCTCAAATGGTCTC

<i>nad1</i> intron2 exon2	GGTTGGGTTAGGGGAACATC	TCTGCAGCTCAAATGGTCTC
<i>nad1</i> exon3-4	AAAAGAGCAGACCCCATTGA	TCCGTTGATCTCCCAGAAG
<i>nad1</i> intron3 exon4	AAAAGAGCAGACCCCATTGA	GGGAGCTGTATGAGCGGTAA
<i>nad1</i> exon4-5	AGCCCAGGATCTCTTGA	TCTTCATGGGTCTGCTC
<i>nad1</i> intron4 exon5	AGCCCAGGATCTCTTGA	ACGGAGCTGCATCCCTACT
<i>nad2</i> exon1-2	GCGAGCAGAACAGGTTAT	GGATCCTCCCACACATGTTTC
<i>nad2</i> intron1 exon2	GCGAGCAGAACAGGTTAT	CCCATTCTAACCAAGTGGAG
<i>nad2</i> exon2-3	AAAGGAACGTGAGTCTTGA	AATATTGATCTTAGGTGCATTTTC
<i>nad2</i> intron2 exon2	CCCGATCCGATAGTTACAA	AATATTGATCTTAGGTGCATTTTC
<i>nad2</i> exon3-4	GCGCAATAGAACAGGAATGCT	CTATGGGTCTACTGGAGCTACCC
<i>nad2</i> intron3 exon4	GCGCAATAGAACAGGAATGCT	GGCGAATTCAAACCTGTGG
<i>nad2</i> exon4-5	CAAAGGAGAGGGTATAGCAA	TATTTGTTCTCGCCGCTTT
<i>nad2</i> intron4exon4	CTTATTCTGGCAACCTTCC	TATTTGTTCTCGCCGCTTT
<i>nad4</i> exon1-2	ATTCTATGTTTCCCAGAAC	GAAAAAACTGATATGCTGCCCTG
<i>nad4</i> intron1 exon2	CCGTATGATCGGAAAGTCTC	GAAAAAACTGATATGCTGCCCTG
<i>nad4</i> exon2-3	AATACTCATGTTCCCAGAAC	TGCTACCTCCAATTCCCTGT
<i>nad4</i> intron2 exon3	GCGGAACGACCAGAAAAATA	TGCTACCTCCAATTCCCTGT
<i>nad4</i> exon3-4	TTCCCTCATAAATTCTCCGATT	TGAAATTGCCATGTTGCAC
<i>nad4</i> intron3 exon4	TCTAGCTGGTTGGAGAGC	TGAAATTGCCATGTTGCAC
<i>nad5</i> exon1-2	TGGACCAAGCTACTTATGGATG	CCATGGATCTCATCGGAAAT
<i>nad5</i> intron1 exon2	TGGACCAAGCTACTTATGGATG	TTCGCAAATAGGTCCGACT
<i>nad5</i> exon2-3	TACCTAACCAATCATCATATC	CTGGCTCTCGGGAGTCTCTT
<i>nad5</i> intron2-exon2	GTACGATCGTGTGGGTGA	CTGGCTCTCGGGAGTCTCTT
<i>nad5</i> exon3-4	AACTCGGATTGGCAAGAA	GATATGATGATTGGTTAGGTA
<i>nad5</i> intron3-exon4	AACTCGGATTGGCAAGAA	GCCGTGTAATAGGCACCA
<i>nad5</i> exon4-5	AACATTGCAAAGGCATAATGA	GTTCCCTCGTTCCGGATATG
<i>nad5</i> intron4 exon5	AACATTGCAAAGGCATAATGA	CCTGTAAACCCCCATGATGT
<i>nad7</i> exon1-2	ACCTAACATCCTGCTGCTC	AAGGTAAAGCTTGAAGATAAGTTTGT
<i>nad7</i> intron1 exon2	ACGGTTTTAGGGGATCTG	AAGGTAAAGCTTGAAGATAAGTTTGT
<i>nad7</i> exon2-3	GAGGGACTGAGAAATTATAGAGTACA	TGGTACCTCGCAATTCAAAA
<i>nad7</i> intron2 exon3	AGTGGGAGAGCCGTGTTATG	TGGTACCTCGCAATTCAAAA
<i>nad7</i> exon3-4	ACTGTCACTGCACAGCAAGC	CATTGCAACATGATCCGAAG
<i>nad7</i> intron3 exon4	TAAAGTGAAGTGGTGGCCT	CATTGCAACATGATCCGAAG
<i>nad7</i> exon4-5	GATCAAAGCCGATGATCGTAA	AGGTGCTTCAACTGCGGTAT
<i>nad7</i> intron4 exon5	CGGCCAAATGACTACAGGAT	AGGTGCTTCAACTGCGGTAT

List of other oligonucleotides used for the analysis of *MISF2* and *RDMI* genes, the construction of *MISF2*-GFP and RACE analyses.

Gene	Forward primer	Reverse primer
<i>MISF2</i> (<i>AT3G22670</i>)	ATGCTCACTAAGCTTAGGATTA	ATTATAAGTATGACCTGAATGCA
<i>MISF2.1</i> (<i>SALK_067654</i>)	ATTTTGCCGATTCGGAAC	ATTATAAGTATGACCTGAATGCA
<i>MISF2.2</i> (<i>SALK_066141</i>)	ATTTTGCCGATTCGGAAC	ATTATAAGTATGACCTGAATGCA
<i>MISF2-GFP</i> (<i>misf2-B1+</i> <i>misf2-B2</i>)	GGGGACAAGTTGTACAAAAAAAGC AGGCTAAATGCTACTAAGCTTAG GATTAGT	GGGGACCACTTGTACAAGAAAG CTGGGTCAAGCGTAACAAGCTTA GATTCTTC
<i>MISF2-RACE_SI</i>	AACAATCGCGACTCCTCTG	AGGATGGAGGATGAGGAGGG
<i>MISF2-promo-B1</i> (<i>F</i>) <i>MISF2-Cpl-B2</i> (<i>R</i>)	GGGGACAAGTTGTACAAAAAAAGCAGGCT AAAGCTTCAATGCTTGAACAAGGTTTC	GGGGACCACTTGTACAAGAAAG CTGGGTCCGAAACAGATAAAGGA CTGTGAG
<i>MISF2-B1</i> (<i>F</i>) <i>MISF2-Cpl-B2</i> (<i>R</i>)	GGGGACAAGTTGTACAAAAAAAGCAGGCT AAAATGCTCACTAAGCTTAGGATTAGT	GGGGACCACTTGTACAAGAAAG CTGGGTCCGAAACAGATAAAGGA CTGTGAG
<i>RDMI-GSP</i>	GATTACGCCAAGCTTCAGCGATGGC TTTCACCGCTCGAT	GATTACGCCAAGCTTCAACGAG CCACGATTGTCGGGAT
<i>RDMI-NGSP</i>	GATTACGCCAAGCTTCTTCACAG AGACGTGCTGATGAAG	GATTACGCCAAGCTTGCTTAATCC AACCCAGTTGTGAACCGGA
<i>RDMI-RT-PCR1</i>	GTCAAAATCTAACTGCTCGACAG	CTCTGCTCTCTCAGCAACGAAC
<i>RDMI-RT-PCR2</i>	GTCATCGTCTCGTACCTTCTGC	CTCTGCTCTCTCAGCAACGAAC
<i>RDMI-RT-PCR3</i>	AAGGAACCTCGTGTGGTCATC	CTCTGCTCTCTCAGCAACGAAC
<i>RDMI-RT-PCR4</i>	CCACTGGTTCTTCACTACATTCCC	CTCTGCTCTCTCAGCAACGAAC

Table S2. List of antibodies used in this study.

Antibody	Protein I.D.	origin	serum	dilution	Reference / source
AtpA	Mitochondrial ATP-synthase subunit α	<i>Zea mays</i>	Mouse (monoclonal)	1/500	Thomas Elthon collection, PM014
AtpB	Mitochondrial ATP-synthase subunit β	<i>Zea mays</i>	Mouse (monoclonal)	1/5,000	(Michael et al. 1993)
CA2	γ -carbonic anhydrase-like subunit 2	<i>Arabidopsis thaliana</i>	Rabbit (polyclonal)	1/1,000	(Perales et al. 2005, Sunderhaus et al. 2006)
Cox2	Cytochrome oxidase subunit-2	<i>Arabidopsis thaliana</i>	Rabbit (polyclonal)	1/5,000	Agrisera antibodies, AS04 053A
Nad1	NADH-dehydrogenase complex subunit-1	<i>Arabidopsis thaliana</i>	Rabbit (polyclonal)	1/1000	Gift of Dr. Etienne Meyer, Halle U.
Nad9	NADH-dehydrogenase complex subunit-9	<i>Triticum spp.</i>	Rabbit (polyclonal)	1/50,000	(Lamattina et al. 1993)
RISP	Rieske iron-sulfur protein	<i>Arabidopsis thaliana</i>	Rabbit (polyclonal)	1/5,000	Gift of Prof. Ian Small, UWA
VDAC (PORIN)	Mitochondrial membrane-associated β -barrel proteins	<i>Zea mays</i>	Mouse (monoclonal)	1/1000	Thomas Elthon collection, PM035
AOX1/2	Alternative oxidase	<i>Sauromatum guttatum</i>	Rabbit (polyclonal)	1/500	Agrisera antibodies, AS04 054
Nad6	NADH-ubiquinone oxidoreductase chain 6	<i>Arabidopsis thaliana</i>	Rabbit (polyclonal)	1:1000	Agrisera (AS15 2926)
Cob	Apocytochrome B	<i>Beta vulgaris</i> ssp. <i>maritima</i>	Rabbit (polyclonal)	1:4000	Meyer et al. 2018
CYTC1	Cytochrome C1	<i>Arabidopsis thaliana</i>	Rabbit (polyclonal)	1:1000	Giégé et al. 2005
CYTC	Cytochrome C	<i>Arabidopsis thaliana</i>	Rabbit (polyclonal)	1:1000	Agrisera (AS08 343A)

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