

Biomimetic Nanosponges Enable the Detoxification of *Vibrio vulnificus* Hemolysin

Shuaijun Zou,^{1,†} Qianqian Wang,^{1,†} Peipei Zhang,^{2,‡} Bo Wang,¹ Guoyan Liu,¹ Fuhai Zhang,¹ Jie Li,¹ Fan Wang,¹ Beilei Wang,^{1,*} Liming Zhang^{1,*}

¹Department of Marine Biomedicine and Polar Medicine, Naval Special Medical Center, Naval Medical University, Shanghai 200433, China

²Department of Marine Biological Injury and Dermatology, Naval Special Medical Center, Naval Medical University, Shanghai 200052, China

[‡] These authors contributed equally to this work

* Corresponding Authors: lilly_wang@126.com (B.W.); lmzhang@smmu.edu.cn (L.Z.).

1. Materials and Methods

Expression and purification of recombinant VvhA (rVvhA)

The oligonucleotides were designed based on the genomic sequence of the *V. vulnificus* strain FJ03-X2 (GenBank KC821520) with the signal peptide sequence removed. The sequence was amplified by PCR (forward primer: 5'-CATATGCAAGAATATGTGCCGATTGTTGAG-3' and reverse primer: 5'-TCTAGACTAGAGTTTGACTTGTGTAATGT-3') and then cloned into the His₆ tag expression vector pCZN1. After verification of the target sequence in the recombinant plasmids, the recombinant vectors were expressed in *Arctic-Express* (DE3) cells (Agilent Technologies). The bacteria were grown in LB-ampicillin medium at 37 °C until the cultures reached an OD₆₀₀ between 0.6 and 0.8. Then, the bacteria were induced with 0.5 mM isopropyl-β-dithiogalactopyranoside (IPTG) for 4 h. Next, the cultures were harvested, lysed and examined by sodium dodecyl sulfate-polyacrylamide gel electrophoresis (SDS-PAGE). A Ni-IDA-Sepharose Cl-6B affinity chromatography column (Sigma-Aldrich) was used for protein purification, and the protein was verified by Western blotting. The residual endotoxin in the purified rVvhA was measured using ToxinSensor™ Endotoxin Test Kit (GenScript, Nanjing, China). Finally, the activity of rVvhA was determined according to the hemolysis protocol in a 2% murine RBC suspension [1].

2. Results

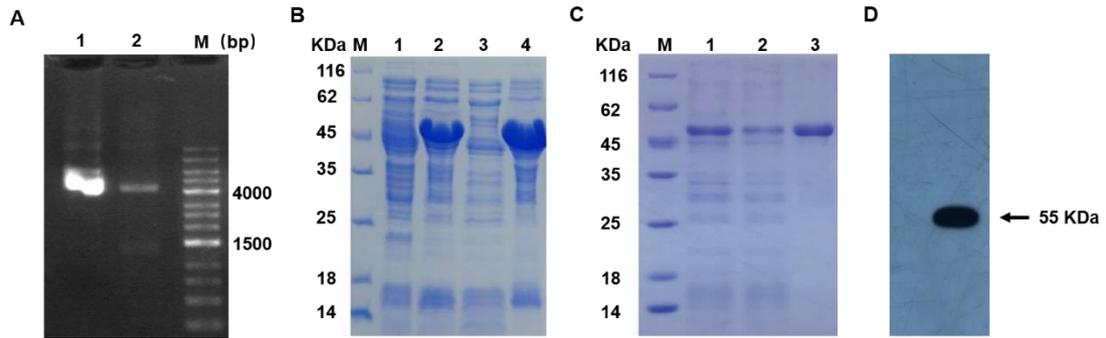


Figure S1. Expression and purification of rVvhA. (A) Determination of recombinant pCZN1 plasmids before and after enzyme digestion at *NdeI-XbaI* sites. (Line 1: plasmid before enzyme digestion, Line 2: plasmid after enzyme digestion, M: marker) (B) Determination of overexpression of rVvhA (M: marker, Line 1: sample from non-induced bacteria, Line 2: sample from IPTG-induced bacteria, Line 3: supernatant of lysed IPTG-induced bacteria, Line 4: sediment of lysed IPTG-induced bacteria). (C) SDS-PAGE analysis of rVvhA purification (M: marker, Line 1: sample harvested from lysed bacteria, Line 2: sample of outflow peak, Line 3: sample of elution peak). (D) Western-blot verification of purified rVvhA.

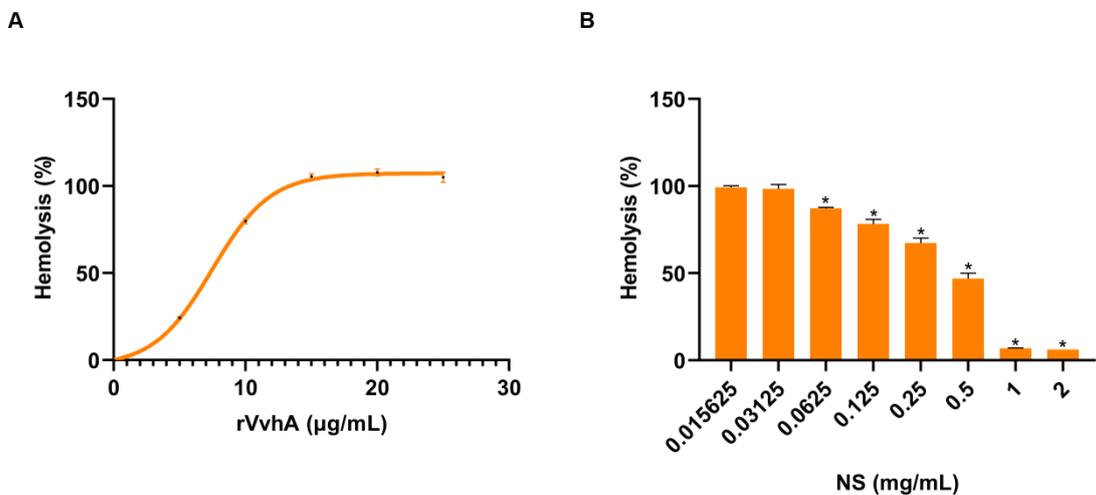


Figure S2. Hemolytic activity of rVvhA and inhibition effect of NSs. (A) Hemolytic activity of rVvhA on 2% RBC suspension *in vitro*. **(B)** Inhibition of NSs on rVvhA-caused hemolysis *in vitro*. (The data represent the means \pm SE. n = 3. * p < 0.05).

References

1. Zohra M.; Fawzia A. Hemolytic activity of different herbal extracts used in Algeria. *Int. J. Pharm. Sci. Res.* **2014**, *5*, 495-500.