

## CAF Proteins Help SOT1 Regulate the Stability of Chloroplast *ndhA* Transcripts

Xiuming Li <sup>1</sup>, Wenzhen Luo <sup>2</sup>, Wen Zhou <sup>3</sup>, Xiaopeng Yin <sup>2</sup>, Xuemei Wang <sup>4</sup>, Xiujin Li <sup>2</sup>, Chenchen Jiang <sup>2</sup>, Qingqing Zhang <sup>2</sup>, Xiaojing Kang <sup>2</sup>, Aihong Zhang <sup>2,\*</sup>, Yi Zhang <sup>2,\*</sup> and Congming Lu <sup>2</sup>

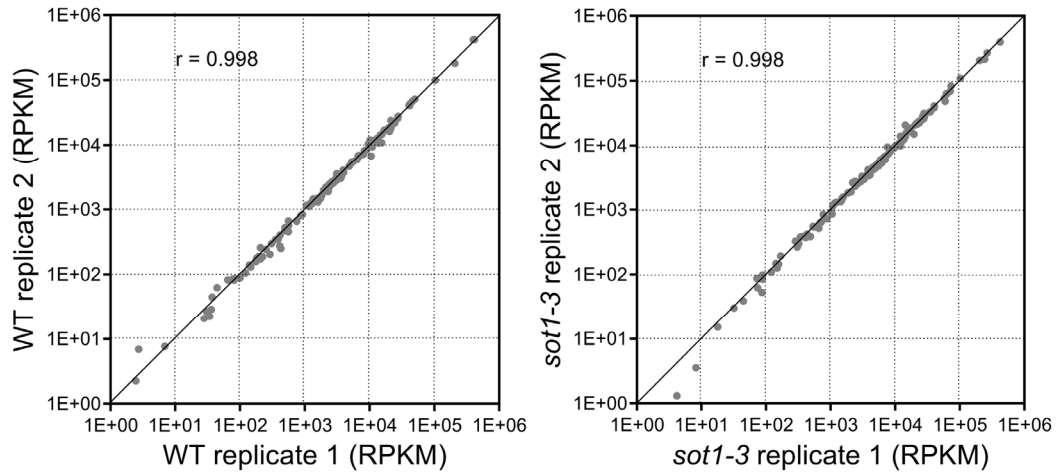
<sup>1</sup> State Key Laboratory of Crop Biology, College of Horticulture Science and Engineering, Shandong Agricultural University, Taian 271018, China; lixiuming@sdaa.edu.cn (X.L.)

<sup>2</sup> State Key Laboratory of Crop Biology, College of Life Sciences, Shandong Agricultural University, Taian 271018, China; wzluo1998@163.com (W.L.); xiaopengy@126.com (X.Y.); xjli1998@126.com (X.L.); cc\_jiang2019@163.com (C.J.); qingqingz2021@163.com (Q.Z.); kxj1201@163.com (X.K.); ahzhang@sdaa.edu.cn (A.Z.); zhangyi@sdaa.edu.cn (Y.Z.); cmlu@sdaa.edu.cn (C.L.)

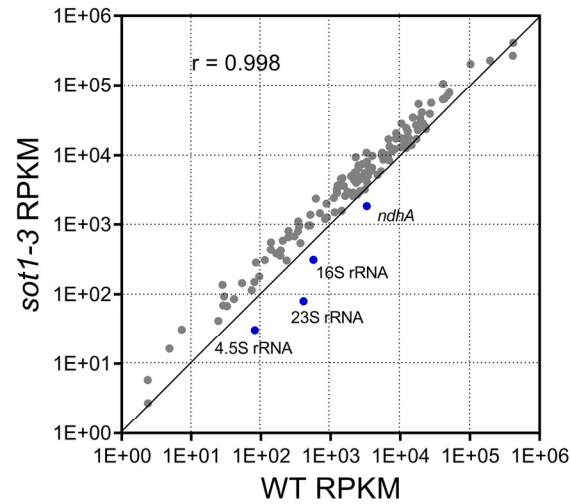
<sup>3</sup> School of Life Science, Department of Biology, Southern University of Science and Technology, Shenzhen 518055, China; zhouw@sustech.edu.cn

<sup>4</sup> Shandong Provincial Key Laboratory of Plant Stress, College of Life Sciences, Shandong Normal University, Jinan 250014, China, wangxuemei2020@sdsu.edu.cn

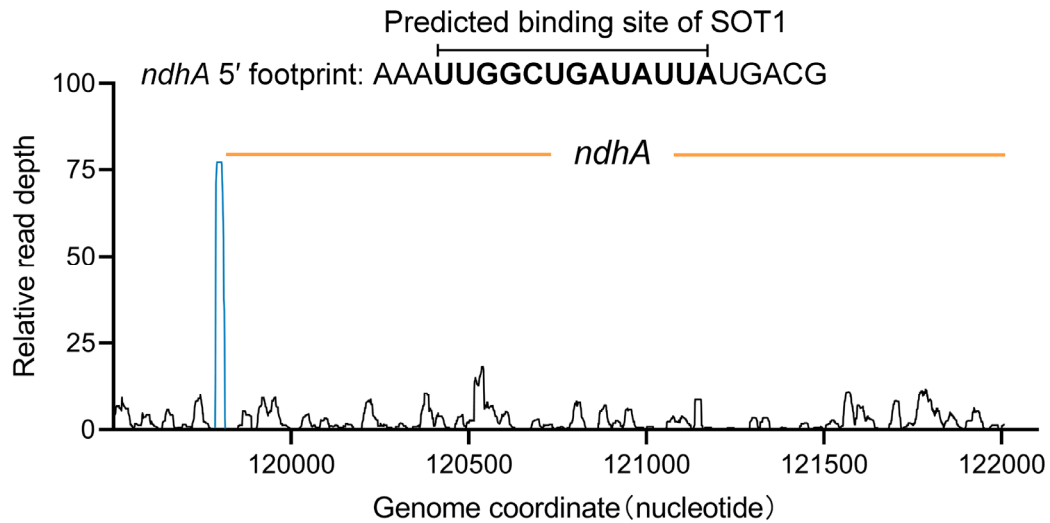
\* Correspondence: ahzhang@sdaa.edu.cn (A.Z.); zhangyi@sdaa.edu.cn (Y.Z.)



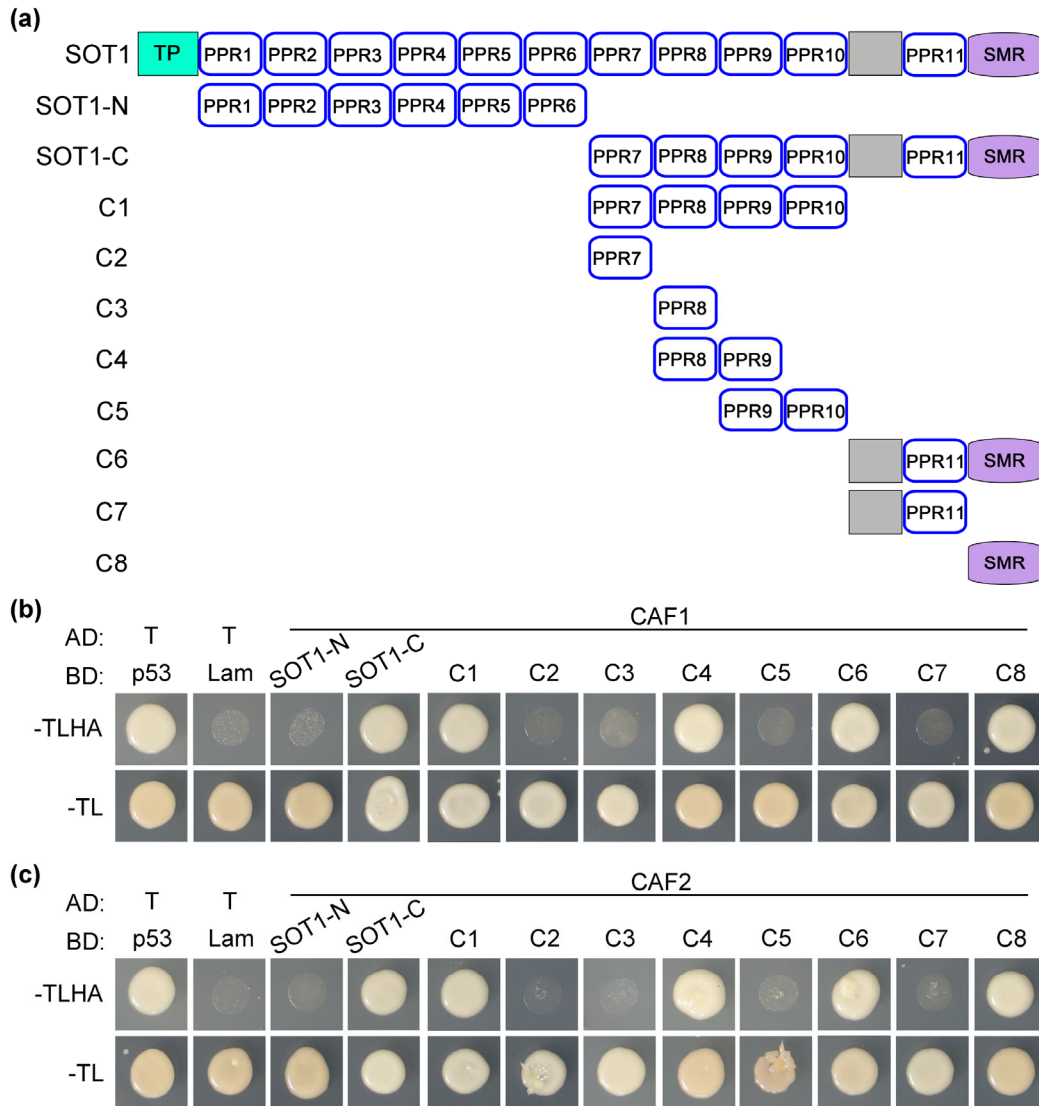
**Supplementary Figure S1.** Reproducibility of gene expression levels between the wild type (WT) and *sot1-3* replicates. Total RNA was isolated from 12-day-old WT and *sot1-3* seedlings and used for strand-specific RNA sequencing. The differential expression of each replicate was compared using the average number of reads per kilobase of transcript per million mapped reads (RPKM) values of total mapped reads from the chloroplast ( $n = 2$ ). Genes in the large inverted repeat of the chloroplast genome were included only once.



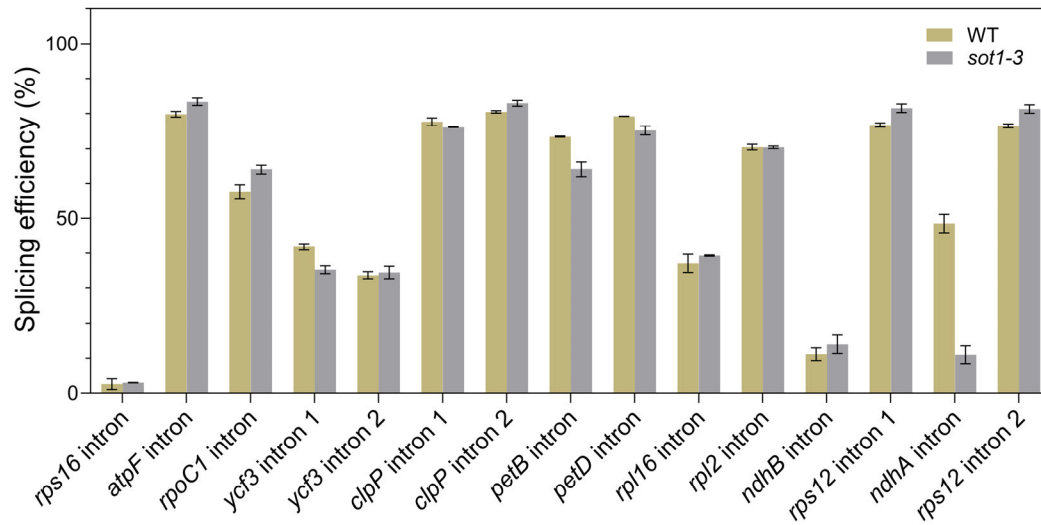
**Supplementary Figure S2.** Differential expression of chloroplast genes between the WT and *sot1-3* seedlings. Total RNA was isolated from 12-day-old WT and *sot1-3* seedlings and used for strand-specific RNA sequencing. The differential expression of chloroplast genes was compared between the WT and *sot1-3* using the average number of reads per kilobase of transcript per million mapped reads (RPKM) values ( $n = 2$ ). Genes in the large inverted repeat of the chloroplast genome were included only once. The genes with a decreased expression in the *sot1-3* seedlings relative to the WT are highlighted in blue.



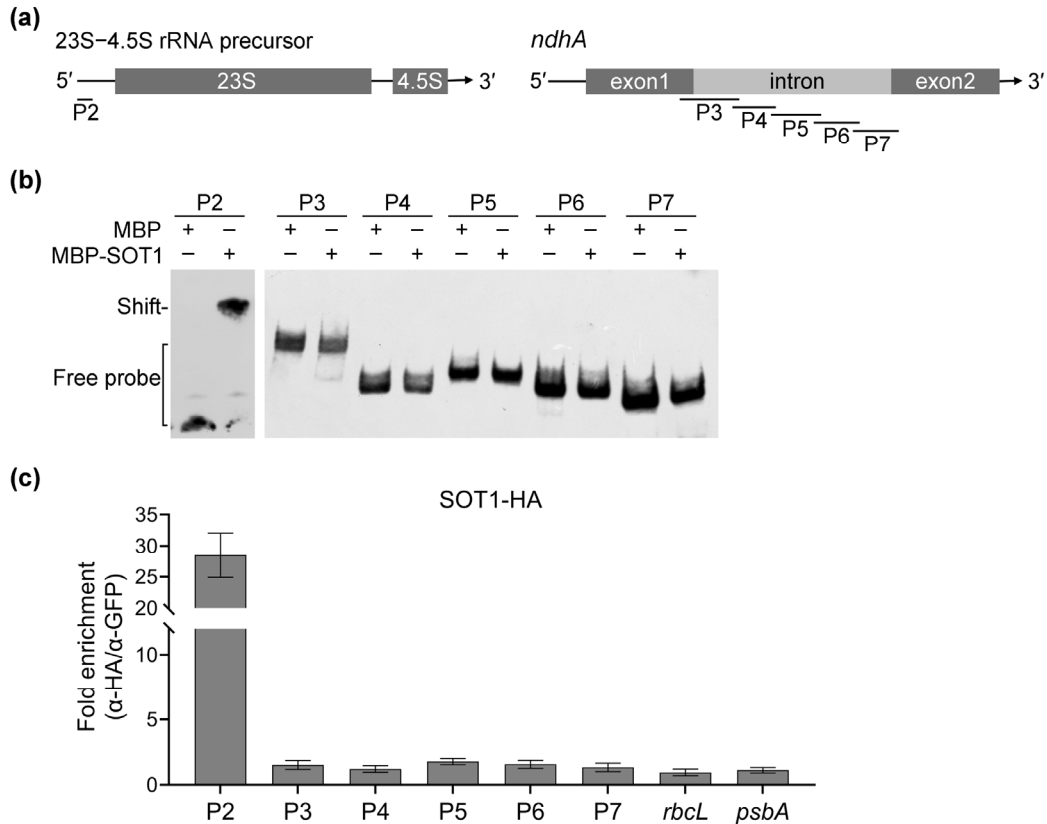
**Supplementary Figure S3.** Read coverage over *ndhA* regions for wild-type plants. Reads derived from small RNA sequencing were mapped to the Arabidopsis chloroplast genome. The graph indicates the read depth at each nucleotide of the *ndhA* region, normalized against the total number of reads for each sample. The RNA 'footprint' that matches the 5'-end upstream sequence of *ndhA* is highlighted in blue.



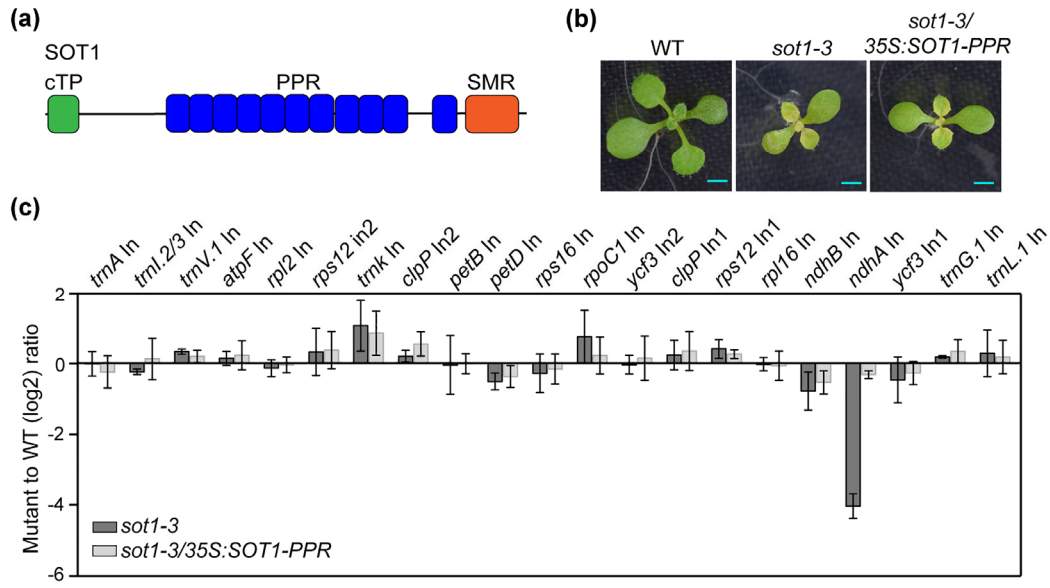
**Supplementary Figure S4.** Identification of the SOT1 domains that interact with CAF proteins. **(a)** Diagram the domain structures of SOT1 and various SOT1 deletions (SOT1-N, SOT1-C, and C1–C8). The turquoise and blue boxes indicate the chloroplast transit peptide (TP) and the single PPR motif (PPR1–11), respectively. The SMR domain is highlighted in purple. **(b)** and **(c)** Identification of the SOT1 domains that interact with **(b)** CAF1 and **(c)** CAF2 using a yeast two-hybrid assay. CAF1 and CAF2 were fused to the prey construct (AD); SOT1 and various SOT1 deletions were fused to the bait construct (BD). The ability to grow on -TLHA dropout plates indicates an interaction between the two proteins. The assay of interaction between SV40 large T-antigen (T) and murine p53 was used as a positive control; the interaction between SV40 large T-antigen (T) and lamin (Lam) was used as a negative control. -TL and -TLHA indicate SD/-Trp-Leu and SD/-Trp-Leu-His-Ade dropout plates, respectively.



**Supplementary Figure S5.** RNA-sequencing (RNA-seq) determination of the RNA splicing efficiencies of chloroplast introns in 12-day-old WT and *sot1-3* seedlings. The splicing efficiency was determined according to Hotto et al. (2015). The splicing efficiency presented is the average of two replicates for both WT and *sot1-3* samples. Since the mature tRNA was discarded in the RNA-seq library preparation, the splicing efficiencies of tRNA introns was omitted. The values represent means  $\pm$  SD ( $n = 2$ ).



**Supplementary Figure S6.** SOT1 does not bind the intron of *ndhA* transcripts. **(a)** Schematic representation of the domain structures of the *ndhA* mRNA and the 23S-4.5S rRNA precursor. The positions of probes P2 to P7, subjected to RNA coimmunoprecipitation assays and electrophoretic mobility shift assay (EMSA), are shown below the models. **(b)** EMSA showing that SOT1 alone exhibits little binding activity to the *ndhA* intron. A total of 150 nM recombinant MBP and MBP-SOT1 proteins were incubated with 10 nM biotin-labeled probes. Three independent experiments were performed, and one representative experiment is shown. **(c)** RNA coimmunoprecipitation assays showing that SOT1 does not bind the *ndhA* intron in vivo. Intact chloroplasts were isolated from 12-day-old complemented *sot1-3/35S:SOT1-HA* plants (*sot1-3/35S:SOT1-HA*). The chloroplast extracts were subjected to immunoprecipitation against HA and GFP antibodies. SOT1 was reported to bind the 5' ends of the 23S-4.5S rRNA precursor (Wu et al., 2016; Zhou et al., 2017); therefore, this interaction served as a positive control. The sample immunoprecipitated using the GFP antibody served as a negative control. The relative RNA enrichment levels were determined using qPCR. Mean values ± SD of the triplicate replicates are shown.



**Supplementary Figure S7.** The defective *ndhA* splicing in the *sot1-3* mutant was recovered in 12-day-old PPR-domain-complemented plants. **(a)** Schematic diagram of the SOT1 protein. The green and blue boxes represent the chloroplast transit peptide (TP) and each PPR motif, respectively. The SMR domain is highlighted in orange. **(b)** Phenotypes of WT, *sot1-3*, and its PPR-domain-complemented (*sot1-3/35S:SOT1-PPR*) plants grown for 12 days. **(c)** qPCR analysis of the splicing efficiency of chloroplast introns in the *sot1-3* and *sot1-3/35S:SOT1-PPR* complemented plants. The values represent the differences in the ratios of spliced to unspliced transcripts in 12-day-old *sot1-3* and *sot1-3/35S:SOT1-PPR* seedlings compared with those of the WT. Four biological replicates were analyzed. Error bars indicate SD.



**Supplementary Table S1.** RNA-seq alignment summary for two WT and *sot1-3* samples.

Sample	Mappable reads <sup>a</sup>	Mapped reads <sup>b</sup>	Mapped/Mappable (%)
WT rep1	47,045,476	22,527,949	47.89
WT rep2	52,133,521	23,977,993	45.99
<i>sot1-3</i> rep1	51,145,460	26,044,674	50.92
<i>sot1-3</i> rep2	47,618,661	22,350,101	46.94

<sup>a</sup>Mappable reads were selected after quality control, and therefore had a minimum length of 60 and a quality score higher than 30.

<sup>b</sup>Mapped reads represent the mappable reads that could be aligned to the Arabidopsis chloroplast genome (TAIR10) using TopHat2.

**Supplementary Table S2.** List of primers and oligonucleotides used in this study.

Purpose	Primer name	Primer sequence (5'-3')
Quantitative PCR	<i>trnH</i> -forward	GCGGATGTAGCCAAGTGGAT
	<i>trnH</i> -reverse	GGCGAACGACGGAATTGAA
	<i>psbA</i> -forward	GGTCGCTTCTGTAAGTGGAT
	<i>psbA</i> -reverse	GTTGCGGTCAATAAGGTAGG
	<i>ndhK</i> -forward	GCCTATGGCCGCTTCTTTAT
	<i>ndhK</i> -reverse	GTCTAGGACTCGATCTTGGTACT
	<i>rps16</i> -forward	ATTGATGTTTCGATCCCGAAGAG
	<i>rps16</i> -reverse	TCTTGTTGGTTGAGCTCCTTT
	<i>trnQ</i> -forward	TGGGGCGTAGCCAAGCGGTA
	<i>trnQ</i> -reverse	CTGGGACGGAAGGATTCGAA
	<i>trnG</i> -forward	TGGCGGAAATAGCTTAATGG
	<i>trnG</i> -reverse	AGCGGAAGGAGGGACTTGAA
	<i>trnR</i> -forward	GCGTCCATTGTCTAATGGAT
	<i>trnR</i> -reverse	TGCGTCCAATAGGATTTGAA
	<i>trnS</i> -forward	GGAGAGATGGCTGAGTGGAC
	<i>trnS</i> -reverse	GGGAAAGAGAGGGATTTCGAA
	<i>atpA</i> -forward	TATTATTTTCGAGACGTTCTG
	<i>atpA</i> -reverse	ATTGTATCTGTGGCTACTGC
	<i>atpF</i> -forward	CGATTCTTTCGTTTACTTGG
	<i>atpF</i> -reverse	TCATTTAACACTCCCTTTCC
	<i>atpH</i> -forward	CACTGGTTTCTGCTGCTTCG
	<i>atpH</i> -reverse	TTCTGCCTCAGGTTGTCTCG
	<i>atpI</i> -forward	CCTTTATTGGAACCTGTTT
	<i>atpI</i> -reverse	ATCATTTCGTTGGTGTCTGCTA
	<i>rps2</i> -forward	GGGCTCGGTGTCATTATGTT
	<i>rps2</i> -reverse	ACGGTTGAATCCCTCTGTCT
	<i>rpoB</i> -forward	TTGATGTGAGGTGGGTTTCAG
	<i>rpoB</i> -reverse	GCATATCCTGTCTAGGCAAA
	<i>rpoC1</i> -forward	GAGTTGAGACCCATCATTCA
	<i>rpoC1</i> -reverse	GTATCCACGGCTTCTTGAC
	<i>rpoC2</i> -forward	CCACTCATGGTGACCTCGTT
	<i>rpoC2</i> -reverse	TTCGGCAGTACCTCCTGTAA
	<i>trnC</i> -forward	GGCGGCATGGCCGAGTGGTA
	<i>trnC</i> -reverse	AGGCGGCACCCGGATTGAA
	<i>ycf6</i> -forward	ATGGATATAGTAAGTCTCGCAT
	<i>ycf6</i> -reverse	CTAGAGTCCACTTCTTCCC
	<i>trnY</i> -forward	GGGTCGATGCCCGAGCGGTT
	<i>trnY</i> -reverse	TGGGCCGAGCTGGATTGAAAC
	<i>trnE</i> -forward	GCCCCATCGTCTAGTGGTT
	<i>trnE</i> -reverse	TACCCCCAGGGGAAGTCGAA
	<i>trnT</i> -forward	GCCCTTTTAACTCAGTGGTA
	<i>trnT</i> -reverse	AGCCCCTTATCGGATTTGAA
	<i>psbC</i> -forward	ACTTCTGGGACCCGAAACTC
	<i>psbC</i> -reverse	AAAGGCACCTACACCTAACA
	<i>psbD</i> -forward	CAAGGGTTTCATAATTGGAC
	<i>psbD</i> -reverse	ATGTATTTGCACCATCACCA
	<i>ycf9</i> -forward	CTTACTGATTAGTGTACCCGTTGT

	<i>ycf9</i> -reverse	AGGATACCCACCAAGAAGACTA
	<i>rps14</i> -forward	CGCGTAATAGTGCACCTACA
	<i>rps14</i> -reverse	CGAAGGATGTGTCCAGATAGTC
	<i>psaA</i> -forward	TGAGTTAGTAGCAGTGGGTG
	<i>psaA</i> -reverse	CAACAGTATCAATACCGTCA
	<i>psaB</i> -forward	ATTCGCTGGAAAGATAAACC
	<i>psaB</i> -reverse	ATCAAGAAAGCCGCATAAGT
	<i>ycf3</i> -forward	GATCTGTCATTACCGTGGAG
	<i>ycf3</i> -reverse	TAAGCGTTATAGCCTGTTTC
	<i>rps4</i> -forward	GCGTTTGGATAACATCCTTT
	<i>rps4</i> -reverse	GTTTGCAGCGATAACTTGGT
	<i>trnL</i> -forward	GGGGATATGGCGGAATTGGT
	<i>trnL</i> -reverse	TGGGGATAGAGGGACTTGAAC
	<i>ndhJ</i> -forward	TTACAAATAAAGCCCGAAGA
	<i>ndhJ</i> -reverse	TGATACACGCTGGCTAAGAG
	<i>ndhC</i> -forward	GTATCCGTGGGCAATGAGTT
	<i>ndhC</i> -reverse	ACCATTCCAATGCTCCTTTT
	<i>trnV</i> -forward	AGGGCTATAGCTCAGTTAGG
	<i>trnV</i> -reverse	TAGGGCTATACGGACTCGAA
	<i>trnM</i> -forward	ACCTACTTAAGTCAGTGGTT
	<i>trnM</i> -reverse	TACCTACTATTGGATTGAA
	<i>atpE</i> -forward	ATTGTTTGGGATTGAGAAGT
	<i>atpE</i> -reverse	CCATTGGTTAGCAAGGCGTA
	<i>atpB</i> -forward	TTGGTCTAGCGGAAACAATT
	<i>atpB</i> -reverse	CCTTCGCAGTAGCTTCATCG
	<i>rbcL</i> -forward	AAACTTGAAGGAGACAGGGAG
	<i>rbcL</i> -reverse	CTGAAGCCACAGGCAGAACA
	<i>accD</i> -forward	ATTATTGCCGAACCCTATGC
	<i>accD</i> -reverse	TAAAGATTGAGCCGCTTGTG
	<i>ycf4</i> -forward	ATATGGAAATTCGAGGTCAA
	<i>ycf4</i> -reverse	GGTACACGCAAGAAGTAAGC
	<i>ycf10</i> -forward	TGGATTACTAATTGGTGGAA
	<i>ycf10</i> -reverse	GTAAATTGGTTTCTGGGTAT
	<i>petA</i> -forward	TCGCTCCATATCTGTCTCAC
	<i>petA</i> -reverse	ACATACAATACGCCCAGTCG
	<i>trnW</i> -forward	ACGCTCTTAGTTCAGTTCGG
	<i>trnW</i> -reverse	CACGCTCTGTAGGATTGAA
	<i>trnP</i> -forward	AGGGATGTAGCGCAGCTTGG
	<i>trnP</i> -reverse	TAGGGATGACAGGATTGAA
	<i>rpl33</i> -forward	GTTTCGAGTAACAATTATTTTGG
	<i>rpl33</i> -reverse	TTATGCCGATTCTTTTGAGT
	<i>rps18</i> -forward	AATCCAAGCGATCTTTTCGT
	<i>rps18</i> -reverse	GTCACTCTATTACCCGTCT
	<i>rpl20</i> -forward	AGCTCGGAGGCGTAGAACAA
	<i>rpl20</i> -reverse	CCCGATGAGCCGAAACTAAA
	<i>rps12</i> -forward	ACGATTAACCTCGGGATTG
	<i>rps12</i> -reverse	GGTTCCTCGAACAATGTGAT
	<i>clpP</i> -forward	CAAAGAACGGGCAAACCTAT
	<i>clpP</i> -reverse	GAACCGCTACAAGATCAACA

	<i>psbB</i> -forward	GTTGTGCTGGAACATATGTG
	<i>psbB</i> -reverse	CTTGTTGAAAGTATCCCTGA
	<i>petB</i> -forward	ATATGTTCCCTCCGCATGTCA
	<i>petB</i> -reverse	ACGGTTGGACGGTAATAAAA
	<i>petD</i> -forward	GCTAAAGGTATGGGTCACAA
	<i>petD</i> -reverse	AAGCCTACGTTACAGGCAAT
	<i>rpoA</i> -forward	ACTCGGACACTACAGTGGA
	<i>rpoA</i> -reverse	AAGTAAAGCTCTTCGCATCG
	<i>rps11</i> -forward	GGTCGGGTGATTTCTTGGTC
	<i>rps11</i> -reverse	CAGCTCGTTGCATACCTTGATC
	<i>rpl36</i> -forward	AAATAAGGGCTTCCGTTTCGT
	<i>rpl36</i> -reverse	TTTGTTTATGCCTCGGGTTG
	<i>rps8</i> -forward	CGACCGGTCTACGAATCTA
	<i>rps8</i> -reverse	GCTTCTCGGTCTGTCATTATACC
	<i>rpl14</i> -forward	TAGTAATCGCCGATATGCTC
	<i>rpl14</i> -reverse	GTCCCATTTGTTACGTTTGAG
	<i>rpl16</i> -forward	CGCAATGACACGAAATGTAC
	<i>rpl16</i> -reverse	TCCTTTCCCAGAACCCATAC
	<i>rpl22</i> -forward	TATGCCTTATCGAGGATGTT
	<i>rpl22</i> -reverse	AGTATTCCTTGGTTCACCT
	<i>rps19</i> -forward	AGAAATCATAATAACTTGGTCC
	<i>rps19</i> -reverse	TATAAACGGGTAAGTGTTC
	<i>rpl2</i> -forward	GCTGTAGCGAAACTGATTGC
	<i>rpl2</i> -reverse	ACTTGTCCGACTGTTGCTGA
	<i>rpl23</i> -forward	GGGTCGAACTCTTCTTTGGT
	<i>rpl23</i> -reverse	AACCCGGTTGAAGCGTAATG
	<i>trnI</i> -forward	GGGCTATTAGCTCAGTGGA
	<i>trnI</i> -reverse	TGGGCCATCCTGGATTTGAA
	<i>ycf2</i> -forward	ACCGATTCCCTAAATACCTT
	<i>ycf2</i> -reverse	GAAATTCTTCGCTTTCTTC
	<i>orf77</i> -forward	ATGCTACTACTGAAACATGGAAGA
	<i>orf77</i> -reverse	GTAGTGAGTAATAGCTCCGGTTG
	<i>ndhB</i> -forward	CTCCCACTCCAGTCGTTGCT
	<i>ndhB</i> -reverse	TCCAGAAGAAGATGCCATTC
	<i>rps7</i> -forward	TAGGCGGGTCAACTCATCAA
	<i>rps7</i> -reverse	CCTTTGGCAGCATCCACTAA
	<i>rrn16S</i> -forward	CGCTAGTAATCGCCGGTCAG
	<i>rrn16S</i> -reverse	CCTCCTTGCGGTTAAGGTAA
	<i>trnA</i> -forward	GGGATATAGCTCAGTTGGT
	<i>trnA</i> -reverse	TGGAGATAAGCGGACTCGAA
	<i>trnL</i> -forward	GGGATATGGCGGAATTGGT
	<i>trnL</i> -reverse	TGGGGATAGAGGGACTTGAAC
	<i>rrn23S</i> -forward	TGGGCGTTAGAGCATTGAGA
	<i>rrn23S</i> -reverse	GTTATCCGCTCCGCACTTGG
	<i>rrn4.5S</i> -forward	GAAGGTCACGGCGAGACGAGCC
	<i>rrn4.5S</i> -reverse	GTTCAAGTCTACCGGTCTGTT
	<i>rrn5S</i> -forward	TCCTCAGTAGCTCAGTGGA
	<i>rrn5S</i> -reverse	CTCCCAAGTAGGATTCGAA
	<i>trnN</i> -forward	CGGTTATGGACGAAGGAGA

	<i>trnN</i> -reverse	CCAATGCTAAATGCAGAGGC
	<i>ycf1</i> -forward	TACTATGGTAGCGGCGGGTAT
	<i>ycf1</i> -reverse	TGAGCAAGAGCTAAAGTGGC
	<i>ndhF</i> -forward	AAAACGTACTTCTATCTCGA
	<i>ndhF</i> -reverse	TGTAGAAAGTGATTTCCCTA
	<i>rpl32</i> -forward	TCCCAGTGGTTAATGATGCA
	<i>rpl32</i> -reverse	CCGACTTTGCGAAATGTAAT
	<i>ycf5</i> -forward	TCCTCAGTAGCTCAGTGGTA
	<i>ycf5</i> -reverse	CTCCCCAAGTAGGATTTCGAA
	<i>ndhD</i> -forward	TTCTTCTAACGACCTACGCT
	<i>ndhD</i> -reverse	TCTATTCCCATTCTCCAGTA
	<i>psaC</i> -forward	CTTGTTTTGAGTGCCTATTT
	<i>psaC</i> -reverse	AACTGCATTGAGTATAAGTT
	<i>ndhE</i> -forward	GCTGCTGCACAACTCCTTAT
	<i>ndhE</i> -reverse	CAAACCAACGAAGTAATCCC
	<i>ndhG</i> -forward	CTGTAAC TATTCAATATCCCTA
	<i>ndhG</i> -reverse	GTTTCCAATTTCCAATCAAC
	<i>ndhI</i> -forward	TTTACCAGAAGCGGAAGAAG
	<i>ndhI</i> -reverse	CAACCGCCTAAGTATAGAAC
	<i>ndhA</i> -forward	TCGTATTGGAGGAATAGCGG
	<i>ndhA</i> -reverse	GGATTTCTGTGAATGAGTTT
	<i>ndhH</i> -forward	TTCACTAATAAGATACGAAGAC
	<i>ndhH</i> -reverse	TTTTGACAAATAAGCCAGCA
	<i>rps15</i> -forward	CTTGTTTTGAGTGCCTATTT
	<i>rps15</i> -reverse	AACTGCATTGAGTATAAGTT
RNA gel blot	probe a-forward	GGATCTTCGTAAAATTGATCG
	probe a-reverse	ATCTATGCTACCGAGTATCGTC
	probe b-forward	AAGTGAGCGGCAATTAGGTAA
	probe b-reverse	CCCGAATTATACCAATGGAATTC
	probe c-forward	GTACAGTTGATATAGTTGAGGC
	probe c-reverse	AGATTACCAAGGGAAATAGG
	probe d-forward	ATGCTTCCTATGATAACCGG
	probe d-reverse	TCAAATTAAAGGGTTTACCCC
	probe e-forward	ATGGATTTGCCTGGACCAA
	probe e-reverse	GACGAGCCACAGAAATTGC
	probe f-forward	ATGATACTCGAACATGTACTTGT
	probe f-reverse	CTTATTTAATAAGGTCGATTGG
	probe g-forward	GTCACATTCAGTAAAAATTTATG
	probe g-reverse	TCAATAAGCTAGACCCATACT
	probe h-forward	AGCAATGTACAGCGGTCAAA
	probe h-reverse	ATTCGAACCAACCCATAGGC
RACE	<i>ndhA</i> 5'	CCTGACATAAGAAGTCCAATAGGAGC
qRT-PCR for splicing	<i>trnA</i> (unspliced)-forward	GGGGATATAGCTCAGTTG
	<i>trnA</i> (spliced)-reverse	TGGAGATAAGCGGACTC
	<i>trnA</i> (unspliced)-reverse	TAGAAAAAGTGAGCCACC
	<i>trnI</i> (spliced)-forward	GGGCTATTAGCTCAGTGGTAG
	<i>trnI</i> (unspliced)-reverse	TGGGCCATCCTGGATTTG
	<i>trnI</i> (unspliced)-forward	CGTTCGGGAAGGATGAATC
	<i>trnV</i> (unspliced)-forward	AGGGCTATAGCTCAGTTAG

<i>trnV</i> (spliced)-reverse	TAGGGCTATACGGACTC
<i>trnV</i> (unspliced)-reverse	GACATCGATTTCTTAATAAGATC
<i>atpF</i> (spliced)-forward	GCAACAAATCCAATAAATCTAAG
<i>atpF</i> (unspliced)-reverse	ATAGCTCCTTCACGCAG
<i>atpF</i> (unspliced)-forward	TTCGGGAAGGGATCATAG
<i>rpl2</i> (spliced)-forward	CAGAGGGGCTATAATTGGAG
<i>rpl2</i> (unspliced)-reverse	CGCTGCTCTAGCTAATTG
<i>rpl2</i> (unspliced)-forward	TGCTTTGGAAGAAGCTTG
<i>rps12-2</i> (unspliced)-forward	CTGTAGTCTTAGTAAGAGGG
<i>rps12-2</i> (spliced)-reverse	TTATTTTGGCTTTTTGACCC
<i>rps12-2</i> (unspliced)-reverse	CTTGATAAGAATCTACAACG
<i>trnK</i> (unspliced)-forward	GGGTTGCTAACTCAACG
<i>trnK</i> (spliced)-reverse	ACTCGAACCCGGAAGTAG
<i>trnK</i> (unspliced)-reverse	GTGGTCTTACAACTCTACC
<i>clpP-2</i> (unspliced)-forward	TTCAATAGCATCCTTTATCC
<i>clpP-2</i> (spliced)-reverse	AAGATAAATTCTCCCGTTG
<i>clpP-2</i> (unspliced)-reverse	CTAATTCATATTTCAAATGGCG
<i>petB</i> (spliced)-forward	CATTGTATATTTCCGGAATATGAG
<i>petB</i> (unspliced)-reverse	TATGTTGACATGCGGAGGAA
<i>petB</i> (unspliced)-forward	TCTTGAGGGGGAGTAACCT
<i>petD</i> (spliced)-forward	GAAGAGATAATGGATTATGGGAGTG
<i>petD</i> (unspliced)-reverse	GGGTTCCCCGTAATAATTGTG
<i>petD</i> (unspliced)-forward	AAAAATTATCATGTCCGGTTCC
<i>rps16</i> (spliced)-forward	ATGGTAAAACTTCGTTTAAAC
<i>rps16</i> (unspliced)-reverse	TAAGATCTCTTCCTTCTCTTCG
<i>rps16</i> (unspliced)-forward	CCGTACGAGGCCAAAAC
<i>rpoC1</i> (spliced)-forward	CGTCTTCCTAGTTATATTGC
<i>rpoC1</i> (unspliced)-reverse	ACCTCGTAATCGTAAGAAAG
<i>rpoC1</i> (unspliced)-forward	GGATGAGAGGAAACTTTCATG
<i>ycf3-2</i> (spliced)-forward	TTCGGGCATTAGAACGAAAC
<i>ycf3-2</i> (unspliced)-reverse	AATACTCAGCGGCTTGAG
<i>ycf3-2</i> (unspliced)-forward	TGAGGTAGGAACTCTCAAGTAC
<i>clpP-1</i> (unspliced)-forward	CCTATTGGCGTTCCAAAAG
<i>clpP-1</i> (spliced)-reverse	ATTCGAGATTTCCGGTATCAACC
<i>clpP-1</i> (unspliced)-reverse	TGGGGAAATCCCATATAGC
<i>rps12-1</i> (unspliced)-forward	AGAAATACAAGACAGCCAATC
<i>rps12-1</i> (spliced)-reverse	TACGTAAAGCAGAGTTTGG
<i>rps12-1</i> (unspliced)-reverse	TCAGTCTATGATCTAAACGAGTC
<i>rpl16</i> (spliced)-forward	TCCTTTGATATAATTGCTATGCTTAGT
<i>rpl16</i> (unspliced)-reverse	CCAAATTTTTCCACCACGTC
<i>rpl16</i> (unspliced)-forward	GAAACTCTCACGTTCAATTCTGT
<i>ndhB</i> (spliced)-forward	TCATCAATGGACTCCTGACG
<i>ndhB</i> (unspliced)-reverse	CCAGAAGAAGATGCCATTCA
<i>ndhB</i> (unspliced)-forward	AGTCTCATGCACGGTTTTGA
<i>ndhA</i> (spliced)-forward	CTGCCCAATCGATTAGTTATG
<i>ndhA</i> (spliced)-reverse	GTTTTTGGGGATGGAATTTG
<i>ndhA</i> (unspliced)-forward	TGAGGCCAAGACCTCATG

	<i>ycf3-1</i> (spliced)-forward	CGAGTCATTCCGACAACCTTC
	<i>ycf3-1</i> (unspliced)-reverse	TGTGGTAAAAAGGGGTTTCG
	<i>ycf3-1</i> (unspliced)-forward	CCTAAAGGAGGAGCCGTATG
	<i>trnG</i> (unspliced)-forward	GCGGGTATAGTTTAGTGG
	<i>trnG</i> (spliced)-reverse	GGTAGCGGGAATCGAAC
	<i>trnG</i> (unspliced)-reverse	GGAAAGGACTAAATCCTTTTAAG
	<i>trnL</i> (spliced)-forward	GGGATATGGCGGAATTG
	<i>trnL</i> (unspliced)-reverse	TGGGGATAGAGGGACTTG
	<i>trnL</i> (unspliced)-forward	GATAGAGTCCCATTTTACATGTC
RT-PCR for splicing	<i>trnA</i> -RT-forward	GGGGATATAGCTCAGTTG
	<i>trnA</i> -RT-reverse	TGGAGATAAGCGGACTC
	<i>trnI</i> -RT-forward	GGGCTATTAGCTCAGTGGTAG
	<i>trnI</i> -RT-reverse	TGGGCCATCCTGGATTTG
	<i>trnV</i> -RT-forward	AGGGCTATAGCTCAGTTAG
	<i>trnV</i> -RT-reverse	TAGGGCTATACGGACTC
	<i>atpF</i> -RT-forward	GCAACAAATCCAATAAATCTAAG
	<i>atpF</i> -RT-reverse	ATAGCTCCTTCACGCAG
	<i>rpl2</i> -RT-forward	CAGAGGGGCTATAATTGGAG
	<i>rpl2</i> -RT-reverse	CGCTGCTCTAGCTAATTG
	<i>rps12-2</i> -RT-forward	CTGTAGTCTTAGTAAGAGGG
	<i>rps12-2</i> -RT-reverse	TTATTTTGGCTTTTIGACCC
	<i>clpP-2</i> -RT-forward	TTCAATAGCATCCTTTATCC
	<i>clpP-2</i> -RT-reverse	AAGATAAATTCTCCCGTTTG
	<i>petB</i> -RT-forward	CATTGTATATTTCCGGAATATGAG
	<i>petB</i> -RT-reverse	TATGTTGACATGCGGAGGAA
	<i>petD</i> -RT-forward	GAAGAGATAATGGATTATGGGAGTG
	<i>petD</i> -RT-reverse	GGGTTCCCCGTAATAATTGTG
	<i>rps16</i> -RT-forward	ATGGTAAAACTTCGTTTAAAAC
	<i>rps16</i> -RT-reverse	TAAGATCTCTTCCTTCTCTTCG
	<i>rpoC1</i> -RT-forward	CGTCTCCTAGTTATATTGC
	<i>rpoC1</i> -RT-reverse	ACCTCGTAATCGTAAGAAAG
	<i>ycf3-2</i> -RT-forward	TTCGGGCATTAGAACGAAAC
	<i>ycf3-2</i> -RT-reverse	AATACTCAGCGGCTTGAG
	<i>clpP-1</i> -RT-forward	CCTATTGGCGTTCCAAAAG
	<i>clpP-1</i> -RT-reverse	ATTGAGATTTCGGTATCAACC
	<i>rps12-1</i> -RT-forward	AGAAATACAAGACAGCCAATC
	<i>rps12-1</i> -RT-reverse	TACGTAAAGCAGAGTTTGG
	<i>rpl16</i> -RT-forward	TCCTTTGATATAATTGCTATGCTTAGT
	<i>rpl16</i> -RT-reverse	CCAAATTTTTCCACCACGTC
	<i>ndhB</i> -RT-forward	TCATCAATGGACTCCTGACG
	<i>ndhB</i> -RT-reverse	CCAGAAGAAGATGCCATTCA
	<i>ndhA</i> -RT-forward	CTGCCCAATCGATTAGTTATG
	<i>ndhA</i> -RT-reverse	GTTTTTGGGGATGGAATTTG
	<i>ycf3-1</i> -RT-forward	CGAGTCATTCCGACAACCTTC
	<i>ycf3-1</i> -RT-reverse	TGTGGTAAAAAGGGGTTTCG
	<i>trnG</i> -RT-forward	GCGGGTATAGTTTAGTGG
	<i>trnG</i> -RT-reverse	GGTAGCGGGAATCGAAC
	<i>trnL</i> -RT-forward	GGGATATGGCGGAATTG
	<i>trnL</i> -RT-reverse	TGGGGATAGAGGGACTTG

Poisoned primer extension analysis	PPE( <i>ndhA</i> )-reverse	AACTGTACTTAAACTGTTAGAT(ddCTP)
	PPE( <i>clpP</i> Intron2)-reverse	GCGGGTTGATGGATCATTACCC(ddCTP)
	PPE( <i>rps12</i> Intron1)-reverse	AGCAGAGTTTGGTTTTTGGGGG(ddCTP)
Y2H assay	CAF1(AD:EcoRI)-forward	GGAATTCCATATGGCCGAACGGTTCGAGCAGCG
	CAF1(AD:BamHI)-reverse	CGCGGATCCTGCAAGTAGTTTAGCTAGTTC
	CAF2(AD:EcoRI)-forward	GGAATTCTCCAATAGAAATCGTAATC
	CAF2(AD:BamHI)-reverse	CGCGGATCCTTTTAGTCTTCCCTCAACTTC
	CFM2(AD:SmaI)-forward	TCCCCCGGGTTGTAGTTCTGCTTCCGGTC
	CFM2(AD:BamHI)-reverse	CGCGGATCCCAAACCACATTCAAGTCTTATGG
	CRS2(AD:EcoRI)-forward	GGAATTCCATATGGCTTCGTTGCCTGTTTCTG
	CRS2(AD:BamHI)-reverse	CGCGGATCCAACCTTTATGAAACTTGTATTCTGTC
	SOT1(BD:NcoI)-forward	CATGCCATGGAGTCCTTGAAACAACCTAAACC
	SOT1(BD:BamHI)-reverse	CGCGGATCCTGCTTGAGAAGTAACTAAAGG
Luciferase Complementation assays	CAF1(CLuc:KpnI)-forward	GGGGTACCATGTCGTTAAAACTCAACACTCC
	CAF1(CLuc:Sall)-reverse	ACGCGTCGACTGCAAGTAGTTTAGCTAGTTCATC
	CAF2(CLuc:KpnI)-forward	GGGGTACCATGGCGATTGTAGCATCACTAA
	CAF2(CLuc:Sall)-reverse	ACGCGTCGACTTTTAGTCTTCCCTCAACTTCC
	CFM2(CLuc:KpnI)-forward	GGGGTACCATGTTGCTTCCACTGTTTC
	CFM2(CLuc:Sall)-reverse	ACGCGTCGACCAAACCACATTCAAGTCTTA TG
	CRS2(CLuc:KpnI)-forward	GGGGTACCATGGTTACAGCGATGTTTTG
	CRS2(CLuc:Sall)-reverse	ACGCGTCGACAACTTTATGAAACTTGTATTCTG
	CRS2(NE:KpnI)-reverse	CGGGGTACCAACTTTATGAAACTTGTATTCTG
	SOT1(NLuc:KpnI)-forward	GGGGTACCATGGCGACTGTTCTTACCAC
	SOT1(NLuc:Sall)-reverse	ACGCGTCGACTGCTTGAGAAGTAACTAAAGG
Co-IP assay	CAF1(pUC19:KpnI)-forward	GGGGTACCATGTCGTTAAAACTCAACACTCC
	CAF1(pUC19:BstBI)-reverse	ACTTCGAATGCAAGTAGTTTAGCTAGTTCATC



	CAF2(pUC19:KpnI) -forward	GGGGTACCATGGCGATTGTAGCATCAC
	CAF2(pUC19:BstBI) -reverse	ACTTCGAATTTTAGTCTTCCCTCAACTTC C
	YCF4(pUC19:KpnI) -forward	GGGGTACCATGAGTTGGCGATCAGAATC TATATG
	YCF4(pUC19:BstBI) -reverse	GCTTCGAAAAATACTTCAATTGGTACAC GC
	SOT1(pUC19:KpnI) -forward	GGGGTACCATGGCGACTGTTCTTACCAC
	SOT1(pUC19:Sall) -reverse	ACGCGTCGACTGCTTGAGAAGTAACTAA AGG
EMSA	probe P1-forward	AGGAGATCAAAGCGGTTTT
	probe P1-reverse	TTTAACGATCAACTTCTCCC
	probe P2-forward	CTCGTGAACCTTAGCCGATAC
	probe P2-reverse	TGGAAAGATCTTATCAACGT
	probe P3-forward	CTGCCCAATCGATTAGTTAT
	probe P3-reverse	CCGTTTAACCTTTGTACATAGG
	probe P4-forward	CCTATGTACAAAGGTTAAACGG
	probe P4-reverse	CCTTTTCAATTTGGTTAATCTC
	probe P5-forward	GAGATTAACCAAATTGAAAAGG
	probe P5-reverse	CTTGTTTCGTTCTATTCTTC
	probe P6-forward	GAAGAATAGGAACGAAACAAG
	probe P6-reverse	CTGTCTGCTCAAATCTAAGA
	probe P7-forward	TCTTAGAATTTGAGCAGACAG
	probe P7-reverse	GCCTCAACTATATCAACTGTAC
	RNA 12	Biotin-UGGCUGAUUUA