



Review

Risk Factors for Retinal Ganglion Cell Distress in Glaucoma and Neuroprotective Potential Intervention

Stefania Vernazza ^{1,*},[†] , Francesco Oddone ^{2,†}, Sara Tirendi ^{1,3} and Anna Maria Bassi ^{1,3} 

¹ Department of Experimental Medicine (DIMES), University of Genoa, 16126 Genoa, Italy; tirendisara@gmail.com (S.T.); Anna.Maria.Bassi@unige.it (A.M.B.)

² IRCCS-Fondazione Bietti, 00198 Rome, Italy; francesco.oddone@fondazionebietti.it

³ Inter-University Center for the Promotion of the 3Rs Principles in Teaching & Research (Centro 3R), 56122 Pisa, Italy

* Correspondence: stefania.vernazza@yahoo.it

† These authors contributed equally to this work.

Abstract: Retinal ganglion cells (RGCs) are a population of neurons of the central nervous system (CNS) extending with their soma to the inner retina and with their axons to the optic nerve. Glaucoma represents a group of neurodegenerative diseases where the slow progressive death of RGCs results in a permanent loss of vision. To date, although Intra Ocular Pressure (IOP) is considered the main therapeutic target, the precise mechanisms by which RGCs die in glaucoma have not yet been clarified. In fact, Primary Open Angle Glaucoma (POAG), which is the most common glaucoma form, also occurs without elevated IOP. This present review provides a summary of some pathological conditions, i.e., axonal transport blockade, glutamate excitotoxicity and changes in pro-inflammatory cytokines along the RGC projection, all involved in the glaucoma cascade. Moreover, neuro-protective therapeutic approaches, which aim to improve RGC degeneration, have also been taken into consideration.

Keywords: neuroprotection; glaucoma; retinal ganglion cells; neuroinflammation; degeneration



Citation: Vernazza, S.; Oddone, F.; Tirendi, S.; Bassi, A.M. Risk Factors for Retinal Ganglion Cell Distress in Glaucoma and Neuroprotective Potential Intervention. *Int. J. Mol. Sci.* **2021**, *22*, 7994. <https://doi.org/10.3390/ijms22157994>

Academic Editors:

Sandra Petrus-Reurer,
Arturo Ortín-Martínez and
Luis Alarcon-Martinez

Received: 23 June 2021

Accepted: 23 July 2021

Published: 27 July 2021

Publisher's Note: MDPI stays neutral with regard to jurisdictional claims in published maps and institutional affiliations.



Copyright: © 2021 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (<https://creativecommons.org/licenses/by/4.0/>).

1. Introduction

Glaucoma represents a group of neurodegenerative diseases characterized by optic nerve damage and the slow progressive death of retinal ganglion cells (RGCs). Indeed, glaucoma is regarded as the second cause of irreversible blindness worldwide and it is estimated that its incidence will increase to more than 112 million cases in the future [1–3].

In connection with an increased susceptibility to glaucoma and the progression of such a disease, several risk factors, including age, Intra Ocular Pressure (IOP), race, severe myopia, genetic background, vascular dysregulation and central corneal thickness, have been identified [4,5]. Such a multitude of risk factors can be explained by glaucoma etiological complexity, as well as by the several glaucoma forms in which pressure does not increase [6]. Indeed, although IOP has been recognized as one of the main glaucoma risk factors due to it being responsible for both mechanical axonal damage and nutrient interruption [5,7], there is a particular glaucoma form, corresponding to 20–25% of glaucomatous optic neuropathy, characterized by IOP within a normal range, i.e., Normal Tension Glaucoma (NTG), which, however, leads to progressive blindness. Moreover, not all people who have elevated IOP suffer from glaucoma [8–10].

From an anatomical point of view all glaucoma forms are classified in two types, i.e., the open angle and closed-angle glaucoma, according to the geometry of iridocorneal angle, the point where the iris and the cornea meet [6]. However, Primary Open Angle Glaucoma (POAG) is the most common glaucoma type, accounting for over 70% of cases.

Currently, clinical treatments for all glaucoma types aim for lowering IOP through topical hypotensive drugs or surgery. However, these approaches are not sufficiently successful for many patients who continue to lose their vision [11]. Therefore, it would

seem evident that RGC death is also driven by different converging molecular pathways, engaged in additional damage more or less closely connected with IOP elevation, which are able to trigger or exacerbate the glaucomatous cascade. The purpose of this present review is to summarize the most recent evidence about some of the possible upstream causes which are responsible for RGC death, as well as neuroprotective strategies to prevent or at least to slow down progression of the retinal distress.

2. Retinal Ganglion Cells

RGCs, in addition to cone photoreceptors and horizontal, amacrine and bipolar cells, as well as Müller glia, all belong to the neural cells of the vertebrate retina [12]. Although they share a common multi-potent retinal progenitor cell (RPC), RGCs are the first to be generated.

During the embryonic phase, by way of Sonic Hedgehog (SHH), RGCs exert a control over different downstream molecular events, including the laminar organization of the retina, the size of successive cell populations, the development of optic stalk neuro-epithelial cells and their morphogenesis [12–15]. In the advanced stages of development, the RGC axons of the optic nerve, which form the retinal nerve fiber layer, are closely involved with the migrating astrocytes. The Roundabout (Robo) single-pass type I transmembrane family receptors are key regulators of commissural axon guidance, providing and controlling the complex neuronal networks [16]. In fact, by way of *Robo 1* and *Robo 2*, RGCs provide directional information to the astrocytes, i.e., the centrifugal trajectory, which is useful for their polarization and for the colonization of the peripheral retina [14,17]. Moreover, RGCs and their axons are responsible for transmitting all the visual information from the eye to the brain (i.e., lateral geniculate nucleus of the thalamus and superior colliculus of the midbrain) through the optic nerve [18].

Amongst all the neuron types, RGCs, due to their morphological features and their high energy requirements, are particularly vulnerable to mitochondrial dysfunction. In fact, a lack of saltatory conduction, due to unmyelinated axons of RGCs within the retina, leads to a higher requirement of more mitochondria in order to respond to the high metabolic demand [19]. Therefore, to provide RGCs with the correct energy contribution, the mitochondria integrity, in terms of their biogenesis, dynamics, transport and degradation (e.g., mitophagy), is required. The presence of mitochondria deficits, triggered by mutations in genes encoding essential mitochondrial functions, as in the case of aging, results in an inadequate energy supply which is involved in RGC degeneration [20,21].

Several diseases can occur as a result of either RGC axon dysfunction or their degeneration, including glaucomatous optic neuropathy, inflammatory optic neuropathy, ischemic optic neuropathy, traumatic optic neuropathy, hereditary optic neuropathy and toxic/nutritional optic neuropathy [22].

In particular, the neuropathy characteristics of different glaucoma types include permanent visual deficits that begin with peripheral vision loss, degenerating towards the so-called “tunnel vision” and, eventually, the loss of sight [23].

Although the precise mechanisms by which RGCs die in glaucoma have not yet been clarified, three main causes of severe and extensive damage have been proposed: (i) axonal transport blockade; (ii) Glutamate excitotoxicity; and (iii) changes in pro-inflammatory cytokine along the RGC projection (Figure 1).

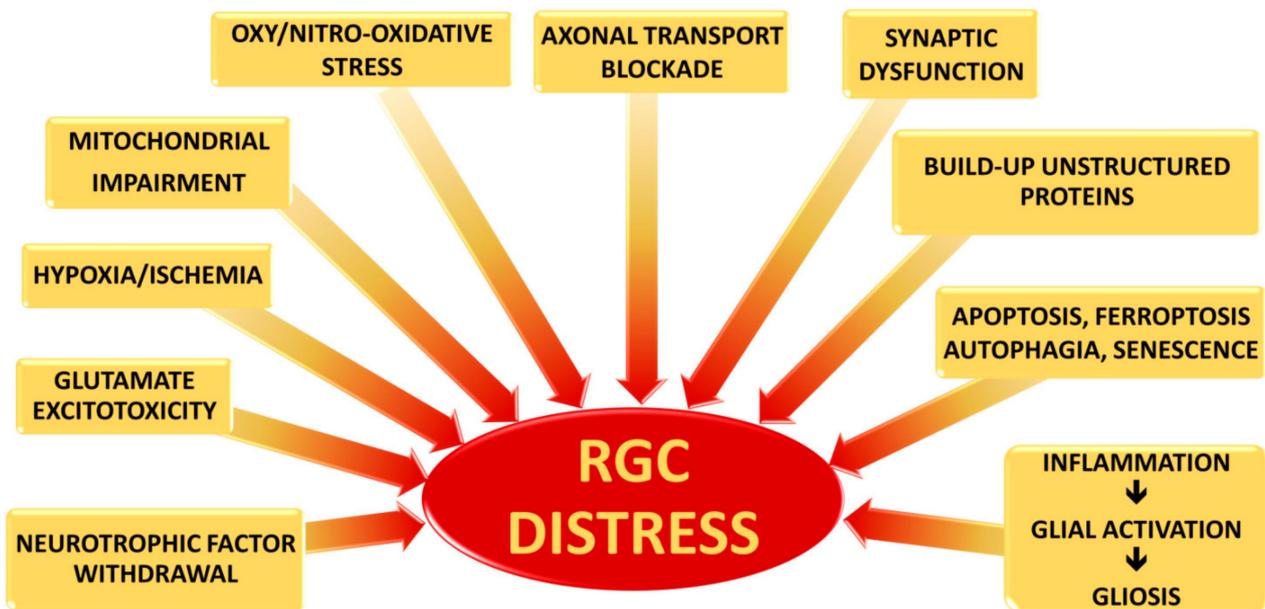


Figure 1. Risk factors contributing to RGC distress in glaucoma: the mechanisms implicated in RGC damage include deficiency in neurotrophic elements, glutamate excitotoxicity, hypoxia and ischemia, impairment of mitochondria functions, oxidative and nitrosative stress, alterations in axonal transport and in synaptic signals, unstructured proteins, apoptosis and other death mechanisms which can lead to premature senescence, and inflammation cascade which induce the activation of glial cells with consequent gliosis. The pathways, triggered by each risk factor, often result to be strictly interrelated, contributing to amplifying RGC distress in an irreversible way.

2.1. Axonal Transport Blockade

Undoubtedly, axonal transport is an essential process for neuron survival and, overall, for the maintenance of both the metabolic balance and intracellular neurotransmission. Moreover, it allows the neuron to respond effectively to trophic signals or stress insults. Such a process is distinguished in anterograde and retrograde axonal transport. The anterograde axonal transport, in addition to providing proteins and lipids from the soma to the distal synapses, also supplies local energy by way of a targeted mitochondria movement [24]. However, the retrograde transport is involved in the misfolded protein removal from the axon and in the intracellular transport of distal trophic signals to the soma in order to avoid toxic aggregate formation [25].

The axonal transport along the microtubule network, which can be divided into fast (0.5–4 $\mu\text{m/s}$) and slow (0.01–0.05 $\mu\text{m/s}$) transport according to the type of transported cargo [24,26], takes place thanks to the so-called molecular motors, i.e., the kinesin and dynein complexes. Although both require ATP to propel their cargo along the microtubule rails, kinesin is involved in the anterograde transport while dynein in the retrograde one [27].

It is now known that a defective axonal transport, related to the malfunctioning of one or more involved components (e.g., molecular motors, microtubules, cargo, or mitochondria), results in a predisposition to neurodegenerative diseases such as Alzheimer's disease (AD) and other dementia, Parkinson's disease (PD), Huntington's disease, Charcot-Marie-Tooth peripheral neuropathy, amyotrophic lateral sclerosis and glaucoma [28,29].

Increasing evidence has highlighted that the neurodegeneration outcome can change according to the different involvement of defects borne by mutations in the motors that drive the anterograde and/or retrograde axon transport [25,30–34].

In particular, in glaucoma neurodegeneration, long-lasting harmful stimuli, such as low levels of oxygen or energy, the presence of neurotoxins, an increased pressure gradient across RGCs axons and pro-inflammatory cytokines, initially promote an anterograde transport deficit and, successively, a retrograde one from the RGC body [7,20]. Although

the presence of cytoskeleton and/or protein abnormalities, as well as metabolic deficits, are all clear signs of a pre-degenerative state, they do not necessarily affect the axonal function immediately [35]. However, the progressive axonal transport blockade, which is responsible for trophic signal deprivation (e.g., neurotrophic factors), plays an essential role in promoting RGC loss [36].

As known, neurotrophic factors (NTFs) are critical for the differentiation and maintenance of the nervous system, as well as for the support of neuronal cell survival. NTFs generally include: the neurophin family, the glial cell-line derived neurotrophic factor (GDNF) family and the family represented by Ciliary Neurotrophic Factor (CNTF) [37,38].

As for neurotrophins (NTs), they are a family of polypeptides with multiple functions, both at peripheral and central nervous system level, ranging from neuronal survival and differentiation to the modulation of synaptic transmission and plasticity [39,40].

There are four NTs, namely Nerve Growth Factor (NGF) [41,42], Brain-Derived Neurotrophic Factor (BDNF) [43], NT-3 [44] and NT-4/5 [45], which are all characterized by homologies in terms of their sequence and structure [46]. These NTs bind two different classes of transmembrane receptors: the p75 NT receptor (p75^{NTR}) and three members of Tropomyosin-Related Kinase (Trk) receptor tyrosine kinases, i.e., TrkA, TrkB and TrkC [47]. Therefore, NTs, through p75^{NTR}, control both the neuron fate, via the NF- κ B or Jun kinase pathways, and the motility of cone growth regulating Rho activity [48,49]. On the other hand, NTs, through Trk receptors, activate pro-survival and/or pro-growth signaling cascades, such as Ras/Raf/MEK/ERK and PI3-K/Akt/mTOR pathways [47,50].

Among NTs, BDNF seems to be particularly crucial for RGC survival because, in addition to being produced by the RGCs themselves and by retinal astrocytes, it also has a role in retrograde axonal transport (from the brain to the retina) [51,52]. Indeed, BDNF, once produced by the superior colliculus or by the lateral geniculate nucleus, binds to the TrkB receptor in order to reach the cell bodies by way of retrograde transport within the microsomal vesicles [53].

Previous studies of induced glaucoma animal models have shown that ocular hypertension is responsible for the obstruction of retrograde axonal transport at the optic nerve head to the point of affecting the distribution of both BDNF and TrkB along the RGCs, thus promoting RGC death [54,55]. In order to support these results, it has been shown that either multiple intravitreal injections of BDNF or intravitreal treatment with a BDNF-expressing adenoviral vector can actually temporarily slow down RGC loss [54,56,57].

However, RGC death, induced by ocular hypertension, can also be explained in another way. Indeed, in some cases, changes in both the mRNA and protein of NT receptors favor the activation of the p75-mediated pro-apoptotic signal, resulting in the alteration of the effects of NTs on RGC survival [58–60].

The GDNF family (i.e., GDNF, neurturin, artemin and persephin) is a distant member of the transforming growth factor- β (TGF- β) superfamily. Given its powerful neuroprotective role on various neurons, it has also been proposed as a target for treating several neurodegenerative diseases (e.g., AD and PD) [61–63].

In addition, it seems that GDNF acts on retinal degeneration both indirectly, stimulating the photoreceptor survival [41,42], and directly, after intravitreal injection of microspheres containing GDNF [64–67].

At a Müller glia level, a further interesting aspect of GDNF is its ability in enhancing the gene expression of the glutamate/aspartate transporter (GLAST), which is essential for RGC protection [68]. GLAST, together with Glutamate Transporter 1 (GLT-1) located in the bipolar cell terminals, and the Excitatory Amino Acids Carrier 1 (EAAC1) in retinal neurons including RGCs, are the only mechanisms for glutamate removal from the extracellular fluid in the retina. Moreover, GLAST is also important for glutathione synthesis, a major cellular antioxidant in the retina.

Although the excitotoxicity of glutamate in glaucoma pathogenesis is still controversial, in a mouse model, in which GLAST and EAAC1 were knocked-out, it has been shown

that their absence promoted RGC death and glaucomatous damage, even without ocular hypertension [65].

Moreover, since CNTF belongs to the neurotrophic cytokine family, it is expressed by different cells in the retina and especially by Müller Glia. In addition to enhancing the survival of both photoreceptors and RGCs, CNTF is capable of stimulating axonal regeneration [69]. The specific binding of CNTF to the receptor complex, which includes the CNTF receptor- α (CNTFR α), gp130 and Leukemia Inhibitory Factor receptor (LIFR), activates Jak-STAT, MEK-ERK and PI3K/Akt signaling pathways [70,71].

The protective role of CNTF in eye-sight maintenance has been elucidated by aqueous humor analysis of POAG patients in which the severity of the disease correlated to the increased reduction in CNTF levels [72]. Moreover, studies on animal models have shown a progressive up-regulation of CNTF levels after optic nerve damage but only during the first weeks and not beyond [73,74].

2.2. Glutamate Excitotoxicity

Glutamate is the most abundant excitatory neurotransmitter in the mammalian CNS and is also involved in most of excitatory neurotransmissions when bound to its specific receptors.

Thus, in the CNS, glutamate is mainly located intracellularly even though, during synaptic transmission when its extracellular concentration increases, its levels in the synaptic cleft at resting conditions are kept low by the glutamate uptake/transporter system, which actively removes the residual glutamate [75,76].

Glutamate binds mainly to ligand-gated ionotropic glutamate receptors (iGluRs), which are classified into two groups: N-methyl D-aspartate (NMDA) receptors and non-NMDA receptors (i.e., AMPA and kainate receptors) [77,78]. These play fundamental roles in synaptic plasticity [79] and, in particular, NMDA receptors are also essential for neuron survival by activating the neuronal survival pathway [80,81]. At resting membrane potentials, glutamate cannot activate the NMDA receptors due to a control mechanism, consisting of magnesium ions (Mg^{2+}), which blocks the intracellular influx of sodium (Na^+) and calcium ions (Ca^{2+}). However, this blockade by Mg^{2+} , being voltage-dependent, is lost after neuron depolarization, enabling NMDA receptors to be activated by glutamate and also allowing the influx of sodium (Na^+) and calcium ions (Ca^{2+}), as well as the efflux of potassium ions (K^+) [82]. Interestingly, on astrocyte and oligodendrocyte membranes, several receptors, belonging to the NMDA type, but with different subunit compositions, have been found. It would appear that their activation occurs without antecedent depolarization because they have not the blockade by Mg^{2+} [83,84].

An increase in extracellular glutamate concentration may be observed under several pathological conditions (e.g., ischemia/reperfusion injury, oxidative stress, inflammation and as a result of aging) due to a reduced activity of the glutamate uptake/transporter system [85–88]. The abundance of glutamate, in turn, results in nervous tissue damage which can alter physiological brain functions. Moreover, since TNF α increases the glutaminase expression on astrocytes, it is also responsible for increasing the glutamate amount [89].

A progressive glutamate accumulation in the extracellular space leads to glutamate excitotoxicity. Therefore, excitotoxicity injury refers to a condition in which the ionic homeostasis is lost due to an NMDA receptor overstimulation by glutamatergic neurotransmission. Since such damage leads to cell death, it has been proposed as a possible mechanism involved in neurodegenerative conditions, including AD and glaucoma [82,90,91]. However, increasing evidence indicates that an excess of glutamate is responsible for the hyperactivation of the glutamate receptors but the resulting massive Ca^{2+} influx into the post synaptic neurons is also due to low ATP levels, which prevent the proper functioning of the ion pumps [92]. Such intracellular accumulation of Ca^{2+} ions represents the trigger for activation of both the protein-kinases and other downstream Ca^{2+} -dependent enzymes, which through a series of molecular modifications, such as cellular membrane damage, ROS release, mitochondrial dysfunction, lead to cell death [91,93].

Although the direct role of glutamate excitotoxicity in glaucoma pathogenesis remains circumstantial [94–97], its role has been demonstrated during both ischemic and hypoxia injuries [98,99]. Therefore, since both conditions have been found in glaucoma, it is conceivable that glutamate excitotoxicity could be involved in RGC death as a possible outcome of extensive damage rather than being the main cause [100].

Under prolonged hypoxic-ischemic conditions, the neural protective mechanisms are lost, leading to cell death and tissue damage. Therefore, under such conditions, neuronal degeneration can result from different detrimental mechanisms triggered by oxygen and substrate deprivation, including glutamate excitotoxicity, free oxygen radicals, and pro-inflammatory cytokines, causing a loss in the function of the blood retinal barrier (BRB) [100].

2.3. Changes in Pro-Inflammatory Cytokine along the RGC Projection

As known, inflammation is defined as an unavoidable cellular response to a prolonged period of noxious stress or tissue malfunction, characterized by a marked release in pro-inflammatory cytokines/chemokines and tissue damage. Although the para-inflammation condition refers to an adaptive-tissue response, necessary for maintaining tissue homeostasis and restoring its functionality, either an increase in its magnitude or its dysfunction leads to the loss of its physiological role, thus resulting in an inflammation state [101].

Increasing evidence suggests that oxidative stress and ocular inflammation are the main factors involved in the pathogenic mechanisms that lead to the neurodegeneration process found in NTG and high tension glaucoma (HTG) [102–104].

Under physiological conditions, structural and functional integrity of the retina is provided by a regular oxygen supply since it is one of the most metabolically-active tissues [105]. However, pathological conditions, such as central retinal artery occlusion and ischemic central retinal vein thrombosis, lead to retinal hypoxia-ischemia, which is involved in several diseases including glaucoma [106–109]. As above-described (see Section Glutamate Excitotoxicity), during retinal hypoxia-ischemia, a series of cellular alterations, such as the increase in the free oxygen radical production, the activation of the inflammatory pathway, the glutamate excitotoxicity and the destruction of the BRB, occur.

In particular, the strong connection between the retinal ischemic episode and the increase in Reactive Oxygen Species (ROS) production is of great interest. In fact, when normal blood-flow is re-established, the ischemic injury gets even worse due to re-oxygenation, which represents an important mechanism of cellular damage and ROS production. In this regard, three phases of ROS generation have been distinguished: the first one is sustained by mitochondria in response to the inhibition of mitochondrial respiration, the second by xanthine oxidase activation, due to ATP depletion, and the third, by the calcium-dependent activation of Nicotinamide Adenine Dinucleotide Phosphate (NADPH) oxidase [110]. This massive increase in ROS stimulates the damaged cells to produce inflammatory cytokines, such as TNF α and interleukins (IL), as well as vascular endothelial growth factor (VEGF) and various chemokines [100,111]. Moreover, the trigger of an acute or chronic inflammation response leads to a further infiltration in the ischemic tissue by pro-inflammatory mediators, by way of the intercellular space between the vascular endothelial cells (i.e., BRB), resulting in a rapid microglial/monocytic response and gliosis [111].

The BRB is a functional neurovascular structure composed of two barriers: the retinal vascular endothelium (inner part) and the retinal pigment epithelium (outer part). Under normal conditions, the BRB, similar to the blood brain barrier (BBB), prevents extracellular fluid accumulation through specific tight cell-cell junctions [112] and maintains the retina as an immune-privileged site, thus stopping the passage of both systemic immune and inflammatory components [113]. Indeed, local immune suppression in the eye is achieved through the blood-aqueous, the BRB, as well as the local production of immunosuppressive cytokines and neuropeptides, which promptly eliminate, by apoptosis, any immune cells which have entered the retina in response to an infection [114].

However, in glaucoma, the presence of either foreign/auto- antigens or an increased IOP lead to neuroinflammation by way of both toll-like receptor (TLR) and tumor necrosis factor receptor (TNFR) signaling-pathways, the glial NF- κ B activation and the assembly of inflammasome [93,115–118].

The resulting increase in inflammatory mediators (i.e., the Complement system) and cytokine secretion (i.e., TNF α) are responsible for the reduced BRB efficacy. The edema formation, in turn, disrupts the functioning of surrounding cells by compression [119]. Moreover, the increase in the Major Histocompatibility Complex Class I and II by VEGF and the Heat Shock Proteins leads to the activation of an adaptive immune response and, consequently, the disruption of the BRB [111,120].

An impairment in both BBB and BRB may also be found in aging, resulting in an inflammatory or immune response. In fact, despite such impairments not being observed in all aging conditions, during age-related neurodegeneration, including glaucoma, there is sufficient evidence that T lymphocyte infiltration causes microglia activation and RGC degeneration [121].

Previous studies have shown differences in T-cell subsets between blood samples of glaucomatous patients and healthy subjects [10,122], leading to the assumption that T-lymphocytes could be involved in the initiation and progression of some glaucomatous lesions [10]. In fact, the lymphokine pattern relating to various T-lymphocyte subsets plays a crucial role in worsening or switching off the immune response (i.e., Th1 secrete IFN γ and IL2, Th2 secrete IL4 and IL10, T suppressor/cytotoxic 1 secrete IFN γ and T suppressor/cytotoxic 2 secrete IL5 and IL10).

At present, new experimental evidence has clearly shown the role played by glial cells, together with dendritic cells and invading monocytes in triggering the adaptive immunity during glaucomatous neuroinflammation [123,124]. Such conditions, favoring the interaction between glia and T-cells, boost the immune response and lead to an immune-degenerative response [10,122]. Furthermore, it would seem that some proteins belonging to the Complement system, including C1q, C3 and C4, are mainly up-regulated at both the optic nerve head (ONH) and the inner plexiform layer in the early stages of glaucoma degeneration [5,125]. In particular, Hubens et al. [126] have shown that the ratio C3a:C3 was significantly increased in both the serum and the AH of only the progressive POAG patients compared to the stable POAG patients. Therefore, the rising levels of anaphylatoxin C3a is related to the increased cleavage of C3 and the consequent release of C3b, which then binds to the C3 convertase (C4bC2a), forming a new enzymatic complex, namely C5 convertase. In turn, the C5 convertase cleaves C5 to bioactive fragments C5a (anaphylatoxin) and C5b, with the latter recruiting the last complement components C6, C7, C8, and C9 to form the membrane-attack-complex (MAC) ring, leading to cell lysis (Figure 2). This effect provokes RGC damage exacerbation at a retinal tissue level [127,128]. The increase in Complement activation has been linked to reduced RGC survival both in humans and in animal glaucoma models, suggesting its crucial role in glaucoma progression [19].

- (1) The classical pathway is activated by the interaction between the antibody-antigen (Ab-Ag, respectively) and C1-complex, which consists of C1q, C1r, and C1s. This interaction leads to the cleavage of C4 and C2 and the complex C4bC2a to form the C3 convertase which cleaves C3 into C3b and C3a. Then C3b associates with C4bC2a to give rise to the C5 convertase which, in turn, cleaves C5 in C5a and C5b. C3a and C5a are anaphylatoxins, acting as vasoactive and chemotactic factors, while C3b is an opsonin inducing phagocytosis. C5b interacts and activates other complement components, namely C6, C7, C8, and C9 to form the membrane-attack-complex (MAC), which lyses targeted surfaces.
- (2) The lectin pathway is activated by the mannose-binding lectin (MBL) recognition of pathogenic carbohydrate motifs. The complex MBL-associated serine protease (MASP) cleaves C2 and C4 and generates the C3 convertase which then merge at the subsequent step of the classical pathway.

- (3) The alternative pathway starts from the spontaneous hydrolysis of C3 to the C3b analog, C3(H₂O), which, in the presence of Factors B and D, forms an alternative C3 convertase, namely C3(H₂O)Bb, which converts C3 into C3b and C3a, acting as the C3 convertase of the classical and lectin pathways. The alternative pathway can also contribute to forming a C5 convertase (C3bBbC3b) merging at C5 of the classical and lectin pathways [129,130].

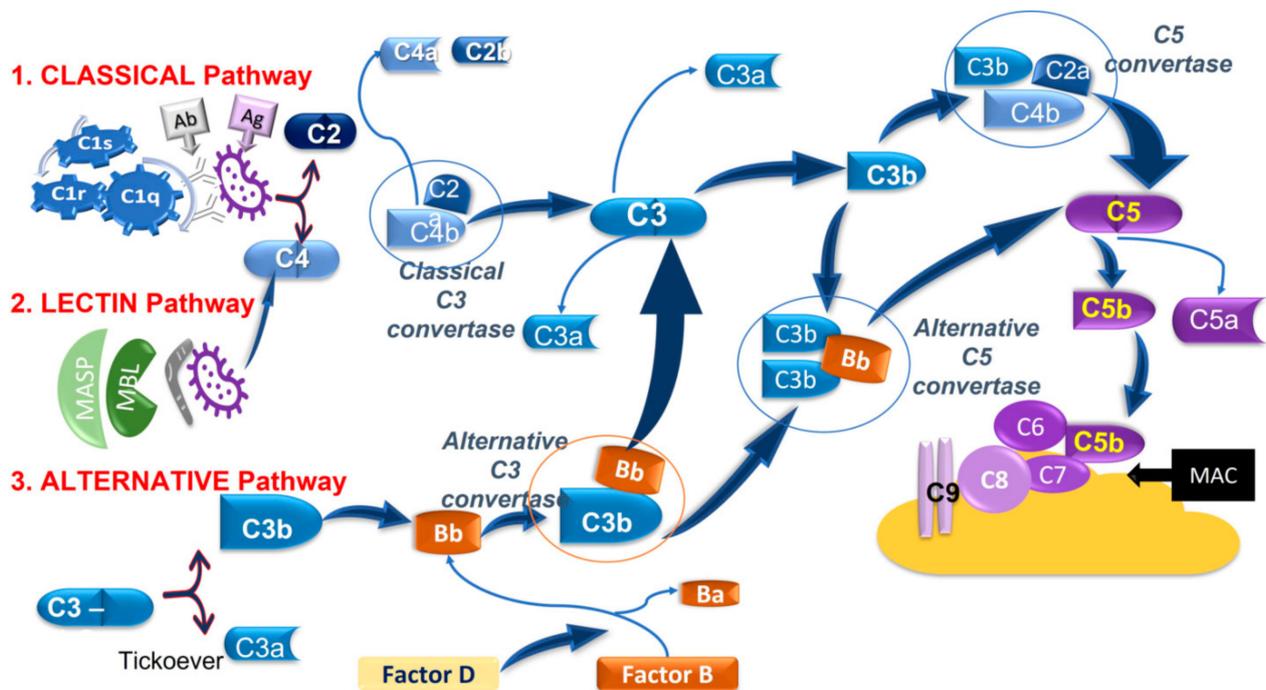


Figure 2. The Complement pathway: three pathways activate the Complement cascade, and they converge within this.

3. Neuroprotection

Retinal damage in glaucoma is due to the gradual degenerative process that results in a worsening of the retina function and the impairment of the visual microenvironment [131]. Indeed, neuro-imaging technologies showed that POAG patients' eyes display abnormalities, including the death of a significant number of RGCs and the loss of axons in the optic nerve [132].

Although IOP management with surgery or drugs is the main clinical approach, the outcome is not always satisfactory. Generally, this is due to the different and complex mechanisms by which RGCs die, such as mitochondrial dysfunction, oxidative stress, NT deficits, glutamate excitotoxicity, acute/chronic ischemia, inflammation, and glial activation [133].

Therefore, in addition to well-known hypotensive drugs, either new local or systemic drugs with neuroprotective mechanisms, are needed in order to prevent the progression of visual deficits. The term "neuroprotection" refers to a therapeutic paradigm which is able to prevent glaucomatous neurodegeneration, or at least to slow down the progression of the RGC death [131]. Indeed, the possibility of counteracting some of the different complex pathways underlying the apoptotic cascade could represent the key to RGC protection [134,135].

In general, also under para-physiological conditions, RGCs may suffer damage, such as oxygen deficiency, leading to an impairment in their mitochondrial function, or to a condition of oxidative stress. Therefore, these risks are counterbalanced by certain defense systems, which, depending on the context of the retinal microenvironment, instead of limiting the damage, may actually contribute to glial cell activation and, as a consequence, lead to the progressive deterioration of retinal function [135,136].

In this regard, the more promising neuroprotective approaches are based on the increase of antioxidant activities given that oxidative stress, like mitochondrial dysfunction,

can be considered common denominators of all glaucoma forms [137]. In fact, the integrity of RGCs can be affected by an imbalance between ROS production and the inefficiency of antioxidant defenses, allowing mitochondrial dysfunction to play both a direct and indirect role in RGC apoptosis [138–141]. Thus, several therapeutic approaches have recently been focused on the protection of mitochondrial functions, as well as on the increasing of antioxidant systems, in order to protect the RGCs from damage and from the consequent risk of apoptosis activation. Since these approaches contribute to the maintenance of mitochondrial integrity, they can be considered a useful tool in the neuroprotection field [142].

3.1. Müller Glia and Neuroprotection

Müller glial cells (MGCs), which together with astrocytes form a part of retinal macroglia, play an important role in maintaining retinal homeostasis by supervising the ionic exchange and the response to neurotrophic molecules [143,144]. Moreover, MGCs exert a real neuroprotective effect by releasing NTs and carrying out antioxidant functions, as well as regulating neuronal signaling by recycling neurotransmitters [145].

The function of NTs has already been discussed in the previous section, but it is important to highlight the role of MGCs in the regulation of such factors, which are selectively produced in response to surrounding stimuli. MGCs control the uptake and clearance of glutamate, preventing neurotoxicity. If an excess of this neurotransmitter occurs in the extracellular environment, MGCs up-regulate BDNF, NGF, NT-3, NT4 and GDNF levels to avoid glutamate toxicity [146]. Furthermore, the release of neurotrophins by MGCs occurs also in the presence of other NTs, inflammatory cytokines and activated macroglia [65,147,148].

MGCs control the physiological antioxidant response through the expression of high levels of both glutathione (GSH) and glutamate transporters [149,150] and, during oxidative stress conditions, they also regulate the nuclear factor erythroid 2-related factor 2 (NRF2) expression [151,152].

In several *in vivo* and *in vitro* studies, it has been demonstrated that kidney, liver, and brain cells express a cystine-glutamate antiporter, which facilitates the exchange between cystine and glutamate, promoting GSH synthesis as a consequence. This exchange is crucial in the retina microenvironment, where, under ischemia or other redox imbalances, the extracellular concentration of glutamate is high, and therefore GSH levels have to be increased in order to counteract the oxidative stress conditions [153–157].

Lastly, another important function of MGCs is the direct control of neuronal-signaling by recycling of both excitatory (i.e., glutamate) and inhibitory (i.e., GABA) neurotransmitters [158–160]. Under pathological conditions, such as ischemia, the loss of extracellular glutamate/GABA regulation leads to the loss of MGC neuroprotective effects.

MGCs, due to their intrinsic role in counteracting stress damage, are also involved in inflammation and cell repair/survival processes. [145,161–165]. Indeed, under pathological processes, the establishing of a reactive gliosis results in the MGC release of vasoactive molecules and pro-inflammatory factors [166]. Therefore, during aging, cell injury and oxidative stress, an impairment of GSH content and an increase in TNF α levels occur, which, in turn, lead to other damaging conditions, such as lipid-peroxidation cascade, ferroptosis, autophagia, senescence and death pathway (i.e., apoptosis or necrosis) [167], exacerbating neuronal dysfunction. Although these outcomes are known to be strictly correlated with one another, their prevalence would seem to be dependent on the particular cell model on which they are analyzed [168].

3.2. Neuroprotective Agents in Glaucoma Management

3.2.1. Antioxidants

To maintain redox homeostasis, multiple therapeutic targets, on which endogenous and exogenous neuroprotective molecules may act, have been proposed.

For instance, Vitamin E (i.e., α -tocopherol, γ -tocopherol and TPGS) has been shown to reduce the edema formation during ischemia-reperfusion injury [169].

Coenzyme Q10 (CoQ10), which is an important component of the mitochondrial respiratory chain, when administered as a supplementary therapy slows or reverses pathological conditions, including ischemia and neurodegeneration and also protects against glutamate excitotoxicity [170].

Moreover, the concomitant topical administration of Vitamin E and CoQ10 reduce both retinal damage and RGC loss, thus preventing mitochondrial permeability transition pore formation and the release of cytochrome c [171].

Furthermore, Nebbioso et al. [172] demonstrated that the treatment with alpha-lipoic acid and superoxide dismutase exerts anti-apoptotic effects and protects against oxidative stress in glaucoma animal models. In addition, intravitreal pre-treatment with recombinant adeno-associated virus expressing SOD2 inhibits RGC death, despite IOP elevation [173].

Plant foods, which are the so-called exogenous antioxidant molecules, are rich antioxidant sources. In this regard, the relation between their dietary intake and glaucoma [174–176] is currently being assessed. In glaucoma patients, the dietary intake of Vitamin A and C showed soothing effects on the disease progression [177]. Moreover, quercetin supplementation has evidenced a protection on the mitochondrial function in a rat chronic ocular hypertension model and in primary-cultured RGCs exposed to hypoxia [178] whereas Ginkgo Biloba extract and anthocyanins improve the general visual function of patients affected by normal tension glaucoma [179]. Lycium barbarum extracts, in rat PC12 neuronal cells, showed a neuroprotective action towards glaucoma onset by modulating the expression of extracellular matrix proteins and polysaccharides, preserving the RGCs from oxidative stress-induced apoptosis [180,181].

Carotenoids also evidenced neuroprotective effects, exerting remarkable anti-inflammatory and antioxidant action in the retina. In particular, among carotenoids, lutein- and axanthin-derived molecules, have been studied in several in-vitro and clinical studies [182–190].

However, despite the large number of available antioxidants, the limits of their therapeutic use are represented by a failure to reach the relevant sites of free radical generation (e.g., mitochondria). The use of specific mitochondrial-targeted peptides with targeted delivery antioxidants to the inner mitochondrial membrane as in the case of Szeto-Schiller peptide 31 (SS-31), can at least partly overcome this problem [191–193]. In fact, SS-31, in reducing ROS-mitochondria levels, exerts neuroprotective effects directly on RGCs and antioxidant effects on the retina [194].

3.2.2. Neurotrophic Factors (NTFs)

In addition to antioxidant molecules, also neurotrophic factors are the focus of basic research and clinical studies due to the fact that a lack of neurotrophic factors contribute to the impairment of the axonal and synaptic functions (see the previous section) [5,135,136,195].

Although NTF-signaling pathways are subjected to a complex regulation, that does not always favor cell survival, different techniques for restoring NTFs have been developed.

It is now known that in HTG, the increase in IOP leads to reduction of NGF content in cerebrospinal fluid and lateral geniculate nucleus as well as a reduced expression of its receptors in brain visual centers [196–198]. Lower NGF levels or a lack thereof are also found in age-related neurodegenerative diseases (e.g., AD neurodegeneration) [199]. Therefore, chronic NGF administration has been proposed to counteract both AD and glaucoma neurodegenerations. In the particular case of glaucoma, it has been shown, both in rat glaucoma models and in three patients with advanced glaucoma, that repeated topical administration of exogenous NGF prevented RGC apoptosis or, at least, resulted in a long-term improvement in their eyesight.

However, glaucoma degeneration results in generalized NT deprivation, which, together with additional obstruction to retrograde transport, contributes to disease progression. In this regard, further studies have been aimed at restoring the loss of other NTs, such as BDNF and CNTF. Indeed, repeated intravitreal injections of purified BDNF or

CNTF exert neuroprotective effects by reducing the amount of RGC loss, even though BDNF was more effective than CNTF since, after four injections, it was seen to increase the RGC survival by about 73–83% compared to the uninjured controls [56,200,201]. Nevertheless, given the issue related to their sustained delivery, this approach has not found a feasible clinical translation for treating glaucoma. In this context, the introduction of slow-release devices, such as microspheres of poly-DL-lactide-coglycolide (PLGA) infused with NTFs, allow for a sustained NTF release after microsphere degradation, even though repeated treatments are necessary. In fact, with this approach, after repeated injections of microspheres, infused with GDNF, the RGC survival of DBA/2J mice was increased up to nine months [202]. Moreover, gene therapy, using a viral vector (e.g., adenovirus), could improve RGC degeneration long term through a transient up-regulation of specific NTFs [57,203].

Even though several pre-clinical studies have demonstrated that NTF supplementation could attenuate the RGC loss, it is necessary to better understand the NTF-signaling pathways, as well as improving the delivery strategies with which NTFs are administered.

3.2.3. Novel Neuroprotective Agents

Cytidine 5'-diphosphocholine, better known as Citicoline, already finds its therapeutic use in systemic neurodegenerations, such as AD, PD and ischemia. In fact, citicoline, since its chemical structure resembles a precursor of phosphatidylcholine, has several action mechanisms, including phospholipid homeostasis, redox homeostasis, mitochondrial dynamics, cholinergic and dopaminergic neurotransmission [204–206]. Therefore, given the similarities between the neurodegeneration pattern of glaucoma and other neurodegenerative diseases (e.g., AD), citicoline has become increasingly used also in glaucoma studies showing its ability to stabilize the plasma membrane of RGC axons and to counteract glutamate excitotoxicity [207], as well as to control oxidative stress.

Synthetic Bile Salts (SBS), i.e., Ursodeoxycholic acid and tauroursodeoxycholic acid, are used for hepatic and cardiovascular diseases, and for neurological signaling both in physiological and pathological conditions [208–211]. Moreover, the biological potential of SBS also includes a role in stem-cell differentiation, in promoting the anti-apoptotic pathway, together with Unfolded Protein Response (UPR) signaling, as well as antioxidant activities [212–217].

Therefore, SBSs should also be considered as neuroprotective agents for their involvement in anti-inflammatory actions, acting as inhibitors of NF- κ B-signaling in glial cells and, at the same time, as promoters of the TGF β -cascade pathway [218,219].

Recently, further innovative retinal neuroprotective approaches, have been targeted to fight the numerous molecular events related to retinal damage. Among these, the antagonist of endothelin receptors [220] and nitric oxide donors [221] have been studied. In addition, also progesterone has been proposed since it plays multiple roles in neuroprotection in terms of axonal transport regulation, increase of NTs, modulation of pro-inflammatory processes, and reduction of GABA-induced neurotoxic effects [222–228].

Furthermore, cannabinoids have shown neuroprotective potential in various neurological disorders by modulating oxidative stress, the inflammatory response, and glutamate-induced excitotoxicity. The neuroprotective action is mediated by their capability to activate the PPAR α signaling, which inhibits the NF- κ B pathway, thus increasing the transcription of anti-inflammatory cytokines, as well as GDNF and BDNF neurotrophins [229–235].

It is important to underline that several other innovative neuroprotective approaches are currently being assessed (Figure 3). However, since almost all studies are based on animal models, attempts to perform clinical trials has resulted in many difficulties due to the results not necessarily reproducing those obtained in animal glaucoma models, which in themselves differ from one species to another. Moreover, some of these potential drugs have shown not only instability and difficulty in their delivery to the retina, but also several adverse outcomes [135,236].

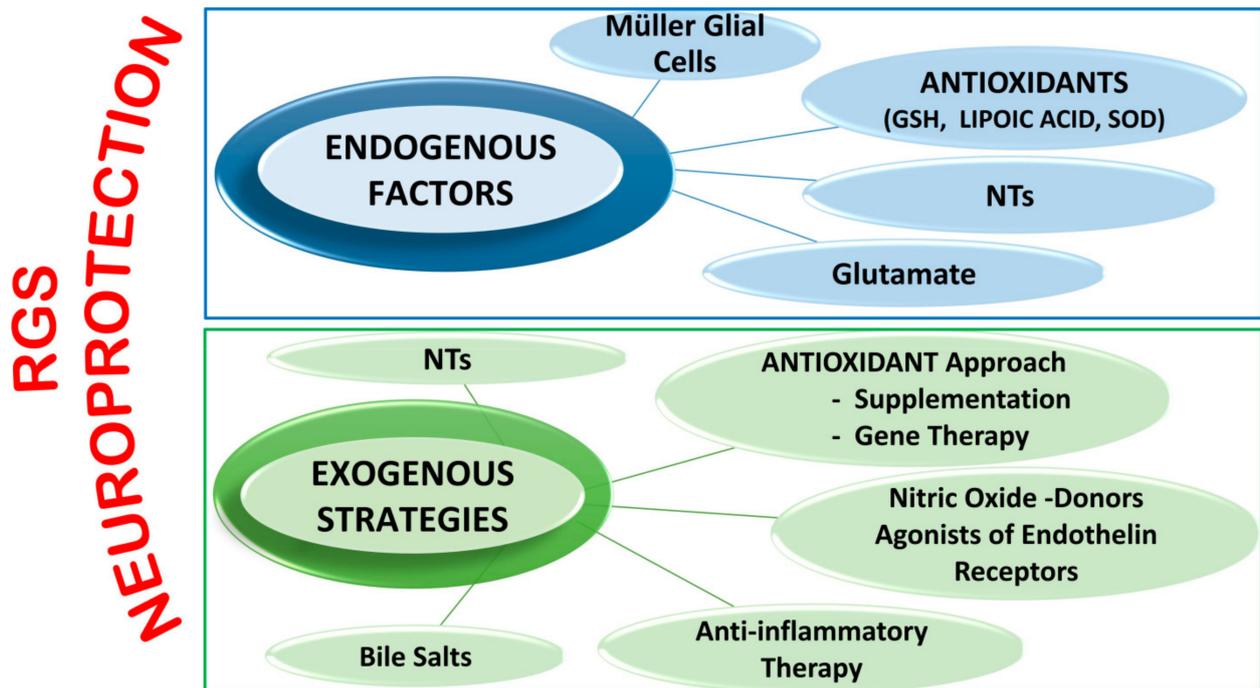


Figure 3. Neuroprotective mechanisms for preventing and slowing down RGC diseases.

Endogenous factors include Müller Glial Cells functions, antioxidant systems, NTs and glutamate.

Exogenous neuroprotective approaches include NTs and antioxidant supplementation or gene therapy, molecular donors of nitric oxide, agonists of endothelin receptors, drugs for counteracting inflammation pathway and bile salts.

4. Conclusions

Although the first-line treatment for patients with glaucoma is lowering the IOP, it should be taken into consideration that glaucoma manifests various and important underlying molecular changes, which, in a certain way, lead to RGC death.

Therefore, it is crucial to increasingly broaden our knowledge of the initial conditions that bring about glaucoma, such as those reported in this present review, to develop new therapeutic approaches.

Funding: Francesco Oddone was supported by the Italian Ministry of Health and by Fondazione Roma, Rome, Italy. Sara Tirendi was supported by Programma Operativo Por FSE, Liguria region 2014–2020, Italy.

Conflicts of Interest: The authors declare no conflict of interest.

References

1. You, M.; Rong, R.; Zeng, Z.; Xia, X.; Ji, D. Transneuronal Degeneration in the Brain During Glaucoma. *Front. Aging Neurosci.* **2021**, *13*, 643685. [[CrossRef](#)]
2. Quigley, H.A.; Broman, A.T. The Number of People with Glaucoma Worldwide in 2010 and 2020. *Br. J. Ophthalmol.* **2006**, *90*, 262–267. [[CrossRef](#)] [[PubMed](#)]
3. Kingman, S. Glaucoma Is Second Leading Cause of Blindness Globally. *Bull. World Health Organ.* **2004**, *82*, 887–888.
4. Calkins, D.J. Adaptive Responses to Neurodegenerative Stress in Glaucoma. *Prog. Retin. Eye Res.* **2021**, 100953. [[CrossRef](#)]
5. Almasieh, M.; Wilson, A.M.; Morquette, B.; Cueva Vargas, J.L.; Di Polo, A. The Molecular Basis of Retinal Ganglion Cell Death in Glaucoma. *Prog. Retin. Eye Res.* **2012**, *31*, 152–181. [[CrossRef](#)] [[PubMed](#)]
6. King, A.; Azuara-Blanco, A.; Tuulonen, A. Glaucoma. *BMJ* **2013**, *346*, f3518. [[CrossRef](#)]
7. Quaranta, L.; Bruttini, C.; Micheletti, E.; Konstas, A.G.P.; Michelessi, M.; Oddone, F.; Katsanos, A.; Sbardella, D.; De Angelis, G.; Riva, I. Glaucoma and Neuroinflammation: An Overview. *Surv. Ophthalmol.* **2021**, *66*, 693–713. [[CrossRef](#)] [[PubMed](#)]

8. Gordon, M.O. The Ocular Hypertension Treatment Study: Baseline Factors That Predict the Onset of Primary Open-Angle Glaucoma. *Arch. Ophthalmol.* **2002**, *120*, 714. [[CrossRef](#)] [[PubMed](#)]
9. Friedman, D.S.; Wilson, M.R.; Liebmann, J.M.; Fechtner, R.D.; Weinreb, R.N. An Evidence-Based Assessment of Risk Factors for the Progression of Ocular Hypertension and Glaucoma. *Am. J. Ophthalmol.* **2004**, *138*, 19–31. [[CrossRef](#)] [[PubMed](#)]
10. Fahmy, I.A.; Amer, A.K.; EL-Ghaffar, N.A. The Role of T-Cell Subsets and Natural Killer Lymphocytes in the Pathogenesis of Primary Open Angle Glaucoma. *Maced. J. Med. Sci.* **2010**, *3*, 307–313. [[CrossRef](#)]
11. Susanna, R.; Moraes, C.G.D.; Cioffi, G.A.; Ritch, R. Why Do People (Still) Go Blind from Glaucoma? *Transl. Vis. Sci. Technol.* **2015**, *4*. [[CrossRef](#)]
12. Wang, Y. Retinal Ganglion Cell-Derived Sonic Hedgehog Locally Controls Proliferation and the Timing of RGC Development in the Embryonic Mouse Retina. *Development* **2005**, *132*, 5103–5113. [[CrossRef](#)]
13. Dakubo, G.D. Retinal Ganglion Cell-Derived Sonic Hedgehog Signaling Is Required for Optic Disc and Stalk Neuroepithelial Cell Development. *Development* **2003**, *130*, 2967–2980. [[CrossRef](#)]
14. Miesfeld, J.B.; Ghiasvand, N.M.; Marsh-Armstrong, B.; Marsh-Armstrong, N.; Miller, E.B.; Zhang, P.; Manna, S.K.; Zawadzki, R.J.; Brown, N.L.; Glaser, T. The *Atoh7* Remote Enhancer Provides Transcriptional Robustness during Retinal Ganglion Cell Development. *Proc. Natl. Acad. Sci. USA* **2020**, *117*, 21690–21700. [[CrossRef](#)]
15. Wang, Y.P.; Dakubo, G.; Howley, P.; Campsall, K.D.; Mazarolle, C.J.; Shiga, S.A.; Lewis, P.M.; McMahon, A.P.; Wallace, V.A. Development of Normal Retinal Organization Depends on Sonic Hedgehog Signaling from Ganglion Cells. *Nat. Neurosci.* **2002**, *5*, 831–832. [[CrossRef](#)] [[PubMed](#)]
16. Bisiak, F.; McCarthy, A.A. Structure and Function of Roundabout Receptors. In *Macromolecular Protein Complexes II: Structure and Function*; Harris, J.R., Marles-Wright, J., Eds.; Subcellular Biochemistry; Springer International Publishing: Cham, Switzerland, 2019; Volume 93, pp. 291–319. ISBN 978-3-030-28150-2.
17. O’Sullivan, M.L.; Puñal, V.M.; Kerstein, P.C.; Brzezinski, J.A.; Glaser, T.; Wright, K.M.; Kay, J.N. Astrocytes Follow Ganglion Cell Axons to Establish an Angiogenic Template during Retinal Development. *Glia* **2017**, *65*, 1697–1716. [[CrossRef](#)]
18. Mead, B.; Tomarev, S. Evaluating Retinal Ganglion Cell Loss and Dysfunction. *Exp. Eye Res.* **2016**, *151*, 96–106. [[CrossRef](#)] [[PubMed](#)]
19. Duarte, J.N. Neuroinflammatory Mechanisms of Mitochondrial Dysfunction and Neurodegeneration in Glaucoma. *J. Ophthalmol.* **2021**, *2021*, 4581909. [[CrossRef](#)]
20. Ito, Y.A.; Di Polo, A. Mitochondrial Dynamics, Transport, and Quality Control: A Bottleneck for Retinal Ganglion Cell Viability in Optic Neuropathies. *Mitochondrion* **2017**, *36*, 186–192. [[CrossRef](#)]
21. Saccà, S.C.; Paluan, F.; Gandolfi, S.; Manni, G.; Cutolo, C.A.; Izzotti, A. Common Aspects between Glaucoma and Brain Neurodegeneration. *Mutat. Res. Mutat. Res.* **2020**, *786*, 108323. [[CrossRef](#)]
22. You, Y.; Gupta, V.K.; Li, J.C.; Klistorner, A.; Graham, S.L. Optic Neuropathies: Characteristic Features and Mechanisms of Retinal Ganglion Cell Loss. *Rev. Neurosci.* **2013**, *24*. [[CrossRef](#)]
23. Sharif, N. Glaucomatous Optic Neuropathy Treatment Options: The Promise of Novel Therapeutics, Techniques and Tools to Help Preserve Vision. *Neural Regen. Res.* **2018**, *13*, 1145. [[CrossRef](#)]
24. Millecamps, S.; Julien, J.-P. Axonal Transport Deficits and Neurodegenerative Diseases. *Nat. Rev. Neurosci.* **2013**, *14*, 161–176. [[CrossRef](#)]
25. Perlson, E.; Maday, S.; Fu, M.; Moughamian, A.J.; Holzbaur, E.L.F. Retrograde Axonal Transport: Pathways to Cell Death? *Trends Neurosci.* **2010**, *33*, 335–344. [[CrossRef](#)] [[PubMed](#)]
26. Lasek, R.J.; Garner, J.A.; Brady, S.T. Axonal Transport of the Cytoplasmic Matrix. *J. Cell Biol.* **1984**, *99*, 212s–221s. [[CrossRef](#)] [[PubMed](#)]
27. Morgan, J.E. Circulation and Axonal Transport in the Optic Nerve. *Eye* **2004**, *18*, 1089–1095. [[CrossRef](#)]
28. De Vos, K.J.; Grierson, A.J.; Ackerley, S.; Miller, C.C.J. Role of Axonal Transport in Neurodegenerative Diseases. *Annu. Rev. Neurosci.* **2008**, *31*, 151–173. [[CrossRef](#)]
29. Fahy, E.T.; Chrysostomou, V.; Crowston, J.G. Impaired Axonal Transport and Glaucoma. *Curr. Eye Res.* **2015**, 1–11. [[CrossRef](#)]
30. Hirokawa, N.; Noda, Y.; Tanaka, Y.; Niwa, S. Kinesin Superfamily Motor Proteins and Intracellular Transport. *Nat. Rev. Mol. Cell Biol.* **2009**, *10*, 682–696. [[CrossRef](#)]
31. LaMonte, B.H.; Wallace, K.E.; Holloway, B.A.; Shelly, S.S.; Ascaño, J.; Tokito, M.; Van Winkle, T.; Howland, D.S.; Holzbaur, E.L.F. Disruption of Dynein/Dynactin Inhibits Axonal Transport in Motor Neurons Causing Late-Onset Progressive Degeneration. *Neuron* **2002**, *34*, 715–727. [[CrossRef](#)]
32. Hafezparast, M. Mutations in Dynein Link Motor Neuron Degeneration to Defects in Retrograde Transport. *Science* **2003**, *300*, 808–812. [[CrossRef](#)]
33. Saha, A.R. Parkinson’s Disease -Synuclein Mutations Exhibit Defective Axonal Transport in Cultured Neurons. *J. Cell Sci.* **2004**, *117*, 1017–1024. [[CrossRef](#)]
34. Trushina, E.; Dyer, R.B.; Badger, J.D.; Ure, D.; Eide, L.; Tran, D.D.; Vrieze, B.T.; Legendre-Guillemain, V.; McPherson, P.S.; Mandavilli, B.S.; et al. Mutant Huntingtin Impairs Axonal Trafficking in Mammalian Neurons In Vivo and In Vitro. *Mol. Cell. Biol.* **2004**, *24*, 8195–8209. [[CrossRef](#)]

35. Dengler-Crish, C.M.; Smith, M.A.; Inman, D.M.; Wilson, G.N.; Young, J.W.; Crish, S.D. Anterograde Transport Blockade Precedes Deficits in Retrograde Transport in the Visual Projection of the DBA/2J Mouse Model of Glaucoma. *Front. Neurosci.* **2014**, *8*. [[CrossRef](#)]
36. Hirooka, K.; Yamamoto, T.; Kiuchi, Y. Dysfunction of Axonal Transport in Normal-Tension Glaucoma: A Biomarker of Disease Progression and a Potential Therapeutic Target. *Neural Regen. Res.* **2021**, *16*, 506. [[CrossRef](#)]
37. Ip, N.Y.; Yancopoulos, G.D. The Neurotrophins and CNTF: Two Families of Collaborative Neurotrophic Factors. *Annu. Rev. Neurosci.* **1996**, *19*, 491–515. [[CrossRef](#)]
38. Kimura, A.; Namekata, K.; Guo, X.; Harada, C.; Harada, T. Neuroprotection, Growth Factors and BDNF-TrkB Signalling in Retinal Degeneration. *Int. J. Mol. Sci.* **2016**, *17*, 1584. [[CrossRef](#)]
39. Bibel, M. Neurotrophins: Key Regulators of Cell Fate and Cell Shape in the Vertebrate Nervous System. *Genes Dev.* **2000**, *14*, 2919–2937. [[CrossRef](#)]
40. Platholi, J.; Lee, F.S. Neurotrophic Factors. In *Handbook of Developmental Neurotoxicology*; Elsevier: Amsterdam, The Netherlands, 2018; pp. 55–64. ISBN 978-0-12-809405-1.
41. Levi-Montalcini, R. The Nerve Growth Factor: Its Mode of Action on Sensory and Sympathetic Nerve Cells. *Harvey Lect.* **1966**, *60*, 217–259. [[PubMed](#)]
42. Levi-Montalcini, R.; Hamburger, V. A Diffusible Agent of Mouse Sarcoma, Producing Hyperplasia of Sympathetic Ganglia and Hyperneurotization of Viscera in the Chick Embryo. *J. Exp. Zool.* **1953**, *123*, 233–287. [[CrossRef](#)]
43. Barde, Y.A.; Edgar, D.; Thoenen, H. Purification of a New Neurotrophic Factor from Mammalian Brain. *EMBO J.* **1982**, *1*, 549–553. [[CrossRef](#)]
44. Maisonpierre, P.C.; Belluscio, L.; Friedman, B.; Alderson, R.F.; Wiegand, S.J.; Furth, M.E.; Lindsay, R.M.; Yancopoulos, G.D. NT-3, BDNF, and NGF in the Developing Rat Nervous System: Parallel as Well as Reciprocal Patterns of Expression. *Neuron* **1990**, *5*, 501–509. [[CrossRef](#)]
45. Ip, N.Y.; Ibanez, C.F.; Nye, S.H.; McClain, J.; Jones, P.F.; Gies, D.R.; Belluscio, L.; Le Beau, M.M.; Espinosa, R.; Squinto, S.P. Mammalian Neurotrophin-4: Structure, Chromosomal Localization, Tissue Distribution, and Receptor Specificity. *Proc. Natl. Acad. Sci. USA* **1992**, *89*, 3060–3064. [[CrossRef](#)] [[PubMed](#)]
46. Lessmann, V.; Gottmann, K.; Malsangio, M. Neurotrophin Secretion: Current Facts and Future Prospects. *Prog. Neurobiol.* **2003**, *69*, 341–374. [[CrossRef](#)]
47. Chao, M.V. Neurotrophins and Their Receptors: A Convergence Point for Many Signalling Pathways. *Nat. Rev. Neurosci.* **2003**, *4*, 299–309. [[CrossRef](#)] [[PubMed](#)]
48. Hempstead, B. The Many Faces of P75NTR. *Curr. Opin. Neurobiol.* **2002**, *12*, 260–267. [[CrossRef](#)]
49. Ibáñez, C.F.; Simi, A. P75 Neurotrophin Receptor Signaling in Nervous System Injury and Degeneration: Paradox and Opportunity. *Trends Neurosci.* **2012**, *35*, 431–440. [[CrossRef](#)] [[PubMed](#)]
50. Reichardt, L.F. Neurotrophin-Regulated Signalling Pathways. *Philos. Trans. R. Soc. B Biol. Sci.* **2006**, *361*, 1545–1564. [[CrossRef](#)] [[PubMed](#)]
51. Fathy, M.; Darweesh, A.M.; Sharaf, S.; El-Hanafi, H.M.; Ghaleb, F.M.; Fahmy, I.A.; Hussein, S.M. Brain-Derived Neurotrophic Factor (BDNF) Gene Polymorphism in a Cohort of Egyptian Primary Open-Angle Glaucoma (POAG) Patients. *Bull. Natl. Res. Cent.* **2020**, *44*, 45. [[CrossRef](#)]
52. Johnson, T.V.; Bull, N.D.; Martin, K.R. Neurotrophic Factor Delivery as a Protective Treatment for Glaucoma. *Exp. Eye Res.* **2011**, *93*, 196–203. [[CrossRef](#)]
53. Martin, K.R.G.; Quigley, H.A.; Zack, D.J.; Levkovitch-Verbin, H.; Kielczewski, J.; Valenta, D.; Baumrind, L.; Pease, M.E.; Klein, R.L.; Hauswirth, W.W. Gene Therapy with Brain-Derived Neurotrophic Factor As a Protection: Retinal Ganglion Cells in a Rat Glaucoma Model. *Investig. Ophthalmol. Vis. Sci.* **2003**, *44*, 4357. [[CrossRef](#)] [[PubMed](#)]
54. Pease, M.E.; McKinnon, S.J.; Quigley, H.A.; Kerrigan-Baumrind, L.A.; Zack, D.J. Obstructed Axonal Transport of BDNF and Its Receptor TrkB in Experimental Glaucoma. *Investig. Ophthalmol. Vis. Sci.* **2000**, *41*, 764–774.
55. Iwabe, S.; Moreno-Mendoza, N.A.; Trigo-Tavera, F.; Crowder, C.; García-Sánchez, G.A. Retrograde Axonal Transport Obstruction of Brain-Derived Neurotrophic Factor (BDNF) and Its TrkB Receptor in the Retina and Optic Nerve of American Cocker Spaniel Dogs with Spontaneous Glaucoma. *Vet. Ophthalmol.* **2007**, *10*, 12–19. [[CrossRef](#)] [[PubMed](#)]
56. Ko, M.-L.; Hu, D.-N.; Ritch, R.; Sharma, S.C.; Chen, C.-F. Patterns of Retinal Ganglion Cell Survival after Brain-Derived Neurotrophic Factor Administration in Hypertensive Eyes of Rats. *Neurosci. Lett.* **2001**, *305*, 139–142. [[CrossRef](#)]
57. Di Polo, A.; Aigner, L.J.; Dunn, R.J.; Bray, G.M.; Aguayo, A.J. Prolonged Delivery of Brain-Derived Neurotrophic Factor by Adenovirus-Infected Muller Cells Temporarily Rescues Injured Retinal Ganglion Cells. *Proc. Natl. Acad. Sci. USA* **1998**, *95*, 3978–3983. [[CrossRef](#)] [[PubMed](#)]
58. Coassin, M.; Lambiase, A.; Sposato, V.; Micera, A.; Bonini, S.; Aloe, L. Retinal P75 and Bax Overexpression Is Associated with Retinal Ganglion Cells Apoptosis in a Rat Model of Glaucoma. *Graefes Arch. Clin. Exp. Ophthalmol.* **2008**, *246*, 1743–1749. [[CrossRef](#)]
59. Sposato, V.; Bucci, M.G.; Coassin, M.; Russo, M.A.; Lambiase, A.; Aloe, L. Reduced NGF Level and TrkA Protein and TrkA Gene Expression in the Optic Nerve of Rats with Experimentally Induced Glaucoma. *Neurosci. Lett.* **2008**, *446*, 20–24. [[CrossRef](#)] [[PubMed](#)]
60. Guo, Y.; Johnson, E.; Cepurna, W.; Jia, L.; Dyck, J.; Morrison, J.C. Does Elevated Intraocular Pressure Reduce Retinal TRKB-Mediated Survival Signaling in Experimental Glaucoma? *Exp. Eye Res.* **2009**, *89*, 921–933. [[CrossRef](#)] [[PubMed](#)]

61. Walton, K.M. GDNF: A Novel Factor with Therapeutic Potential for Neurodegenerative Disorders. *Mol. Neurobiol.* **1999**, *19*, 43–59. [[CrossRef](#)]
62. Kramer, E.R.; Liss, B. GDNF-Ret Signaling in Midbrain Dopaminergic Neurons and Its Implication for Parkinson Disease. *FEBS Lett.* **2015**, *589*, 3760–3772. [[CrossRef](#)] [[PubMed](#)]
63. Budni, J.; Bellettini-Santos, T.; Mina, F.; Lima Garcez, M.; Ioppi Zugno, A. The Involvement of BDNF, NGF and GDNF in Aging and Alzheimer's Disease. *Aging Dis.* **2015**, *6*, 331. [[CrossRef](#)]
64. Frasson, M.; Picaud, S.; Léveillard, T.; Simonutti, M.; Mohand-Said, S.; Dreyfus, H.; Hicks, D.; Sahel, J. Glial Cell Line-Derived Neurotrophic Factor Induces Histologic and Functional Protection of Rod Photoreceptors in the Rd/Rd Mouse. *Investig. Ophthalmol. Vis. Sci.* **1999**, *40*, 2724–2734.
65. Harada, C.; Harada, T.; Quah, H.-M.A.; Maekawa, F.; Yoshida, K.; Ohno, S.; Wada, K.; Parada, L.F.; Tanaka, K. Potential Role of Glial Cell Line-Derived Neurotrophic Factor Receptors in Müller Glial Cells during Light-Induced Retinal Degeneration. *Neuroscience* **2003**, *122*, 229–235. [[CrossRef](#)]
66. Ward, M.S.; Khoobehi, A.; Lavik, E.B.; Langer, R.; Young, M.J. Neuroprotection of Retinal Ganglion Cells in DBA/2J Mice With GDNF-Loaded Biodegradable Microspheres. *J. Pharm. Sci.* **2007**, *96*, 558–568. [[CrossRef](#)]
67. Checa-Casalengua, P.; Jiang, C.; Bravo-Osuna, I.; Tucker, B.A.; Molina-Martínez, I.T.; Young, M.J.; Herrero-Vanrell, R. Retinal Ganglion Cells Survival in a Glaucoma Model by GDNF/Vit E PLGA Microspheres Prepared According to a Novel Microencapsulation Procedure. *J. Control. Release* **2011**, *156*, 92–100. [[CrossRef](#)]
68. Koeberle, P.D.; Bähr, M. The Upregulation of GLAST-1 Is an Indirect Antiapoptotic Mechanism of GDNF and Neurturin in the Adult CNS. *Cell Death Differ.* **2008**, *15*, 471–483. [[CrossRef](#)] [[PubMed](#)]
69. Fischer, D.; Leibinger, M. Promoting Optic Nerve Regeneration. *Prog. Retin. Eye Res.* **2012**, *31*, 688–701. [[CrossRef](#)] [[PubMed](#)]
70. Wang, Y.; Rhee, K.-D.; Pellegrini, M.; Yang, X.-J. Impacts of Ciliary Neurotrophic Factor on the Retinal Transcriptome in a Mouse Model of Photoreceptor Degeneration. *Sci. Rep.* **2020**, *10*, 6593. [[CrossRef](#)] [[PubMed](#)]
71. Ernst, M. Acquiring Signalling Specificity from the Cytokine Receptor Gp130. *Trends Genet.* **2004**, *20*, 23–32. [[CrossRef](#)]
72. Shpak, A.A.; Guekht, A.B.; Druzhkova, T.A.; Kozlova, K.I.; Gulyaeva, N.V. Ciliary Neurotrophic Factor in Patients with Primary Open-Angle Glaucoma and Age-Related Cataract. *Mol. Vis.* **2017**, *23*, 799–809.
73. Valter, K.; Bisti, S.; Gargini, C.; Di Loreto, S.; Maccarone, R.; Cervetto, L.; Stone, J. Time Course of Neurotrophic Factor Upregulation and Retinal Protection against Light-Induced Damage after Optic Nerve Section. *Investig. Ophthalmol. Vis. Sci.* **2005**, *46*, 1748. [[CrossRef](#)] [[PubMed](#)]
74. Cai, J.; Cheng, J.; Huang, X.; Li, Y.; Ma, X.; Li, Y.; Wei, R. Pathologic Changes in Chronic Intraorbital Optic Nerve Damage in Rabbits. *Brain Res.* **2009**, *1267*, 103–115. [[CrossRef](#)] [[PubMed](#)]
75. Bouvier, M.; Szatkowski, M.; Amato, A.; Attwell, D. The Glial Cell Glutamate Uptake Carrier Countertransports PH-Changing Anions. *Nature* **1992**, *360*, 471–474. [[CrossRef](#)] [[PubMed](#)]
76. Clements, J.; Lester, R.; Tong, G.; Jahr, C.; Westbrook, G. The Time Course of Glutamate in the Synaptic Cleft. *Science* **1992**, *258*, 1498–1501. [[CrossRef](#)]
77. Atoji, Y.; Sarkar, S. Localization of AMPA, Kainate, and NMDA Receptor MRNAs in the Pigeon Cerebellum. *J. Chem. Neuroanat.* **2019**, *98*, 71–79. [[CrossRef](#)]
78. Girling, K.D.; Demers, M.-J.; Laine, J.; Zhang, S.; Wang, Y.T.; Graham, R.K. Activation of Caspase-6 and Cleavage of Caspase-6 Substrates Is an Early Event in NMDA Receptor-Mediated Excitotoxicity. *J. Neurosci. Res.* **2018**, *96*, 391–406. [[CrossRef](#)] [[PubMed](#)]
79. Riedel, G. Glutamate Receptor Function in Learning and Memory. *Behav. Brain Res.* **2003**, *140*, 1–47. [[CrossRef](#)]
80. Hardingham, G.E. Pro-Survival Signalling from the NMDA Receptor. *Biochem. Soc. Trans.* **2006**, *34*, 936–938. [[CrossRef](#)]
81. Hetman, M.; Kharebava, G. Survival Signaling Pathways Activated by NMDA Receptors. *Curr. Top. Med. Chem.* **2006**, *6*, 787–799. [[CrossRef](#)]
82. Casson, R.J. Possible Role of Excitotoxicity in the Pathogenesis of Glaucoma. *Clin. Experiment. Ophthalmol.* **2006**, *34*, 54–63. [[CrossRef](#)]
83. Lalo, U. NMDA Receptors Mediate Neuron-to-Glia Signaling in Mouse Cortical Astrocytes. *J. Neurosci.* **2006**, *26*, 2673–2683. [[CrossRef](#)]
84. Salter, M.G.; Fern, R. NMDA Receptors Are Expressed in Developing Oligodendrocyte Processes and Mediate Injury. *Nature* **2005**, *438*, 1167–1171. [[CrossRef](#)]
85. Bylicky, M.A.; Mueller, G.P.; Day, R.M. Mechanisms of Endogenous Neuroprotective Effects of Astrocytes in Brain Injury. *Oxid. Med. Cell. Longev.* **2018**, *2018*, 6501031. [[CrossRef](#)]
86. Volterra, A.; Trotti, D.; Racagni, G. Glutamate Uptake Is Inhibited by Arachidonic Acid and Oxygen Radicals via Two Distinct and Additive Mechanisms. *Mol. Pharmacol.* **1994**, *46*, 986–992.
87. Harvey, B.K.; Airavaara, M.; Hinzman, J.; Wires, E.M.; Chiocco, M.J.; Howard, D.B.; Shen, H.; Gerhardt, G.; Hoffer, B.J.; Wang, Y. Targeted Over-Expression of Glutamate Transporter 1 (GLT-1) Reduces Ischemic Brain Injury in a Rat Model of Stroke. *PLoS ONE* **2011**, *6*, e22135. [[CrossRef](#)] [[PubMed](#)]
88. Zhang, L.-N.; Hao, L.; Guo, Y.-S.; Wang, H.-Y.; Li, L.; Liu, L.-Z.; Li, W.-B. Are Glutamate Transporters Neuroprotective or Neurodegenerative during Cerebral Ischemia? *J. Mol. Med.* **2019**, *97*, 281–289. [[CrossRef](#)] [[PubMed](#)]
89. Milewski, K.; Bogacińska-Karaś, M.; Hilgier, W.; Albrecht, J.; Zielińska, M. TNF α Increases STAT3-Mediated Expression of Glutaminase Isoform KGA in Cultured Rat Astrocytes. *Cytokine* **2019**, *123*, 154774. [[CrossRef](#)] [[PubMed](#)]
90. Kemp, J.A.; McKernan, R.M. NMDA Receptor Pathways as Drug Targets. *Nat. Neurosci.* **2002**, *5*, 1039–1042. [[CrossRef](#)] [[PubMed](#)]

91. Wang, R.; Reddy, P.H. Role of Glutamate and NMDA Receptors in Alzheimer's Disease. *J. Alzheimers Dis.* **2017**, *57*, 1041–1048. [[CrossRef](#)]
92. Kumagai, A.; Sasaki, T.; Matsuoka, K.; Abe, M.; Tabata, T.; Itoh, Y.; Fuchino, H.; Wugangerile, S.; Suga, M.; Yamaguchi, T.; et al. Monitoring of Glutamate-Induced Excitotoxicity by Mitochondrial Oxygen Consumption: XXXX. *Synapse* **2019**, *73*, e22067. [[CrossRef](#)]
93. Vernazza, S.; Tirendi, S.; Bassi, A.M.; Traverso, C.E.; Saccà, S.C. Neuroinflammation in Primary Open-Angle Glaucoma. *J. Clin. Med.* **2020**, *9*, 3172. [[CrossRef](#)]
94. Dreyer, E.B. Elevated Glutamate Levels in the Vitreous Body of Humans and Monkeys with Glaucoma. *Arch. Ophthalmol.* **1996**, *114*, 299. [[CrossRef](#)] [[PubMed](#)]
95. Dreyer, E.B. A Proposed Role for Excitotoxicity in Glaucoma. *J. Glaucoma* **1998**, *7*, 62–67. [[CrossRef](#)] [[PubMed](#)]
96. Honkanen, R.A. Vitreous Amino Acid Concentrations in Patients with Glaucoma Undergoing Vitrectomy. *Arch. Ophthalmol.* **2003**, *121*, 183. [[CrossRef](#)] [[PubMed](#)]
97. Wamsley, S. Vitreous Glutamate Concentration and Axon Loss in Monkeys with Experimental Glaucoma. *Arch. Ophthalmol.* **2005**, *123*, 64. [[CrossRef](#)]
98. Belov Kirdajova, D.; Kriska, J.; Tureckova, J.; Anderova, M. Ischemia-Triggered Glutamate Excitotoxicity from the Perspective of Glial Cells. *Front. Cell. Neurosci.* **2020**, *14*, 51. [[CrossRef](#)]
99. Dallas, M.; Boycott, H.E.; Atkinson, L.; Miller, A.; Boyle, J.P.; Pearson, H.A.; Peers, C. Hypoxia Suppresses Glutamate Transport in Astrocytes. *J. Neurosci.* **2007**, *27*, 3946–3955. [[CrossRef](#)]
100. Kaur, C. Hypoxia-Ischemia and Retinal Ganglion Cell Damage. *Clin. Ophthalmol.* **2008**, 879. [[CrossRef](#)]
101. Medzhitov, R. Origin and Physiological Roles of Inflammation. *Nature* **2008**, *454*, 428–435. [[CrossRef](#)]
102. Kimura, A.; Namekata, K.; Guo, X.; Noro, T.; Harada, C.; Harada, T. Targeting Oxidative Stress for Treatment of Glaucoma and Optic Neuritis. *Oxid. Med. Cell. Longev.* **2017**, *2017*, 2817252. [[CrossRef](#)]
103. Qi, Y.; Zhao, M.; Bai, Y.; Huang, L.; Yu, W.; Bian, Z.; Zhao, M.; Li, X. Retinal Ischemia/Reperfusion Injury Is Mediated by Toll-like Receptor 4 Activation of NLRP3 Inflammasomes. *Investig. Ophthalmol. Vis. Sci.* **2014**, *55*, 5466. [[CrossRef](#)] [[PubMed](#)]
104. Yoneda, S.; Tanihara, H.; Kido, N.; Honda, Y.; Goto, W.; Hara, H.; Miyawaki, N. Interleukin-1 β Mediates Ischemic Injury in the Rat Retina. *Exp. Eye Res.* **2001**, *73*, 661–667. [[CrossRef](#)]
105. Cohen, L. Relationships between Visual Function and Metabolism. In *Biochemistry of the Retina*; Graymore, C.N., Ed.; Academic Press: New York, NY, USA, 1965; pp. 36–50.
106. Tielsch, J.M.; Katz, J.; Sommer, A.; Quigley, H.A.; Javitt, J.C. Hypertension, Perfusion Pressure, and Primary Open-Angle Glaucoma: A Population-Based Assessment. *Arch. Ophthalmol.* **1995**, *113*, 216–221. [[CrossRef](#)]
107. Flammer, J. The Vascular Concept of Glaucoma. *Surv. Ophthalmol.* **1994**, *38*, S3–S6. [[CrossRef](#)]
108. Chung, H.S.; Harris, A.; Evans, D.W.; Kagemann, L.; Garzozzi, H.J.; Martin, B. Vascular Aspects in the Pathophysiology of Glaucomatous Optic Neuropathy. *Surv. Ophthalmol.* **1999**, *43*, S43–S50. [[CrossRef](#)]
109. Osborne, N.N.; Casson, R.J.; Wood, J.P.; Chidlow, G.; Graham, M.; Melena, J. Retinal Ischemia: Mechanisms of Damage and Potential Therapeutic Strategies. *Prog. Retin. Eye Res.* **2004**, *23*, 91–147. [[CrossRef](#)] [[PubMed](#)]
110. Abramov, A.Y.; Scorziello, A.; Duchon, M.R. Three Distinct Mechanisms Generate Oxygen Free Radicals in Neurons and Contribute to Cell Death during Anoxia and Reoxygenation. *J. Neurosci.* **2007**, *27*, 1129–1138. [[CrossRef](#)] [[PubMed](#)]
111. Vohra, R.; Tsai, J.C.; Kolko, M. The Role of Inflammation in the Pathogenesis of Glaucoma. *Surv. Ophthalmol.* **2013**, *58*, 311–320. [[CrossRef](#)]
112. Casson, R.J.; Chidlow, G.; Crowston, J.G.; Williams, P.A.; Wood, J.P.M. Retinal Energy Metabolism in Health and Glaucoma. *Prog. Retin. Eye Res.* **2020**, 100881. [[CrossRef](#)]
113. Yang, X.; Yu, X.-W.; Zhang, D.-D.; Fan, Z.-G. Blood-Retinal Barrier as a Converging Pivot in Understanding the Initiation and Development of Retinal Diseases. *Chin. Med. J.* **2020**, *133*, 2586–2594. [[CrossRef](#)] [[PubMed](#)]
114. Jiang, S.; Kametani, M.; Chen, D.F. Adaptive Immunity: New Aspects of Pathogenesis Underlying Neurodegeneration in Glaucoma and Optic Neuropathy. *Front. Immunol.* **2020**, *11*, 65. [[CrossRef](#)] [[PubMed](#)]
115. Yan, X. Matrix Metalloproteinases and Tumor Necrosis Factor α in Glaucomatous Optic Nerve Head. *Arch. Ophthalmol.* **2000**, *118*, 666. [[CrossRef](#)] [[PubMed](#)]
116. Tezel, G.; Li, L.Y.; Patil, R.V.; Wax, M.B. TNF- α and TNF- α Receptor-1 in the Retina of Normal and Glaucomatous Eyes. *Investig. Ophthalmol. Vis. Sci.* **2001**, *42*, 1787–1794.
117. Tezel, G. TNF- α signaling in glaucomatous neurodegeneration. In *Progress in Brain Research*; Elsevier: Amsterdam, The Netherlands, 2008; Volume 173, pp. 409–421. ISBN 978-0-444-53256-5.
118. Yang, X.; Luo, C.; Cai, J.; Powell, D.W.; Yu, D.; Kuehn, M.H.; Tezel, G. Neurodegenerative and Inflammatory Pathway Components Linked to TNF- α /TNFR1 Signaling in the Glaucomatous Human Retina. *Investig. Ophthalmol. Vis. Sci.* **2011**, *52*, 8442. [[CrossRef](#)]
119. Luna, J.D.; Chan, C.-C.; Derevjani, N.L.; Mahlow, J.; Chiu, C.; Peng, B.; Tobe, T.; Campochiaro, P.A.; Vinoses, S.A. Blood-Retinal Barrier (BRB) Breakdown in Experimental Autoimmune Uveoretinitis: Comparison with Vascular Endothelial Growth Factor, Tumor Necrosis Factor α , and Interleukin-1 β -Mediated Breakdown. *J. Neurosci. Res.* **1997**, *49*, 268–280. [[CrossRef](#)]
120. Rolle, T.; Ponzetto, A.; Malinverni, L. The Role of Neuroinflammation in Glaucoma: An Update on Molecular Mechanisms and New Therapeutic Options. *Front. Neurol.* **2021**, *11*, 612422. [[CrossRef](#)] [[PubMed](#)]

121. Chen, H.; Cho, K.-S.; Vu, T.H.K.; Shen, C.-H.; Kaur, M.; Chen, G.; Mathew, R.; McHam, M.L.; Fazelat, A.; Lashkari, K.; et al. Commensal Microflora-Induced T Cell Responses Mediate Progressive Neurodegeneration in Glaucoma. *Nat. Commun.* **2018**, *9*, 3209. [[CrossRef](#)] [[PubMed](#)]
122. Yang, X.; Zeng, Q.; Göktas, E.; Gopal, K.; Al-Aswad, L.; Blumberg, D.M.; Cioffi, G.A.; Liebmann, J.M.; Tezel, G. T-Lymphocyte Subset Distribution and Activity in Patients with Glaucoma. *Investig. Ophthalmol. Vis. Sci.* **2019**, *60*, 877. [[CrossRef](#)]
123. Schlereth, S.L.; Kremers, S.; Schrödl, F.; Cursiefen, C.; Heindl, L.M. Characterization of Antigen-Presenting Macrophages and Dendritic Cells in the Healthy Human Sclera. *Investig. Ophthalmol. Vis. Sci.* **2016**, *57*, 4878–4885. [[CrossRef](#)] [[PubMed](#)]
124. Howell, G.R.; Soto, I.; Zhu, X.; Ryan, M.; Macalinao, D.G.; Sousa, G.L.; Caddle, L.B.; MacNicol, K.H.; Barbay, J.M.; Porciatti, V. Radiation Treatment Inhibits Monocyte Entry into the Optic Nerve Head and Prevents Neuronal Damage in a Mouse Model of Glaucoma. *J. Clin. Investig.* **2012**, *122*, 1246–1261. [[CrossRef](#)] [[PubMed](#)]
125. Williams, P.A.; Marsh-Armstrong, N.; Howell, G.R.; Bosco, A.; Danias, J.; Simon, J.; Di Polo, A.; Kuehn, M.H.; Przedborski, S.; Raff, M.; et al. Neuroinflammation in Glaucoma: A New Opportunity. *Exp. Eye Res.* **2017**, *157*, 20–27. [[CrossRef](#)] [[PubMed](#)]
126. Hubens, W.H.G.; Beckers, H.J.M.; Gorgels, T.G.M.F.; Webers, C.A.B. Increased Ratios of Complement Factors C3a to C3 in Aqueous Humor and Serum Mark Glaucoma Progression. *Exp. Eye Res.* **2021**, *204*, 108460. [[CrossRef](#)]
127. Samelska, K.; Zaleska-Żmijewska, A.; Bałan, B.; Grąbczewski, A.; Szaflik, J.; Kubiak, A.; Skopiński, P. Immunological and Molecular Basics of the Primary Open Angle Glaucoma Pathomechanism. *Cent. Eur. J. Immunol.* **2021**. [[CrossRef](#)] [[PubMed](#)]
128. Kuehn, M.H.; Kim, C.Y.; Ostojic, J.; Bellin, M.; Alward, W.L.M.; Stone, E.M.; Sakaguchi, D.S.; Grozdanic, S.D.; Kwon, Y.H. Retinal Synthesis and Deposition of Complement Components Induced by Ocular Hypertension. *Exp. Eye Res.* **2006**, *83*, 620–628. [[CrossRef](#)] [[PubMed](#)]
129. Noris, M.; Remuzzi, G. Overview of Complement Activation and Regulation. *Semin. Nephrol.* **2013**, *33*, 479–492. [[CrossRef](#)] [[PubMed](#)]
130. Ling, M.; Murali, M. Analysis of the Complement System in the Clinical Immunology Laboratory. *Clin. Lab. Med.* **2019**, *39*, 579–590. [[CrossRef](#)] [[PubMed](#)]
131. Chang, E.E.; Goldberg, J.L. Glaucoma 2.0: Neuroprotection, Neuroregeneration, Neuroenhancement. *Ophthalmology* **2012**, *119*, 979–986. [[CrossRef](#)] [[PubMed](#)]
132. Chen, W.W.; Wang, N.; Cai, S.; Fang, Z.; Yu, M.; Wu, Q.; Tang, L.; Guo, B.; Feng, Y.; Jonas, J.B.; et al. Structural Brain Abnormalities in Patients with Primary Open-Angle Glaucoma: A Study with 3T MR Imaging. *Investig. Ophthalmol. Vis. Sci.* **2013**, *54*, 545–554. [[CrossRef](#)]
133. Shen, J.; Wang, Y.; Yao, K. Protection of Retinal Ganglion Cells in Glaucoma: Current Status and Future. *Exp. Eye Res.* **2021**, *205*, 108506. [[CrossRef](#)] [[PubMed](#)]
134. Chen, S.; Wang, L.; Zhang, X. Neuroprotection in Glaucoma: Present and Future. *Chin. Med. J.* **2013**, *126*, 1567–1577. [[CrossRef](#)]
135. Naik, S.; Pandey, A.; Lewis, S.A.; Rao, B.S.S.; Mutalik, S. Neuroprotection: A Versatile Approach to Combat Glaucoma. *Eur. J. Pharmacol.* **2020**, *881*, 173208. [[CrossRef](#)] [[PubMed](#)]
136. Pardue, M.T.; Allen, R.S. Neuroprotective Strategies for Retinal Disease. *Prog. Retin. Eye Res.* **2018**, *65*, 50–76. [[CrossRef](#)] [[PubMed](#)]
137. Osborne, N.N.; Núñez-Álvarez, C.; Joglar, B.; Del Olmo-Aguado, S. Glaucoma: Focus on Mitochondria in Relation to Pathogenesis and Neuroprotection. *Eur. J. Pharmacol.* **2016**, *787*, 127–133. [[CrossRef](#)] [[PubMed](#)]
138. Ju, W.-K.; Kim, K.-Y.; Lindsey, J.D.; Angert, M.; Patel, A.; Scott, R.T.; Liu, Q.; Crowston, J.G.; Ellisman, M.H.; Perkins, G.A.; et al. Elevated Hydrostatic Pressure Triggers Release of OPA1 and Cytochrome C, and Induces Apoptotic Cell Death in Differentiated RGC-5 Cells. *Mol. Vis.* **2009**, *15*, 120–134. [[PubMed](#)]
139. Wang, Y.X.; Jonas, J.B.; Wang, N.; You, Q.S.; Yang, D.; Xie, X.B.; Xu, L. Intraocular Pressure and Estimated Cerebrospinal Fluid Pressure. The Beijing Eye Study 2011. *PLoS ONE* **2014**, *9*, e104267. [[CrossRef](#)]
140. Jonas, J.B.; Yang, D.; Wang, N. Einfluss des Liquordrucks auf die glaukomatöse Schädigung des Nervus opticus. *Ophthalmol.* **2014**, *111*, 181–190. [[CrossRef](#)] [[PubMed](#)]
141. Izzotti, A.; Bagnis, A.; Saccà, S.C. The Role of Oxidative Stress in Glaucoma. *Mutat. Res. Mutat. Res.* **2006**, *612*, 105–114. [[CrossRef](#)]
142. Costa, V.P.; Harris, A.; Stefánsson, E.; Flammer, J.; Krieglstein, G.K.; Orzalesi, N.; Heijl, A.; Renard, J.-P.; Serra, L.M. The Effects of Antiglaucoma and Systemic Medications on Ocular Blood Flow. *Prog. Retin. Eye Res.* **2003**, *22*, 769–805. [[CrossRef](#)]
143. García-Bermúdez, M.Y.; Freude, K.K.; Mouhammad, Z.A.; van Wijngaarden, P.; Martin, K.K.; Kolko, M. Glial Cells in Glaucoma: Friends, Foes, and Potential Therapeutic Targets. *Front. Neurol.* **2021**, *12*, 624983. [[CrossRef](#)] [[PubMed](#)]
144. Chao, T.I.; Grosche, J.; Friedrich, K.J.; Biedermann, B.; Francke, M.; Pannicke, T.; Reichelt, W.; Wulst, M.; Mühle, C.; Pritz-Hohmeier, S.; et al. Comparative Studies on Mammalian Müller (Retinal Glial) Cells. *J. Neurocytol.* **1997**, *26*, 439–454. [[CrossRef](#)] [[PubMed](#)]
145. Eastlake, K.; Luis, J.; Limb, G.A. Potential of Müller Glia for Retina Neuroprotection. *Curr. Eye Res.* **2020**, *45*, 339–348. [[CrossRef](#)]
146. Taylor, S.; Srinivasan, B.; Wordinger, R.J.; Roque, R.S. Glutamate Stimulates Neurotrophin Expression in Cultured Müller Cells. *Mol. Brain Res.* **2003**, *111*, 189–197. [[CrossRef](#)]
147. Wahlin, K.J.; Campochiaro, P.A.; Zack, D.J.; Adler, R. Neurotrophic Factors Cause Activation of Intracellular Signaling Pathways in Muller Cells and Other Cells of the Inner Retina, but Not Photoreceptors. *Investig. Ophthalmol. Vis. Sci.* **2000**, *41*, 927–936.
148. Boss, J.D.; Singh, P.K.; Pandya, H.K.; Tosi, J.; Kim, C.; Tewari, A.; Juzych, M.S.; Abrams, G.W.; Kumar, A. Assessment of Neurotrophins and Inflammatory Mediators in Vitreous of Patients with Diabetic Retinopathy. *Investig. Ophthalmol. Vis. Sci.* **2017**, *58*, 5594. [[CrossRef](#)]

149. Huster, D.; Reichenbach, A.; Reichelt, W. The Glutathione Content of Retinal Müller (Glial) Cells: Effect of Pathological Conditions. *Neurochem. Int.* **2000**, *36*, 461–469. [[CrossRef](#)]
150. Pow, D.V.; Crook, D.K. Immunocytochemical Evidence for the Presence of High Levels of Reduced Glutathione in Radial Glial Cells and Horizontal Cells in the Rabbit Retina. *Neurosci. Lett.* **1995**, *193*, 25–28. [[CrossRef](#)]
151. Inoue, Y.; Shimazawa, M.; Noda, Y.; Nagano, R.; Otsuka, T.; Kuse, Y.; Nakano, Y.; Tsuruma, K.; Nakagami, Y.; Hara, H. RS9, a Novel Nrf2 Activator, Attenuates Light-Induced Death of Cells of Photoreceptor Cells and Müller Glia Cells. *J. Neurochem.* **2017**, *141*, 750–765. [[CrossRef](#)] [[PubMed](#)]
152. Xu, Z.; Wei, Y.; Gong, J.; Cho, H.; Park, J.K.; Sung, E.-R.; Huang, H.; Wu, L.; Eberhart, C.; Handa, J.T.; et al. NRF2 Plays a Protective Role in Diabetic Retinopathy in Mice. *Diabetologia* **2014**, *57*, 204–213. [[CrossRef](#)] [[PubMed](#)]
153. Kato, S.; Ishita, S.; Sugawara, K.; Mawatari, K. Cystine/Glutamate Antiporter Expression in Retinal Müller Glial Cells: Implications for α -Aminoacidopate Toxicity. *Neuroscience* **1993**, *57*, 473–482. [[CrossRef](#)]
154. Tomi, M.; Funaki, T.; Abukawa, H.; Katayama, K.; Kondo, T.; Ohtsuki, S.; Ueda, M.; Obinata, M.; Terasaki, T.; Hosoya, K.-I. Expression and Regulation of L-Cystine Transporter, System Xc⁻, in the Newly Developed Rat Retinal Müller Cell Line (TR-MUL). *Glia* **2003**, *43*, 208–217. [[CrossRef](#)]
155. Lewerenz, J.; Klein, M.; Methner, A. Cooperative Action of Glutamate Transporters and Cystine/Glutamate Antiporter System Xc⁻ Protects from Oxidative Glutamate Toxicity. *J. Neurochem.* **2006**, *98*, 916–925. [[CrossRef](#)] [[PubMed](#)]
156. Martis, R.M.; Knight, L.J.; Donaldson, P.J.; Lim, J.C. Identification, Expression, and Roles of the Cystine/Glutamate Antiporter in Ocular Tissues. *Oxid. Med. Cell. Longev.* **2020**, *2020*, 4594606. [[CrossRef](#)]
157. Albrecht, P.; Lewerenz, J.; Dittmer, S.; Noack, R.; Maher, P.; Methner, A. Mechanisms of Oxidative Glutamate Toxicity: The Glutamate/Cystine Antiporter System Xc⁻ as a Neuroprotective Drug Target. *CNS Neurol. Disord.-Drug Targets* **2010**, *9*, 373–382. [[CrossRef](#)]
158. Derouiche, A.; Rauen, T. Coincidence of L-Glutamate/L-Aspartate Transporter (GLAST) and Glutamine Synthetase (GS) Immunoreactions in Retinal Glia: Evidence for Coupling of GLAST and GS in Transmitter Clearance. *J. Neurosci. Res.* **1995**, *42*, 131–143. [[CrossRef](#)] [[PubMed](#)]
159. Biedermann, B.; Bringmann, A.; Reichenbach, A. High-Affinity GABA Uptake in Retinal Glial (Müller) Cells of the Guinea Pig: Electrophysiological Characterization, Immunohistochemical Localization, and Modeling of Efficiency: Gaba Uptake in Müller Glial Cells. *Glia* **2002**, *39*, 217–228. [[CrossRef](#)] [[PubMed](#)]
160. Bringmann, A.; Grosche, A.; Pannicke, T.; Reichenbach, A. GABA and Glutamate Uptake and Metabolism in Retinal Glial (Müller) Cells. *Front. Endocrinol.* **2013**, *4*. [[CrossRef](#)]
161. Uga, S.; Smelser, G.K. Comparative Study of the Fine Structure of Retinal Müller Cells in Various Vertebrates. *Investig. Ophthalmol.* **1973**, *12*, 434–448.
162. Tsacopoulos, M.; Poitry-Yamate, C.L.; MacLeish, P.R.; Poitry, S. Trafficking of Molecules and Metabolic Signals in the Retina. *Prog. Retin. Eye Res.* **1998**, *17*, 429–442. [[CrossRef](#)]
163. Poitry-Yamate, C.L.; Poitry, S.; Tsacopoulos, M. Lactate Released by Müller Glial Cells Is Metabolized by Photoreceptors from Mammalian Retina. *J. Neurosci. Off. J. Soc. Neurosci.* **1995**, *15*, 5179–5191. [[CrossRef](#)]
164. Poitry-Yamate, C.L.; Tsacopoulos, M. Glucose Metabolism in Freshly Isolated Müller Glial Cells from a Mammalian Retina. *J. Comp. Neurol.* **1992**, *320*, 257–266. [[CrossRef](#)] [[PubMed](#)]
165. Reichenbach, A.; Bringmann, A. New Functions of Müller Cells. *Glia* **2013**, *61*, 651–678. [[CrossRef](#)] [[PubMed](#)]
166. Liu, Y.; Wang, C.; Su, G. Cellular Signaling in Müller Glia: Progenitor Cells for Regenerative and Neuroprotective Responses in Pharmacological Models of Retinal Degeneration. *J. Ophthalmol.* **2019**, *2019*, 5743109. [[CrossRef](#)] [[PubMed](#)]
167. Sun, Y.; Zheng, Y.; Wang, C.; Liu, Y. Glutathione Depletion Induces Ferroptosis, Autophagy, and Premature Cell Senescence in Retinal Pigment Epithelial Cells. *Cell Death Dis.* **2018**, *9*, 753. [[CrossRef](#)] [[PubMed](#)]
168. Paasche, G.; Huster, D.; Reichenbach, A. The Glutathione Content of Retinal Müller (Glial) Cells: The Effects of Aging and of Application of Free-Radical Scavengers. *Ophthalmic Res.* **1998**, *30*, 351–360. [[CrossRef](#)] [[PubMed](#)]
169. Aydemir, O.; Celebi, S.; Yilmaz, T.; Yekeler, H.; Kükner, A.S. Protective Effects of Vitamin E Forms (Alpha-Tocopherol, Gamma-Tocopherol and d-Alpha-Tocopherol Polyethylene Glycol 1000 Succinate) on Retinal Edema during Ischemia-Reperfusion Injury in the Guinea Pig Retina. *Int. Ophthalmol.* **2004**, *25*, 283–289. [[CrossRef](#)] [[PubMed](#)]
170. Nucci, C.; Martucci, A.; Giannini, C.; Morrone, L.A.; Bagetta, G.; Mancino, R. Neuroprotective Agents in the Management of Glaucoma. *Eye* **2018**, *32*, 938–945. [[CrossRef](#)] [[PubMed](#)]
171. Russo, R.; Cavaliere, F.; Rombolà, L.; Gliozzi, M.; Cerulli, A.; Nucci, C.; Fazzi, E.; Bagetta, G.; Corasaniti, M.T.; Morrone, L.A. Rational Basis for the Development of Coenzyme Q10 as a Neurotherapeutic Agent for Retinal Protection. *Prog. Brain Res.* **2008**, *173*, 575–582.
172. Nebbioso, M.; Scarsella, G.; Tafani, M.; Pescosolido, N. Mechanisms of Ocular Neuroprotection by Antioxidant Molecules in Animal Models. *J. Biol. Regul. Homeost. Agents* **2013**, *27*, 197–209. [[PubMed](#)]
173. Jiang, W.; Tang, L.; Zeng, J.; Chen, B. Adeno-Associated Virus Mediated SOD Gene Therapy Protects the Retinal Ganglion Cells from Chronic Intraocular Pressure Elevation Induced Injury via Attenuating Oxidative Stress and Improving Mitochondrial Dysfunction in a Rat Model. *Am. J. Transl. Res.* **2016**, *8*, 799–810. [[PubMed](#)]
174. Perez, C.I.; Singh, K.; Lin, S. Relationship of Lifestyle, Exercise, and Nutrition with Glaucoma. *Curr. Opin. Ophthalmol.* **2019**, *30*, 82–88. [[CrossRef](#)] [[PubMed](#)]

175. Ramdas, W.D. The Relation between Dietary Intake and Glaucoma: A Systematic Review. *Acta Ophthalmol.* **2018**, *96*, 550–556. [[CrossRef](#)] [[PubMed](#)]
176. Braakhuis, A.; Raman, R.; Vaghefi, E. The Association between Dietary Intake of Antioxidants and Ocular Disease. *Diseases* **2017**, *5*, 3. [[CrossRef](#)]
177. Ramdas, W.D.; Schouten, J.S.A.G.; Webers, C.A.B. The Effect of Vitamins on Glaucoma: A Systematic Review and Meta-Analysis. *Nutrients* **2018**, *10*, 359. [[CrossRef](#)] [[PubMed](#)]
178. Gao, F.-J.; Zhang, S.-H.; Xu, P.; Yang, B.-Q.; Zhang, R.; Cheng, Y.; Zhou, X.-J.; Huang, W.-J.; Wang, M.; Chen, J.-Y.; et al. Quercetin Declines Apoptosis, Ameliorates Mitochondrial Function and Improves Retinal Ganglion Cell Survival and Function in In Vivo Model of Glaucoma in Rat and Retinal Ganglion Cell Culture In Vitro. *Front. Mol. Neurosci.* **2017**, *10*, 285. [[CrossRef](#)] [[PubMed](#)]
179. Shim, S.H.; Kim, J.M.; Choi, C.Y.; Kim, C.Y.; Park, K.H. Ginkgo Biloba Extract and Bilberry Anthocyanins Improve Visual Function in Patients with Normal Tension Glaucoma. *J. Med. Food* **2012**, *15*, 818–823. [[CrossRef](#)] [[PubMed](#)]
180. Xu, S.; Liu, S.; Yan, G. Lycium Barbarum Exerts Protection against Glaucoma-Like Injury Via Inhibition of MMP-9 Signaling In Vitro. *Med. Sci. Monit. Int. Med. J. Exp. Clin. Res.* **2019**, *25*, 9794–9800. [[CrossRef](#)]
181. Liu, L.; Sha, X.-Y.; Wu, Y.-N.; Chen, M.-T.; Zhong, J.-X. Lycium Barbarum Polysaccharides Protects Retinal Ganglion Cells against Oxidative Stress Injury. *Neural Regen. Res.* **2020**, *15*, 1526–1531. [[CrossRef](#)] [[PubMed](#)]
182. Bernstein, P.S.; Li, B.; Vachali, P.P.; Gorusupudi, A.; Shyam, R.; Henriksen, B.S.; Nolan, J.M. Lutein, Zeaxanthin, and Meso-Zeaxanthin: The Basic and Clinical Science Underlying Carotenoid-Based Nutritional Interventions against Ocular Disease. *Prog. Retin. Eye Res.* **2016**, *50*, 34–66. [[CrossRef](#)] [[PubMed](#)]
183. Hammond, B.R.; Fletcher, L.M.; Roos, F.; Wittwer, J.; Schalch, W. A Double-Blind, Placebo-Controlled Study on the Effects of Lutein and Zeaxanthin on Photostress Recovery, Glare Disability, and Chromatic Contrast. *Investig. Ophthalmol. Vis. Sci.* **2014**, *55*, 8583–8589. [[CrossRef](#)] [[PubMed](#)]
184. Age-Related Eye Disease Study 2 Research Group Lutein + Zeaxanthin and Omega-3 Fatty Acids for Age-Related Macular Degeneration: The Age-Related Eye Disease Study 2 (AREDS2) Randomized Clinical Trial. *JAMA* **2013**, *309*, 2005–2015. [[CrossRef](#)]
185. Davey, P.G.; Henderson, T.; Lem, D.W.; Weis, R.; Amonoo-Monney, S.; Evans, D.W. Visual Function and Macular Carotenoid Changes in Eyes with Retinal Drusen-An Open Label Randomized Controlled Trial to Compare a Micronized Lipid-Based Carotenoid Liquid Supplementation and AREDS-2 Formula. *Nutrients* **2020**, *12*, 3271. [[CrossRef](#)] [[PubMed](#)]
186. Lem, D.W.; Gierhart, D.L.; Davey, P.G. *Management of Diabetic Eye Disease Using Carotenoids and Nutrients*; IntechOpen: London, UK, 2021; ISBN 978-1-83968-865-2.
187. Choi, J.-S.; Kim, D.; Hong, Y.-M.; Mizuno, S.; Joo, C.-K. Inhibition of NNOS and COX-2 Expression by Lutein in Acute Retinal Ischemia. *Nutr. Burbank Los Angel. Cty. Calif* **2006**, *22*, 668–671. [[CrossRef](#)] [[PubMed](#)]
188. Fung, F.K.C.; Law, B.Y.K.; Lo, A.C.Y. Lutein Attenuates Both Apoptosis and Autophagy upon Cobalt (II) Chloride-Induced Hypoxia in Rat Müller Cells. *PLoS ONE* **2016**, *11*, e0167828. [[CrossRef](#)] [[PubMed](#)]
189. Li, S.-Y.; Fung, F.K.C.; Fu, Z.J.; Wong, D.; Chan, H.H.L.; Lo, A.C.Y. Anti-Inflammatory Effects of Lutein in Retinal Ischemic/Hypoxic Injury: In Vivo and in Vitro Studies. *Investig. Ophthalmol. Vis. Sci.* **2012**, *53*, 5976–5984. [[CrossRef](#)]
190. Zhang, C.; Wang, Z.; Zhao, J.; Li, Q.; Huang, C.; Zhu, L.; Lu, D. Neuroprotective Effect of Lutein on NMDA-Induced Retinal Ganglion Cell Injury in Rat Retina. *Cell. Mol. Neurobiol.* **2016**, *36*, 531–540. [[CrossRef](#)]
191. Szeto, H.H. Cell-Permeable, Mitochondrial-Targeted, Peptide Antioxidants. *AAPS J.* **2006**, *8*, E277–E283. [[CrossRef](#)] [[PubMed](#)]
192. Rocha, M.; Hernandez-Mijares, A.; Garcia-Malpartida, K.; Bañuls, C.; Bellod, L.; Victor, V.M. Mitochondria-Targeted Antioxidant Peptides. *Curr. Pharm. Des.* **2010**, *16*, 3124–3131. [[CrossRef](#)] [[PubMed](#)]
193. Zhao, K.; Zhao, G.-M.; Wu, D.; Soong, Y.; Birk, A.V.; Schiller, P.W.; Szeto, H.H. Cell-Permeable Peptide Antioxidants Targeted to Inner Mitochondrial Membrane Inhibit Mitochondrial Swelling, Oxidative Cell Death, and Reperfusion Injury. *J. Biol. Chem.* **2004**, *279*, 34682–34690. [[CrossRef](#)] [[PubMed](#)]
194. Wu, X.; Pang, Y.; Zhang, Z.; Li, X.; Wang, C.; Lei, Y.; Li, A.; Yu, L.; Ye, J. Mitochondria-Targeted Antioxidant Peptide SS-31 Mediates Neuroprotection in a Rat Experimental Glaucoma Model. *Acta Biochim. Biophys. Sin.* **2019**, *51*, 411–421. [[CrossRef](#)] [[PubMed](#)]
195. Vasudevan, S.K.; Gupta, V.; Crowston, J.G. Neuroprotection in Glaucoma. *Indian J. Ophthalmol.* **2011**, *59*, S102–S113. [[CrossRef](#)] [[PubMed](#)]
196. Sposato, V.; Parisi, V.; Manni, L.; Antonucci, M.T.; Fausto, V.D.; Sornelli, F.; Aloe, L. Glaucoma Alters the Expression of NGF and NGF Receptors in Visual Cortex and Geniculate Nucleus of Rats: Effect of Eye NGF Application. *Vis. Res.* **2009**, *49*, 54–63. [[CrossRef](#)]
197. Lambiase, A.; Aloe, L.; Centofanti, M.; Parisi, V.; Mantelli, F.; Colafrancesco, V.; Manni, G.L.; Bucci, M.G.; Bonini, S.; Levi-Montalcini, R. Experimental and Clinical Evidence of Neuroprotection by Nerve Growth Factor Eye Drops: Implications for Glaucoma. *Proc. Natl. Acad. Sci. USA* **2009**, *106*, 13469–13474. [[CrossRef](#)] [[PubMed](#)]
198. Wang, H.; Wang, R.; Thrimawithana, T.; Little, P.J.; Xu, J.; Feng, Z.-P.; Zheng, W. The Nerve Growth Factor Signaling and Its Potential as Therapeutic Target for Glaucoma. *BioMed Res. Int.* **2014**, *2014*, 759473. [[CrossRef](#)]
199. Capsoni, S.; Tiveron, C.; Vignone, D.; Amato, G.; Cattaneo, A. Dissecting the Involvement of Tropomyosin-Related Kinase A and P75 Neurotrophin Receptor Signaling in NGF Deficit-Induced Neurodegeneration. *Proc. Natl. Acad. Sci. USA* **2010**, *107*, 12299–12304. [[CrossRef](#)]
200. Ko, M.-L.; Hu, D.-N.; Ritch, R.; Sharma, S.C. The Combined Effect of Brain-Derived Neurotrophic Factor and a Free Radical Scavenger in Experimental Glaucoma. *Investig. Ophthalmol. Vis. Sci.* **2000**, *41*, 2967–2971.

201. Ji, J.-Z.; Elyaman, W.; Yip, H.K.; Lee, V.W.; Yick, L.-W.; Hugon, J.; So, K.-F. CNTF Promotes Survival of Retinal Ganglion Cells after Induction of Ocular Hypertension in Rats: The Possible Involvement of STAT3 Pathway. *Eur. J. Neurosci.* **2004**, *19*, 265–272. [[CrossRef](#)]
202. Jiang, C.; Moore, M.J.; Zhang, X.; Klassen, H.; Langer, R.; Young, M. Intravitreal Injections of GDNF-Loaded Biodegradable Microspheres Are Neuroprotective in a Rat Model of Glaucoma. *Mol. Vis.* **2007**, *13*, 1783–1792. [[PubMed](#)]
203. Schmeer, C.; Straten, G.; Kügler, S.; Gravel, C.; Bähr, M.; Isenmann, S. Dose-Dependent Rescue of Axotomized Rat Retinal Ganglion Cells by Adenovirus-Mediated Expression of Glial Cell-Line Derived Neurotrophic Factorin Vivo. *Eur. J. Neurosci.* **2002**, *15*, 637–643. [[CrossRef](#)]
204. Bhattacharya, A. Lipid Metabolism in Plants Under High Temperature. In *Effect of High Temperature on Crop Productivity and Metabolism of Macro Molecules*; Elsevier: Amsterdam, The Netherlands, 2019; pp. 311–389. ISBN 978-0-12-817562-0.
205. Oddone, F.; Rossetti, L.; Parravano, M.; Sbardella, D.; Coletta, M.; Ziccardi, L.; Roberti, G.; Carnevale, C.; Romano, D.; Manni, G.; et al. Citicoline in Ophthalmological Neurodegenerative Disease: A Comprehensive Review. *Pharmaceuticals* **2021**, *14*, 281. [[CrossRef](#)]
206. Matteucci, A.; Varano, M.; Gaddini, L.; Mallozzi, C.; Villa, M.; Pricci, F.; Malchiodi-Albedi, F. Neuroprotective Effects of Citicoline in Vitro Models of Retinal Neurodegeneration. *Int. J. Mol. Sci.* **2014**, *15*, 6286–6297. [[CrossRef](#)]
207. Skopiński, P.; Magdalena Radomska-Leśniewska, D.; Izdebska, J.; Kamińska, A.; Kupis, M.; Kubiak, A.; Samelska, K. New Perspectives of Immunomodulation and Neuroprotection in Glaucoma. *Cent. Eur. J. Immunol.* **2021**. [[CrossRef](#)] [[PubMed](#)]
208. Lazaridis, K.N.; Gores, G.J.; Lindor, K.D. Ursodeoxycholic Acid “Mechanisms of Action and Clinical Use in Hepatobiliary Disorders”. *J. Hepatol.* **2001**, *35*, 134–146. [[CrossRef](#)]
209. Duan, W.-M.; Rodrigues, C.M.P.; Zhao, L.-R.; Steer, C.J.; Low, W.C.; Rodrigues, C.M.P. Tauroursodeoxycholic Acid Improves the Survival and Function of Nigral Transplants in a Rat Model of Parkinson’s Disease. *Cell Transplant.* **2002**, *11*, 195–205. [[CrossRef](#)]
210. Keene, C.D.; Rodrigues, C.M.P.; Eich, T.; Chhabra, M.S.; Steer, C.J.; Low, W.C. Tauroursodeoxycholic Acid, a Bile Acid, Is Neuroprotective in a Transgenic Animal Model of Huntington’s Disease. *Proc. Natl. Acad. Sci. USA* **2002**, *99*, 10671–10676. [[CrossRef](#)] [[PubMed](#)]
211. Win, A.; Delgado, A.; Jadeja, R.N.; Martin, P.M.; Bartoli, M.; Thounaojam, M.C. Pharmacological and Metabolic Significance of Bile Acids in Retinal Diseases. *Biomolecules* **2021**, *11*, 292. [[CrossRef](#)] [[PubMed](#)]
212. Yoon, Y.M.; Lee, J.H.; Yun, S.P.; Han, Y.-S.; Yun, C.W.; Lee, H.J.; Noh, H.; Lee, S.-J.; Han, H.J.; Lee, S.H. Tauroursodeoxycholic Acid Reduces ER Stress by Regulating of Akt-Dependent Cellular Prion Protein. *Sci. Rep.* **2016**, *6*, 39838. [[CrossRef](#)] [[PubMed](#)]
213. Soares, R.; Ribeiro, F.F.; Xapelli, S.; Genebra, T.; Ribeiro, M.F.; Sebastião, A.M.; Rodrigues, C.M.P.; Solá, S. Tauroursodeoxycholic Acid Enhances Mitochondrial Biogenesis, Neural Stem Cell Pool, and Early Neurogenesis in Adult Rats. *Mol. Neurobiol.* **2018**, *55*, 3725–3738. [[CrossRef](#)]
214. Vang, S.; Longley, K.; Steer, C.J.; Low, W.C. The Unexpected Uses of Urso- and Tauroursodeoxycholic Acid in the Treatment of Non-Liver Diseases. *Glob. Adv. Health Med.* **2014**, *3*, 58–69. [[CrossRef](#)]
215. Omura, T.; Asari, M.; Yamamoto, J.; Oka, K.; Hoshina, C.; Maseda, C.; Awaya, T.; Tasaki, Y.; Shiono, H.; Yonezawa, A.; et al. Sodium Tauroursodeoxycholate Prevents Paraquat-Induced Cell Death by Suppressing Endoplasmic Reticulum Stress Responses in Human Lung Epithelial A549 Cells. *Biochem. Biophys. Res. Commun.* **2013**, *432*, 689–694. [[CrossRef](#)]
216. Gaspar, J.M.; Martins, A.; Cruz, R.; Rodrigues, C.M.P.; Ambrósio, A.F.; Santiago, A.R. Tauroursodeoxycholic Acid Protects Retinal Neural Cells from Cell Death Induced by Prolonged Exposure to Elevated Glucose. *Neuroscience* **2013**, *253*, 380–388. [[CrossRef](#)]
217. Oveson, B.C.; Iwase, T.; Hackett, S.F.; Lee, S.Y.; Usui, S.; Sedlak, T.W.; Snyder, S.H.; Campochiaro, P.A.; Sung, J.U. Constituents of Bile, Bilirubin and TUDCA, Protect against Oxidative Stress-Induced Retinal Degeneration. *J. Neurochem.* **2011**, *116*, 144–153. [[CrossRef](#)]
218. Noailles, A.; Fernández-Sánchez, L.; Lax, P.; Cuenca, N. Microglia Activation in a Model of Retinal Degeneration and TUDCA Neuroprotective Effects. *J. Neuroinflamm.* **2014**, *11*. [[CrossRef](#)]
219. Romero-Ramírez, L.; Nieto-Sampedro, M.; Barreda-Manso, M.A. Integrated Stress Response as a Therapeutic Target for CNS Injuries. *BioMed Res. Int.* **2017**, *2017*, 6953156. [[CrossRef](#)] [[PubMed](#)]
220. Howell, G.R.; MacNicoll, K.H.; Braine, C.E.; Soto, I.; Macalinao, D.G.; Sousa, G.L.; John, S.W.M. Combinatorial Targeting of Early Pathways Profoundly Inhibits Neurodegeneration in a Mouse Model of Glaucoma. *Neurobiol. Dis.* **2014**, *71*, 44–52. [[CrossRef](#)] [[PubMed](#)]
221. He, S.; Stankowska, D.L.; Ellis, D.Z.; Krishnamoorthy, R.R.; Yorio, T. Targets of Neuroprotection in Glaucoma. *J. Ocul. Pharmacol. Ther. Off. J. Assoc. Ocul. Pharmacol. Ther.* **2018**, *34*, 85–106. [[CrossRef](#)] [[PubMed](#)]
222. Gonzalez Deniselle, M.C.; Garay, L.; Gonzalez, S.; Guennoun, R.; Schumacher, M.; De Nicola, A.F. Progesterone Restores Retrograde Labeling of Cervical Motoneurons in Wobbler Mouse Motoneuron Disease. *Exp. Neurol.* **2005**, *195*, 518–523. [[CrossRef](#)] [[PubMed](#)]
223. González, S.L.; Coronel, M.F.; Raggio, M.C.; Labombarda, F. Progesterone Receptor-Mediated Actions and the Treatment of Central Nervous System Disorders: An up-Date of the Known and the Challenge of the Unknown. *Steroids* **2020**, *153*, 108525. [[CrossRef](#)]
224. Drew, P.D.; Chavis, J.A. Female Sex Steroids: Effects upon Microglial Cell Activation. *J. Neuroimmunol.* **2000**, *111*, 77–85. [[CrossRef](#)]
225. Allen, R.S.; Sayeed, I.; Oumarbaeva, Y.; Morrison, K.C.; Choi, P.H.; Pardue, M.T.; Stein, D.G. Progesterone Treatment Shows Greater Protection in Brain vs. Retina in a Rat Model of Middle Cerebral Artery Occlusion: Progesterone Receptor Levels May Play an Important Role. *Restor. Neurol. Neurosci.* **2016**, *34*, 947–963. [[CrossRef](#)]

226. Ishrat, T.; Sayeed, I.; Atif, F.; Hua, F.; Stein, D.G. Progesterone and Allopregnanolone Attenuate Blood-Brain Barrier Dysfunction Following Permanent Focal Ischemia by Regulating the Expression of Matrix Metalloproteinases. *Exp. Neurol.* **2010**, *226*, 183–190. [[CrossRef](#)]
227. Guo, L.; Salt, T.E.; Maass, A.; Luong, V.; Moss, S.E.; Fitzke, F.W.; Cordeiro, M.F. Assessment of Neuroprotective Effects of Glutamate Modulation on Glaucoma-Related Retinal Ganglion Cell Apoptosis in vivo. *Investig. Ophthalmol. Vis. Sci.* **2006**, *47*, 626–633. [[CrossRef](#)]
228. González, S.L.; Labombarda, F.; González Deniselle, M.C.; Guennoun, R.; Schumacher, M.; De Nicola, A.F. Progesterone Up-Regulates Neuronal Brain-Derived Neurotrophic Factor Expression in the Injured Spinal Cord. *Neuroscience* **2004**, *125*, 605–614. [[CrossRef](#)] [[PubMed](#)]
229. Kokona, D.; Georgiou, P.-C.; Kounenidakis, M.; Kiagiadaki, F.; Thermos, K. Endogenous and Synthetic Cannabinoids as Therapeutics in Retinal Disease. *Neural Plast.* **2016**, *2016*, 8373020. [[CrossRef](#)]
230. Rapino, C.; Tortolani, D.; Scipioni, L.; Maccarrone, M. Neuroprotection by (Endo)Cannabinoids in Glaucoma and Retinal Neurodegenerative Diseases. *Curr. Neuropharmacol.* **2018**, *16*, 959–970. [[CrossRef](#)] [[PubMed](#)]
231. Nucci, C.; Tartaglione, R.; Cerulli, A.; Mancino, R.; Spanò, A.; Cavaliere, F.; Rombolà, L.; Bagetta, G.; Corasaniti, M.T.; Morrone, L.A. Retinal Damage Caused by High Intraocular Pressure-Induced Transient Ischemia is Prevented by Coenzyme Q10 in Rat. In *International Review of Neurobiology*; Neuroinflammation in Neuronal Death and Repair; Academic Press: Cambridge, MA, USA, 2007; Volume 82, pp. 397–406.
232. Drew, P.D.; Xu, J.; Storer, P.D.; Chavis, J.A.; Racke, M.K. Peroxisome Proliferator-Activated Receptor Agonist Regulation of Glial Activation: Relevance to CNS Inflammatory Disorders. *Neurochem. Int.* **2006**, *49*, 183–189. [[CrossRef](#)] [[PubMed](#)]
233. Araújo, D.S.M.; Miya-Coreixas, V.S.; Pandolfo, P.; Calaza, K.C. Cannabinoid Receptors and TRPA1 on Neuroprotection in a Model of Retinal Ischemia. *Exp. Eye Res.* **2017**, *154*, 116–125. [[CrossRef](#)] [[PubMed](#)]
234. Hohmann, U.; Pelzer, M.; Kleine, J.; Hohmann, T.; Ghadban, C.; Dehghani, F. Opposite Effects of Neuroprotective Cannabinoids, Palmitoylethanolamide, and 2-Arachidonoylglycerol on Function and Morphology of Microglia. *Front. Neurosci.* **2019**, *13*, 1180. [[CrossRef](#)] [[PubMed](#)]
235. Kim, J.; Choi, J.Y.; Seo, J.; Choi, I.S. Neuroprotective Effect of Cannabidiol Against Hydrogen Peroxide in Hippocampal Neuron Culture. *Cannabis Cannabinoid Res.* **2020**, *6*, 40–47. [[CrossRef](#)] [[PubMed](#)]
236. Sharif, N.A. IDrugs and IDevelopments Discovery Research: Preclinical Assays, Techniques, and Animal Model Studies for Ocular Hypotensives and Neuroprotectants. *J. Ocul. Pharmacol. Ther.* **2018**, *34*, 7–39. [[CrossRef](#)] [[PubMed](#)]