

Article

# Characterization of two NMN deamidase mutants as possible probes for an NMN biosensor

Alessandra Camarca<sup>1</sup>, Gabriele Minazzato<sup>2</sup>, Angela Pennacchio<sup>1</sup>, Alessandro Capo<sup>1</sup>, Adolfo Amici<sup>3</sup>, Sabato D'Auria<sup>1,4\*</sup> and Nadia Raffaelli<sup>2\*</sup>

<sup>1</sup> Institute of Food Sciences, National Research Council, Via Roma 64, 83100 Avellino, Italy

<sup>2</sup> Department of Agricultural, Food and Environmental Sciences, Polytechnic University of Marche, Via Breccie Bianche, 60131 Ancona, Italy

<sup>3</sup> Department of Clinical Sciences DISCO, Section of Biochemistry, Polytechnic University of Marche, Via Breccie Bianche, 60131 Ancona, Italy

<sup>4</sup> Department of Biology, Agriculture and Food Science, CNR, Piazzale Aldo Moro, 7, 00125 Rome, Italy

\* Correspondence to: N. Raffaelli, Department of Agricultural, Food and Environmental Sciences, Via Breccie Bianche, 60131 Ancona, Italy, Fax: +39 71 2204 677, Tel: +39 71 2204 682, E-mail: [n.raffaelli@staff.univpm.it](mailto:n.raffaelli@staff.univpm.it);

S. D'Auria, Department of Biology, Agriculture and Food Science, CNR, Piazzale Aldo Moro, 7, 00125 Rome, Italy, Tel. +39-3683422770, [sabato.dauria@cnr.it](mailto:sabato.dauria@cnr.it)

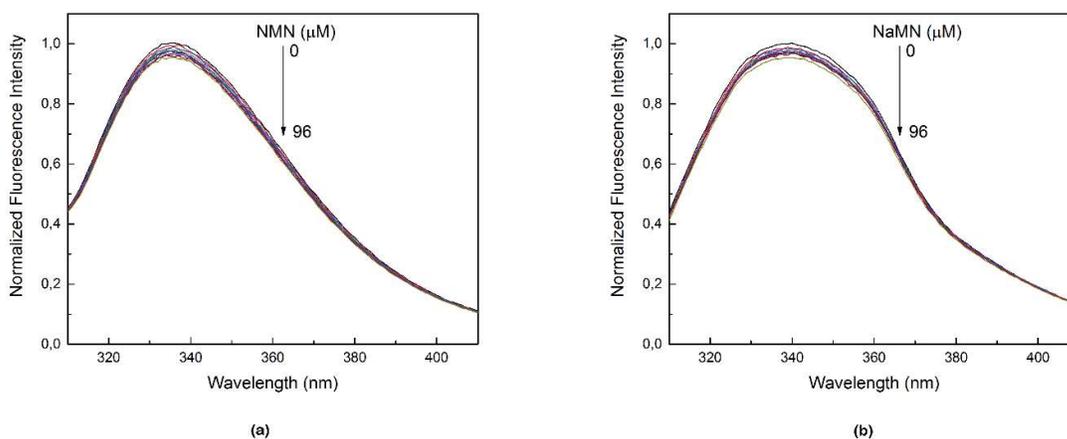
**Supplementary Table 1.** S29A PncC fluorescence quenching (%) vs buffer

	Nucleotides concentration [ $\mu\text{M}$ ]									
	0.185	0.375	0.75	1.5	3	6	12	24	48	96
<b>NMN</b>	0.48	0.58	2.58	10.1	17.42	17.55	27.06	37.52	46.79	52.82
<b>NaNM</b>	1.79	2.56	6.47	20.83	30.74	30.87	40.52	47.93	52.28	55.20
<b>Na</b>	0	0	0	0	0	0	0	0.16	0.62	0.24
<b>Nam</b>	0.08	0.18	0.48	1.49	2.51	2.65	3.29	3.29	3.71	5.01
<b>NR</b>	0	0.12	0.80	0.96	1.84	1.98	2.29	3.13	3.51	4.48
<b>NAD</b>	0.52	0.32	0.80	0	0.19	0.32	1.35	2.54	4.58	7.97
<b>NADP</b>	0	0	0.67	1.60	2.74	2.87	3.89	4.31	6.36	9.50
<b>NaAD</b>	0	0	0	0	0	0	0	0	1.97	6.69

**Supplementary Table 2.** K61Q PncC fluorescence quenching (%) vs buffer

	Nucleotides concentration [ $\mu\text{M}$ ]									
	0.185	0.375	0.75	1.5	3	6	12	24	48	96
<b>NMN</b>	1.09	1.84	3.19	4.87	6.26	8.11	12.12	20.07	38.03	53.35
<b>NaNM</b>	0.71	1.11	2.05	2.12	2.82	2.80	4.14	4.9	7.14	10.68
<b>Na</b>	0.49	1.2	2.11	2.66	2.89	2.81	3.26	2.74	3.69	4.46
<b>Nam</b>	0	0.34	0.87	0.68	0.91	0.26	1.15	1.04	1.14	1.57
<b>NR</b>	2.13	2.97	3.37	3.71	3.13	2.36	2.7	2.1	1.79	1.49
<b>NAD</b>	0.63	1.39	1.57	0.80	0.85	0	0.65	0.36	0.72	1.58
<b>NADP</b>	0	0.06	0.29	0.55	0.42	0	1.20	1.59	2.90	5.10
<b>NaAD</b>	0.65	1.02	1.68	2.19	2.25	2.39	3.72	4.13	5.77	8.28

**Supplementary table 1 and table 2:** S29A PncC and K61Q PncC fluorescence quenching (%) at increasing concentration of nucleotides. The percentages of reduction of fluorescence intensities compared to buffer, were calculated as follow: (normalized fluorescence in presence of analyte at a given [ $\mu\text{M}$ ]/normalized fluorescence in presence of corresponding volume of buffer) \*100.



**GlnBP fluorescence reduction (%) vs buffer**

	Nucleotide concentration [ $\mu\text{M}$ ]								
	0,375	0,75	1,5	3	6	12	24	48	96
<b>NMN</b>	1,21	1,34	0,689	0,882	2,354	2,657	4,212	3,842	4,362
<b>NaNM</b>	1,463	0,699	0,375	0,567	2,273	1,958	3,227	3,267	4,358

(c)

**Figure S1:** Effects of NMN and NaMN on steady-state fluorescence emission of GlnBP. GlnPB ( $3\mu\text{M}$ ) was incubated with increasing concentration of NMN, NaMN or buffer (not shown), as described for the PncC proteins. **(a)** and **(b)** steady-state fluorescence emission representative spectra **(c)** Percentages of reduction of fluorescence intensities compared to buffer, were calculated as follow: (normalized fluorescence in presence of analyte at a given [ $\mu\text{M}$ ]/normalized fluorescence in presence of corresponding volume of buffer) \*100.