

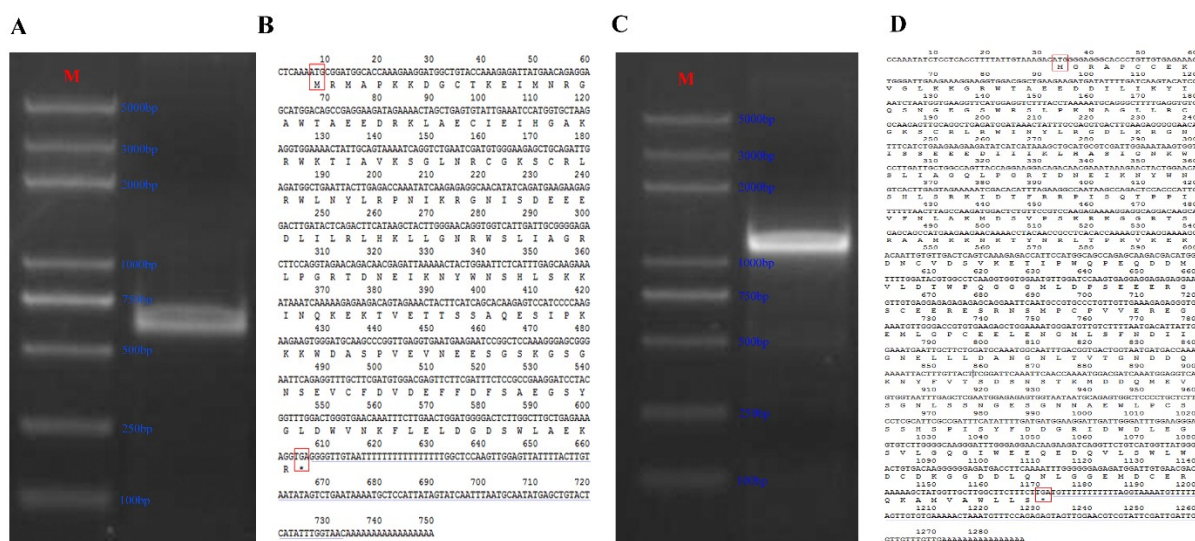


Figure S1. Full-length cDNA amplification of the *PsMYB114L* and *PsMYB12L* genes. **(A)** Results of PCR detection of the *PsMYB114L* gene. **(B)** The full-length cDNA sequence of the *PsMYB114L* gene. **(C)** Results of PCR detection of the *PsMYB12L* gene. **(D)** The full-length cDNA sequence of the *PsMYB12L* gene. ATG (start codon) and TAA (termination codon) are shown in the red box. M, Marker. The sequence of the 3' UTR is underlined in blue.



Figure S2. Flowers of *P. suffruticosa* 'Shima Nishiki' at five developmental stages. S1, flower bud emerging stage; S2, small bell-like flower-bud stage; S3, large bell-like flower-bud stage; S4, bell-like flower-bud extending stage; S5, color exposing stage.

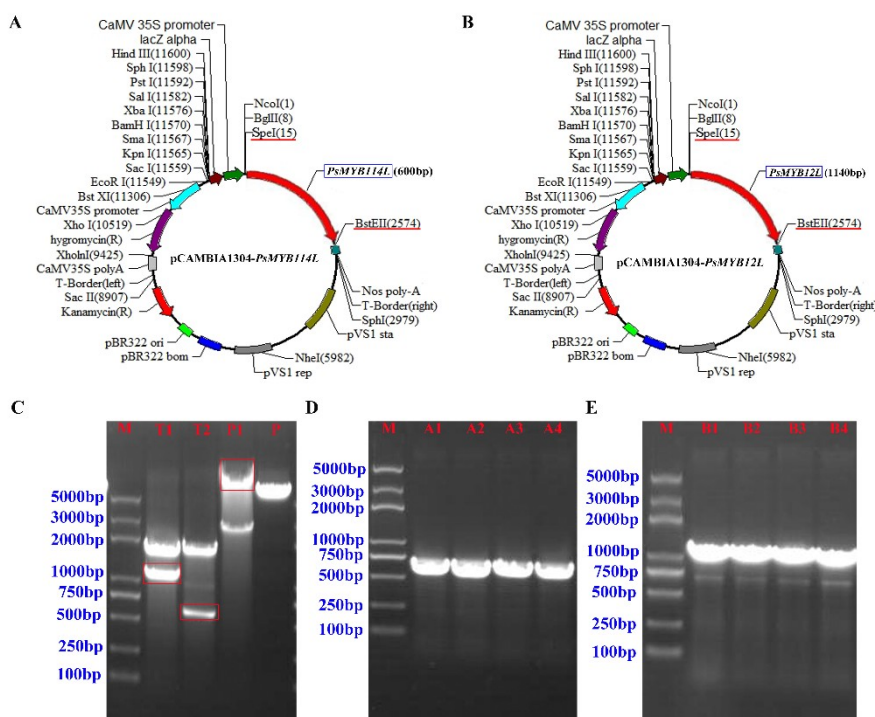


Figure S3. Construction of the recombinant expression vectors of pCambia1304-*PsMYB114L* and pCambia1304-*PsMYB12L*. Maps of the recombinant vectors pCambia1304-*PsMYB114L* (**A**) and pCambia1304-*PsMYB12L* (**B**). The restriction sites (*Spe* I and *BstE* II) are underlined in red. (**C**) Double digestion of the vector and target gene. P, The empty vector of pCambia1304; P1, Double digestion of the pCambia1304 empty vector; T1, Double digestion of the pTOPO-Blunt Simple vector carrying the *PsMYB12L* gene; T2, Double digestion of the pTOPO-Blunt Simple vector carrying the *PsMYB114L* gene. The fragments used to construct the recombinant vector are shown in the red box. (**D**) PCR verification of the recombinant pCambia1304-*PsMYB114L* vector. A1, A2, A3, and A4 are the four replicates used for PCR amplification. (**E**) PCR verification of the recombinant pCambia1304-*PsMYB12L* vector. B1, B2, B3, and B4 are the four replicates used for PCR amplification.

Table S1. Primers used in this study.

Primer name	Sequence (5'-3')	Description
<i>PsMYB114L</i> -1-F	GAGCAACATATCAGATGAAGAAGAGG	Primers used for 3'RACE amplification
<i>PsMYB114L</i> -2-F	CTACTTCATCAGCACAAGAGTCCATCC	
<i>PsMYB12L</i> -1-F	AATGGCAATTTGACGGTGACTGGT	
<i>PsMYB12L</i> -2-F	TGATGGAAGGATTGATTGGGATTTG	
B26	GACTCGAGTCGACATCGATTTTTTTTTTTTTTTTTT	Primers used for Full-length cDNA amplification
<i>PsMYB114L</i> -F1	ATGCGGATGGCACCAAGAAG	
<i>PsMYB114L</i> -R1	TCACCTTTTCTCAGCAAGCCAAG	
<i>PsMYB12L</i> -F1	ATGGGGAGGGCACCCTGTTGT	
<i>PsMYB12L</i> -R1	TCAAGAAAGAAGCCAAGCAACCAT	

<i>PsMYB114L</i> -GFPF	<u>GGTACCAT</u> GC GGATGGCACCAAAGAA	Primers used for subcellular localization vector construction
<i>PsMYB114L</i> -GFPR	<u>TCTAGAC</u> CTTTTCTCAGCAAGCCAAG	
<i>PsMYB12L</i> -GFPF	<u>GGTACCAT</u> TGGGGAGGGCACCTGTTGT	
<i>PsMYB12L</i> -GFPR	<u>TCTAGA</u> AAGAAAGAAGCCAAGCAACCA	
<i>PsMYB114L</i> -F2	<u>ACTAGT</u> ATGC GGATGGCACCAAAGAAG	Primers used for overexpression vector construction
<i>PsMYB114L</i> -R2	<u>GGTCACCT</u> CACCTTTTCTCAGCAAGCCAAG	
<i>PsMYB12L</i> -F2	<u>ACTAGT</u> ATGGGGAGGGCACCTGTTGT	
<i>PsMYB12L</i> -R2	<u>GGTCACCT</u> CAAGAAAGAAGCCAAGCAACCAT	
1304Ve-F	GGACTCTTGACCATGGTAGATCTG	Primers used for qRT-PCR
<i>PsCHS</i> -F	AGCAGAGAACAACAAAGGGTCACG	
<i>PsCHS</i> -R	TCAGCACCGACAATAACCGCAG	
<i>PsCHI</i> -F	TCCACCTGGTTCTTCTA	
<i>PsCHI</i> -R	AACTCTGCTTTGCTTCCG	
<i>PsF3H</i> -F	CCCAAGGTAGCCTACAACCAA	
<i>PsF3H</i> -R	GAAAATCCCCCAGTCTTCACA	
<i>PsF3'H</i> -F	AACTTGTTACGGCAGGGACT	
<i>PsF3'H</i> -R	GGCTTGGGCTAGGATTTTAGG	
<i>PsDFR</i> -F	TTCATCGGTTTCATGGCTTGTC	
<i>PsDFR</i> -R	AATGGGTATCCGCTTTTGGC	
<i>PsANS</i> -F	GCCCTCACTTTCATCCTCCACAAC	
<i>PsANS</i> -R	AAAAGTGGCCACGAAATCCTTACCT	
<i>PsFLS</i> -F	GAATGAACAGCCTGCAATCA	
<i>PsFLS</i> -R	TCTCGACTAGCCTCCACCAT	
<i>PsANR</i> -F	ATGGCTGGTCCCTTCACTTACT	
<i>PsANR</i> -R	CGAAATGGATATGGAACCTGA	
<i>PsMYB114L</i> -F	GGGAGCGGGAATTTCAGAGGTT	
<i>PsMYB114L</i> -R	AGCCAAGAGTCCCCATCCAG	
<i>PsMYB12L</i> -F	ATGGCAGCCAGAGCAAGACG	
<i>PsMYB12L</i> -R	TCAACAACAGGGCACGGCAT	
<i>PsUbiquitin</i> -F	GACCTATACCAAGCCGAAG	
<i>PsUbiquitin</i> -R	CGTTCCAGCACCACAATC	
<i>AtCHS</i> -F	ACGGACATTTGAGGGAAGTTGG	
<i>AtCHS</i> -R	AGGGTGGGCTATCCAGAAGAGG	
<i>AtCHI</i> -F	GCGGTTCTGGAATCTATCATCG	
<i>AtCHI</i> -R	TCGTCCTTGTTCTTCATCATTAGC	
<i>AtF3H</i> -F	CTGACCCTGGAACCTTACCTTG	
<i>AtF3H</i> -R	CAGCATTCCTTGAACCTCCCATT	
<i>AtF3'H</i> -F	ACACCGATGGAGACTGTTGAGAA	
<i>AtF3'H</i> -R	GCGTTAGCGTTCCAACCTCTTC	

<i>AtDFR-F</i>	GCCAAACGCCAAGACGCTA
<i>AtDFR-R</i>	CATTCACTGTCTGGCTTTATCACTTC
<i>AtANS-F</i>	AGGTTAGGATTTCTTGGGCTGTG
<i>AtANS-R</i>	CCGTGGAGGAACTTAGCCG
<i>AtFLS-F</i>	ACCGTTTGCTTTCAAGGATTACA
<i>AtFLS-R</i>	AAATGATCACCGATTTTTTTCCGTG
<i>AtANR-F</i>	TTCAAGAACTTGGCGACCTG
<i>AtANR-R</i>	CGGAGTTGCGACATGGAAGA
<i>AtActin2-F</i>	TGCTGGATTCTGGTGATGGT
<i>AtActin2-R</i>	AAGGTCAAGACGGAGGATGG
<i>MdCHS-F</i>	GGCAAGTGCTGTCGGATT
<i>MdCHS-R</i>	CCCAAAGAAATAACCACAAG
<i>MdCHI-F</i>	GCTACAAATGCGGTGATAG
<i>MdCHI-R</i>	CGCCTCCACTACAACCTCC
<i>MdF3H-F</i>	GCCGATCACCTACACCGAG
<i>MdF3H-R</i>	GTACAAGAAGTGGAAGGC
<i>MdF3'H-F</i>	TTGCGGATGGTGTCTCTGATG
<i>MdF3'H-R</i>	GCCTCGTCCATGTTCAACTTCTC
<i>MdDFR-F</i>	GTTGAGGGAGATAGGGTTTGAG
<i>MdDFR-R</i>	GGTAAATGTAAAACAATAGAGAGG
<i>MdANS-F</i>	GGAGAAGATCATCCTTAAGCCA
<i>MdANS-R</i>	CTAAGATATATCATACCAACTATGCC
<i>MdFLS-F</i>	GGATAAGACAAGAATCTCATGGC
<i>MdFLS-R</i>	CACACCACTCACAACTTTACC
<i>MdANR-F</i>	TCAACAAAAGATACCCCGAG
<i>MdANR-R</i>	GATAGCTAGCTCGATACATGC
<i>MdActin-F</i>	ACACGGGGAGGTAGTGACAA
<i>MdActin-R</i>	CCTCCAATGGATCCTCGTTA
