



Review

Chemical Basis of Reactive Oxygen Species Reactivity and Involvement in Neurodegenerative Diseases

Fabrice Collin

Laboratoire des IMRCP, Université de Toulouse, CNRS UMR 5623, Université Toulouse III-Paul Sabatier, 118 Route de Narbonne, 31062 Toulouse CEDEX 09, France; fabrice.collin@univ-tlse3.fr

Received: 26 April 2019; Accepted: 13 May 2019; Published: 15 May 2019

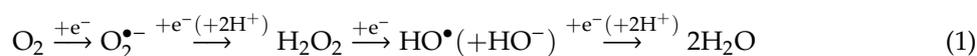


Abstract: Increasing numbers of individuals suffer from neurodegenerative diseases, which are characterized by progressive loss of neurons. Oxidative stress, in particular, the overproduction of Reactive Oxygen Species (ROS), play an important role in the development of these diseases, as evidenced by the detection of products of lipid, protein and DNA oxidation *in vivo*. Even if they participate in cell signaling and metabolism regulation, ROS are also formidable weapons against most of the biological materials because of their intrinsic nature. By nature too, neurons are particularly sensitive to oxidation because of their high polyunsaturated fatty acid content, weak antioxidant defense and high oxygen consumption. Thus, the overproduction of ROS in neurons appears as particularly deleterious and the mechanisms involved in oxidative degradation of biomolecules are numerous and complexes. This review highlights the production and regulation of ROS, their chemical properties, both from kinetic and thermodynamic points of view, the links between them, and their implication in neurodegenerative diseases.

Keywords: reactive oxygen species; superoxide anion; hydroxyl radical; hydrogen peroxide; hydroperoxides; neurodegenerative diseases; NADPH oxidase; superoxide dismutase

1. Introduction

Reactive Oxygen Species (ROS) are radical or molecular species whose physical-chemical properties are well-known both on thermodynamic and kinetic points of view. They are produced from molecular oxygen, during the successive 4 steps of 1-electron reduction (reaction (1)). The reaction occurs in particular in the mitochondrial respiratory chain, where 85% of O₂ is metabolized and where partially reduced O₂ intermediates are produced in low quantity [1].



The three primary species, i.e., the superoxide anion (O₂^{•−}), hydrogen peroxide (H₂O₂) and the hydroxyl radical (HO[•]), are called reactive oxygen species because they are oxygen-containing compounds with reactive properties. O₂^{•−} and HO[•] are commonly referred to as “free radicals”. They can react with organic substrates and lead to intermediate species able to further produce other ROS. For instance, H atom abstraction by HO[•] free radicals on a C-H bond leads to a carbon-centered radical, that further reacts rapidly with O₂ to give a peroxy radical RO₂[•] (Figure 1) [2]. The latter may react with another substrate to give a new carbon-centered radical and a hydroperoxide ROOH, which may decompose into alkoxy radical RO[•] in a reaction catalyzed by redox competent metal cations such as iron or copper (as occurring with heme proteins [3]). These “secondary” species are all ROS and share a similarity in structure and reactivity with the three primary species O₂^{•−}, H₂O₂ and HO[•]. Among them, H₂O₂ (and hydroperoxides) is a molecular species and is supposed to be less reactive than the other radical short-lived species that are able to react with a range of targets (an

exception may apply for $O_2^{\bullet-}$). However, its toxicity can be exerted via Fenton reaction in the presence of redox metal ions such as iron or copper (Figure 1), or via Haber–Weiss reaction in the presence of $O_2^{\bullet-}$ [4].

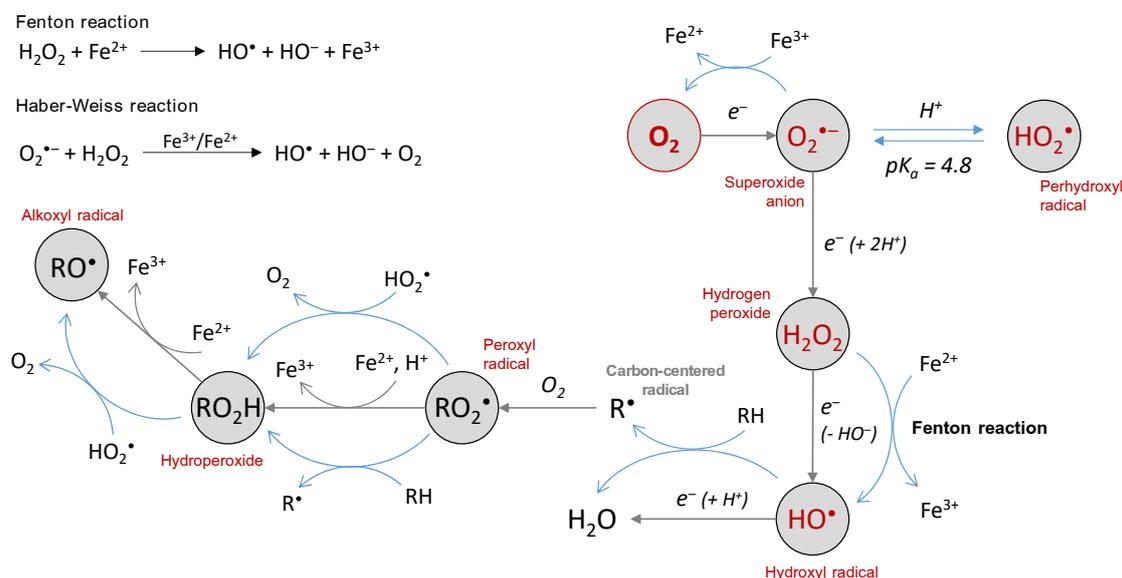


Figure 1. The chemical basis of Reactive Oxygen Species (ROS) generation—primary radical and molecular species are produced by incomplete reduction of molecular oxygen and can further react with an organic substrate to generate substrate-derived ROS. Metal ions are engaged in electron transfer (through metalloenzymes *in vivo*), but also involved in both Fenton and Haber–Weiss reactions, and in the reduction of hydroperoxide into alkoxyl radical.

2. Production of ROS

2.1. Production of ROS *In Vivo*, Regulation and Oxidative Stress

ROS can be deleterious for biomolecules and lead to oxidative damages involved in several pathologies (neurodegenerative diseases, atherosclerosis, cancer and other disorders). However, they play, above all, an important role in homeostasis, cell signaling, regulation of metabolism, or memory formation via DNA methylation [5,6]. As recently reviewed, oxidative stress may be a key modulator in neurodegenerative diseases [7]. In mammalian cells, ROS are essentially produced by enzymes and are from different origins: mainly from the cytoplasmic membrane NADPH oxidase and from the enzyme complex of the mitochondrial respiratory chain, but also from sources of other organelles such as xanthine oxidase (XO), lipo- and cyclo-oxygenase, cytochromes P450 (endoplasmic reticulum) and peroxisomes. NADPH oxidase catalyzes the mono-electronic reduction of molecular oxygen, thus producing $O_2^{\bullet-}$ [8,9] that is released either outside the cell (for phagocytic cells) or inside the cell (for non-phagocytic cells) [10]. In mitochondria, ROS are produced during ATP biosynthesis which is accompanied by electron and proton transfers, with molecular oxygen as the final target. Electron leaks, which represent around 1–3% of the total electron production, may occur in complex I (NADH-ubiquinone oxidoreductase) and complex III (ubiquinol-cytochrome c oxidoreductase) of the electron transport chain and leads to the production of $O_2^{\bullet-}$ [11]. Because of the high activity of the mitochondrial respiratory chain in aerobic organisms, such a leak is the major source of ROS production in cells, more important than NADPH oxidase (except during the activation of phagocytic cells) and XO [1]. The latter is a molybdenum enzyme, essentially located in the cytosol, that catalyzes the oxidation of hypoxanthine into xanthine and produces $O_2^{\bullet-}$, which might be further converted into H_2O_2 by XO (and oxidation of xanthine into uric acid) or by cytosolic Superoxide Dismutase (SOD) [12]. Xanthine oxidase is also able to convert nitrite into nitric oxide, and is thus a potential source of peroxynitrite [13]. Lipoxygenases and cyclooxygenases, which oxidize arachidonic acid into

leukotrienes and prostanoids (including thromboxanes and prostaglandins), respectively, are other potential sources of ROS [14,15]. In the endoplasmic reticulum, enzymes belonging to the family of cytochromes P450 play a key role in the metabolism of drugs and other xenobiotics [16]. They reduce molecular oxygen to generate $O_2^{\bullet-}$ and H_2O_2 , the latter being involved in the redox regulation of some essential functions of the endoplasmic reticulum [17].

In mitochondria, $O_2^{\bullet-}$ and H_2O_2 participate in redox signaling [18], but their production is significantly enhanced during oxidative stress conditions, as, for instance, in response to various diseases or stimuli. Oxidative stress reflects an imbalance between the production of ROS and the action of the antioxidant defense system in charge of their neutralization. They include enzymes, namely SOD that reduce $O_2^{\bullet-}$ into H_2O_2 [19,20], and catalase, glutathione peroxidases and thioredoxin reductase that regulate levels of H_2O_2 by converting it into H_2O and O_2 [21,22]. The selenoproteins glutathione peroxidases, among which the most abundant is the cytoplasmic and mitochondrial GPx1 [23], are also able to reduce hydroperoxides into alcohols. In addition to the enzymatic systems of defense, the regulation of the oxidative balance in vivo and the protection against oxidative attacks are also carried out by a myriad of non-enzymatic antioxidant systems, among which some are endogenous (glutathione, bilirubin, coenzyme Q, lipoic acid, melatonin, uric acid, etc.) and other ones are exogenous (α -tocopherol, ascorbic acid, carotenoids, etc.). Thus, under oxidative stress conditions, biomolecules may undergo the attack of ROS and get oxidized. Most of the time, such phenomena are deleterious for cells, but in some case, inducing an overproduction of ROS can help kill cells such as cancer cells [24].

2.2. Production of ROS In Vitro

Several commonly used methods are available for producing ROS in vitro, either based on metal-catalyzed production or not, and capable for some to selectively produce ROS. Water radiolysis is one of them and consists of irradiating water with γ -rays of ^{60}Co or ^{137}Cs (or X-ray). The initial energy deposition leads in situ to the generation of the primary radical and molecular species HO^{\bullet} , H^{\bullet} , e_{aq}^- (solvated electron), H_2O_2 , H_2 and H^+ , with well-known radiolytic yields of production [25]. The cumulated amount of ROS produced is directly linked to the radiation dose (expressed in Gy), which is dependent on the time the sample is exposed to the radiation source: the longer the exposure, the higher the radiation dose. Thus, it is easy to modulate the amount of ROS produced. A second advantage lies in the possibility of selecting ROS for a specific attack on a substrate: in aerated solutions ($[O_2] \approx 2 \times 10^{-4} \text{ mol L}^{-1}$ in water), $O_2^{\bullet-}$, HO^{\bullet} and H_2O_2 are generated [26,27], whereas $O_2^{\bullet-}$ or HO^{\bullet} are selectively produced (along with H_2O_2) in 0.1 M sodium formate aqueous solution [28,29] or N_2O -saturated water [30,31], respectively. For the diluted solution (below $10^{-2} \text{ mol}\cdot\text{L}^{-1}$), no direct interaction of radiation with the substrate occurs [25] and the latter is only oxidized by the ROS produced by water radiolysis. The production of ROS in vitro may also be achieved through the xanthine/xanthine oxidase system, an enzymatic way of selectively producing $O_2^{\bullet-}$ [32]. The selective production of HO^{\bullet} is usually obtained by the Fenton reaction where Fe^{2+} reduces H_2O_2 into HO^{\bullet} and HO^- (Figure 1). In this case, ROS are generated by a metal-catalyzed reaction and the resulting oxidative damages are often site-directed, in particular when biomolecules are able to coordinate metal ions [33]. The same applies when ROS are produced by the Cu^{2+} /ascorbate system, able to successively generate $O_2^{\bullet-}$, H_2O_2 and HO^{\bullet} [34–36]. For such systems, and unlike gamma radiolysis, the reaction continues as long as there are reagents, although it can be stopped in some cases [36,37]. The modulation of the production of ROS is more difficult to implement.

3. Chemical Properties and Reactivity of ROS

3.1. The Superoxide Anion

The superoxide anion is generated by the first 1-electron reduction of oxygen. At low pH, it is protonated and called perhydroxyl radical, with $pK_a(HO_2^{\bullet}/O_2^{\bullet-}) = 4.8$ [38] (Figure 1). There are two

redox standard potentials for $O_2^{\bullet-}$, showing that it can act as a reductant ($E^{\circ'}(O_2/O_2^{\bullet-}) = -0.33$ V) or as an oxidant ($E^{\circ'}(O_2^{\bullet-}/H_2O_2) = 0.93$ V) [39]. The 1-electron reduction of oxygen is not thermodynamically favored compared to its complete reduction (4 electrons, $E^{\circ'}(O_2/H_2O) = 0.81$ V). Redox potentials also show that $O_2^{\bullet-}$ disproportionation and reduction of H_2O_2 by $O_2^{\bullet-}$ [40] (Haber–Weiss reaction, Figure 2) are thermodynamically spontaneous reactions.

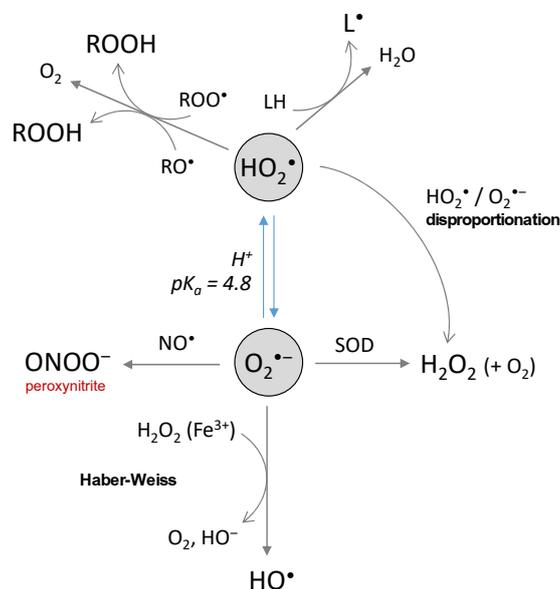


Figure 2. The chemical reactions of the superoxide and the perhydroxyl radicals.

Despite the relatively high values of its redox potential, $O_2^{\bullet-}$ is not a good reductant nor a good oxidant towards most of the biological substrates because of low rate constant values (usually below 10^2 $L \cdot mol^{-1} \cdot s^{-1}$) [38]. Some exception applies as $O_2^{\bullet-}$ is able to react with a few favored targets, with the rate constant ranging from 10^5 to 10^9 $L \cdot mol^{-1} \cdot s^{-1}$ [1]: cytochrome c, ascorbate and SOD (for which $O_2^{\bullet-}$ is the substrate). Recently, cytochrome c was used as a probe to demonstrate that $O_2^{\bullet-}$ was produced as an intermediate by the system $Cu(I)-A\beta/O_2$ [41]. The perhydroxyl radical is more reactive ($E^{\circ'}(HO_2^{\bullet}/H_2O_2) = 1.48$ V) and able to oxidize polyunsaturated fatty acids such as linoleic, linolenic or arachidonic acids ($k = 1.18 \times 10^3$, 1.70×10^3 and 3.05×10^3 $L \cdot mol^{-1} \cdot s^{-1}$) [42]. It is also engaged in the conversion of the peroxy radical to hydroperoxide (Figure 2) and then to the alkoxy radical [43]. The protonated form of $O_2^{\bullet-}$ could thus be the reactive one even if it is present at low concentrations at physiological pH. The toxicity of $O_2^{\bullet-}$ in a biological context is rather indirect since it is involved in the generation of highly-reactive secondary species. In the Haber–Weiss reaction (Figure 2), $O_2^{\bullet-}$ reacts with H_2O_2 to produce HO^{\bullet} radicals. The reaction is thermodynamically favored, but not kinetically [40,44,45], and needs to be catalyzed by iron. Disproportionation of perhydroxyl and of perhydroxyl/superoxide radicals (Figure 2) also represent a part of indirect toxicity of the superoxide anion as a potential source of H_2O_2 . The rate constant is 6×10^5 $L \cdot mol^{-1} \cdot s^{-1}$ at pH 7, thus, the reaction is relevant under physiological conditions. The disproportionation of $O_2^{\bullet-}$, while thermodynamically spontaneous, is not kinetically favored [38]. Finally, the reaction of $O_2^{\bullet-}$ with $\bullet NO$ ($k = 1.9 \times 10^{10}$ $L \cdot mol^{-1} \cdot s^{-1}$) [46] to generate the highly-reactive peroxynitrite $ONOO^-$ is another reaction conferring an indirect toxicity to $O_2^{\bullet-}$, in particular towards DNA, proteins and lipids [47,48]. Peroxynitrite is able to nitrate tyrosine or tryptophan residues, or to oxidize methionine residues [49–51].

The reactivity of the superoxide anion does not always lead to a deleterious effect towards biomolecules as it is also able to help to fight against oxidative damages. Recently, Muñoz-Rugeles et al. [52] have shown that the superoxide anion is able to repair oxidized DNA by transferring one electron to the guanosyl radical of a single-stranded DNA. However, such an involvement in unusual chemical processes remains almost unexplored.

3.2. The Hydroxyl Radical

The hydroxyl radical is the most powerful oxidant among the ROS, with a potential of $E^\circ(\text{HO}^\bullet/\text{H}_2\text{O}) = 2.34 \text{ V}$ [39]. At very low pH, HO^\bullet converts into its conjugate base $\text{O}^{\bullet-}$ ($\text{pK}_a(\text{HO}^\bullet/\text{O}^{\bullet-}) = 11.9$), the oxide radical, which is less reactive [53] but not relevant at physiological pH. Reactions of HO^\bullet radicals with most substrates are diffusion-controlled (rate constants of $10^{10} \text{ L}\cdot\text{mol}^{-1}\cdot\text{s}^{-1}$) as, for example, with biological molecules such as DNA bases, aromatic amino acids, albumin, hemoglobin, linoleate or ascorbate [53–55]. Thus, HO^\bullet radicals are engaged in fast reactions, with an activation energy close to zero, meaning that they are not able to diffuse, have a very short lifetime (few 10^{-6} s) and free course (few 10^{-8} m), and are weakly selective towards molecular targets. A side consequence of this high reactivity is that the disproportionation of HO^\bullet radicals, even if kinetically favored ($k \approx 5 \times 10^9 \text{ L}\cdot\text{mol}^{-1}\cdot\text{s}^{-1}$) [54], remains a rather infrequent event in biological conditions, the probability of collision between two hydroxyl radicals being very low.

There are three ways of action for the HO^\bullet radical: electron abstraction, hydrogen abstraction and double bond addition (Figure 3). The HO^\bullet radical is electrophilic and has a strong affinity for electron-rich sites of molecules, in particular for aromatic or sulfur-containing molecules. This is illustrated by the rate constants of reaction with amino acids, ranging from $10^7 \text{ L}\cdot\text{mol}^{-1}\cdot\text{s}^{-1}$ for Gly to $10^{10} \text{ L}\cdot\text{mol}^{-1}\cdot\text{s}^{-1}$ for His, Trp or Cys [53]. Addition reactions are usually faster than H atom abstraction [56], except with Cys where H abstraction from the thiol group is faster [57,58]. HO^\bullet addition is commonly involved in biomolecule oxidation, such as oxidation of guanine into 8-oxoguanine [59], of histidine into 2-oxohistidine [60,61] or of tryptophan into N-formylkynurenine and kynurenine [43,62]. Hydroxyl radical addition may also occur on the sulfur atom, in particular, from methionine residue, leading to the hydroxysulfuranyl radical as the intermediate species and, finally, to methionine sulfoxide and sulfone as the end-products [63]. Methionine also undergoes electron abstraction when reacting with HO^\bullet , thus generating the sulfuranyl radical cation that is able to further evolve. In this case, oxidation leads to irreversible biological damages, contrary to the oxidation of methionine into methionine sulfoxide for which reversibility is ensured through methionine sulfoxide reductases (MsrA and MsrB) [64]. Electron abstraction is also observed with inorganic substrates such as ferrous ions or halides, with high rate constants [55]. The last pathway for HO^\bullet reaction is H-abstraction, for which numerous and various biomolecules are targets as, for instance, polyunsaturated fatty acids such as linoleate [65,66] or arachidonate [67,68], sulfur-containing, basic and aromatic amino acid residues from protein and peptides [69,70], or 2-deoxyribose and DNA bases [71]. Most of the time, H abstraction leads to a carbon-centered radical that either further reacts fast with molecular oxygen to generate a peroxy radical or, in the absence of oxygen, is engaged in a biradical reaction generating a carbon-carbon bond [72]. However, abstraction may also occur on the hydroxyl or thiol functional groups, leading to oxygen- or sulfur-centered radicals [73]. Such mechanisms are observed for protein and peptide cross-linking via bityrosine formation [70,74] or disulfide bridge formation [75].

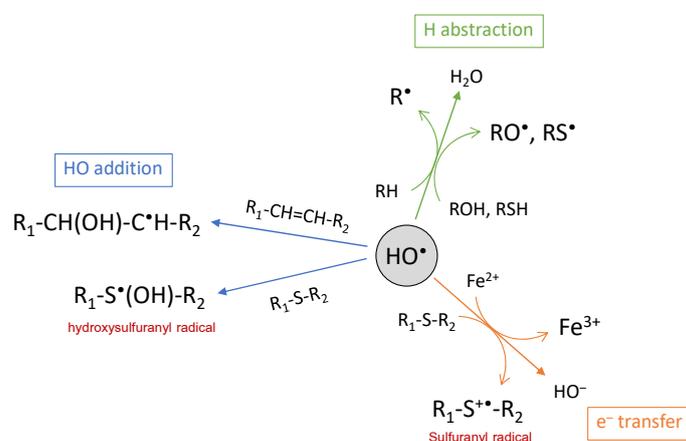


Figure 3. The chemical reactions of the hydroxyl radical.

3.3. Hydrogen Peroxide

Hydrogen peroxide is produced by the two-electron reduction of molecular oxygen. Its conjugate base HOO^- is a strong nucleophile but not relevant at physiological pH because of a high pKa value ($\text{pKa}(\text{H}_2\text{O}_2/\text{HOO}^-) = 11.6$). H_2O_2 is either a reductant or an oxidant in one-electron transfer reactions. The latter is not thermodynamically favored in biological conditions ($E^\circ(\text{O}_2^{\bullet-}/\text{H}_2\text{O}_2) = 0.93$ V and $E^\circ(\text{H}_2\text{O}_2/\text{HO}^\bullet) = 0.30$ V) [39] but H_2O_2 can act as an oxidant if catalysis by metal ions takes place (Fenton and Haber–Weiss reactions). It is rather engaged in two-electron transfer reactions, with a high potential ($E^\circ(\text{H}_2\text{O}_2/\text{H}_2\text{O}) = 1.32$ V) in physiological conditions. It is more oxidizing than hypochlorous acid and peroxyntrite ($E^\circ(\text{ClO}^-/\text{Cl}^-) = 1.28$ V and $E^\circ(\text{ONOO}^-/\text{NO}_2^-) = 1.20$ V). However, it reacts only poorly with most biological molecules because of a high activation energy barrier, oxidation by H_2O_2 being kinetically driven. Thus, the strongest oxidizing power of hydrogen peroxide comes indirectly from its metal-catalyzed conversion into HO^\bullet radicals by the Fenton and Haber–Weiss reactions (Figure 4).

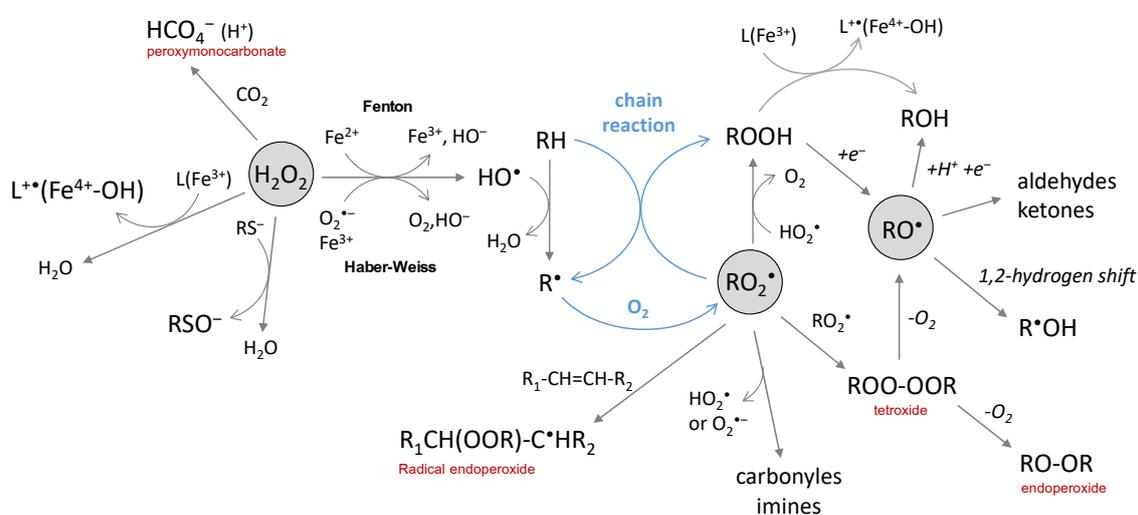


Figure 4. The chemical reactions of hydrogen peroxide, peroxy and alkoxy radicals.

In proteins, H_2O_2 reacts as a two-electron oxidant towards sulfur-containing residues (cysteine and methionine) but with a low rate constant ($k = 2.9$ $\text{L}\cdot\text{mol}^{-1}\cdot\text{s}^{-1}$ for cysteine) [76]. For thiols, the reaction is exclusive to the thiolate anion, thus the reactivity at physiological pH is dependent on pKa values. It leads to sulfenic acid (RSOH) as the initial product, able to be oxidized one more time by H_2O_2 into sulfinic acid (RSO_2H) or to react with thiols to form disulfides. The highest rate constants are observed for the thiol proteins peroxiredoxins and glutathione peroxidases that react with H_2O_2 several orders of magnitude faster ($\sim 10^7$ $\text{L}\cdot\text{mol}^{-1}\cdot\text{s}^{-1}$) [77]. Such a difference is explained by the polarization of the O–O bond of H_2O_2 by hydrogen bonding into the protein that facilitates the electrophilic attack on the thiolate. Pyruvate oxidation in acetate and carbon dioxide by H_2O_2 is also biologically relevant because of a rate constant of 2.2 $\text{L}\cdot\text{mol}^{-1}\cdot\text{s}^{-1}$ [78] and a pyruvate intracellular concentration of 0.1 – 0.5 mM (competitive with most thiols).

The toxicity of H_2O_2 can also be expressed indirectly. The reaction with bicarbonate leads to peroxymonocarbonate (HCO_4^-) species that react approx. 300 times faster than H_2O_2 with thiols and sulfide (Figure 4) [79–81]. Only a few percent of H_2O_2 is present as peroxymonocarbonate in a physiological bicarbonate buffer since the reaction is an equilibrium ($K = 0.32$) [81]; it can be accelerated by carbonic anhydrase [79], thus enhancing the physiological relevance of the reaction. However, the most deleterious effect of H_2O_2 comes from its reaction with transition metals able to generate highly reactive radical species or activated metal complexes. The widely-known example is the production of hydroxyl radicals from hydrogen peroxide by the Fenton reaction (Figure 4), which involves iron ions as the metal catalyst. The reaction may be catalyzed by other redox competent metal ions (and, in this case,

is called a “Fenton-like” reaction) as, for example, by copper complexed to the amyloid-beta peptide (A β) [82]. With the Fe³⁺ of heme proteins, the reaction of H₂O₂ is fast ($k = 10^7$ – 10^8 L·mol⁻¹·s⁻¹) [83] and give rise to Fe⁴⁺-oxoferryl porphyrin radical cation, able to transfer one electron to the surrounding protein [3,84], resulting in a formation of a protein radical that further evolves. The question of whether H₂O₂ is directly converted into HO• radicals or whether intermediates of higher oxidation states of the metal are produced has been debated in the past years. Both are possibly involved, depending on circumstances, but they are all strong oxidative species and the products resulting from their reaction with a substrate should be similar. When metal ions are coordinated to a biological molecule, the reaction may be different from Fenton chemistry since metal-catalyzed oxidation (MCO) is site-directed. Such a case is observed, for instance, for protein, DNA or the A β peptide in iron- or copper-catalyzed oxidation in the presence of ascorbate [36,85,86].

3.4. Peroxyl Radicals, Hydroperoxides and Alkoxy Radicals

Peroxyl radicals are secondary species generated by the addition of molecular oxygen on carbon-centered radicals ($[O_2] \approx 2 \times 10^{-4}$ mol·L⁻¹ in aerated aqueous solution), whose rate constant usually range between 10^8 and 10^9 L·mol⁻¹·s⁻¹ [2]. They can also be produced in the absence of oxygen by metal-induced conversion of hydroperoxides [87]. Hydroperoxides are generated from peroxyl radicals by reaction with HO₂• or by H abstraction from another molecule, and may further react with HO₂• or a metal ion to generate alkoxy radicals (Figure 4). The latter can also be generated from peroxyl radicals via a tetroxide. Peroxyl and alkoxy radicals are oxidant species, with relatively high redox standard potentials of $E^{\circ'}(RO_2^{\bullet}/RO_2H) = 1.00$ V and $E^{\circ'}(RO^{\bullet}/ROH) = 1.60$ V, respectively [88].

Peroxyl radicals react faster than the superoxide anion with numerous biological substrates (DNA, lipids, proteins); rate constants [2] ranging from 10^2 to 10^8 L·mol⁻¹·s⁻¹. Even if they are much less reactive than the hydroxyl radical, they share some similarity in their mode of reaction as they are able to either be engaged in electron abstraction, H atom abstraction or addition on double-bonds (Figure 4) [89]. In the latter case, intra- or intermolecular reactions lead to the formation of the radical endoperoxide ROOR• species. Peroxyl radicals with the α -hydroxyl or α -amino groups can also undergo rapid unimolecular elimination of HO₂•/O₂•⁻, leading to carbonyl or imine group formation [2,90,91]. Peroxyl radicals ROO• can undergo dimerization with other peroxyl radicals R'OO• and yield tetroxide species ROO-OOR'; the reaction is also possible between peroxyl radicals and HO₂•, as observed for thymine [59]. Tetroxide is an unstable species and their subsequent decomposition yields carbonyl groups and alcohol, accompanied by the loss of molecular oxygen [3].

Among the very diverse reactions that peroxyl radicals can initiate, some are of particular importance because they contribute to the degradation of cell membranes induced by lipid peroxidation. Once a carbon-centered radical has been generated on a fatty acid moiety, it reacts fast with molecular oxygen to yield a peroxyl radical, able to abstract an H atom from another fatty acid moiety to give birth to another carbon-centered radical. This H abstraction is facilitated by the proximity of the two fatty acid chains within the lipid bilayers of cell membranes. In such a condition, a chain reaction starts and is propagated by the R• and RO₂• radicals. The chain reaction stops either when there are no more lipids, no more oxygen or when peroxide radicals react with a lipid-soluble antioxidant, such as α -tocopherol or carotenoids [92,93].

Primary and secondary alkoxy radicals undergo a rapid 1,2-hydrogen shift, resulting in the generation of α -hydroxyalkyl radicals, in competition with the intramolecular 1,5-hydrogen shift and the formation of alcohol by intermolecular H abstraction [94,95]. In some cases, in particular, when a 1,2-hydrogen shift is not possible (tertiary alkoxy radicals), β -fragmentation reactions occur and yield aldehydes and ketones [96] with relatively high rate constants in aqueous solutions ($k > 10^6$ s⁻¹) [97,98].

4. The Implication of ROS in Neurodegenerative Diseases

The high consumption of molecular oxygen and the high content of polyunsaturated fatty acid, strongly sensitive to peroxidation, make the brain a particularly vulnerable tissue to oxidative

stress [99]. The latter is a modulator of neurodegenerative diseases (recently reviewed in Reference [7]). Peroxidation products of fatty acids are among the biomarkers of oxidative stress in neurodegenerative diseases such as Alzheimer's disease (AD), Parkinson's disease (PD) and amyotrophic lateral sclerosis (ALS), along with protein carbonylation and nitration, DNA and RNA oxidative damages [100–104]. Neurodegenerative diseases are commonly associated with abnormal protein aggregation. In AD, the A β peptide is found aggregated in senile plaques (composed of A β fibrils and metal ions) and hyperphosphorylated Tau in neurofibrillary tangles. In PD, aggregation of α -synuclein leads to Lewy bodies inclusions, while the aggregation of the huntingtin protein and copper/zinc superoxide dismutase are involved in Huntington's disease (HD) and ALS, respectively. Such abnormal protein aggregation is able to induce oxidative stress via mitochondria dysfunction and ROS production [105–107], leading to chronic inflammation, and play an important role in neurodegeneration.

In AD, an imbalance between the production of ROS and the reduced activity of enzymes responsible for ROS scavenging leads to oxidative damages on biomolecules, and on the A β peptide itself [108–110]. The link between oxidative stress and the amyloid beta peptide has been recently reviewed [111]. Because copper is present in relatively high levels in the brain and because of the ability of the A β peptide to chelate metal ions, A β -copper is a potential direct source of ROS in the presence of ascorbate and molecular oxygen. Reybier et al. [41] have shown that the superoxide anion is generated as an intermediate during H₂O₂ production by A β -copper. No direct link has yet been established between the production of ROS by A β -copper and oxidation of biological material *in vivo*. However, increased levels of lipids, protein and DNA oxidation have been reported to be associated with elevated levels of A β , whereas low A β -content brain regions do not present high concentrations of oxidative stress markers [112–115]. Lipid peroxidation is one of the events associated with AD, which might be involved in the phospholipid imbalance observed in the brain of AD patients [116,117]. Malondialdehyde (MDA) and 4-hydroxynonenal (4-HNE) are two aldehydes commonly found in high levels in AD brains [118]. The compound 4-HNE is toxic for neurons by causing apoptosis or by altering the microtubule structure [119,120], but is also prone to react with lipid acid [121] and to form adducts with proteins (target amino acids are cysteine, histidine and lysine) [122] detected in AD brains [123]. In particular, adducts with Tau were found to modify its conformation and to favor neurofibrillary tangles formation [124]. The compound 4-HNE is generated by the non-enzymatic oxidation of polyunsaturated omega-6 fatty acids, such as arachidonate or linoleate. It is a direct consequence of the peroxidation of lipids by ROS since it is generated by the degradation of lipid hydroperoxides [125]—hydroperoxyoctadecadienoate (HPODE) from linoleate or hydroperoxyeicosatetraenoate (HPETE) from arachidonate (Figure 5). F₂- and F₄-isoprostanes, which are generated by peroxidation of arachidonate, are other markers of oxidative stress in AD and found in elevated levels in the brain of AD patients [126–128].

Protein oxidation has been evidenced by high levels of carbonylated proteins in the brain areas the most involved in AD (i.e., hippocampus and parietal cortex) [114,129]. Several molecular mechanisms have been proposed for protein carbonylation, some of them being induced by direct ROS attacks and leading to protein cleavage via an alkoxy radical formation (Figure 5). The target proteins are, among others, those involved in glucose metabolism and ATP synthesis [130], such as ATP synthase [131], pyruvate kinase, phosphoglucose mutase, α -enolase, malate dehydrogenase or glyceraldehyde-3-phosphate dehydrogenase (see Reference [132] for a review). Modifications detected include carbonylation, nitration and HNE-adducts formation. Like protein carbonylation, oxidative damages of DNA bases may result from a direct attack of ROS. Increased levels of 8-oxo-2-dehydroguanine, 8-hydroxyadenine and 5-hydroxyuracil have been reported in the temporal, parietal and frontal lobes of AD brains [133,134], along with 8-hydroxyguanine in the hippocampus of patients with preclinical stages of AD [135]. The high levels of oxidized DNA bases are detected in neurons where lipids and protein oxidation are also increased [136].

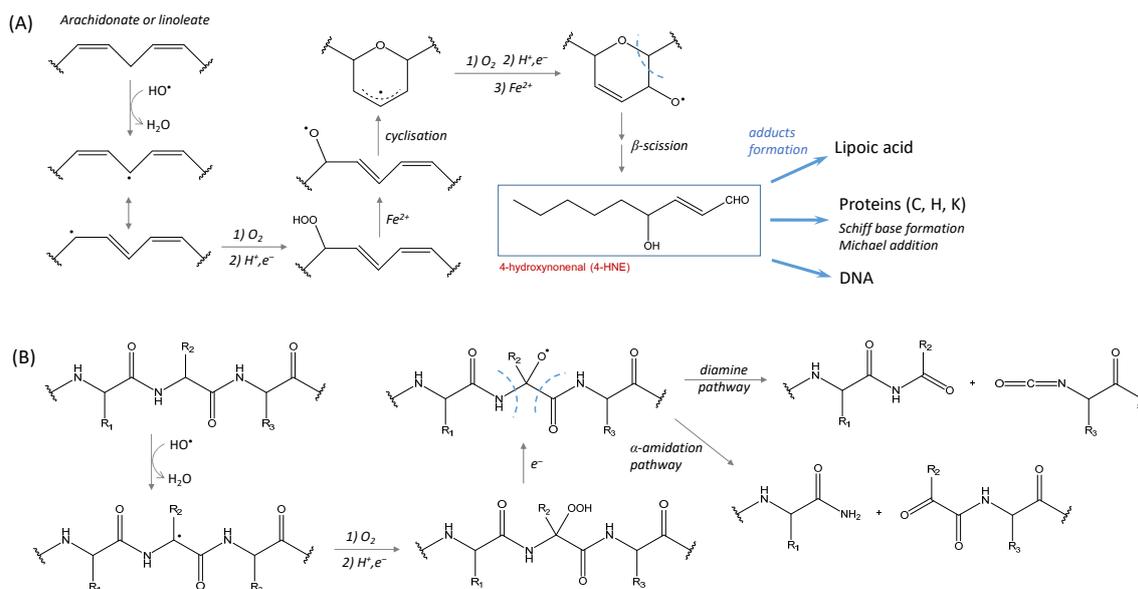


Figure 5. The direct involvement of ROS in lipid peroxidation and protein carbonylation. **(A)** the mechanism of 4-HNE formation from ROS-induced polyunsaturated omega-6 fatty acid peroxidation (from Pryor and Porter [125]); 4-HNE is able to form adducts with lipoic acid, proteins (C, H and K residues) and DNA bases; **(B)** ROS-induced protein carbonylation and cleavage (from Stadtman and Levine [43]).

In PD, the involvement of ROS and oxidative stress might be one of the major factors causing the disease. Dopaminergic neurons of the substantia nigra, where the basal level of free radicals is important [137], are particularly sensitive to degeneration. Elevated levels of oxidized lipids and proteins have been detected in the substantia nigra of PD patients [138,139]. Additionally, an increase of 8-hydroxy-2'-deoxyguanosine and 8-hydroxyguanine levels, two markers of DNA oxidation, was observed [140]. As in AD, 4-HNE-modified proteins have also been detected in PD [141]. Thus, AD and PD share some similarities regarding the biomarkers of the oxidative stress detected. In PD, metal ion release (e.g., Fe²⁺) would be an important mechanism of neurodegeneration, through ROS production and dopamine oxidation. High levels of iron ions, in conjunction with the production of H₂O₂ via dopamine oxidation (enzymatically by monoamine oxidases, would lead to an overproduction of ROS and thus to oxidative stress conditions. Non-enzymatic oxidation of dopamine is involved in free radicals production and in elevation of free iron levels in dopaminergic cells [142–144]. Oxidative modification of proteins in PD may also have an impact on their propensity to aggregate. Surgucheva et al. [145] have shown that oxidation of γ -synuclein enhanced the formation of annular oligomers that accumulate in cells and that can initiate α -synuclein aggregation.

In other diseases such as ALS or HD, the link with oxidative stress is also evidenced, even if the mechanisms involved in their etiology are not fully understood. In ALS, oxidative stress is evidenced by elevated levels of MDA, 4-HNE, advanced oxidation protein products, isoprostanoids and 8-hydroxy-2'-deoxyguanosine [146–150]. Oxidative stress is also coupled to mitochondrial damages and dysfunction, each exacerbating the other, and to RNA dysmetabolism and unfolded protein aggregates formation [151,152]. As in other neurodegenerative diseases, copper and iron homeostasis is disturbed in ALS and elevated levels of these redox-competent metal ions could participate in ROS production [153]. Strong evidence exists also for early oxidative stress in HD, coupled with mitochondrial dysfunction, but it is still not clear whether oxidative stress is a cause or a consequence of HD. As for ALS, metal dyshomeostasis, evidenced by high levels of iron and copper in post-mortem brain tissues of HD patients [154], would participate in ROS production via Fenton chemistry. Increases of nuclear and mitochondrial DNA 8-hydroxy-2'-deoxyguanosine were detected in the blood and serum of HD patients [155,156], along with DNA double-strand breaks, a potential

result of free radical damage [157,158]. Lipid peroxidation (high levels of MDA and cytoplasmic lipofuscin), protein carbonylation and nitration (an increase of 3-nitrotyrosine levels) are also observed in HD [159,160].

5. Concluding Remarks

An important feature shared by most of the neurodegenerative diseases is the presence of oxidative damages that link them to oxidative stress. The latter is supposed to be an early event in the etiology of some diseases since the biomarkers of oxidation appear early in their development [132]. An overproduction of ROS is considered to have a major contribution in oxidative damages undergone by biomolecules, including lipids, proteins and DNA. The intrinsic chemical properties of ROS make them formidable weapons against most biomolecules. Among them, because of its diffusion-controlled reactivity with most of the biological material, HO• may be considered as a nuclear weapon compared to O₂•⁻ and H₂O₂. These last two react directly only with few specific targets (e.g., SOD and catalase). However, they are strong deleterious species because of (i) their ability to be engaged in Fenton and Haber-Weiss reactions, two metal-catalyzed reactions that lead to HO• production, (ii) their lack of direct reactivity which gives them the possibility to spread to areas where metal levels are high. So, most of the time, final oxidative damages on biomolecules could be considered as resulting from HO• attacks. This could particularly apply to neurodegenerative diseases where metal dyshomeostasis takes place and where elevated levels of redox-competent metal ions—such as iron or copper—are observed. In this context, better control of ROS homeostasis would be important for neuron survival. This could be achieved, among others, by developing antioxidant-based strategies. This is the reason why many studies have focused and are still focusing on possible therapeutic approaches based on antioxidant strategies, either by the administration of antioxidant in the form of plant extracts or nutraceuticals [161,162] or by reinforcing the antioxidant defense system in vivo [163]. Antioxidant therapy-based strategies to fight against neurodegenerative diseases have shown promising results in preclinical trials but only a few clinical trials have been conducted and the benefit of such a therapy is still under debate [164]. In this context, and because most of the mechanisms underlying the etiology of neurodegenerative diseases have still not been elucidated, all efforts to better understand, through basic research, the causes of disease development will increase the global knowledge and will help to develop novel therapeutic strategies.

Funding: This research received no external funding.

Conflicts of Interest: The author declares no conflict of interest.

References

1. Halliwell, B.; Gutteridge, J.M.C. *Free Radicals in Biology and Medicine*, 3rd ed.; Oxford science Publications: Oxford, UK, 1999.
2. Neta, P.; Huie, R.E.; Ross, A.B. Rate constants for reactions of peroxy radicals in fluid solutions. *J. Phys. Chem. Ref. Data* **1990**, *19*, 413–513. [[CrossRef](#)]
3. Hawkins, C.L.; Davies, M.J. Generation and propagation of radical reactions on proteins. *Biochim. Et Biophys. Acta* **2001**, *1504*, 196–219. [[CrossRef](#)]
4. Haber, F.; Weiss, J. The catalytic decomposition of hydrogen peroxide by iron salts. *Proc. R. Soc. Lond.* **1934**, *147*, 332–351.
5. Rhee, S.G. Redox signaling: Hydrogen peroxide as intracellular messenger. *Exp. Mol. Med.* **1999**, *31*, 53–59. [[CrossRef](#)] [[PubMed](#)]
6. Zhou, X.; Zhuang, Z.; Wang, W.; He, L.; Wu, H.; Cao, Y.; Pan, F.; Zhao, J.; Hu, Z.; Sekhar, C.; et al. OGG1 is essential in oxidative stress induced DNA demethylation. *Cell Signal.* **2016**, *28*, 1163–1171. [[CrossRef](#)]
7. Singh, A.; Kukreti, R.; Saso, L.; Kukreti, S. Oxidative Stress: A Key Modulator in Neurodegenerative Diseases. *Molecules* **2019**, *24*. [[CrossRef](#)]
8. Vignais, P.V. The superoxide-generating NADPH oxidase: Structural aspects and activation mechanism. *Cell. Mol. Life Sci.* **2002**, *59*, 1428–1459. [[CrossRef](#)]

9. Finkel, T. Redox-dependent signal transduction. *FEBS Lett.* **2000**, *476*, 52–54. [[CrossRef](#)]
10. Souza, H.P.; Laurindo, F.R.; Ziegelstein, R.C.; Berlowitz, C.O.; Zweier, J.L. Vascular NAD(P)H oxidase is distinct from the phagocytic enzyme and modulates vascular reactivity control. *Am. J. Physiol. Heart Circ. Physiol.* **2001**, *280*, H658–H667. [[CrossRef](#)]
11. Turrens, J.F. Mitochondrial formation of reactive oxygen species. *J. Physiol.* **2003**, *552*, 335–344. [[CrossRef](#)]
12. Schmidt, H.M.; Kelley, E.E.; Straub, A.C. The impact of xanthine oxidase (XO) on hemolytic diseases. *Redox Biol.* **2019**, *21*, 101072. [[CrossRef](#)]
13. Godber, B.L.; Doel, J.J.; Sapkota, G.P.; Blake, D.R.; Stevens, C.R.; Eisenthal, R.; Harrison, R. Reduction of nitrite to nitric oxide catalyzed by xanthine oxidoreductase. *J. Biol. Chem.* **2000**, *275*, 7757–7763. [[CrossRef](#)]
14. Bonizzi, G.; Piette, J.; Merville, M.P.; Bours, V. Cell type-specific role for reactive oxygen species in nuclear factor-kappaB activation by interleukin-1. *Biochem. Pharmacol.* **2000**, *59*, 7–11. [[CrossRef](#)]
15. van der Donk, W.A.; Tsai, A.L.; Kulmacz, R.J. The cyclooxygenase reaction mechanism. *Biochemistry* **2002**, *41*, 15451–15458. [[CrossRef](#)]
16. Estabrook, R.W. A passion for P450s (rememberances of the early history of research on cytochrome P450). *Drug Metab. Dispos. Biol. Fate Chem.* **2003**, *31*, 1461–1473. [[CrossRef](#)]
17. Coon, M.J.; Ding, X.X.; Pernecky, S.J.; Vaz, A.D. Cytochrome P450: Progress and predictions. *Faseb J. Off. Publ. Fed. Am. Soc. Exp. Biol.* **1992**, *6*, 669–673. [[CrossRef](#)]
18. Brand, M.D. Mitochondrial generation of superoxide and hydrogen peroxide as the source of mitochondrial redox signaling. *Free Radic. Biol. Med.* **2016**, *100*, 14–31. [[CrossRef](#)]
19. Hsu, J.L.; Hsieh, Y.; Tu, C.; O'Connor, D.; Nick, H.S.; Silverman, D.N. Catalytic properties of human manganese superoxide dismutase. *J. Biol. Chem.* **1996**, *271*, 17687–17691. [[CrossRef](#)]
20. Oury, T.D.; Crapo, J.D.; Valnickova, Z.; Enghild, J.J. Human extracellular superoxide dismutase is a tetramer composed of two disulphide-linked dimers: A simplified, high-yield purification of extracellular superoxide dismutase. *Biochem. J.* **1996**, *317* (Pt 1), 51–57. [[CrossRef](#)]
21. Valko, M.; Leibfritz, D.; Moncol, J.; Cronin, M.T.; Mazur, M.; Telser, J. Free radicals and antioxidants in normal physiological functions and human disease. *Int. J. Biochem. Cell Biol.* **2007**, *39*, 44–84. [[CrossRef](#)]
22. Stanley, B.A.; Sivakumaran, V.; Shi, S.; McDonald, I.; Lloyd, D.; Watson, W.H.; Aon, M.A.; Paolocci, N. Thioredoxin reductase-2 is essential for keeping low levels of H₂O₂ emission from isolated heart mitochondria. *J. Biol. Chem.* **2011**, *286*, 33669–33677. [[CrossRef](#)] [[PubMed](#)]
23. Ursini, F.; Maiorino, M.; Brigelius-Flohe, R.; Aumann, K.D.; Roveri, A.; Schomburg, D.; Flohe, L. Diversity of glutathione peroxidases. *Methods Enzymol.* **1995**, *252*, 38–53.
24. Lee, C.M.; Lee, J.; Nam, M.J.; Choi, Y.S.; Park, S.H. Tomentosin Displays Anti-Carcinogenic Effect in Human Osteosarcoma MG-63 Cells via the Induction of Intracellular Reactive Oxygen Species. *Int. J. Mol. Sci.* **2019**, *20*. [[CrossRef](#)]
25. Spinks, J.W.T.; Woods, R.J. Water and inorganic aqueous systems. In *Introduction to Radiation Chemistry*; John Wiley & Sons: New York, NY, USA, 1990; pp. 243–313.
26. Bielski, B.H.J. Re-Evaluation of Spectral and Kinetic-Properties of HO₂ and (O₂)₂ Free-Radicals. *Photochem. Photobiol.* **1978**, *28*, 645–649. [[CrossRef](#)]
27. Thomas, J.K. The rate constants for H atom reactions in aqueous solutions 1. *J. Phys. Chem.* **1963**, *67*, 2593–2595. [[CrossRef](#)]
28. Matthews, R.W.; Sangster, D.F. Measurement by Benzoate Radiolytic Decarboxylation of Relative Rate Constants for Hydroxyl Radical Reactions. *J. Phys. Chem.* **1965**, *69*, 1938–1946. [[CrossRef](#)]
29. Rabani, J.; Matheson, M.S. The Pulse Radiolysis of Aqueous Solutions of Potassium Ferrocyanide. *J. Phys. Chem.* **1966**, *70*, 761–769. [[CrossRef](#)]
30. Janata, E.; Schuler, R.H. Rate constant for scavenging eaq⁻ in nitrous oxide-saturated solutions. *J. Phys. Chem.* **1982**, *86*, 2078–2084. [[CrossRef](#)]
31. Asmus, K.D.; Fendler, J.H. Reaction of sulfur hexafluoride with hydrated electrons. *J. Phys. Chem.* **1968**, *72*, 4285–4289. [[CrossRef](#)]
32. Hille, R.; Hall, J.; Basu, P. The mononuclear molybdenum enzymes. *Chem. Rev.* **2014**, *114*, 3963–4038. [[CrossRef](#)]
33. Peng, Y.; Wang, C.; Xu, H.H.; Liu, Y.N.; Zhou, F. Binding of alpha-synuclein with Fe(III) and with Fe(II) and biological implications of the resultant complexes. *J. Inorg. Biochem.* **2010**, *104*, 365–370. [[CrossRef](#)]

34. Miotto, M.C.; Rodriguez, E.E.; Valiente-Gabioud, A.A.; Torres-Monserrat, V.; Binolfi, A.; Quintanar, L.; Zweckstetter, M.; Griesinger, C.; Fernandez, C.O. Site-specific copper-catalyzed oxidation of alpha-synuclein: Tightening the link between metal binding and protein oxidative damage in Parkinson's disease. *Inorg. Chem.* **2014**, *53*, 4350–4358. [[CrossRef](#)] [[PubMed](#)]
35. Cheignon, C.; Faller, P.; Testemale, D.; Hureau, C.; Collin, F. Metal-catalyzed oxidation of Abeta and the resulting reorganization of Cu binding sites promote ROS production. *Metallomics* **2016**, *8*, 1081–1089. [[CrossRef](#)]
36. Cassagnes, L.E.; Herve, V.; Nepveu, F.; Hureau, C.; Faller, P.; Collin, F. The catalytically active copper-amyloid-Beta state: Coordination site responsible for reactive oxygen species production. *Angew. Chem. Int. Ed. Engl.* **2013**, *52*, 11110–11113. [[CrossRef](#)]
37. Cheignon, C.; Hureau, C.; Collin, F. Real-time evolution of A beta(40) metal-catalyzed oxidation reveals Asp1 as the main target and a dependence on metal binding site. *Inorg. Chim. Acta* **2018**, *472*, 111–118. [[CrossRef](#)]
38. Bielski, B.H.J.; Cabelli, D.E.; Arudi, R.L.; Ross, A.B. Reactivity of HO₂/O₂⁻ Radicals in Aqueous Solution. *J. Phys. Chem. Ref. Data* **1985**, *14*, 1041–1100. [[CrossRef](#)]
39. Wardman, P. Reduction Potentials of One-Electron Couples Involving Free-Radicals in Aqueous-Solution. *J. Phys. Chem. Ref. Data* **1989**, *18*, 1637–1755. [[CrossRef](#)]
40. Ferradini, C.; Foos, J.; Houee, C.; Pucheault, J. Reaction between Superoxide Anion and Hydrogen-Peroxide. *Photochem. Photobiol.* **1978**, *28*, 697–700. [[CrossRef](#)]
41. Reybier, K.; Ayala, S.; Alies, B.; Rodrigues, J.V.; Bustos Rodriguez, S.; La Penna, G.; Collin, F.; Gomes, C.M.; Hureau, C.; Faller, P. Free Superoxide is an Intermediate in the Production of H₂O₂ by Copper(I)-Abeta Peptide and O₂. *Angew. Chem. Int. Ed.* **2016**, *55*, 1085–1089. [[CrossRef](#)] [[PubMed](#)]
42. Bielski, B.H.J.; Arudi, R.L.; Sutherland, M.W. A Study of the Reactivity of HO₂/O₂⁻ with Unsaturated Fatty-Acids. *J. Biol. Chem.* **1983**, *258*, 4759–4761. [[PubMed](#)]
43. Stadtman, E.R.; Levine, R.L. Free radical-mediated oxidation of free amino acids and amino acid residues in proteins. *Amino Acids* **2003**, *25*, 207–218. [[CrossRef](#)] [[PubMed](#)]
44. Rigo, A.; Stevanato, R.; Finazzi-Agro, A.; Rotilio, G. An attempt to evaluate the rate of the Haber-Weiss reaction by using OH radical scavengers. *FEBS Lett.* **1977**, *80*, 130–132. [[CrossRef](#)]
45. Weinstein, J.; Bielski, B.H.J. Kinetics of the Interaction of HO₂ and O₂⁻ Radicals with Hydrogen-Peroxide - Haber-Weiss Reaction. *J. Am. Chem. Soc.* **1979**, *101*, 58–62. [[CrossRef](#)]
46. Kissner, R.; Nauser, T.; Bugnon, P.; Lye, P.G.; Koppenol, W.H. Formation and properties of peroxyxynitrite as studied by laser flash photolysis, high-pressure stopped-flow technique, and pulse radiolysis. *Chem. Res. Toxicol.* **1997**, *10*, 1285–1292. [[CrossRef](#)] [[PubMed](#)]
47. Squadrito, G.L.; Pryor, W.A. The formation of peroxyxynitrite in vivo from nitric oxide and superoxide. *Chem. Biol. Interact.* **1995**, *96*, 203–206. [[CrossRef](#)]
48. Packer, M.A.; Porteous, C.M.; Murphy, M.P. Superoxide production by mitochondria in the presence of nitric oxide forms peroxyxynitrite. *Biochem. Mol. Biol. Int.* **1996**, *40*, 527–534. [[CrossRef](#)]
49. Kato, Y.; Uchida, K.; Kawakishi, S. Oxidative fragmentation of collagen and prolyl peptide by Cu(II)/H₂O₂. Conversion of proline residue to 2-pyrrolidone. *J. Biol. Chem.* **1992**, *267*, 23646–23651. [[PubMed](#)]
50. Pryor, W.A.; Squadrito, G.L. The chemistry of peroxyxynitrite: A product from the reaction of nitric oxide with superoxide. *Am. J. Physiol.* **1995**, *268*, L699–L722. [[CrossRef](#)] [[PubMed](#)]
51. Berlett, B.S.; Friguier, B.; Yim, M.B.; Chock, P.B.; Stadtman, E.R. Peroxyxynitrite-mediated nitration of tyrosine residues in Escherichia coli glutamine synthetase mimics adenylylation: Relevance to signal transduction. *Proc. Natl. Acad. Sci. USA* **1996**, *93*, 1776–1780. [[CrossRef](#)]
52. Munoz-Rugeles, L.; Galano, A.; Alvarez-Idaboy, J.R. The other side of the superoxide radical anion: Its ability to chemically repair DNA oxidized sites. *Chem. Commun.* **2018**, *54*, 13710–13713. [[CrossRef](#)]
53. Buxton, G.V.; Greenstock, C.L.; Helman, W.P.; Ross, A.B. Critical-Review of Rate Constants for Reactions of Hydrated Electrons, Hydrogen-Atoms and Hydroxyl Radicals (.OH/O⁻) in Aqueous-Solution. *J. Phys. Chem. Ref. Data* **1988**, *17*, 513–886. [[CrossRef](#)]
54. Farhataziz, R.A.; Ross, A. Selected specific rates of reactions of transients from water in aqueous solution. III. *Hydroxyl Radic. Perhydroxyl Radic. Radic. Ions Nsrds-Nbs* **1977**, *59*.
55. Dorfman, L.M.; Adams, G.E. Reactivity of the hydroxyl radical in aqueous solutions; DTIC Document: 1973.
56. Von Sonntag, C. *The Chemical Basis of Radiation Biology*; Taylor and Francis: Abingdon, UK, 1987; p. 515.

57. Von Sonntag, C. Free-radical reactions involving thiols and disulphides. In *Sulfur-Centered Reactive Intermediates in Chemistry and Biology*; Chatgililoglu, C., Asmus, K.-D., Eds.; Plenum Press: New York, NY, USA, 1990; pp. 359–366.
58. Armstrong, D.A. Application of pulse radiolysis for the study of short-lived sulphur species. In *Sulfur-Centered Reactive Intermediates in Chemistry and Biology*; Chatgililoglu, K.-D.A., Ed.; Plenum Press: New York, NY, USA, 1990; pp. 121–134.
59. Cadet, J.; Delatour, T.; Douki, T.; Gasparutto, D.; Pouget, J.P.; Ravanat, J.L.; Sauvaigo, S. Hydroxyl radicals and DNA base damage. *Mutat. Res.* **1999**, *424*, 9–21. [[CrossRef](#)]
60. Schoneich, C. Mechanisms of metal-catalyzed oxidation of histidine to 2-oxo-histidine in peptides and proteins. *J. Pharm. Biomed. Anal.* **2000**, *21*, 1093–1097. [[CrossRef](#)]
61. Uchida, K.; Kawakishi, S. 2-Oxo-histidine as a novel biological marker for oxidatively modified proteins. *FEBS Lett.* **1993**, *332*, 208–210. [[CrossRef](#)]
62. Berlett, B.S.; Levine, R.L.; Stadtman, E.R. Comparison of the effects of ozone on the modification of amino acid residues in glutamine synthetase and bovine serum albumin. *J. Biol. Chem.* **1996**, *271*, 4177–4182. [[CrossRef](#)]
63. Schoneich, C.; Pogocki, D.; Hug, G.L.; Bobrowski, K. Free radical reactions of methionine in peptides: Mechanisms relevant to beta-amyloid oxidation and Alzheimer's disease. *J. Am. Chem. Soc.* **2003**, *125*, 13700–13713. [[CrossRef](#)]
64. Hoshi, T.; Heinemann, S. Regulation of cell function by methionine oxidation and reduction. *J. Physiol.* **2001**, *531*, 1–11. [[CrossRef](#)]
65. Hasegawa, K.; Patterson, L.K. Pulse-Radiolysis Studies in Model Lipid Systems: Formation and Behavior of Peroxy Radicals in Fatty-Acids. *Photochem. Photobiol.* **1978**, *28*, 817–823. [[CrossRef](#)]
66. Gardner, H.W. Oxygen radical chemistry of polyunsaturated fatty acids. *Free Radic. Biol. Med.* **1989**, *7*, 65–86. [[CrossRef](#)]
67. Yin, H.; Havrilla, C.M.; Gao, L.; Morrow, J.D.; Porter, N.A. Mechanisms for the formation of isoprostane endoperoxides from arachidonic acid. "Dioxetane" intermediate versus beta-fragmentation of peroxy radicals. *J. Biol. Chem.* **2003**, *278*, 16720–16725. [[CrossRef](#)]
68. Greco, A.; Minghetti, L.; Levi, G. Isoprostanes, novel markers of oxidative injury, help understanding the pathogenesis of neurodegenerative diseases. *Neurochem. Res.* **2000**, *25*, 1357–1364. [[CrossRef](#)]
69. Stadtman, E.R. Protein oxidation in aging and age-related diseases. *Ann. N. Y. Acad. Sci.* **2001**, *928*, 22–38. [[CrossRef](#)]
70. Davies, K.J.; Delsignore, M.E.; Lin, S.W. Protein damage and degradation by oxygen radicals. II. Modification of amino acids. *J. Biol. Chem.* **1987**, *262*, 9902–9907. [[PubMed](#)]
71. Breen, A.P.; Murphy, J.A. Reactions of oxyl radicals with DNA. *Free Radic. Biol. Med.* **1995**, *18*, 1033–1077. [[CrossRef](#)]
72. Stadtman, E.R. Oxidation of free amino acids and amino acid residues in proteins by radiolysis and by metal-catalyzed reactions. *Annu. Rev. Biochem.* **1993**, *62*, 797–821. [[CrossRef](#)] [[PubMed](#)]
73. Lal, M. Radiation-Induced Oxidation of Sulfhydryl Molecules in Aqueous-Solutions - a Comprehensive Review. *Radiat. Phys. Chem.* **1994**, *43*, 595–611. [[CrossRef](#)]
74. Atwood, C.S.; Perry, G.; Zeng, H.; Kato, Y.; Jones, W.D.; Ling, K.Q.; Huang, X.; Moir, R.D.; Wang, D.; Sayre, L.M.; et al. Copper mediates dityrosine cross-linking of Alzheimer's amyloid-beta. *Biochemistry* **2004**, *43*, 560–568. [[CrossRef](#)]
75. Garrison, W.M. Reaction-Mechanisms in the Radiolysis of Peptides, Polypeptides, and Proteins. *Chem. Rev.* **1987**, *87*, 381–398. [[CrossRef](#)]
76. Winterbourn, C.C.; Metodiewa, D. Reactivity of biologically important thiol compounds with superoxide and hydrogen peroxide. *Free Radic. Biol. Med.* **1999**, *27*, 322–328. [[CrossRef](#)]
77. Winterbourn, C.C.; Hampton, M.B. Thiol chemistry and specificity in redox signaling. *Free Radic. Biol. Med.* **2008**, *45*, 549–561. [[CrossRef](#)]
78. Vasquez-Vivar, J.; Denicola, A.; Radi, R.; Augusto, O. Peroxynitrite-mediated decarboxylation of pyruvate to both carbon dioxide and carbon dioxide radical anion. *Chem. Res. Toxicol.* **1997**, *10*, 786–794. [[CrossRef](#)]
79. Bakhmutova-Albert, E.V.; Yao, H.; Denevan, D.E.; Richardson, D.E. Kinetics and mechanism of peroxydicarbonate formation. *Inorg. Chem.* **2010**, *49*, 11287–11296. [[CrossRef](#)] [[PubMed](#)]

80. Trindade, D.F.; Cerchiaro, G.; Augusto, O. A role for peroxydicarbonate in the stimulation of biothiols peroxidation by the bicarbonate/carbon dioxide pair. *Chem. Res. Toxicol.* **2006**, *19*, 1475–1482. [[CrossRef](#)] [[PubMed](#)]
81. Richardson, D.E.; Yao, H.R.; Frank, K.M.; Bennett, D.A. Equilibria, kinetics, and mechanism in the bicarbonate activation of hydrogen peroxide: Oxidation of sulfides by peroxydicarbonate. *J. Am. Chem. Soc.* **2000**, *122*, 1729–1739. [[CrossRef](#)]
82. Kowalik-Jankowska, T.; Ruta, M.; Wisniewska, K.; Lankiewicz, L.; Dyba, M. Products of Cu(II)-catalyzed oxidation in the presence of hydrogen peroxide of the 1-10, 1-16 fragments of human and mouse beta-amyloid peptide. *J. Inorg. Biochem.* **2004**, *98*, 940–950. [[CrossRef](#)]
83. Davies, M.J.; Hawkins, C.L.; Pattison, D.I.; Rees, M.D. Mammalian heme peroxidases: From molecular mechanisms to health implications. *Antioxid. Redox Signal.* **2008**, *10*, 1199–1234. [[CrossRef](#)]
84. Davies, M.J.; Dean, R.T. *Radical-Mediated Protein Oxidation: From Chemistry to Medicine*; Oxford University Press: Oxford, UK, 1997.
85. Chevion, M. A site-specific mechanism for free radical induced biological damage: The essential role of redox-active transition metals. *Free Radic. Biol. Med.* **1988**, *5*, 27–37. [[CrossRef](#)]
86. Stadtman, E.R. Metal ion-catalyzed oxidation of proteins: Biochemical mechanism and biological consequences. *Free Radic. Biol. Med.* **1990**, *9*, 315–325. [[CrossRef](#)]
87. Davies, M.J.; Fu, S.; Dean, R.T. Protein hydroperoxides can give rise to reactive free radicals. *Biochem. J.* **1995**, *305*, 643–649. [[CrossRef](#)]
88. Buettner, G.R.; Jurkiewicz, B.A. Catalytic metals, ascorbate and free radicals: Combinations to avoid. *Radiat. Res.* **1996**, *145*, 532–541. [[CrossRef](#)]
89. Von Sonntag, C. Peroxyl Radicals in Aqueous Media. In *Oxygen Radicals in Biology and Medicine*; Simic, M.G., Taylor, K.A., Ward, J.F., von Sonntag, C., Eds.; Springer: Berlin/Heidelberg, Germany, 1988; Volume 49.
90. Von Sonntag, C.; Schuchmann, H.-P. Peroxyl radicals in aqueous solution. In *Peroxyl Radicals*; Alfassi, Z.B., Ed.; John Wiley and Sons: Chichester, UK, 1997; pp. 173–234.
91. Bothe, E.; Schuchmann, M.N.; Schultefrohlinde, D.; Vonsonntag, C. HO₂ Elimination from Alpha-Hydroxyalkylperoxyl Radicals in Aqueous-Solution. *Photochem. Photobiol.* **1978**, *28*, 639–644. [[CrossRef](#)]
92. Niki, E. Role of vitamin E as a lipid-soluble peroxyl radical scavenger: In vitro and in vivo evidence. *Free Radic. Biol. Med.* **2014**, *66*, 3–12. [[CrossRef](#)]
93. El-Agamey, A.; McGarvey, D.J. Peroxyl radical reactions with carotenoids in microemulsions: Influence of microemulsion composition and the nature of peroxyl radical precursor. *Free Radic. Biol. Med.* **2016**, *90*, 75–84. [[CrossRef](#)] [[PubMed](#)]
94. Gilbert, B.C.; Holmes, R.G.G.; Laue, H.A.H.; Norman, R.O.C. Electron-Spin Resonance Studies. 50. Reactions of Alkoxy Radicals Generated from Alkyl Hydroperoxides and Titanium(III) Ion in Aqueous-Solution. *J. Chem. Soc. Perkin Trans.* **1976**, *2*, 1047–1052. [[CrossRef](#)]
95. Gilbert, B.C.; David, P.; Marshall, R.; Norman, R.O.C.; Pineda, N.; Williams, P.S. Electron-Spin Resonance Studies. 61. The Generation and Reactions of the Tert-Butoxyl Radical in Aqueous-Solution. *J. Chem. Soc. Perkin Trans.* **1981**, *2*, 1392–1400. [[CrossRef](#)]
96. Headlam, H.A.; Davies, M.J. Beta-scission of side-chain alkoxy radicals on peptides and proteins results in the loss of side-chains as aldehydes and ketones. *Free Radic. Biol. Med.* **2002**, *32*, 1171–1184. [[CrossRef](#)]
97. Bors, W.; Tait, D.; Michel, C.; Saran, M.; Erbenruss, M. Reactions of Alkoxy Radicals in Aqueous-Solutions. *Isr. J. Chem.* **1984**, *24*, 17–24. [[CrossRef](#)]
98. Erbenruss, M.; Michel, C.; Bors, W.; Saran, M. Absolute Rate Constants of Alkoxy Radical Reactions in Aqueous-Solution. *J. Phys. Chem.* **1987**, *91*, 2362–2365. [[CrossRef](#)]
99. Subczynski, W.K.; Hyde, J.S. Concentration of oxygen in lipid bilayers using a spin-label method. *Biophys. J.* **1983**, *41*, 283–286. [[CrossRef](#)]
100. Beal, M.F.; Ferrante, R.J.; Browne, S.E.; Matthews, R.T.; Kowall, N.W.; Brown, R.H., Jr. Increased 3-nitrotyrosine in both sporadic and familial amyotrophic lateral sclerosis. *Ann. Neurol.* **1997**, *42*, 644–654. [[CrossRef](#)]
101. Ferrante, R.J.; Browne, S.E.; Shinobu, L.A.; Bowling, A.C.; Baik, M.J.; MacGarvey, U.; Kowall, N.W.; Brown, R.H., Jr.; Beal, M.F. Evidence of increased oxidative damage in both sporadic and familial amyotrophic lateral sclerosis. *J. Neurochem.* **1997**, *69*, 2064–2074. [[CrossRef](#)]

102. Giasson, B.I.; Duda, J.E.; Murray, I.V.; Chen, Q.; Souza, J.M.; Hurtig, H.I.; Ischiropoulos, H.; Trojanowski, J.Q.; Lee, V.M. Oxidative damage linked to neurodegeneration by selective alpha-synuclein nitration in synucleinopathy lesions. *Science* **2000**, *290*, 985–989. [[CrossRef](#)]
103. Hensley, K.; Maidt, M.L.; Yu, Z.; Sang, H.; Markesbery, W.R.; Floyd, R.A. Electrochemical analysis of protein nitrotyrosine and dityrosine in the Alzheimer brain indicates region-specific accumulation. *J. Neurosci.* **1998**, *18*, 8126–8132. [[CrossRef](#)]
104. Butterfield, D.A.; Reed, T.; Newman, S.F.; Sultana, R. Roles of amyloid beta-peptide-associated oxidative stress and brain protein modifications in the pathogenesis of Alzheimer's disease and mild cognitive impairment. *Free Radic. Biol. Med.* **2007**, *43*, 658–677. [[CrossRef](#)]
105. Selfridge, J.E.; E., L.; Lu, J.; Swerdlow, R.H. Role of mitochondrial homeostasis and dynamics in Alzheimer's disease. *Neurobiol. Dis.* **2013**, *51*, 3–12. [[CrossRef](#)]
106. Zhao, Y.; Zhao, B. Oxidative stress and the pathogenesis of Alzheimer's disease. *Oxid. Med. Cell. Longev.* **2013**, *2013*, 316523. [[CrossRef](#)]
107. Hands, S.; Sajjad, M.U.; Newton, M.J.; Wyttenbach, A. In vitro and in vivo aggregation of a fragment of huntingtin protein directly causes free radical production. *J. Biol. Chem.* **2011**, *286*, 44512–44520. [[CrossRef](#)]
108. Markesbery, W.R. Oxidative Stress Hypothesis in Alzheimer's Disease. *Free Radic. Biol. Med.* **1997**, *23*, 134–147. [[CrossRef](#)]
109. Butterfield, D.A.; Drake, J.; Pocernich, C.; Castegna, A. Evidence of oxidative damage in Alzheimer's disease brain: Central role for amyloid β -peptide. *Trends Mol. Med.* **2001**, *7*, 548–554. [[CrossRef](#)]
110. Naslund, J.; Schierhorn, A.; Hellman, U.; Lannfelt, L.; Roses, A.D.; Tjernberg, L.O.; Silberring, J.; Gandy, S.E.; Winblad, B.; Greengard, P.; et al. Relative abundance of Alzheimer A beta amyloid peptide variants in Alzheimer disease and normal aging. *Proc. Natl. Acad. Sci. USA* **1994**, *91*, 8378–8382. [[CrossRef](#)]
111. Cheignon, C.; Tomas, M.; Bonnefont-Rousselot, D.; Faller, P.; Hureau, C.; Collin, F. Oxidative stress and the amyloid beta peptide in Alzheimer's disease. *Redox Biol.* **2018**, *14*, 450–464. [[CrossRef](#)]
112. Butterfield, D.A.; Lauderback, C.M. Lipid peroxidation and protein oxidation in Alzheimer's disease brain: Potential causes and consequences involving amyloid beta-peptide-associated free radical oxidative stress. *Free Radic. Biol. Med.* **2002**, *32*, 1050–1060. [[CrossRef](#)]
113. Sultana, R.; Boyd-Kimball, D.; Poon, H.F.; Cai, J.; Pierce, W.M.; Klein, J.B.; Merchant, M.; Markesbery, W.R.; Butterfield, D.A. Redox proteomics identification of oxidized proteins in Alzheimer's disease hippocampus and cerebellum: An approach to understand pathological and biochemical alterations in AD. *Neurobiol. Aging* **2006**, *27*, 1564–1576. [[CrossRef](#)]
114. Hensley, K.; Hall, N.; Subramaniam, R.; Cole, P.; Harris, M.; Aksenov, M.; Aksenova, M.; Gabbita, S.P.; Wu, J.F.; Carney, J.M.; et al. Brain regional correspondence between Alzheimer's disease histopathology and biomarkers of protein oxidation. *J. Neurochem.* **1995**, *65*, 2146–2156. [[CrossRef](#)]
115. Butterfield, D.A.; Yatin, S.M.; Varadarajan, S.; Koppal, T. Amyloid beta-peptide-associated free radical oxidative stress, neurotoxicity, and Alzheimer's disease. *Methods Enzymol.* **1999**, *309*, 746–768.
116. Nitsch, R.M.; Blusztajn, J.K.; Pittas, A.G.; Slack, B.E.; Growdon, J.H.; Wurtman, R.J. Evidence for a membrane defect in Alzheimer disease brain. *Proc. Natl. Acad. Sci. USA* **1992**, *89*, 1671–1675. [[CrossRef](#)]
117. Svennerholm, L.; Gottfries, C.G. Membrane lipids, selectively diminished in Alzheimer brains, suggest synapse loss as a primary event in early-onset form (type I) and demyelination in late-onset form (type II). *J. Neurochem.* **1994**, *62*, 1039–1047. [[CrossRef](#)]
118. Markesbery, W.R.; Lovell, M.A. Four-hydroxynonenal, a product of lipid peroxidation, is increased in the brain in Alzheimer's disease. *Neurobiol. Aging* **1998**, *19*, 33–36. [[CrossRef](#)]
119. Neely, M.D.; Sidell, K.R.; Graham, D.G.; Montine, T.J. The lipid peroxidation product 4-hydroxynonenal inhibits neurite outgrowth, disrupts neuronal microtubules, and modifies cellular tubulin. *J. Neurochem.* **1999**, *72*, 2323–2333. [[CrossRef](#)]
120. Keller, J.N.; Pang, Z.; Geddes, J.W.; Begley, J.G.; Germeyer, A.; Waeg, G.; Mattson, M.P. Impairment of glucose and glutamate transport and induction of mitochondrial oxidative stress and dysfunction in synaptosomes by amyloid beta-peptide: Role of the lipid peroxidation product 4-hydroxynonenal. *J. Neurochem.* **1997**, *69*, 273–284. [[CrossRef](#)]
121. Hardas, S.S.; Sultana, R.; Clark, A.M.; Beckett, T.L.; Szwedda, L.I.; Murphy, M.P.; Butterfield, D.A. Oxidative modification of lipoic acid by HNE in Alzheimer disease brain. *Redox Biol.* **2013**, *1*, 80–85. [[CrossRef](#)]

122. Gegotek, A.; Skrzydlewska, E. Biological effect of protein modifications by lipid peroxidation products. *Chem. Phys. Lipids* **2019**, *221*, 46–52. [[CrossRef](#)]
123. Sayre, L.M.; Zelasko, D.A.; Harris, P.L.; Perry, G.; Salomon, R.G.; Smith, M.A. 4-Hydroxynonenal-derived advanced lipid peroxidation end products are increased in Alzheimer's disease. *J. Neurochem.* **1997**, *68*, 2092–2097. [[CrossRef](#)]
124. Liu, Q.; Smith, M.A.; Avila, J.; DeBernardis, J.; Kansal, M.; Takeda, A.; Zhu, X.; Nunomura, A.; Honda, K.; Moreira, P.I.; et al. Alzheimer-specific epitopes of tau represent lipid peroxidation-induced conformations. *Free Radic. Biol. Med.* **2005**, *38*, 746–754. [[CrossRef](#)]
125. Pryor, W.A.; Porter, N.A. Suggested mechanisms for the production of 4-hydroxy-2-nonenal from the autoxidation of polyunsaturated fatty acids. *Free Radic. Biol. Med.* **1990**, *8*, 541–543. [[CrossRef](#)]
126. Montine, T.J.; Markesbery, W.R.; Morrow, J.D.; Roberts, L.J., 2nd. Cerebrospinal fluid F2-isoprostane levels are increased in Alzheimer's disease. *Ann. Neurol.* **1998**, *44*, 410–413. [[CrossRef](#)]
127. Nourooz-Zadeh, J.; Liu, E.H.; Yhlen, B.; Anggard, E.E.; Halliwell, B. F4-isoprostanes as specific marker of docosahexaenoic acid peroxidation in Alzheimer's disease. *J. Neurochem.* **1999**, *72*, 734–740. [[CrossRef](#)]
128. Reich, E.E.; Markesbery, W.R.; Roberts, L.J., 2nd; Swift, L.L.; Morrow, J.D.; Montine, T.J. Brain regional quantification of F-ring and D-/E-ring isoprostanes and neuroprostanes in Alzheimer's disease. *Am. J. Pathol.* **2001**, *158*, 293–297. [[CrossRef](#)]
129. Smith, M.A.; Perry, G.; Richey, P.L.; Sayre, L.M.; Anderson, V.E.; Beal, M.F.; Kowall, N. Oxidative damage in Alzheimer's. *Nature* **1996**, *382*, 120–121. [[CrossRef](#)] [[PubMed](#)]
130. Tramutola, A.; Lanzillotta, C.; Perluigi, M.; Butterfield, D.A. Oxidative stress, protein modification and Alzheimer disease. *Brain Res. Bull.* **2016**. [[CrossRef](#)] [[PubMed](#)]
131. Terni, B.; Boada, J.; Portero-Otin, M.; Pamplona, R.; Ferrer, I. Mitochondrial ATP-synthase in the entorhinal cortex is a target of oxidative stress at stages I/II of Alzheimer's disease pathology. *Brain Pathol.* **2010**, *20*, 222–233. [[CrossRef](#)]
132. Butterfield, D.A.; Boyd-Kimball, D. Redox proteomics and amyloid beta-peptide: Insights into Alzheimer disease. *J. Neurochem.* **2018**. [[CrossRef](#)] [[PubMed](#)]
133. Mecocci, P.; MacGarvey, U.; Beal, M.F. Oxidative damage to mitochondrial DNA is increased in Alzheimer's disease. *Ann. Neurol.* **1994**, *36*, 747–751. [[CrossRef](#)]
134. Gabbita, S.P.; Lovell, M.A.; Markesbery, W.R. Increased nuclear DNA oxidation in the brain in Alzheimer's disease. *J. Neurochem.* **1998**, *71*, 2034–2040. [[CrossRef](#)]
135. Lovell, M.A.; Soman, S.; Bradley, M.A. Oxidatively modified nucleic acids in preclinical Alzheimer's disease (PCAD) brain. *Mech. Ageing Dev.* **2011**, *132*, 443–448. [[CrossRef](#)]
136. Nunomura, A.; Perry, G.; Pappolla, M.A.; Wade, R.; Hirai, K.; Chiba, S.; Smith, M.A. RNA oxidation is a prominent feature of vulnerable neurons in Alzheimer's disease. *J. Neurosci.* **1999**, *19*, 1959–1964. [[CrossRef](#)]
137. Jenner, P. Oxidative stress in Parkinson's disease. *Ann. Neurol.* **2003**, *53* (Suppl. 3), S26–S38. [[CrossRef](#)]
138. Bosco, D.A.; Fowler, D.M.; Zhang, Q.; Nieva, J.; Powers, E.T.; Wentworth, P., Jr.; Lerner, R.A.; Kelly, J.W. Elevated levels of oxidized cholesterol metabolites in Lewy body disease brains accelerate alpha-synuclein fibrilization. *Nat. Chem. Biol.* **2006**, *2*, 249–253. [[CrossRef](#)]
139. Alam, Z.I.; Daniel, S.E.; Lees, A.J.; Marsden, D.C.; Jenner, P.; Halliwell, B. A generalised increase in protein carbonyls in the brain in Parkinson's but not incidental Lewy body disease. *J. Neurochem.* **1997**, *69*, 1326–1329. [[CrossRef](#)] [[PubMed](#)]
140. Nakabeppu, Y.; Tsuchimoto, D.; Yamaguchi, H.; Sakumi, K. Oxidative damage in nucleic acids and Parkinson's disease. *J. Neurosci. Res.* **2007**, *85*, 919–934. [[CrossRef](#)]
141. Selley, M.L. (E)-4-hydroxy-2-nonenal may be involved in the pathogenesis of Parkinson's disease. *Free Radic. Biol. Med.* **1998**, *25*, 169–174. [[CrossRef](#)]
142. Halliwell, B. The wanderings of a free radical. *Free Radic. Biol. Med.* **2009**, *46*, 531–542. [[CrossRef](#)]
143. Jellinger, K.; Kienzl, E.; Rumpelmair, G.; Riederer, P.; Stachelberger, H.; Ben-Shachar, D.; Youdim, M.B. Iron-melanin complex in substantia nigra of parkinsonian brains: An x-ray microanalysis. *J. Neurochem.* **1992**, *59*, 1168–1171. [[CrossRef](#)]
144. Fahn, S.; Cohen, G. The oxidant stress hypothesis in Parkinson's disease: Evidence supporting it. *Ann. Neurol.* **1992**, *32*, 804–812. [[CrossRef](#)]
145. Surgucheva, I.; Sharov, V.S.; Surguchov, A. gamma-Synuclein: Seeding of alpha-synuclein aggregation and transmission between cells. *Biochemistry* **2012**, *51*, 4743–4754. [[CrossRef](#)]

146. Mendez, E.F.; Sattler, R. Biomarker development for C9orf72 repeat expansion in ALS. *Brain Res.* **2015**, *1607*, 26–35. [[CrossRef](#)]
147. Smith, R.G.; Henry, Y.K.; Mattson, M.P.; Appel, S.H. Presence of 4-hydroxynonenal in cerebrospinal fluid of patients with sporadic amyotrophic lateral sclerosis. *Ann. Neurol.* **1998**, *44*, 696–699. [[CrossRef](#)]
148. Bogdanov, M.; Brown, R.H.; Matson, W.; Smart, R.; Hayden, D.; O'Donnell, H.; Flint Beal, M.; Cudkowicz, M. Increased oxidative damage to DNA in ALS patients. *Free Radic. Biol. Med.* **2000**, *29*, 652–658. [[CrossRef](#)]
149. Miller, E.; Morel, A.; Saso, L.; Saluk, J. Isoprostanes and neuroprostanes as biomarkers of oxidative stress in neurodegenerative diseases. *Oxid. Med. Cell. Longev.* **2014**, *2014*, 572491. [[CrossRef](#)]
150. Mitsumoto, H.; Santella, R.M.; Liu, X.; Bogdanov, M.; Zipprich, J.; Wu, H.C.; Mahata, J.; Kilty, M.; Bednarz, K.; Bell, D.; et al. Oxidative stress biomarkers in sporadic ALS. *Amyotroph. Lateral Scler.* **2008**, *9*, 177–183. [[CrossRef](#)]
151. Bozzo, F.; Mirra, A.; Carri, M.T. Oxidative stress and mitochondrial damage in the pathogenesis of ALS: New perspectives. *Neurosci. Lett.* **2017**, *636*, 3–8. [[CrossRef](#)]
152. Blokhuis, A.M.; Groen, E.J.; Koppers, M.; van den Berg, L.H.; Pasterkamp, R.J. Protein aggregation in amyotrophic lateral sclerosis. *Acta Neuropathol.* **2013**, *125*, 777–794. [[CrossRef](#)]
153. Lovejoy, D.B.; Guillemin, G.J. The potential for transition metal-mediated neurodegeneration in amyotrophic lateral sclerosis. *Front. Aging Neurosci.* **2014**, *6*, 173. [[CrossRef](#)]
154. Rosas, H.D.; Chen, Y.I.; Doros, G.; Salat, D.H.; Chen, N.K.; Kwong, K.K.; Bush, A.; Fox, J.; Hersch, S.M. Alterations in brain transition metals in Huntington disease: An evolving and intricate story. *Arch. Neurol.* **2012**, *69*, 887–893. [[CrossRef](#)]
155. Hersch, S.M.; Gevorkian, S.; Marder, K.; Moskowitz, C.; Feigin, A.; Cox, M.; Como, P.; Zimmerman, C.; Lin, M.; Zhang, L.; et al. Creatine in Huntington disease is safe, tolerable, bioavailable in brain and reduces serum 8OH2'dG. *Neurology* **2006**, *66*, 250–252. [[CrossRef](#)]
156. Chen, C.M.; Wu, Y.R.; Cheng, M.L.; Liu, J.L.; Lee, Y.M.; Lee, P.W.; Soong, B.W.; Chiu, D.T. Increased oxidative damage and mitochondrial abnormalities in the peripheral blood of Huntington's disease patients. *Biochem. Biophys. Res. Commun.* **2007**, *359*, 335–340. [[CrossRef](#)]
157. Stack, E.C.; Matson, W.R.; Ferrante, R.J. Evidence of oxidant damage in Huntington's disease: Translational strategies using antioxidants. *Ann. N. Y. Acad. Sci.* **2008**, *1147*, 79–92. [[CrossRef](#)]
158. Dragunow, M.; Faull, R.L.; Lawlor, P.; Beilharz, E.J.; Singleton, K.; Walker, E.B.; Mee, E. In situ evidence for DNA fragmentation in Huntington's disease striatum and Alzheimer's disease temporal lobes. *Neuroreport* **1995**, *6*, 1053–1057. [[CrossRef](#)]
159. Browne, S.E.; Ferrante, R.J.; Beal, M.F. Oxidative stress in Huntington's disease. *Brain Pathol.* **1999**, *9*, 147–163. [[CrossRef](#)]
160. Tunes, I.; Sanchez-Lopez, F.; Aguera, E.; Fernandez-Bolanos, R.; Sanchez, F.M.; Tasset-Cuevas, I. Important role of oxidative stress biomarkers in Huntington's disease. *J. Med. Chem.* **2011**, *54*, 5602–5606. [[CrossRef](#)]
161. LeBars, P.L.; Katz, M.M.; Berman, N.; Itil, T.M.; Freedman, A.M.; Schatzberg, A.F. A placebo-controlled, double-blind, randomized trial of an extract of Ginkgo biloba for dementia. *J. Am. Med. Assoc.* **1997**, *278*, 1327–1332.
162. Purushothuman, S.; Nandasena, C.; Peoples, C.L.; El Massri, N.; Johnstone, D.M.; Mitrofanis, J.; Stone, J. Saffron Pre-Treatment Offers Neuroprotection to Nigral and Retinal Dopaminergic Cells of MPTP-Treated mice. *J. Parkinson Dis.* **2013**, *3*, 77–83.
163. Barkats, M.; Millecamps, S.; Abrioux, P.; Geoffroy, M.C.; Mallet, J. Overexpression of glutathione peroxidase increases the resistance of neuronal cells to Abeta-mediated neurotoxicity. *J. Neurochem.* **2000**, *75*, 1438–1446. [[CrossRef](#)]
164. Liu, Z.; Zhou, T.; Ziegler, A.C.; Dimitrion, P.; Zuo, L. Oxidative Stress in Neurodegenerative Diseases: From Molecular Mechanisms to Clinical Applications. *Oxid. Med. Cell. Longev.* **2017**, *2017*, 2525967. [[CrossRef](#)]

