Supporting Information for

Palladacyclic conjugate group promotes hybridization of short oligonucleotides

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S2





















S8









S10



Figure S1. IE-HPLC trace of oligonucleotide ON1b; $ProSwift^{TM}$ SAX-1S column (50 × 1 mm, monolithic); flow rate = 0.20 mL min⁻¹; linear gradient (0.10 to 0.90 M over 15 min) of NaCl in 10 mM TRIS•HCl buffer (pH 7.6).



Figure S2. IE-HPLC trace of oligonucleotide ON2b; ProSwiftTM SAX-1S column (50 \times 1 mm, monolithic); flow rate = 0.20 mL min⁻¹; linear gradient (0.10 to 0.90 M over 15 min) of NaCl in 10 mM TRIS•HCl buffer (pH 7.6).



Figure S3. IE-HPLC trace of oligonucleotide ON3b; ProSwiftTM SAX-1S column (50 \times 1 mm, monolithic); flow rate = 0.20 mL min⁻¹; linear gradient (0.10 to 0.90 M over 15 min) of NaCl in 10 mM TRIS•HCl buffer (pH 7.6).



Figure S4. IE-HPLC trace of oligonucleotide ON4b; ProSwiftTM SAX-1S column (50 \times 1 mm, monolithic); flow rate = 0.20 mL min⁻¹; linear gradient (0.10 to 0.90 M over 15 min) of NaCl in 10 mM TRIS•HCl buffer (pH 7.6).



Figure S5. RP-HPLC traces of oligonucleotides A) ON1b-Pd, B) ON2b-Pd, C) ON3b-Pd and D) ON4b-Pd; Hypersil ODS C18 column ($250 \times 4.6 \text{ mm}$, 5 µm); flow rate = 1.0 mL min⁻¹; linear gradient (0 to 30% over 25 min) of MeCN in 50 mM aq. triethylammonium acetate.



Figure S6. Mass spectrum of oligonucleotide ON1b.



Figure S7. Mass spectrum of oligonucleotide ON2b.



Figure S8. Mass spectrum of oligonucleotide ON3b.



Figure S9. Mass spectrum of oligonucleotide ON4b.



Figure S10. Mass spectrum of oligonucleotide ON1b-Pd.



Figure S11. Mass spectrum of oligonucleotide ON2b-Pd.



Figure S12. Mass spectrum of oligonucleotide ON3b-Pd.



Figure S13. Mass spectrum of oligonucleotide ON4b-Pd.

Table S1. Melting temperatures of duplexes formed by ON1a, ON1b, ON1b-Pd, ON2a, ON2b, ON2	D-
Pd, ON3a, ON3b, ON3b-Pd, ON4a, ON4b and ON4b-Pd with ON5a, ON5c, ON5g and ON5t; pH = 7	.4
(20 mM cacodylate buffer); [oligonucleotides] = 3.0μ M; I(NaClO ₄) = 0.10μ M.	

	T _m / °C				
	ON5a	ON5c	ON5g	ON5t	
ON1a	40.4 ± 0.7	n.a. ¹	17.6 ± 0.4	17.9 ± 0.3	
ON1b	35.8 ± 0.6	n.a. ¹	16.0 ± 0.7	16.9 ± 0.4	
ON1b-Pd	41.0 ± 0.1	n.a. ¹	22.6 ± 0.4	22.4 ± 0.7	
ON2a	31.8 ± 1.3	n.a. ¹	n.a. ¹	n.a. 1	
ON2b	30.5 ± 1.3	n.a. ¹	n.a. ¹	n.a. ¹	
ON2b-Pd	32.7 ± 1.0	n.a. ¹	n.a. ¹	n.a. ¹	
ON3a	21.0 ± 0.9	n.a. ¹	n.a. ¹	n.a. ¹	
ON3b	20.5 ± 0.8	n.a. ¹	n.a. ¹	n.a. ¹	
ON3b-Pd	21.2 ± 1.2	n.a. ¹	n.a. ¹	n.a. ¹	
ON4a	16.1 ± 0.4	n.a. ¹	n.a. ¹	n.a. ¹	
ON4b	15.4 ± 0.8	n.a. ¹	n.a. ¹	n.a. ¹	
ON4b-Pd	15.7 ± 0.8	n.a. ¹	n.a. ¹	n.a. ¹	

¹ No sigmoidal melting curve was obtained.

Table S2. Melting temperatures of duplexes formed by ON1a, ON1b, ON1b-Pd, ON2a, ON2b, ON2b-Pd, ON3a, ON3b, ON3b-Pd, ON4a, ON4b and ON4b-Pd with ON5a, ON5c, ON5g and ON5t in the presence of 2-mercaptoethanol; pH = 7.4 (20 mM cacodylate buffer); [oligonucleotides] = 3.0μ M; [2-mercaptoethanol] = 100μ M; I(NaClO₄) = 0.10 M.

	T _m / °C				
	ON5a	ON5c	ON5g	ON5t	
ON1a	40.8 ± 0.6	n.a. ¹	16.2 ± 0.7	15.5 ± 0.3	
ON1b	34.9 ± 1.1	n.a. ¹	16.8 ± 0.6	15.5 ± 1.1	
ON1b-Pd	38.3 ± 0.9	n.a. 1	17.3 ± 0.8	15.3 ± 0.7	
ON2a	30.9 ± 1.2	n.a. 1	n.a. ¹	n.a. ¹	
ON2b	29.3 ± 0.8	n.a. 1	n.a. ¹	n.a. ¹	
ON2b-Pd	30.2 ± 0.1	n.a. 1	n.a. ¹	n.a. ¹	
ON3a	19.0 ± 0.5	n.a. ¹	n.a. ¹	n.a. ¹	
ON3b	20.0 ± 0.9	n.a. 1	n.a. ¹	n.a. ¹	
ON3b-Pd	19.3 ± 0.3	n.a. ¹	n.a. ¹	n.a. ¹	
ON4a	17.3 ± 1.0	n.a. 1	n.a. ¹	n.a. ¹	
ON4b	17.0 ± 0.8	n.a. 1	n.a. ¹	n.a. ¹	
ON4b-Pd	17.6 ± 0.9	n.a. ¹	n.a. ¹	n.a. ¹	

¹ No sigmoidal melting curve was obtained.



Figure S14. CD spectra of ON1b-Pd•ON5a, recorded at 10 °C intervals between 10 and 90 °C; pH = 7.4 (20 mM cacodylate buffer); [oligonucleotides] = 3.0μ M; I(NaClO₄) = 0.10 M. Spectra acquired at extreme temperatures are indicated by thicker lines and thermal shifts of the minima and maxima by arrows.



Figure S15. CD spectra of ON1b-Pd•ON5c, recorded at 10 °C intervals between 10 and 90 °C; pH = 7.4 (20 mM cacodylate buffer); [oligonucleotides] = 3.0μ M; I(NaClO₄) = 0.10 M. Spectra acquired at extreme temperatures are indicated by thicker lines and thermal shifts of the minima and maxima by arrows.



Figure S16. CD spectra of ON1b-Pd•ON5g, recorded at 10 °C intervals between 10 and 90 °C; pH = 7.4 (20 mM cacodylate buffer); [oligonucleotides] = 3.0μ M; I(NaClO₄) = 0.10 M. Spectra acquired at extreme temperatures are indicated by thicker lines and thermal shifts of the minima and maxima by arrows.



Figure S17. CD spectra of ON1b-Pd•ON5t, recorded at 10 °C intervals between 10 and 90 °C; pH = 7.4 (20 mM cacodylate buffer); [oligonucleotides] = 3.0μ M; I(NaClO₄) = 0.10 M. Spectra acquired at extreme temperatures are indicated by thicker lines and thermal shifts of the minima and maxima by arrows.



Figure S18. CD spectra of ON2b-Pd•ON5a, recorded at 10 °C intervals between 10 and 90 °C; pH = 7.4 (20 mM cacodylate buffer); [oligonucleotides] = 3.0μ M; I(NaClO₄) = 0.10 M. Spectra acquired at extreme temperatures are indicated by thicker lines and thermal shifts of the minima and maxima by arrows.



Figure S19. CD spectra of ON2b-Pd•ON5c, recorded at 10 °C intervals between 10 and 90 °C; pH = 7.4 (20 mM cacodylate buffer); [oligonucleotides] = 3.0μ M; I(NaClO₄) = 0.10 M. Spectra acquired at extreme temperatures are indicated by thicker lines and thermal shifts of the minima and maxima by arrows.



Figure S20. CD spectra of ON2b-Pd•ON5g, recorded at 10 °C intervals between 10 and 90 °C; pH = 7.4 (20 mM cacodylate buffer); [oligonucleotides] = 3.0μ M; I(NaClO₄) = 0.10 M. Spectra acquired at extreme temperatures are indicated by thicker lines and thermal shifts of the minima and maxima by arrows.



Figure S21. CD spectra of ON2b-Pd•ON5t, recorded at 10 °C intervals between 10 and 90 °C; pH = 7.4 (20 mM cacodylate buffer); [oligonucleotides] = 3.0μ M; I(NaClO₄) = 0.10 M. Spectra acquired at extreme temperatures are indicated by thicker lines and thermal shifts of the minima and maxima by arrows.



Figure S22. CD spectra of ON3b-Pd•ON5a, recorded at 10 °C intervals between 10 and 90 °C; pH = 7.4 (20 mM cacodylate buffer); [oligonucleotides] = 3.0μ M; I(NaClO₄) = 0.10 M. Spectra acquired at extreme temperatures are indicated by thicker lines and thermal shifts of the minima and maxima by arrows.



Figure S23. CD spectra of ON3b-Pd•ON5c, recorded at 10 °C intervals between 10 and 90 °C; pH = 7.4 (20 mM cacodylate buffer); [oligonucleotides] = 3.0μ M; I(NaClO₄) = 0.10 M. Spectra acquired at extreme temperatures are indicated by thicker lines and thermal shifts of the minima and maxima by arrows.



Figure S24. CD spectra of ON3b-Pd•ON5g, recorded at 10 °C intervals between 10 and 90 °C; pH = 7.4 (20 mM cacodylate buffer); [oligonucleotides] = 3.0μ M; I(NaClO₄) = 0.10 M. Spectra acquired at extreme temperatures are indicated by thicker lines and thermal shifts of the minima and maxima by arrows.



Figure S25. CD spectra of ON3b-Pd•ON5t, recorded at 10 °C intervals between 10 and 90 °C; pH = 7.4 (20 mM cacodylate buffer); [oligonucleotides] = 3.0μ M; I(NaClO₄) = 0.10 M. Spectra acquired at extreme temperatures are indicated by thicker lines and thermal shifts of the minima and maxima by arrows.



Figure S26. CD spectra of ON4b-Pd•ON5a, recorded at 10 °C intervals between 10 and 90 °C; pH = 7.4 (20 mM cacodylate buffer); [oligonucleotides] = 3.0μ M; I(NaClO₄) = 0.10 M. Spectra acquired at extreme temperatures are indicated by thicker lines and thermal shifts of the minima and maxima by arrows.



Figure S27. CD spectra of ON4b-Pd•ON5c, recorded at 10 °C intervals between 10 and 90 °C; pH = 7.4 (20 mM cacodylate buffer); [oligonucleotides] = 3.0μ M; I(NaClO₄) = 0.10 M. Spectra acquired at extreme temperatures are indicated by thicker lines and thermal shifts of the minima and maxima by arrows.



Figure S28. CD spectra of ON4b-Pd•ON5g, recorded at 10 °C intervals between 10 and 90 °C; pH = 7.4 (20 mM cacodylate buffer); [oligonucleotides] = 3.0μ M; I(NaClO₄) = 0.10 M. Spectra acquired at extreme temperatures are indicated by thicker lines and thermal shifts of the minima and maxima by arrows.



Figure S29. CD spectra of ON4b-Pd•ON5t, recorded at 10 °C intervals between 10 and 90 °C; pH = 7.4 (20 mM cacodylate buffer); [oligonucleotides] = 3.0μ M; I(NaClO₄) = 0.10 M. Spectra acquired at extreme temperatures are indicated by thicker lines and thermal shifts of the minima and maxima by arrows.