

Supplementary Information

Table S1. Enzyme capacities (units g⁻¹ FW) of superoxide dismutase (SOD), catalase (CAT), guaiacol peroxidase (GPX) and syringaldazine peroxidase (SPX) of *Arabidopsis thaliana* roots and SOD and ascorbate peroxidase (APX) capacities of *Arabidopsis thaliana* leaves exposed to different U concentrations for 3 days at pH 7.5. Values represent the mean ± S.E. of at least 4 biological replicates. Data points with different letters are significantly different ($p < 0.05$).

Organ	Enzyme	0 µM U	6.25 µM U	12.5 µM U	25 µM U	50 µM U	75 µM U	100 µM U
ROOTS	SOD	124 ± 14 ^A	130 ± 19 ^A	135 ± 25 ^A	123 ± 23 ^A	99 ± 12 ^A	154 ± 26 ^A	128 ± 21 ^A
	CAT ($\times 10^{-3}$)	45 ± 8 ^A	66 ± 10 ^A	57 ± 11 ^A	81 ± 9 ^A	54 ± 8 ^A	77 ± 12 ^A	60 ± 4 ^A
	GPX	8.9 ± 0.9 ^A	12.58 ± 1.3 ^A	12.7 ± 1.6 ^A	14.7 ± 1.2 ^A	13.1 ± 1.5 ^A	15.9 ± 1.7 ^A	17.0 ± 3.2 ^A
	SPX	0.57 ± 0.21 ^A	1.33 ± 0.38 ^A	0.78 ± 0.19 ^A	1.81 ± 0.36 ^A	1.07 ± 0.15 ^A	1.24 ± 0.44 ^A	1.12 ± 0.47 ^A
LEAVES	SOD	134 ± 7 ^a	159 ± 12 ^a	154 ± 25 ^a	159 ± 12 ^a	153 ± 17 ^a	171 ± 23 ^a	169 ± 12 ^a
	APX	11.4 ± 1.6 ^a	15.3 ± 2.8 ^a	16.7 ± 3.1 ^a	13.8 ± 2.3 ^a	10.5 ± 1.4 ^a	16.8 ± 2.1 ^a	14.2 ± 3.2 ^a

Table S2. Element concentrations (mg g^{-1} DW) in roots and leaves of *Arabidopsis thaliana* seedlings after exposure to different uranium concentrations for 3 days. Results represent mean \pm S.E. of at least 3 biological replicates. Data points with different letters are significantly different ($p < 0.05$). N/A: metal content in plants exposed to 0 μM U was not determined. n.d.: not detectable. *: value based on one measurement.

Organ	Element	0 μM U	6.25 μM U	12.5 μM U	25 μM U	50 μM U	75 μM U	100 μM U
ROOTS	Fe	N/A	$495.83 \pm 41.80^{\text{AB}}$	$441.59 \pm 74.31^{\text{A}}$	$516.85 \pm 48.47^{\text{AB}}$	$685.25 \pm 43.26^{\text{B}}$	$418.44 \pm 14.57^{\text{AB}}$	$409.41 \pm 20.63^{\text{AB}}$
	Cu	N/A	n.d.	n.d.	$19.21 \pm 0.82^{\text{A}}$	$29.87 \pm 1.89^{\text{B}}$	$40.47 \pm 1.12^{\text{C}}$	$41.62 \pm 0.74^{\text{C}}$
LEAVES	Fe	N/A	$49.58 \pm 1.68^{\text{a}}$	$48.40 \pm 2.39^{\text{a}}$	$48.17 \pm 1.64^{\text{a}}$	$45.60 \pm 0.73^{\text{ab}}$	$41.32 \pm 1.22^{\text{b}}$	$41.63 \pm 1.48^{\text{b}}$
	Cu	N/A	$6.50 \pm 0.41^{\text{a}}$	$6.95 \pm 0.72^{\text{a}}$	$6.85 \pm 0.18^{\text{a}}$	$5.71^{\text{a}*}$	n.d.	n.d.

Table S3. Sequences of forward and reverse primers used in gene expression analysis.

Locus	Gene	Forward Primer	Reverse Primer
Reference Genes			
<i>At2g28390</i>	<i>SAND family protein expressed</i>	AACTCTATGCAGCATTGATCCACT	TGATTGCATATCTTATGCCATC
<i>At4g26410</i>		GAGCTGAAGTGGCTTCATGAC	GGTCCGACATAACCATGATCC
<i>At4g34270</i>	<i>TIP41-like</i>	GTGAAAACGTGGAGAGAAGCAA	TCAACTGGATACCCCTTCGCA
<i>At5g08290</i>	<i>Mitosis protein YSL8</i>	TTACTGTTCGGGTGTCTCCATT	CACTGAATCATGTTGAAGCAAGT
<i>At5g15710</i>	<i>F-box protein</i>	TTTCGGCTGAGAGGTTGAGT	GATTCCAAGACGTAAAGCAGATCAA
<i>At5g25760</i>	<i>UBC</i>	CTGCGACTCAGGAAATCTTCTAA	TTGTGCCATTGAATTGAACCC
<i>At5g55840</i>	<i>PPR gene</i>	AAGACAGTGAAGGTGCAACCTTACT	AGTTTTGAGTTGATTGTCAGAGAAAG
<i>At4g05320</i>	<i>UBQ10</i>	GGCCTTGATAATCCCTGATGAATAAG	AAAGAGATAACAGGAACGGAAACATAGT
Other Genes			
<i>At1g07890</i>	<i>APX1</i>	TGCCACAAGGATAGGTCTGG	CCTTCCTTCTCTCGCTCAA
<i>At1g20630</i>	<i>CAT1</i>	AAGTGCTTCATCGGGAAGGA	CTTCAACAAAACGCTTCACGA
<i>At4g35090</i>	<i>CAT2</i>	AACTCCTCCATGACCGTTGGA	TCCGTTCCCTGTCGAAATTG
<i>At1g20620</i>	<i>CAT3</i>	TCTCCAACAAACATCTTCCCTCA	GTGAAATTAGCAACCTCTCGATCA
<i>At1g08830</i>	<i>CSD1</i>	TCCATGCAGACCCGTATGAC	CCTGGAGACCAATGATGCC
<i>At2g28190</i>	<i>CSD2</i>	GAGCCTTGTGGTTACGAG	CACACCACATGCAATCTCC
<i>At5g18100</i>	<i>CSD3</i>	GTTGTTGTGCATGCGGATCC	CACATCCAACCTCTCGAGCCTG
<i>At4g25100</i>	<i>FSD1</i>	CTCCCAATGCTGTGAATCCC	TGGTCTCGGTTCTGGAAGTC
<i>At5g51100</i>	<i>FSD2</i>	TTGGAAAGGTTCAAGTCGGCT	CATTGCAACGTCAAGTCTATTG
<i>At5g23310</i>	<i>FSD3</i>	AACGGGAATCCTTACCCGA	TGTCTCCACCACCAAGGTTGC
<i>At3g24170</i>	<i>GR1</i>	CTCAAGTGTGGAGCAACCAAAG	ATCGTCTGGTCACACTG
<i>At3g54660</i>	<i>GR2</i>	GCCCAGATGGATGGAACAGAT	TAGGGTTGGAGAATGTTGGCG
<i>At4g23100</i>	<i>GSH1</i>	CCCTGGTGAAC TGCCCTCA	CATCAGCACCTCTCATCTCCA
<i>At5g2738</i>	<i>GSH2</i>	GGACTCGTCGTTGGTACAA	TCTGGGAATGCAGTTGGTAGC
<i>At1g55020</i>	<i>LOX1</i>	TTGGCTAACGGCTTTGTCGG	GTGGCAATCACAAACGGTTC
<i>At3g10920</i>	<i>MSD1</i>	ATGTTGGGAGCACGCCTAC	AACCTCGCTTGCATATTCCA
<i>At5g44070</i>	<i>PCS1</i>	TGGTGTGAATGCTTTCTATCG	GGTCGCAGCAATCCAACAT
<i>At5g14545</i>	<i>pri-miRNA398b</i>	AGTAATCAACGGCTGTAATGACGCTAC	TGACCTGAGAACACATGAAAACGAGAG
<i>At5g14565</i>	<i>pri-miRNA398c</i>	TCGAAACTCAAACGTAAACAGTCC	ATTGGTAAATGAATAGAACGCCACGGGCCACG
<i>At5g51060</i>	<i>RBOHC</i>	TCACCAAGAGACTGGCACAATAAA	GATGCTCGACCTGAATGCTC
<i>At5g47910</i>	<i>RBOHD</i>	TATGCATCGGAGAGGGCTGCT	TAGAGACAACACGTTCCCGGG
<i>At1g64060</i>	<i>RBOHF</i>	GGTGTGATGAACGAAGTTGCA	AATGAGAGCAGAACGAGCATCA