



## Review

# Walnut By-Products and Elderberry Extracts—Sustainable Alternatives for Human and Plant Health

Anca Sandu-Bălan (Tăbăcariu)<sup>1</sup>, Irina-Loredana Ifrim<sup>2,\*</sup> , Oana-Irina Patriciu<sup>2,\*</sup> , Ioana-Adriana Ștefănescu<sup>2</sup> and Adriana-Luminița Fînaru<sup>2</sup>

<sup>1</sup> Doctoral School in Environmental Engineering, “Vasile Alecsandri” University of Bacau, 157 Marasesti Str., 600115 Bacau, Romania; anca\_tabacariu@yahoo.com

<sup>2</sup> Department of Chemical and Food Engineering, “Vasile Alecsandri” University of Bacau, 157 Marasesti Str., 600115 Bacau, Romania; adrianaf@ub.ro (A.-L.F.)

\* Correspondence: irinaifrim@ub.ro (I.-L.I.); oana.patriciu@ub.ro (O.-I.P.)

**Abstract:** A current alternative for sustainable development through green chemistry is the replacement of synthetic compounds with natural ones through the superior capitalization of natural resources, with numerous applications in different fields. The benefits of walnuts (*Juglans regia* L.) and elderberries (*Sambucus nigra* L.) have been known since ancient times, due to the presence of phytochemicals such as flavonoids, polyphenols, carotenoids, alkaloids, nitrogen-containing compounds, tannins, steroids, anthocyanins, etc. These active compounds have multiple biological activities for human health, including benefits that are antibacterial, antioxidant, anti-inflammatory, antidiabetic, hepatoprotective, antihypertensive, neuroprotective, etc. Like other medicinal plants, the walnut and the elderberry possess important phytosanitary properties (antibacterial, antifungal, and insecticidal) and their extracts can also be used as environmentally safe biopesticides, with the result that they constitute a viable and cheap alternative to environmentally harmful synthetic products. During recent years, walnut by-products and elderberries have attracted the attention of researchers, and investigations have focused on the species’ valuable constituents and active properties. Comparing the information from the literature regarding the phytochemical profile and biological activities, it is highlighted that, apart from the predominant specific compounds, the walnut and the elderberry have common bioactive compounds, which come from six classes (phenols and derivatives, flavonoids, hydroxycinnamic acids, tannins, triterpenoids, and phytosteroids), and act on the same microorganisms. From this perspective, the aim of this review is to provide an overview of the bioactive compounds present in the different constitutive parts of walnut by-products and elderberries, which present a specific or common activity related to human health and the protection of agricultural crops in the context of sustainable development.

**Keywords:** walnut; elderberry; extracts; green chemistry; biological activities; phytosanitary properties



**Citation:** Sandu-Bălan, A.; Ifrim, I.-L.; Patriciu, O.-I.; Ștefănescu, I.-A.; Fînaru, A.-L. Walnut By-Products and Elderberry Extracts—Sustainable Alternatives for Human and Plant Health. *Molecules* **2024**, *29*, 498. <https://doi.org/10.3390/molecules29020498>

Academic Editor: Ângelo Luís

Received: 20 December 2023

Revised: 11 January 2024

Accepted: 15 January 2024

Published: 19 January 2024



**Copyright:** © 2024 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (<https://creativecommons.org/licenses/by/4.0/>).

## 1. Introduction

Maintaining the health of the population, as well as the quality of the environment in which we live, requires the correct management of economic relations, social resources, and, even more importantly, everything that can be considered as natural resources.

In this regard, special interest is given to plants that have been and are used in traditional medicine due to the rich content of their different compounds with bioactive properties. In significant proportions, plants are used in the pharmaceutical and nutritional supplement industry as primary sources for the extraction of natural compounds with beneficial effects, especially for preventive but also therapeutic purposes [1–4].

Among the fundamental needs of modern industry, because of overexploitation and globalization, are sustainability and the so-called “green chemistry”, which involves replacing synthetic compounds with natural ones from plants, in order to capitalize and develop

natural extracts, with numerous applications in different fields, such as plant culture, the pharmaceutical industry, food, textiles, the cosmetic industry, etc. [5].

The walnut (*Juglans regia* L.) has been cultivated in Romania since ancient times, and it is known that the Geto-Dacians used walnut oil in their diet. Developed over the centuries, this tradition has propelled Romania to being among the largest walnut-producing countries in the world. Until 1900, the walnut occupied large areas of forests and plantations on the forest floor [6]. Romania remains one of the largest producers of walnuts in the European Union, according to data published by the European Commission. For 2021, Eurostat data indicate a walnut production in Romania of 60,820 tons [7]. In 2022, approximately 2200 ha of walnuts were harvested in Romania [8]. According to the National Institute of Statistics' 2022 report, it is noteworthy that, regarding the regional distribution, in the northeast region of the country, the areas of Bacău, Iași, and Suceava produce the largest quantities (1703–1873 tons/year). Significant productions are also found in the areas of Buzău and Vrancea (in the southeast region) and Bihor and Maramureș (in the northwest region), according to the same report [9]. There is also a major interest in the development of sustainable organic walnut production throughout Romania [10,11].

At the same time, the elderberry has an abundant presence in the spontaneous flora of Romania, especially in the northeast region of the country, and can represent a valuable and cheap resource for bioactive principles. The most widespread species here (there are around 20 in total) is the black elder (*Sambucus nigra* L.). The specialized literature also talks about three autochthonous varieties: Ina, Nora, and Brădet. Being a humidity-loving species, in the spontaneous flora, the elderberry grows on land with groundwater on the surface (on terraces of flowing waters) or on the edge of forests, generally in more shaded areas [12–14].

Various studies have shown that walnuts and elderberries, which are popular all over the world, are two medicinal plants that, thanks to their phytochemical compounds (proteins, flavonoids, phenolic acids, polyphenols, vitamins, and minerals), have beneficial properties for the human body (antibacterial, antioxidant, antiviral, analgesic, anti-inflammatory, etc.) and also possess phytosanitary properties (antibacterial, antifungal, and insecticidal) with benefits on other plants [12–18].

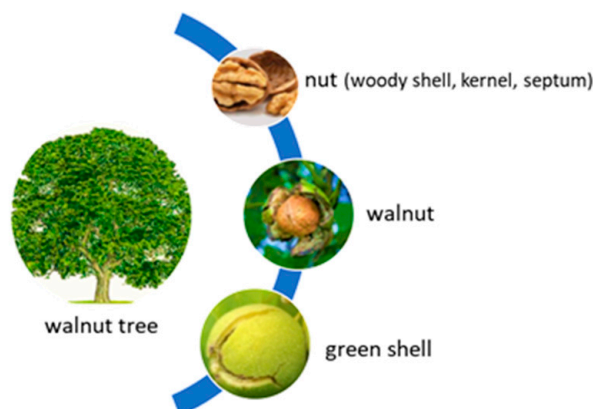
The purpose of this review is to highlight the importance of natural compounds from walnut by-products and elderberry: their biological activities in terms of human health and their phytosanitary properties that could also include the potential for the protection of agricultural crops, respectively. In this context, the available data regarding the predominant bioactive compounds specific to each species and, on the other hand, the common phytochemicals that act on the same microorganisms were critically examined and synthesized, suggesting, in the context of sustainable development, the possibility of exploiting them in a mixture of walnut by-products and elderberry extracts.

## 2. Walnut (*Juglans regia* L.) and Elderberry (*Sambucus nigra* L.) Characterization

Due to the great diversity of extraction and analysis methods, there are numerous studies in the scientific literature that indicate both the chemical composition of walnuts and elderberries and their biopharmaceutical properties and applications in various fields.

### 2.1. General Description

*Juglans regia*, a tree belonging to the Juglandaceae family, is also known as the common walnut [19]. The walnut tree is a vigorous tree that can reach 30 m in height, has strong branches, and has a very wide and rich crown. The leaves are large, composed of 5–9 elliptical leaflets, with whole margins. The fruit includes four main parts: the kernel, the septum, the woody shell, and the green shell (Figure 1). The kernel is the edible part of the fruit, being widely consumed by humans. The other parts of the walnut (the green shell, the dry woody shell, the septum, the bark, the branches, and the leaves) are used for various purposes [16,20].



**Figure 1.** The main components of the walnut.

The green shell is used in traditional Chinese medicine to fight cancer due to its antioxidant properties, as well as for the treatment of pain and for inflammation [21]. The leaves are used to relieve minor inflammatory skin disorders [22]. The walnut shell that results from the processing of walnuts through a successful process of valorization can be another source of active compounds [23,24]. The ripe nut fruit is used in confectionery, while the young form of the fruit is mainly used in the production of liquor [25].

The elder, black elder, or elderberry (*Sambucus nigra* L.) is used as a medicinal and ornamental plant. In the Middle Ages, among the ancient Germans, the elderberry was considered a holy plant, also being used in traditional medicine [12]. In popular tradition, the elderberry, a species of plants in the group of shrubs in the Adoxaceae family (Figure 2), is considered the number one antiviral medicine [13,14]. The hermaphrodite flowers are fragrant, while the leaves have an unpleasant odor when rubbed in the hands. These flowers are followed by clusters of small black fruits. Elderberries are a powerful source of essential vitamins and minerals [13].



**Figure 2.** The component parts of elderberry.

## 2.2. Chemical Composition

The chemical composition varies within very wide limits due to factors related to the species, the variety, pedoclimatic conditions, the harvest period, the degree of ripening, and extraction conditions, as well as the analysis and identification methods.

### 2.2.1. Chemical Composition of Walnut

The benefits of the walnut, which have been known since ancient times, are due to the presence of some bioactive compounds such as flavonoids, polyphenols, juglones, tannins, steroids, etc. [15–17,26].

- Chemical composition of the walnut kernel

The walnut kernel, which mainly has economic importance, contains 4% water, 65% fats, 15% proteins, and 14% carbohydrates, vitamins, and minerals [16].

As described by Slatner et al. [27], who in his work used 2.5 g of ground walnut kernel mixed with 22.5 mL of CH<sub>3</sub>OH/H<sub>2</sub>O (*v/v*, 60/40) in a chilled water bath (0 °C) with sonication for 60 min and analyzed extracts via HPLC, and also in other studies [28–34], the main bioactive compounds present in walnut kernel are tannins (glansreginin A and ellagic acid derivatives), flavonoids (procyanidin trimer),  $\gamma$ -tocopherol (vitamin E), steroids ( $\beta$ -sitosterol), triterpenes, and fatty acids (Table 1).

**Table 1.** Main bioactive compounds of walnut kernel.

Compounds Class	Compound Name	References
Tannins	Glansreginin A	[27]
	Ellagic acid derivatives	[27,29,30]
Flavonoids	Procyanidin dimer 1	[27,34]
	Procyanidin trimer	[27,29,30,34]
	Catechin	[27,34]
Hydroxybenzoic acids	Gallic acid	[27,29,30]
Hydroxycinnamic acids	3-O-p-Coumaroylquinic acid	[27,29,30,34]
	Ferulic acid glucoside	
	Chlorogenic acid	
	(3-O-Caffeoylquinic acid)	
Tocopherols	$\alpha$ -Tocopherol	[28,31,34]
	$\gamma$ -Tocopherol	
	$\delta$ -Tocopherol	
Phytosteroids	Campesterol	[28,32,34]
	$\beta$ -Sitosterol	
	Stigmasterol	
	Avenasterol	
Triterpenes derivatives	Cycloartenol	[33,34]
	2,4-Methyl cycloartenol	
Aliphatic alcohols	Docosanol	
	Tetracosanol	
	Hexacosanol	
Fatty acids	Linoleic acid	[34]
	Oleic acid	
	Linolenic acid	
	Palmitic acid	

- Chemical composition of walnut by-products

The extracts from walnut by-products contain a variety of useful compounds compared to the extracts from the walnut kernel.

#### Chemical Composition of Walnut Leaves

According to Medic et al. [35], who performed the extraction of 0.25 g of leaf powder with methanol in an extract ratio of 1:30 (*w/v*) leaves: methanol using ultrasonication for 60 min and analyzed the extracts via HPLC–MS, and other research [36–42], walnut leaves contain a large amount of compounds from the class of hydroxycinnamic acids (neochlorogenic acid and 3-p-coumaroylquinic acid), flavonoids ((+) catechin, procyanidin dimers, and quercetin derivatives), and naphthoquinones (hydrojuglone derivatives, juglone, and sdihydroxytetralone hexoside) (Table 2).

**Table 2.** Main bioactive compounds of walnut leaf.

Compounds Class	Compound Name	References
Hydroxycinnamic acids	Neochlorogenic acid (5- <i>O</i> -Caffeoylquinic acid)	[35–40,42]
	3- <i>p</i> -Cumaroylquinic acid	
	Ferulic acid	
	Caffeic acid	
	<i>p</i> -Coumaric acid derivatives	
Flavonoids	(+) Catechin	[34,35,41,42]
	(-) Epicatechin	
	Santin	
	Myricetin derivatives	
	Quercetin derivatives	
	Quercetin	
Naphthoquinones	Kaempferol derivatives	[34,35,41,42]
	Juglone (5-Hydroxy-1,4-naphthoquinone)	
	Hydrojuglone	
	Hydrojuglone derivatives	
	1,4-Naphthoquinone	

### Chemical Composition of Walnut Shell

Green walnut shell extracts can be obtained using several extraction methods. For example, the green walnut shell extract can be prepared through maceration by stirring the raw material (1 g) in an aqueous ethanol solution (80% ethanol, *v/v*; 30 mL) at 25 °C for 60 min [43]. Ultrasonic assisted extraction (UAE) and supercritical CO<sub>2</sub> extraction were used to increase the extraction yield. For the UAE, the optimal conditions were a temperature of 60 °C, an extraction time of 30 min, and a mixture of 60% ethanol–water [20]. For the supercritical CO<sub>2</sub> extraction, the optimal conditions were established as 68 °C and 20% ethanol [44]. The total phenol content was determined with the Folin–Ciocâlteu method, using phenolic acids and juglone with ultra-high performance liquid chromatography (UHPLC) [45]. Jahanban-Esfahlan et al. [20] presented a summary of the compounds present in the green walnut shell extract.

Furthermore, the dry walnut shell contains compounds from the class of hydroxybenzoic acid derivatives, sterols, mono- and triglycerides, minerals and (calcium, potassium, and iron). For example, Queirós et al. [46] used 2 g of dry crushed walnut shell for Soxhlet extraction with different solvents (dichloromethane, ethanol, and water) and performed the extract analysis via a UV-coupled high-pressure ion exclusion chromatography (HIPCE-UV).

In agreement with this study and others [43,47–49], the walnut shell contains the following bioactive compounds from the classes of naphthoquinones (juglone and 1,4-naphthoquinone), tannins (tannic acid), hydroxybenzoic acids, hydroxycinnamic acids (sinapic acid and rosmarinic acid), and flavonoids ((–) epicatechin, myricetin, and apigenin) in larger amounts (Table 3). It may also be noted that the walnut shell contains six times more juglone than walnut leaves.

**Table 3.** Main bioactive compounds of walnut shell.

Compounds Class	Compound Name	References
Tannins	Ellagic acid Tannic acid	[20,43–46,49]
Naphthoquinones	Juglone 3-Methoxy-juglone 3-Ethoxy-juglone 8-Hydroxyquinoline 1,4-Naphthoquinone 5,8-Dihydroxy-1,4-naphthoquinone 2-Hidroxi-1,4-naphthoquinone	[20,43–46,48]
Naphthoquinone glycosides	1,4,5-Trihydroxynaphthalene-1,4-di- $\beta$ -D-glucopyranoside 1,4,5-Trihydroxynaphthalene-1,5-di-O- $\beta$ -D-glucopyranoside 1,4,8-Trihydroxynaphthalene-1-O- $\beta$ -D-glucopyranoside	
Naphthalenone	(4 <i>R</i> )-3,4-Dihydro-4-butoxy-5-hydroxy-naphthalen-1(2 <i>H</i> )-one	
Tetralones	Regiolona 5,8-Dihydroxy-4-methoxy- $\alpha$ -tetralone 4,5-Dihydroxy- $\alpha$ -tetralone Sclerone Juglanone	[20,43–46]
Hydroxybenzoic acids	Gallic acid Vanillic acid Syringic acid 2,3-Dihydroxybenzoic acid Tyrosol	[20,43–49]
Hydroxycinnamic acids	Ferulic acid Sinapic acid Rosmarinic acid	
Flavonoids	(+) Catechin Quercetin (–) Epicatechin Myricetin Apigenin Rutin	[20,43–46,48] [20,43–46,48]
Phytosteroids	$\beta$ -Sitosterol Stigmasterol Daucosterol Campesterol	[19,20,43–48]
Triterpenoids	Olenolic acid Oleanolic acid Corosolic acid Ursolic acid	[20,43–46,48]
Sesquiterpenes	(+) Dehydrovomifoliol Blumenol A	
Vitamins	Ascorbic acid $\alpha$ -Tocopherol	[20,49]

### Chemical Composition of the Walnut Septum

As reported by Genovese et al. [50], after the extraction of the walnut septum through ethanolic maceration (10 g of powdered walnut septum in 50 mL of 96% ethanol for 48 h at room temperature) and analysis via HPLC with an ionization mass spectrometry



(HPLC/ESI-MS), and several other studies [51–54], the walnut septum contains an important amount of compounds from the classes of tannins (ellagic acid) and flavonoids (quercitrin) (Table 4).

**Table 4.** Main bioactive compounds of walnut septum.

Compounds Class	Compound Name	References
Hydroxybenzoic acids	Gallic acid	[50–54]
	Protocatechuic acid	
	Vanillic acid	
Tannins	2-Galloyl hexose	[50–53]
	Ellagic acid	
	Ellagic acid hexoside	
Flavonoids	Epigallocatechin	[50,51,54]
	Catechin	
	Epicatechin	
	Epigallocatechin gallate	
	Quercetin-3-O-glucoside	
	Quercitrin (Quercetin-3-O-rhamnoside)	
Hydroxycinnamic acids	p-Coumaric acid	[50–54]

#### 2.2.2. Chemical Composition of Elderberry

The chemical composition of the elderberry indicates the presence of many bioactive substances [12–14]. Raw elderberries contain, per 100 g, 18.4 g of carbohydrates, 7 g of fiber, 0.5 g of fat, 0.66 g of protein, 30 µg of vitamin A, 0.07 mg of thiamin (B1), 0.06 mg of riboflavin (B2), 0.5 mg of niacin (B3), 0.14 mg of pantothenic acid (B5), 0.23 mg of vitamin B6, 36 mg of vitamin C, and minerals (calcium—38 mg; iron—1.6 mg; magnesium—5 mg; phosphorus—39 mg; potassium—280 mg; and zinc—0.11 mg) [55–57]. The fruits of the *Sambucus* tree also contain rutin and isoquercetin; anthocyanins; acids (citric, quinic, and malic); tannins; volatile oils; and mucilage. Elderberries that are fully ripe, which are very attractive from a nutritional point of view, can be consumed as such or in juice form. The elderflowers contain rutoside (rutin) (3%), small amounts of volatile oil (0.03%), sugars, vitamin C, etc. In the elder bark, the presence of choline, sugars, and tannins were observed. The leaves of elder contain glycolic aldehyde, oxalates, appreciable amounts of vitamin C, etc. Sambunigrin, a cyanogenic glycoside, which is missing in the other species of the genus, is present in the leaves, bark, seeds, flowers, and unripe fruits [58,59].

- Chemical composition of elderberry leaves

According to several studies [12–14,18,58–63], elderberry leaves contain a large amount of phenolic compounds (caffeic acid and chlorogenic acid), flavonoids (quercetin-rutinoside), and anthocyanins (cyanidin-glucoside) (Table 5). For example, Kiprovski et al. [62] studied the changes that occurred during ripening in the leaves and fruits of wild and cultivated *Sambucus nigra* through sonication and analyzed the extracts using HPLC-DAD-ESI-MS.

**Table 5.** Main bioactive compounds of elderberry leaves.

Compounds Class	Compound Name	References
Hydroxycinnamic acids	5-p-Coumaroylquinic acid	[12–14,18,58–62]
	Caffeic acid	
	Chlorogenic acid	
	Neochlorogenic acid	
Flavonoids	Rutinoside-3-izorhamnetin	
	Rutinoside-3-kaempferol	
	Quercetin acetyl glucoside	
	Quercetin-rutinoside	
Anthocyanins	Cyanidin-3-p-coumaroyl sambubioside	[58,59]
	Cyanidin-glucoside	
	Pelargonidin-rutinoside	
Cyanogenic glycosides	Sambunigrin	[58,59]

- Chemical composition of elderberry flowers

Vujanović et al. [64], after performing the extraction through different methods (maceration, sonication, and microwave-assisted extraction) using different solvents and analyzing the extracts using the LC–MS/MS technique, found that microwave-assisted extraction provides the highest number of bioactive compounds. According to this study and others [18,58,60,64,65], the elderberry flowers contain a large amount of compounds from the classes of phenols (quinic acid), flavonoids (naringenin, 5-*O*-caffeoylquinic acid, and rutin), and terpenes (ursolic acid) (Table 6).

**Table 6.** Main bioactive compounds of elderberry flowers.

Compounds Class	Compound Name	References
Phenols and derivatives	p-Hydroxybenzoic acid	[18,58,64,65]
	Protocatechuic acid	
	Gentisic acid	
	p-Coumaric acid	
	Quinic acid	
	Ferulic acid	
Flavonoids	Naringenin	[58,60,64,65]
	Catechin	
	Epicatechin	
	Quercetin	
	Isorhamnetin	
	Neochlorogenic acid	
	Kaempferol-3- <i>O</i> -glucoside	
	Quercetin-3- <i>O</i> -hexoside	
Tannins	Kaempferol	[18,60,64]
	Rutin	
Terpenes	Ellagic acid	[64]

- Chemical composition of elderberries

Avula et al. [66], Kiproviski et al. [62], Salvador et al. [67], Pinho et al. [60], Sidor and Gramza-Michałowska [18], Mocanu and Amariei [12], Pascariu and Israel-Roming [13], and Haş et al. [14] showed that *Sambucus* fruits contain a number of phytochemical compounds, such as phenolic acid (chlorogenic acid), flavonoids (quercetin-rutinoside), anthocyanins (cyanidin-glucoside), and triterpenoids (ursolic acid and oleanolic acid) (Table 7). In addition, Kiproviski et al. [62] found that cultivated elderberries had a higher content of phenolics compared to wild edible plants.



**Table 7.** Main bioactive compounds of elderberries.

Compounds Class	Compound Name	References
Phenols	Chlorogenic acid Neochlorogenic acid 4-Caffeoylquinic acid	
Flavonoids	Izorhamnetin-3-glucoside Izorhamnetin-3-rutinoside Quercetin-acetyl-hexoside Quercetin-glucoside Quercetin-rutinoside	
Anthocyanins	Cyanidin-sambubioside Cyanidin-diglucoside Cyanidin-glucoside Cyanidin-3-p-coumaroil sambubioside-5-glucoside	[12–14,18,60,62,66,67]
Fatty acids	Hexadecenoic acid Octadecanoic acid Eicosanoic acid	
Phytosterols	Campesterol Stigmasterol $\beta$ -Sitosterol	
Triterpenoids	$\beta$ -Amyrin Oleanolic acid Ursolic acid	

### 3. Health Benefits and Applications of Walnut By-Products (*Juglans regia* L.) and Elderberry (*Sambucus nigra* L.) Extracts

#### 3.1. Health Benefits and Applications of Walnut By-Products (*Juglans regia* L.) Extracts

The walnut is a crop of great economic interest. Walnut trees are among the most important hardwood species in the northern hemisphere from an ecological and economic point of view. They are mainly cultivated for timber and nut production but are also attractive ornamental trees in parks. All the component parts are used in various industrial branches, including the food industry. The tasty part of the fruit (the kernel) is eaten fresh or roasted and alone or mixed with other edible products. Abdulwahid et al. [47] have shown that the walnut shell is a low-cost and abundant raw material, making it an attractive option for producing building materials. This can help to reduce the cost of construction and to make environmentally friendly building materials more accessible. According to Fordos et al. [48], the walnut shell is extensively used in agriculture, industrial fields, medicine, cosmetics, and environmental fields. This by-product can be a sustainable, ecological, and low-cost alternative to expensive toxic chemicals.

The benefits of the walnut, due to the presence of numerous bioactive compounds such as polyphenols, flavonoids, naphthoquinones, tannins, tocopherols, fatty acids, alkaloids, etc., are reflected not only on our health (with antibacterial, antioxidant, and anti-inflammatory benefits) but also on plants (the phytosanitary properties—antifungal and insecticidal). Like other medicinal plants, the walnut can be used due to its phytosanitary properties in the form of biopesticides, constituting a safe and cheap alternative compared to synthetic products that can be unfriendly to the environment.

##### 3.1.1. Antibacterial Activity

Studies have shown that all parts of the *J. regia* plant have antibacterial properties with beneficial effects on the human body. The green shell extracts of *J. regia* possess a good antibacterial activity on the species *E. coli*, *B. subtilis*, *P. aeruginosa*, and *S. aureus* [68]. Jaiswal et al. [69] showed that juglone inhibits three key enzymes in *H. pylori*. Antibacterial properties were enhanced if the dried walnut shell was modified with silver nanoparticles [70].

The antibacterial activity also depends on the method of extraction, the type of solvent, and the mixing ratio [24,43]. An ethanolic extract with a concentration of 5 mg/mL showed a high antibacterial potential for *S. aureus*. A concentration of 10 mg/mL was effective against a Gram-positive species (*L. monocytogenes*) and a Gram-negative species (*E. coli*). At a concentration of 5 mg/mL, the inhibition rates of *S. aureus*, *B. subtilis*, and *E. coli* bacteria were 85.31–90.26%. Aqueous extracts inhibited Gram-positive bacteria (*P. aeruginosa*) at a concentration of 100 mg/mL and *S. aureus* at a concentration of 0.1 mg/mL [71]. The walnut shell and leaf extracts also had an antibacterial effect against *S. mutans* dental caries [72]. Gomes et al. [73] studied the antibacterial activity of *J. regia*, individually and combined with *Eucalyptus globulus* hydromethanolic extract, and highlighted that the combination of both extracts was most effective against *S. aureus* from cow's milk. An increase in the antimicrobial effect on *E. coli*, *P. aeruginosa* and *A. baumannii* was observed when using a walnut leaf extract mixed with ZnO nanoparticles [74]. Other studies presented the influences of the concentration of the same type of extract and the nature of the solvent used for extraction on the antibacterial activity on Gram-positive bacteria such as *E. faecalis* and *L. monocytogenes* [37]; *S. epidermidis*; *B. subtilis*; *S. aureus* [75]; *B. cereus* [38]; *S. epidermidis* [39]; and *S. arizonae* [76]. Dolatabadi et al. [77] determined that the aqueous extract had a better inhibition on *P. aeruginosa*. Huo et al. [78] showed that the walnut root extract had a strong antibacterial effect against *P. aeruginosa*, *S. aureus*, *E. coli*, and *P. mirabilis*. Extracts from the kernel and peel were proven to have an inhibitory effect on the strains of *E. faecium*, *E. coli*, *K. pneumoniae*, *P. aeruginosa* [79], and coagulase-negative staphylococci (MIC 3.60–461.75 µg/mL) [80]. Kavuncuoğlu et al. [81] explained that the mathematical model based on Artificial Neural Networks (ANN) can be used to determine the inhibition zone diameter of the walnut extract for twelve bacterial species. Duda-Seiman et al. [82] showed that the alcoholic extract of the walnut kernel had a strong antimicrobial effect on the bacteria *S. pyogenes*, *S. aureus*, and *P. mirabilis* and was completely ineffective at the concentration tested against the Gram-negative bacterium *P. aeruginosa*. On the other hand, the study of Perreira et al. explained that aqueous extracts at low concentrations (0.1–1 mg/mL) inhibit Gram-positive bacteria (*B. cereus*, *B. subtilis*, *S. aureus*), and at higher concentrations (10–100 mg/mL) they inhibit the Gram-negative ones (*P. aeruginosa*, *E. coli*, and *K. pneumoniae*) [83]. Alcoholic septum extracts, according to the study of Genovese et al. [50], exhibited a stronger antibacterial effect on Gram-positive bacteria (*S. aureus*, *S. epidermidis*, *E. faecalis*, and *E. faecium*) than on Gram-negative strains (*E. coli*, *K. pneumoniae*, *P. aeruginosa*, and *P. mirabilis*), a fact indicated by lower MIC values. Ethyl acetate extracts from the walnut bark presented a high degree of antimicrobial activity for *S. mutans* and *S. salivarius* [84]; the alcoholic extract seemed to be more effective for *S. aureus* bacteria and less for *S. typhi* (bacterium Gram-negative) [85]; the aqueous extract, compared to the acetonic one, had a stronger inhibitory effect on the growth of the oral microbial flora [86,87]; and the chloroformic extract showed in vitro antimicrobial activity against dental caries microorganisms (*S. mutans*, *S. sobrinus*, *A. viscosus*) [41], as well as for *S. aureus*, *E. coli*, and *P. aeruginosa* [88]. The alcoholic, acetonic, and benzene extracts with the minimum inhibitory concentration of 50 µg/mL to 300 µg/mL inhibited Gram-positive bacteria (*B. subtilis* and *S. aureus*) [89]. It was also shown that chloroformic and aqueous walnut extracts had microbicidal activity against air microorganisms and the leaf extract was very effective in treating skin acne against the *Propioni bacteria* [16].

### 3.1.2. Antifungal Activity

Pathogenic fungi are responsible for crop damage. Attempts are being made to replace fungicidal treatments that use synthetic, polluting chemical substances with more environmentally friendly plant extracts. Studies were carried out on extracts of the woody and green shells of walnuts, and their antifungal activity on cultivated plants, vegetables, and fruits was observed [90–92].

In the case of walnut bark extracts, Upadhyay et al. [26] demonstrated that the methanolic extract had significant activity against *A. niger*, the acetonic extract signifi-

cantly inhibited the growth of *A. alternata*, and the chloroformic extract inhibited *T. virens* and *F. solani*. Ameziane et al. [93] observed a complete inhibition of the mycelial growth of *G. candidum* on citrus with methanolic and chloroformic bark extracts at a 10% concentration (*w/v*).

Bennacer et al. [40] presented that in vitro the tannic extract from walnut leaves showed a high antifungal activity against *A. terreus*, *A. ochraceus*, and *A. brasiliensis* with an inhibition percentage of 77% for a concentration of 40 mg/mL. Wianowska et al. [94] demonstrated that walnut green shell extracts contain a substance called juglone, which exhibits an inhibitory activity for *A. alternata* of 45%. Walnut extracts had little effect on the *R. solani* that affects the rice sheath [95].

### 3.1.3. Insecticidal Activity

More and more, plant extracts are used as an alternative to combat crop plant pests. Walnut extracts have insecticidal properties. Walnut leaves can be introduced as an effective insecticide against the rice weevil (*S. oryzae*). On different days, after treatment, a concentration of 0.1 g/mL of extract was the most effective (66.66%) against the rice weevil [96]. Aqueous extracts of walnut leaves at a concentration of 5% caused 100% mortality in the nematode species *H. bacteriophora*, *S. carpocapsae*, *S. feltiae*, *S. kraussei*, and *P. hermaphrodita* [97]. Walnut extracts prevented 79.94% of tomato moth (*T. absoluta*) oviposition at a concentration of 20% [98] and have shown greater insecticidal activity on *T. griseus* beetle larvae compared to the synthetic insecticide Tanalith C [99]. Nevertheless, it was ineffective against the bean weevil (*A. obtectus*) [100]. Islam and Widhalm [101] observed that juglone had toxic effects on phytophagous insects such as melon, pepper, and tomato aphids (*A. gossypii*); the cabbage moth (*Trichoplusia ni*); the corn moth (*H. armigera*); the red mite (*T. urticae*), the cabbage butterfly (*P. rapae*), and the vinegar midge (*D. melanogaster*). The WCPI (Walnut Cysteine Protease Inhibitor) that is isolated from walnuts can inhibit fungal proteases, demonstrating its biopesticide potential, being an alternative to many chemical pesticides [102].

## 3.2. Health Benefits and Applications of Elderberry (*Sambucus nigra* L.) Extracts

In Europe, elderberry fruits and flowers are used both in food and in folk medicine. The elderflowers can be used in the form of elderberry juice or elderberry syrup, and also in the form of tea, after drying. Although the flowers and berries of the elder have many health benefits, elder leaves, branches, and roots contain substances that are toxic to the body, so they should be avoided. Terzic et al. showed the potential of wine obtained from elderberries as a new food product [103]. Elderberries are used in the treatment of many diseases due to their antioxidant, antitumoral, immunostimulatory, antiallergic, and antiviral benefits; their impact on obesity and metabolic disfunctions; their antidepressant potential; and their antidiabetic and antibacterial properties [12–14,65].

Due to the bioactive compounds, in addition to the health implications, there is a growing interest in the use of the elderberry in various fields, such as as natural additives in the food industry or as biopesticides in the growth of crop plants.

Antibacterial activity with action on the human body as well as phytosanitary activity with benefits on cultivated plants are presented.

### 3.2.1. Antibacterial Activity

Studies have indicated that the elderberry exhibits an antibacterial action that is found in the flowers, fruits, and leaves. Ferreira-Santos et al. [65] showed that aqueous extracts of the elderflowers have an antimicrobial activity against the Gram-positive bacteria *S. aureus* and *S. epidermidis*, and Ramanauskiene et al. [104] proved that ethanolic extracts had in vitro antimicrobial effects against *S. aureus* and *B. cereus*. Álvarez et al. [105] investigated the antibacterial role of peptide extracts from the elderflowers against various Gram-negative bacteria (*A. salmonicida*, *E. psychrophilum*, *V. anguillarum*, and *V. ordalii*). Antolak et al. [106] tested the effect of the elderflower extracts on six strains of *A. lannensis* and

*A. bogorensis* bacteria and observed that the culture medium in the presence of elderberry is less favorable to the bacteria. Pinho et al. indicated a reduced antibacterial action on *K. pneumoniae* [60]. Methanolic extracts of the elderflowers did not inhibit the growth of *B. subtilis*, *P. aeruginosa*, and *S. aureus*, whereas ethanol extracts slightly inhibited the growth of *S. aureus* but did not inhibit *E. coli* [107]. The addition of 10% and 20% of elderberry extract to a culture medium inhibited Gram-positive (*Streptococcus* group G and group C) and Gram-negative (*B. catarrhalis*) bacterial growth by 70 and 99%. An infusion of elderberry leaves presented an inhibitory effect on the growth of bacteria (*B. subtilis*, *B. megaterium*, *E. coli*, and *S. aureus*) and yeasts (*D. hansenii*, *C. shehatae*, and *C. tropicalis*). The antimicrobial activity of extracts from the flowers was higher compared to extracts from the fruits [18]. Elderflower extracts were more toxic to Gram-positive (*Staphylococcus* sp. and *B. cereus*) and Gram-negative (*S. poona* and *P. aeruginosa*) bacteria compared to fruit extracts [108]. Salamon et al. showed that elderberry anthocyanins had no antimicrobial activity on *S. aureus* and *E. faecalis* [109] and that extracts from the leaves and fruits (aqueous 16%, ethanolic 5%, and methanolic 10%) had no antibacterial activity on *Streptococcus* sp., *E. coli*, *P. aeruginosa*, *S. typhimurium*, *S. marcescens*, *P. vulgaris*, *E. cloacae*, and *K. pneumoniae* [110]. Elderberry extracts had less inhibition for Gram-negative (*S. typhimurium*, *S. enteritidis*, and *E. coli*) than Gram-positive (*S. aureus*) [111]. Other studies indicated the antibacterial effect of elderberry extracts on Gram-positive (*E. faecalis*) and Gram-negative (*E. coli* and *P. fluorescens*) bacteria [112] and against *M. luteus*, *P. mirabilis*, and *P. fragii* bacteria [58]. Elderberry extracts (0.625–15 µg/mL) also inhibited the growth of *B. subtilis*, *S. aureus*, *P. aeruginosa*, *S. typhi*, and *E. coli* bacteria [113]. Młynarczyk et al. observed that elderflower extracts had a greater antimicrobial effect against *P. aeruginosa*, aqueous extracts of elder leaves had moderate activity against *B. cereus* and *S. marcescens*, and the fruits presented a superior potential against the bacteria that cause respiratory tract infections (a concentration of 20% of the fruit extract inhibited the growth in bacteria by 99%) [59]. Kačaniová et al. [114] and Allison et al. [115] established that the extracts from elder leaves (8–10 µg/mL) had an effect against the bacteria *S. enterica* and *S. pneumoniae*, but they did not show inhibitory activity against the growth of *E. coli*.

### 3.2.2. Antifungal Activity

Rodino et al. demonstrated that the ethyl alcohol extract obtained from dried elderberry fruits showed the most intense inhibitory activity against the in vitro growth of *P. infestans* (potato mealybug) compared to the extracts from the elderflowers and fresh fruits, respectively [116]. Puia et al. established that an 8% ethanol extract of elderberry was effective against *Chaetomium* with a percentage of 96% and *Penicillium* sp. with a percentage of 85.69%, but was ineffective against *Fusarium* sp. [117].

### 3.2.3. Insecticidal Activity

In the literature, insecticidal activity has been mainly studied on extracts from elder leaves. Jankowska and Wojciechowicz-Żytka showed that aqueous extracts from the elderberry had insecticidal effects on cabbage aphids, with a percentage of 83.34% [118]. Elderberry extract produced a 20% mortality on *T. castaneum* (the reddish flour beetle) at a concentration of 100 mg/2 mL [119]. The insecticidal effect against the horn fly (*H. irritans*) was observed in vitro. Jovanović et al. observed that ethanolic extracts of elder leaves against the bean weevil were ineffective [100], and Ertürk determined that the toxic effect against the third and fourth stage larvae of the moth was low (60% of the larvae did not develop) [120].

## 4. Common Applications—Perspectives

Considering the data presented in this review, it is noted that some bioactive compounds are predominantly found only in walnuts belonging to different classes, such as juglone;  $\alpha$ -hydrojuglone and derivatives (hydrojuglone  $\beta$ -D-glucopyranoside, hydrojuglone derivate pentoside, and hydrojuglone rutoside); 1,4-naphthoquinone from naph-

thoquinones; regiolone, juglanone, and 4,5-dihydroxy- $\alpha$ -tetralone from tetralones; tannic acid from tannins; rosmarinic, 3-p-cumaroylquinic acids, and tyrosol from phenols and derivatives; apigenin from flavonoids; and  $\gamma$ -tocopherol from tocopherols.

The antimicrobial action of the different parts of the walnut is demonstrated by the increased concentrations of naphthoquinones (juglone—4940 mg/100 g and 1,4-naphthoquinone—3680 mg/100 g, respectively). Juglone is reported to have antimicrobial, antifungal, oxidizing, and especially anti-proliferative actions [121,122]. At concentrations of 100–200  $\mu$ g/mL in the growth medium, juglone irreversibly destroys the cell membranes for *Listeria* sp., which explains its bactericidal effect [123]. 1,4-Naphthoquinone exhibits antibacterial activity at concentrations of 2–10  $\mu$ g/mL in the culture medium. A direct application on potato tubers in a concentration of 2 mg/mL inhibits the development of *E. carotovora* [124], a bacterium that produces black rot in a series of vegetables, such as carrots, red beets, onions, peppers, etc. The mechanism of the action of 1,4-naphthoquinone consists in the production of redox reactions that cause the appearance of reactive oxygen species, with a toxic effect on cell membranes [124].

Tetralones such as regiolone, juglanone, and 4,5-dihydroxy- $\alpha$ -tetralone also show antibacterial and antifungal activity against pathogens such as *E. coli*, *S. aureus*, *A. niger*, and *C. albicans* [125].

From the class of flavonoids, the walnut has significant contents of apigenin, which also exhibits antimicrobial actions specific to flavonoids in general [126].

Tannic acid presents bacteriostatic and antifungal actions on pathogens such as *L. monocytogenes* [127], *E. coli*, *Salmonella*, *S. aureus*, *S. epidermidis*, *C. albicans*, and *P. aeruginosa* [128].

Rosmarinic, 3-p-cumaroylquinic acids, and tyrosol are included in the class of phenols and their derivatives. Numerous studies report that rosmarinic acid is a powerful antibacterial and antifungal agent, stopping the growth of pathogens such as *B. cereus*, *S. aureus*, and *E. coli* [129].

Similarly, bioactive compounds predominantly found only in the elderberry are quinic acid from phenols and derivatives; naringenin and rutin from flavonoids; cyanidin-glucoside from anthocyanins; oleanolic acid from triterpenoids; and sambunigrin from cyanogenic glycosides.

From the class of flavonoids, naringenin and rutin stand out with significant concentrations. These compounds demonstrate actions similar to compounds common to the walnut and elderberry, which could lead to a potentiation of the antimicrobial action.

In the composition of the elderberry, there are compounds from the class of triterpenoids, such as oleanolic acid. This compound has been reported to have an antibacterial and antioxidant action on pathogens like *B. cereus*, *S. aureus*, *E. coli*, *S. typhimurium*, and *C. albicans* [130].

The specialized literature reports the antifungal and antibacterial action of the elderberry, mentioning sambunigrin in the composition of this plant, but a direct correlation between the presence of this compound and these activities has not yet been shown [13,58].

Besides these major compounds, *Juglans regia* and *Sambucus nigra* have several common bioactive compounds (gallic acid, caffeic acid, p-coumaric acid, ferulic acid, vanillic acid, quercetin, (+) catechin, (-) epicatechin, ursolic acid, ellagic acid, chlorogenic acid, neochlorogenic acid, and  $\beta$ -sitosterol) in varying amounts that act on the same microorganisms.

Gallic acid shows antifungal and antimicrobial activity [131]. It possesses antifungal action on *Alternaria* sp. [132], *Candida* sp. [133], *A. niger* [134], and *F. solani* [135] and bacterial activity on Gram-positive and Gram-negative bacteria [136]. Studies have indicated decreases of 33, 75, and 81% in the growth rate, depending on the concentration of gallic acid. Treatments with gallic acid can be applied to vegetables affected by diseases caused by these fungi. The action of gallic acid consists in improving the activity of chitinase and peroxidase, thus promoting plant growth [135].

Caffeic acid has antifungal, antibacterial, and antibiotic properties on some pathogens, both for plants and humans. It is used with an antifungal and antibacterial role for human



pathogens such as *C. albicans*, *S. aureus*, and *E. coli* [137]. This study also presents that gallic and caffeic acids can be used to improve the activity of some antimicrobial drugs. Concerning the applicability in agriculture, caffeic acid has an antifungal action on *F. graminearum*, a pathogen found in gramineae and corn. Its high pathogenicity consists not only in the decrease of productivity but also in the accumulation of some  $\beta$ -type mycotoxins in the plant mass [138]. Practically, caffeic acid has a negative impact on fungal growth and mycotoxin production.

Like the previous compounds, p-coumaric acid is part of the class of phenols and is reported in the specialized literature to have an antifungal and antibacterial action, both on human and agricultural pathogens [139–141].

Ferulic acid, found in many plant sources in varying amounts, is another phenolic compound common to the walnut and elderberries. It was applied with good results in combating the proliferation of *Listeria* sp. and *Salmonella* sp., where its inhibitory effect consisted in blocking the bacterial peptidoglycan [142]. It also shows antifungal activity on species such as *Candida* sp. and *Fusarium* sp. [143].

The antifungal action of vanillic acid on *L. theobromae*, *C. gloeosporioides*, *F. oxysporum*, *P. parasiticum*, *P. italicum*, *A. niger*, *A. alternate*, and *Trichoderma* sp. recorded lower effects compared to ferulic acid and p-coumaric acid [144]. Regarding the antibacterial activity, vanillic acid influences both human bacterial pathogens and those from agriculture or food.

Another compound common to the walnut and elderberry that has attracted attention is quercetin, a compound from the class of flavonoids. Jaisinghani reports the antibacterial action of quercetin on *S. aureus*, *E. coli*, *S. flexneri*, *P. vulgaris*, *P. aeruginosa*, and *L. casei* var. *Shirota* via the broth dilution method. The quercetin inhibited the action of these bacteria in a range of 20–500  $\mu\text{g/mL}$  [145]. Numerous authors have reported the inhibitory effect of quercetin on *Candida* sp., *Aspergillus* sp., and *Fusarium* sp. [146–148]. From the class of flavonoids, the two species have significant contents of catechin and epicatechin with antimicrobial activity specific to flavonoids in general [126].

Ursolic acid, which is part of the class of triterpenes, also has antifungal and antibacterial action [130,149].

Ellagic acid, which belongs to the class of tannins, exhibits antibacterial and antifungal activity. Extracts in ethanol and chloroform have an inhibitory effect on *B. subtilis* and *S. aureus* [150]. The inhibition of pathogenic bacteria consists in the anti-enzymatic and antioxidant action [151].

Chlorogenic acid has a similar action to caffeic, rosmarinic, and ellagic acids with an antifungal and antibacterial role for human pathogens such as *C. albicans*, *S. aureus*, and *E. coli* [137,152].

$\beta$ -Sitosterol, a compound from the phytosteroids class, in addition to other actions, possesses antifungal and antibacterial activities [153].

Considering the structure–biological activity relationship of the compounds, studying the two plants together can lead to finding a synergy between them by cumulating the bioactive effect of all the compounds, in order to find inexpensive and non-polluting alternatives for both human and plant health.

## 5. Conclusions

Taking into account the areas cultivated with walnuts and the adaptability of the elderberry (in the spontaneous flora) in the eastern part of Romania, the two crops could constitute a continuous raw material for the extraction of bioactive compounds. Most of the compounds of the two plants have antifungal and antibacterial effects on both plant and human pathogens. This aspect leads to the idea of using them as enhancers for some medicines as well as biopesticides in the context of promoting environmentally friendly technologies and products.

From the presented chemical compositions for both the walnut and the elderberry, it appears that all their components are rich in bioactive compounds. Both the elderberry and

especially the walnut have been intensively studied for their individual, multiple biological activities both for human health and phytosanitary properties.

This review particularly highlights the antimicrobial activity on some pathogenic microorganisms that manifest their action on plant, leguminous, and fruit crops, which raises the hypothesis of the separate or combined use of walnut and elderberry extracts to prevent or stop the proliferation of pathogens. From this perspective, we have already initiated a research series with the aim of verifying the combined action of the phytochemicals of the two species, in order to obtain additional benefits in anticipation of a synergistic effect.

**Author Contributions:** Conceptualization, A.-L.F., I.-L.I. and O.-I.P.; writing—original draft preparation, A.S.-B. and I.-A.S.; writing—review and editing, I.-L.I. and O.-I.P.; visualization, A.S.-B., I.-A.S., I.-L.I. and O.-I.P.; supervision, A.-L.F.; project administration, A.-L.F.; funding acquisition, A.-L.F. All authors have read and agreed to the published version of the manuscript.

**Funding:** This study received no external funding.

**Institutional Review Board Statement:** Not applicable.

**Informed Consent Statement:** Not applicable.

**Data Availability Statement:** Not applicable.

**Conflicts of Interest:** The authors declare no conflicts of interest.

## References

1. Ramawat, K.G.; Mérillon, J.-M. (Eds.) *Bioactive Molecules and Medicinal Plants*; Springer: Berlin/Heidelberg, Germany, 2008.
2. Tringali, C. (Ed.) *Bioactive Compounds from Natural Sources. Isolation, Characterisation and Biological Properties*; Taylor & Francis: London, UK, 2001.
3. Colegate, S.M.; Molyneux, R.J. (Eds.) *Bioactive Natural Products: Detection, Isolation, and Structural Determination*; CRC Press, Taylor & Francis Group: Boca Raton, FL, USA, 2008.
4. Suteu, D.; Rusu, L.; Zaharia, C.; Badeanu, M.; Daraban, G.M. Challenge of utilization vegetal extracts as natural plant protection products. *Appl. Sci.* **2020**, *10*, 8913. [\[CrossRef\]](#)
5. Li, Y.; Chemat, F. *Plant Based “Green Chemistry 2.0”*; Springer: Singapore, 2019.
6. Nicolescu, V.-N.; Rédei, K.; Vor, T.; Bastien, J.-C.; Brus, R.; Benčat, T.; Đodan, M.; Cvjetkovic, B.; Andrašev, S.; La Porta, N.; et al. A review of black walnut (*Juglans nigra* L.) ecology and management in Europe. *Trees* **2020**, *34*, 1087–1112. [\[CrossRef\]](#)
7. Available online: <https://agrobiznes.ro/2023-01-31-romania-ramane-cel-mai-mare-producator-de-nuci-din-ue-circa-20-000-de-ha-urmeaza-sa-intre-pe-rod/> (accessed on 19 September 2023).
8. Available online: <https://www.indexbox.io/search/production-walnut-romania/> (accessed on 5 October 2023).
9. Institutul Național de Statistică. *Producția Vegetală la Principalele Culturi în Anul 2021*; Editura Institutului Național de Statistică: Bucharest, Romania, 2022.
10. Popa, R.-G.; Bălăcescu, A.; Popescu, L.G. Organic walnut cultivation in intensive and super-intensive system—Sustainable investment. Case study: Gorj County, Romania. *Sustainability* **2023**, *15*, 1244. [\[CrossRef\]](#)
11. Lozan, A.; Arndt, C. *Report on the Status of Organic Agriculture and Industry in Romania*; EkoConnect: Desden, Germany, 2022.
12. Mocanu, M.L.; Amariei, S. Elderberries—A source of bioactive compounds with antiviral action. *Plants* **2022**, *11*, 740. [\[CrossRef\]](#) [\[PubMed\]](#)
13. Pascariu, O.-E.; Israel-Roming, F. Bioactive compounds from elderberry: Extraction, health benefits, and food applications. *Processes* **2022**, *10*, 2288. [\[CrossRef\]](#)
14. Haș, I.M.; Teleky, B.-E.; Szabo, K.; Simon, E.; Ranga, F.; Diaconeasa, Z.M.; Purza, A.L.; Vodnar, D.-C.; Tit, D.M.; Nițescu, M. Bioactive potential of elderberry (*Sambucus nigra* L.): Antioxidant, antimicrobial activity, bioaccessibility and prebiotic potential. *Molecules* **2023**, *28*, 3099. [\[CrossRef\]](#) [\[PubMed\]](#)
15. Zhan, Y.; Ma, M.; Chen, Z.; Ma, A.; Li, S.; Xia, J.; Jia, Y. A review on extracts, chemical composition and product development of walnut *Diaphragma juglandis* fructus. *Foods* **2023**, *12*, 3379. [\[CrossRef\]](#)
16. Sharma, M.; Sharma, M.; Sharma, M. A comprehensive review on ethnobotanical, medicinal and nutritional potential of walnut (*Juglans regia* L.). *Proc. Indian Natl. Sci. Acad.* **2022**, *88*, 601–616. [\[CrossRef\]](#)
17. Bourais, I.; Elmarrkechy, S.; Taha, D.; Mourabit, Y.; Bouyahya, A.; El Yadini, M.; Machich, O.; El Hajjaji, S.; El Boury, H.; Dakka, N.; et al. A review on medicinal uses, nutritional value, and antimicrobial, antioxidant, anti-inflammatory, antidiabetic, and anticancer potential related to bioactive compounds of *J. regia*. *Food Rev. Int.* **2023**, *39*, 6199–6249. [\[CrossRef\]](#)
18. Sidor, A.; Gramza-Michałowska, A. Advanced research on the antioxidant and health benefit of elderberry (*Sambucus nigra*) in food—A review. *J. Funct. Food.* **2015**, *18*, 941–958. [\[CrossRef\]](#)
19. Du, H.; Li, C.; Wen, Y.; Tu, Y.; Zhong, Y.; Yuan, Z.; Li, Y.; Liang, B. Secondary metabolites from pericarp of *Juglans regia*. *Biochem. Syst. Ecol.* **2014**, *54*, 88–91. [\[CrossRef\]](#)



20. Jahanban-Esfahlan, A.; Ostadrahimi, A.; Tabibiazar, M.; Amarowicz, R. A comprehensive review on the chemical constituents and functional uses of walnut (*Juglans* spp.) husk. *Int. J. Mol. Sci.* **2019**, *20*, 3920. [\[CrossRef\]](#) [\[PubMed\]](#)
21. Jahanban-Esfahlan, A.; Davaran, S.; Moosavi-Movahedi, A.; Dastmalchi, S. Investigating the interaction of juglone (5-hydroxy-1, 4-naphthoquinone) with serum albumins using spectroscopic and in silico methods. *J. Iran. Chem. Soc.* **2017**, *14*, 1527–1540. [\[CrossRef\]](#)
22. Tsasi, G.; Milošević-Ifantis, T.; Skaltsa, H. Phytochemical study of *Juglans regia* L. pericarps from Greece with a chemotaxonomic approach. *Chem. Biodivers.* **2016**, *13*, 1636–1640. [\[CrossRef\]](#) [\[PubMed\]](#)
23. Maleita, C.; Esteves, I.; Chim, R.; Fonseca, L.; Braga, M.E.M.; Abrantes, I.; de Sousa, H.C. Naphthoquinones from walnut husk residues show strong nematicidal activities against the root-knot nematode *Meloidogyne hispanica*. *ACS Sustain. Chem. Eng.* **2017**, *5*, 3390–3398. [\[CrossRef\]](#)
24. Jahanban-Esfahlan, A.; Amarowicz, R. Walnut (*Juglans regia* L.) shell pyroligneous acid: Chemical constituents and functional applications. *RSC Adv.* **2018**, *8*, 22376. [\[CrossRef\]](#)
25. Stampar, F.; Solar, A.; Hudina, M.; Veberic, R.; Colaric, M.; Fabcic, J. Phenolics in walnut liqueur. *Acta Hortic.* **2007**, *744*, 451–454. [\[CrossRef\]](#)
26. Upadhyay, V.; Kambhoja, S.; Harshaleena, A. Antifungal activity and preliminary phytochemical analysis of stem bark extracts of *Juglans regia* linn. *Int. J. Pharm. Biol. Sci. Arch.* **2010**, *1*, 442–447.
27. Slatnar, A.; Mikulic-Petkovsek, M.; Stampar, F.; Veberic, R.; Solar, A. Identification and quantification of phenolic compounds in kernels, oil and bagasse pellets of common walnut (*Juglans regia* L.). *Food Res. Int.* **2015**, *67*, 255–263. [\[CrossRef\]](#)
28. Gao, P.; Liu, R.; Jin, Q.; Wang, X. Comparative study of chemical compositions and antioxidant capacities of oils obtained from two species of walnut: *Juglans regia* and *Juglans sigillata*. *Food Chem.* **2019**, *279*, 279–287. [\[CrossRef\]](#)
29. Gómez-Caravaca, A.M.; Verardo, V.; Segura-Carretero, A.; Caboni, M.F.; Fernández-Gutiérrez, A. Development of a rapid method to determine phenolic and other polar compounds in walnut by capillary electrophoresis–electrospray ionization time-of-flight mass spectrometry. *J. Chromatogr. A* **2008**, *1209*, 238–245. [\[CrossRef\]](#) [\[PubMed\]](#)
30. Regueiro, J.; Sánchez-González, C.; Vallverdú-Queralt, A.; Simal-Gándara, J.; Lamuela-Raventós, R.; Izquierdo-Pulido, M. Comprehensive identification of walnut polyphenols by liquid chromatography coupled to linear ion trap-Orbitrap mass spectrometry. *Food Chem.* **2014**, *152*, 340–348. [\[CrossRef\]](#) [\[PubMed\]](#)
31. Gharibzadeh, S.M.T.; Mousavi, S.M.; Hamed, M.; Khodaiyan, F. Determination and characterization of kernel biochemical composition and functional compounds of Persian walnut oil. *J. Food Sci. Technol.* **2014**, *51*, 34–42. [\[CrossRef\]](#)
32. Miraliakbari, H.; Shahidi, F. Lipid class compositions, tocopherols and sterols of tree nut oils extracted with different solvents. *J. Food Lipids* **2008**, *15*, 81–96. [\[CrossRef\]](#)
33. Abdallah, I.B.; Tlili, N.; Martinez-Force, E.; Pérez Rubio, A.G.; Pérez Camino, M.C.; Albouchi, A.; Boukhchina, S. Content of carotenoids, tocopherols, sterols, triterpenic and aliphatic alcohols, and volatile compounds in six walnuts (*Juglans regia* L.) varieties. *Food Chem.* **2015**, *173*, 972–978. [\[CrossRef\]](#) [\[PubMed\]](#)
34. Rébua, C.; Artaud, J.; Le Dréau, Y. Walnut (*Juglans regia* L.) oil chemical composition depending on variety, locality, extraction process and storage conditions: A comprehensive review. *J. Food Compos. Anal.* **2015**, *110*, 104534. [\[CrossRef\]](#)
35. Medic, A.; Jakopic, J.; Hudina, M.; Solar, A.; Veberic, R. Identification and quantification of major phenolic constituents in *Juglans regia* L. leaves: Healthy vs. infected leaves with *Xanthomonas campestris* pv. *juglandis* using HPLC-MS/MS. *J. King Saud Univ. Sci.* **2022**, *34*, 101890. [\[CrossRef\]](#)
36. Nour, V.; Trandafir, I.; Cosmulescu, S. HPLC determination of phenolic acids, flavonoids and juglone in walnut leaves. *J. Chromatogr. Sci.* **2013**, *51*, 883–890. [\[CrossRef\]](#)
37. Vieira, V.; Pereira, C.; Pires, T.C.S.P.; Calhelha, R.C.; Alves, M.J.; Ferreira, O.; Barros, L.; Ferreira, I.C.F.R. Phenolic profile, antioxidant and antibacterial properties of *Juglans regia* L. (walnut) leaves from the Northeast of Portugal. *Ind. Crop. Prod.* **2019**, *134*, 347–355. [\[CrossRef\]](#)
38. Pereira, J.A.; Oliveira, I.; Sousa, A.; Valentao, P.; Andrade, P.B.; Ferreira, I.C.F.R.; Ferreres, F.; Bento, A.; Seabra, R.; Estevinho, L. Walnut (*Juglans regia* L.) leaves: Phenolic compounds, antibacterial activity and antioxidant potential of different cultivars. *Food Chem. Toxicol.* **2007**, *45*, 2287–2295. [\[CrossRef\]](#)
39. Nicu, A.I.; Pirvu, L.; Stoian, G.; Vamanu, A. Antibacterial activity of ethanolic extracts from *Fagus sylvatica* L. and *Juglans regia* L. leaves. *Farmacia* **2018**, *66*, 483–486. [\[CrossRef\]](#)
40. Bennacer, A.; Sahir-Halouane, F.; Aitlimane-Aitkaki, S.; Oukali, Z.; Oliveira, I.V.; Rahmouni, N.; Aissaoui, M. Structural characterization of phytochemical content, antibacterial, and antifungal activities of *Juglans regia* L. leaves cultivated in Algeria. *Biocatal. Agric. Biotechnol.* **2022**, *40*, 102304. [\[CrossRef\]](#)
41. Nancy, P.; Manasi, M.; Varghese, A. Antiplatelet activity of *Juglans regia* L. and characterization of juglone from *Juglans regia* L. *Am. J. Biochem. Biotechnol.* **2011**, *7*, 29–31. [\[CrossRef\]](#)
42. Tociu, M.; Manolache, F.; Bălanucă, B.; Moroșan, A.; Stan, R. Superior valorisation of *Juglans regia* L. leaves of different maturity through the isolation of bioactive compounds. *Molecules* **2023**, *28*, 7328. [\[CrossRef\]](#)
43. Vieira, V.; Pereira, C.; Abreu, R.M.V.; Calhelha, R.C.; Alves, M.J.; Coutinho, J.A.P.; Ferreira, O.; Barros, L.; Ferreira, I.C.F.R. Hydroethanolic extract of *Juglans regia* L. green husks: A source of bioactive phytochemicals. *Food Chem. Toxicol.* **2020**, *137*, 111189. [\[CrossRef\]](#)

44. Wenzel, J.; Storer Samaniego, C.; Wang, L.; Burrows, L.; Tucker, E.; Dwarshuis, N.; Ammerman, M.; Zand, A. Antioxidant potential of *Juglans nigra*, black walnut, husks extracted using supercritical carbon dioxide with an ethanol modifier. *Food Sci. Nutr.* **2017**, *5*, 223–232. [CrossRef]
45. Gogoi, R.; Loring, R.; Sarma, N.; Munda, S.; Pandey, S.K.; Lal, M. A comparative study on antioxidant, anti-inflammatory, genotoxicity, antimicrobial activities and chemical composition of fruit and leaf essential oils of *Litsea cubeba* Pers from North-east India. *Ind. Crops Prod.* **2018**, *125*, 131–139. [CrossRef]
46. Queirós, C.S.G.P.; Cardoso, S.; Lourenço, A.; Ferreira, J.; Miranda, I.; Lourenço, M.J.; Pereira, H. Characterization of walnut, almond, and pine nut shells regarding chemical composition and extract composition. *Biomass Convers. Biorefinery* **2020**, *10*, 175–188. [CrossRef]
47. Abdulwahid, M.Y.; Akinwande, A.A.; Kamarou, A.; Kamarou, M.; Romanovski, V.; Al-Qasem, I.A. The production of environmentally friendly building materials out of recycling walnut shell waste: A brief review. *Biomass Convers. Biorefinery* **2023**. [CrossRef]
48. Fordos, S.; Abid, N.; Gulzar, M.; Pasha, I.; Oz, F.; Shahid, A.; Khan, M.K.I.; Khaneghah, A.M.; Aadil, R.M. Recent development in the application of walnut processing by-products (walnut shell and walnut husk). *Biomass Convers. Biorefinery* **2023**, *13*, 14389–14411. [CrossRef]
49. Kizatova, M.; Sultanova, M.; Baikenov, A.; Saduakas, A.; Akzhanov, N. Revealing the features of the composition of the walnut shell from the point of view of the possibility of its use in the food industry. *East.-Eur. J. Enterp. Technol.* **2022**, *1*, 49–55. [CrossRef]
50. Genovese, C.; Cambria, M.T.; D’angeli, F.; Addamo, A.P.; Malfa, G.A.; Siracusa, L.; Pulvirenti, L.; Anfusio, C.D.; Lupo, G.; Salmeri, M. The double effect of walnut septum extract (*Juglans regia* L.) counteracts A172 glioblastoma cell survival and bacterial growth. *Int. J. Oncol.* **2020**, *57*, 1129–1144. [CrossRef] [PubMed]
51. Mateş, L.; Rusu, M.E.; Popa, D.-S. Phytochemicals and biological activities of walnut septum: A systematic review. *Antioxidants* **2023**, *12*, 604. [CrossRef]
52. Wang, D.; Mu, Y.; Dong, H.; Yan, H.; Hao, C.; Wang, X.; Zhang, L. Chemical constituents of the ethyl acetate extract from *Diaphragma juglandis* fructus and their inhibitory activity on nitric oxide production in vitro. *Antioxidants* **2018**, *23*, 72. [CrossRef] [PubMed]
53. Rusu, M.E.; Gheldiu, A.-M.; Mocan, A.; Moldovan, C.; Popa, D.-S.; Tomuta, I.; Vlase, L. Process optimization for improved phenolic compounds recovery from walnut (*Juglans regia* L.) septum: Phytochemical profile and biological activities. *Molecules* **2018**, *23*, 2814. [CrossRef] [PubMed]
54. Liu, R.; Zhao, Z.; Dai, S.; Che, X.; Liu, W. Identification and quantification of bioactive compounds in *Diaphragma juglandis* fructus by UHPLC-Q-Orbitrap HRMS and UHPLC-MS/MS. *J. Agric. Food Chem.* **2019**, *67*, 3811–3825. [CrossRef] [PubMed]
55. Available online: <https://fdc.nal.usda.gov/fdc-app.html#/food-details/171727/nutrients> (accessed on 5 October 2023).
56. Mitroi, C.L.; Simescu, M.; Simescu, R.; Cugorean, M.I.; Cugorean, I.D.; Ştefan, I.D.; Popescu, S.G.; Velciov, A.B. Elderberry-functional product (review). *J. Agroaliment. Process. Technol.* **2022**, *28*, 268–272.
57. Kołodziej, B.; Maksymiec, N.; Drożdżal, K.; Antonkiewicz, J. Effect of traffic pollution on chemical composition of raw elderberry (*Sambucus nigra* L.). *J. Elem.* **2012**, *17*, 67–78. [CrossRef]
58. Przybylska-Balcerek, A.; Szablewski, T.; Szwajkowska-Michalek, L.; Świerk, D.; Cegielska-Radziejewska, R.; Krejpcio, Z.; Suchowilska, E.; Tomczyk, Ł.; Stuper-Szablewska, K. *Sambucus nigra* extracts—natural antioxidants and antimicrobial compounds. *Molecules* **2021**, *26*, 2910. [CrossRef]
59. Młynarczyk, K.; Walkowiak-Tomczak, D.; Łysiak, G.P. Bioactive properties of *Sambucus nigra* L. as a functional ingredient for food and pharmaceutical industry. *J. Funct. Food.* **2018**, *40*, 377–390. [CrossRef]
60. Pinho, E.; Ferreira, I.C.F.R.; Barros, L.; Carvalho, A.M.; Soares, G.; Henriques, M. Antibacterial potential of northeastern Portugal wild plant extracts and respective phenolic compounds. *BioMed Res. Int.* **2014**, *2014*, 814590. [CrossRef] [PubMed]
61. Waswa, E.N.; Li, J.; Mkala, E.M.; Wanga, V.O.; Mutinda, E.S.; Nanjala, C.; Odago, W.O.; Katumo, D.M.; Gichua, M.K.; Gituru, R.W.; et al. Ethnobotany, phytochemistry, pharmacology, and toxicology of the genus *Sambucus* L. (Viburnaceae). *J. Ethnopharmacol.* **2022**, *292*, 115102. [CrossRef] [PubMed]
62. Kiprovski, B.; Malenčić, D.; Ljubojević, M.; Ognjanov, V.; Veberic, R.; Hudina, M.; Mikulic-Petkovsek, M. Quality parameters change during ripening in leaves and fruits of wild growing and cultivated elderberry (*Sambucus nigra*) genotypes. *Sci. Hortic.* **2021**, *277*, 109792. [CrossRef]
63. Sandu Bălan (Tăbăcariu), A.; Patriciu, O.-I.; Ştefănescu, I.-A.; Ifrim, I.-L.; Finaru, A.-L. *Sambucus nigra* L.: Characterization of Elderberry Extracts Obtained by Various Methods. In Proceedings of the Conference Proceedings Abstracts of the 18th International Conference of Constructive Design and Technological Optimization in Machine Building Field—OPROTEH 2023, Bacău, Romania, 11–13 May 2023.
64. Vujanović, M.; Majkić, T.; Zengin, G.; Beara, I.; Cvetanović, A.; Mahomoodally, F.M.; Radojković, M. Advantages of contemporary extraction techniques for the extraction of bioactive constituents from black elderberry (*Sambucus nigra* L.) flowers. *Ind. Crop. Prod.* **2019**, *136*, 93–101. [CrossRef]
65. Ferreira-Santos, P.; Badim, H.; Salvador, Â.C.; Silvestre, A.J.D.; Santos, S.A.O.; Rocha, S.M.; Sousa, A.M.; Pereira, M.O.; Pereira Wilson, C.; Rocha, C.M.R.; et al. Chemical characterization of *Sambucus nigra* L. flowers aqueous extract and its biological implications. *Biomolecules* **2021**, *11*, 1222. [CrossRef] [PubMed]

66. Avula, B.; Katragunta, K.; Wang, Y.H.; Ali, Z.; Srivedavyasasri, R.; Gafner, S.; Slimestad, R.; Khan, I.A. Chemical profiling and UHPLC-QToF analysis for the simultaneous determination of anthocyanins and flavonoids in *Sambucus* berries and authentication and detection of adulteration in elderberry dietary supplements using UHPLC-PDA-MS. *J. Food Compos. Anal.* **2022**, *110*, 104584. [\[CrossRef\]](#)
67. Salvador, A.C.; Rocha, S.M.; Silvestre, A.J.D. Lipophilic phytochemicals from elderberries (*Sambucus nigra* L.): Influence of ripening, cultivar and season. *Ind. Crop. Prod.* **2015**, *71*, 15–23. [\[CrossRef\]](#)
68. Sharma, P.; Ravikumar, G.; Kalaiselvi, M.; Gomathi, D.; Uma, C. In vitro antibacterial and free radical scavenging activity of green hull of *Juglans regia*. *J. Pharm. Anal.* **2013**, *3*, 298–302. [\[CrossRef\]](#)
69. Jaiswal, B.S.; Tailang, M. *Juglans regia*: A review of its traditional uses phytochemistry and pharmacology. *Indo Am. J. Pharm. Res.* **2017**, *7*, 390–398. [\[CrossRef\]](#)
70. Mohan, S.; Panneerselvam, K. An investigation on antibacterial filler property of silver nanoparticles generated from Walnut shell powder by in situ process. *Mater. Today Proc.* **2021**, *39*, 368–372. [\[CrossRef\]](#)
71. Oliveira, I.; Sousa, A.; Ferreira, I.C.F.R.; Bento, A.; Estevinho, L.; Pereira, J.A. Total phenols, antioxidant potential and antimicrobial activity of walnut (*Juglans regia* L.) green husks. *Food Chem. Toxicol.* **2008**, *46*, 2326–2331. [\[CrossRef\]](#) [\[PubMed\]](#)
72. Abdullah, Z.G.; Swilaiman, S.; Ahmad, S.H.; Darogha, S.N. Antibacterial effect of *Juglans regia* against dental caries *Streptococcus mutans*. *Pak. J. Med. Health Sci.* **2020**, *14*, 1313–1317.
73. Gomes, F.; Martins, N.; Ferreira, I.C.F.R.; Henriques, M. Anti-biofilm activity of hydromethanolic plant extracts against *Staphylococcus aureus* isolates from bovine mastitis. *Heliyon* **2019**, *5*, e01728. [\[CrossRef\]](#) [\[PubMed\]](#)
74. Darvishi, E.; Kahrizi, D.; Arkan, E. Comparison of different properties of zinc oxide nanoparticles synthesized by the green (using *Juglans regia* L. leaf extract) and chemical methods. *J. Mol. Liq.* **2019**, *286*, 110831. [\[CrossRef\]](#)
75. Rather, A.M.; Dar, B.A.; Dar, M.Y.; Wani, B.A.; Shah, W.A.; Bhat, B.A.; Ganai, B.A.; Bhat, K.A.; Anand, R.; Qurishi, M.A. Chemical composition, antioxidant and antibacterial activities of the leaf essential oil of *Juglans regia* L. and its constituents. *Phytomedicine* **2012**, *19*, 1185–1190. [\[CrossRef\]](#) [\[PubMed\]](#)
76. Jabli, M.; Sebeia, N.; Boulares, M.; Faïdi, K. Chemical analysis of the characteristics of Tunisian *Juglans regia* L. fractions: Antibacterial potential, gas chromatography–mass spectroscopy and a full investigation of their dyeing properties. *Ind. Crop. Prod.* **2017**, *108*, 690–699. [\[CrossRef\]](#)
77. Dolatabadi, S.; Moghadam, H.N.; Mahdavi-Ourtakand, M. Evaluating the anti-biofilm and antibacterial effects of *Juglans regia* L. extracts against clinical isolates of *Pseudomonas aeruginosa*. *Microb. Pathog.* **2018**, *118*, 285–289. [\[CrossRef\]](#)
78. Huo, J.; Zhao, Z.; Hua, Z.; Fan, J.; Du, J.; Guo, B. Evaluation of *Juglans regia* L., root for wound healing via antioxidant, antimicrobial and anti-inflammatory activity. *Indian J. Biochem. Biophys.* **2020**, *57*, 304–311.
79. D'Angeli, F.; Malfa, G.A.; Garozzo, A.; Li Volti, G.; Genovese, C.; Stivala, A.; Nicolosi, D.; Attanasio, F.; Bellia, F.; Ronsisvalle, S.; et al. Antimicrobial, antioxidant, and cytotoxic activities of *Juglans regia* L. pellicle extract. *Antibiotics* **2021**, *10*, 159. [\[CrossRef\]](#)
80. Acquaviva, R.; D'Angeli, F.; Malfa, G.A.; Ronsisvalle, S.; Garozzo, A.; Stivala, A.; Ragusa, S.; Nicolosi, D.; Salmeri, M.; Genovese, C. Antibacterial and anti-biofilm activities of walnut pellicle extract (*Juglans regia* L.) against coagulase-negative staphylococci. *Nat. Prod. Res.* **2019**, *35*, 2076–2081. [\[CrossRef\]](#)
81. Kavuncuoglu, H.; Kavuncuoglu, E.; Karatas, S.M.; Benli, B.; Sagdic, O.; Yalcin, H. Prediction of the antimicrobial activity of walnut (*Juglans regia* L.) kernel aqueous extracts using artificial neural network and multiple linear regression. *J. Microbiol. Methods* **2018**, *148*, 78–86. [\[CrossRef\]](#) [\[PubMed\]](#)
82. Duda-Seiman, C.; Sinitean, A.; Negrea, P.; Negrea, A.-G.; Dugășescu, D.; Duda-Seiman, D.; Avram, S.; Putz, A.-M.; Bumbăcilă, B.; Irimia, G.-L.; et al. Extraction and antibacterial activity of natural compounds from walnuts (*Juglans regia*). *New Front. Chem.* **2017**, *26*, S4\_P3.
83. Pereira, J.A.; Oliveira, I.; Sousa, A.; Ferreira, I.C.F.R.; Bento, A.; Estevinho, L. Bioactive properties and chemical composition of six walnut (*Juglans regia* L.) cultivars. *Food Chem. Toxicol.* **2008**, *46*, 2103–2111. [\[CrossRef\]](#)
84. Kheddouma, A.; Cherraben, Y. In vitro antimicrobial activity of *Salvadora persica* and *Juglans regia* extracts against microbial strains from oral cavity. *Biocatal. Agric. Biotechnol.* **2021**, *33*, 102003. [\[CrossRef\]](#)
85. Jafer, F.N.; Naser, L. The biological activity of aqueous and methanolic extracts of *Juglans regia* on yeasts and pathologic bacteria. *Arch. Clin. Microbiol.* **2020**, *11*, 113. [\[CrossRef\]](#)
86. Aldawood, T.; Alyousef, A.; Alyousef, S.; Aldosari, N.; Hussam, S.; Alhadad, A.; Bhaian, F.; Eldeen, D.S.; Abdul, N.S. Antibacterial effect of *Juglans regia* L bark extract at different concentrations against human salivary microflora. *J. Oral Med. Oral Surg. Oral Pathol. Oral Radiol.* **2017**, *3*, 214–217. [\[CrossRef\]](#)
87. Mutha, M.; Deshpande, R.R.; Shep, S.; Deshpande, N.R.; Torne, R. Comparative evaluation of antibacterial properties of different extracts of *Juglans regia* (walnut) & *Erethia laevis* (Ajaan Vruksh) against salivary microflora. *Int. J. Pharm. Clin. Res.* **2015**, *7*, 151–153.
88. Alkhawajah, A.M. Studies on the antimicrobial activity of *Juglans regia*. *Am. J. Chin. Med.* **1997**, *25*, 175–180. [\[CrossRef\]](#)
89. Upadhyay, V.; Kambhoja, S.; Harsha, L.K. Antibacterial activity and preliminary phytochemical analysis of stem bark extract of *Juglans regia* linn. *Pharmacologyonline* **2010**, *3*, 274–279.
90. Liu, S.; Cheng, S.; Jia, J.; Cui, J. Resource efficiency and environmental impact of juglone in *Pericarpium Juglandis*: A review. *Front. Environ. Sci.* **2022**, *10*, 999059. [\[CrossRef\]](#)



91. Sandu Bălan (Tăbăcariu), A.; Patriciu, O.-I.; Ștefănescu, I.-A.; Ifrim, I.-L.; Fînar, A.-L. Walnut (*Juglans regia* L.): By-Products Analysis and Capitalization. In Book of Abstracts of Conferința Națională de Chimie, ed., XXXVI, Călimănești-Căciulata, Romania, 4–7 October 2022. Available online: <http://administrare.chimie.upb.ro/schr/doc/evenimente/2022/10/04/1/volum-de-rezumat/doc.pdf> (accessed on 10 January 2024).
92. Sandu Bălan (Tăbăcariu), A.; Patriciu, O.-I.; Ștefănescu, I.-A.; Ifrim, I.-L.; Fînar, A.-L. Walnut (*Juglans regia* L.)—Present and Perspectives on Phytosanitary Activity. In Book of Abstracts of 5th International Conference of the Doctoral School—Excellence in Doctoral Studies through Innovation, Convergence and Interdisciplinarity, “Gheorghe Asachi” Technical University of Iasi, Iasi, Romania, 18–20 May 2022. Available online: [https://conferinta-csd.tuiasi.ro/wp-content/uploads/2022/05/Book\\_of\\_Abstracts\\_CSD2022\\_WEB-1.pdf](https://conferinta-csd.tuiasi.ro/wp-content/uploads/2022/05/Book_of_Abstracts_CSD2022_WEB-1.pdf) (accessed on 10 January 2024).
93. Ameziane, N.; Boubaker, H.; Boudyach, H.; Msanda, F.; Jilal, A.; Ait Benaoumar, A. Antifungal activity of Moroccan plants against citrus fruit pathogens. *Agron. Sustain. Dev.* **2007**, *27*, 273–277. [CrossRef]
94. Wianowska, D.; Garbaczewska, S.; Cieniecka-Roslonkiewicz, A.; Dawidowicz, A.L.; Jankowska, A. Comparison of antifungal activity of extracts from different *Juglans regia* cultivars and juglone. *Microb. Pathog.* **2016**, *100*, 263–267. [CrossRef] [PubMed]
95. Sehajpal, A.; Arora, S.; Kaur, P. Evaluation of plant extracts against *Rhizoctonia solani* causing sheath blight of rice. *J. Plant Prot. Sci.* **2009**, *1*, 25–30.
96. Sadeghnezhad, R.; Enayati, A.; Ebrahimzadeh, M.A.; Azarnoosh, M.; Fazeli-Dinan, M. Toxicity and anti-feeding effects of walnut (*Juglans regia* L.) extract on *Sitophilus oryzae* L. (Coleoptera: Curculionidae). *Fresenius Environ. Bull.* **2020**, *29*, 325–331.
97. Petrikovszki, R.; Tóthné Bogdányi, F.; Tóth, F.; Nagy, P. First report on the effect of aqueous extracts of Hungarian organic mulch materials on entomopathogenic and slug-parasitic nematodes. *Acta Phytopathol. Entomol. Hung.* **2019**, *54*, 279–288. [CrossRef]
98. Erdogan, P. Oviposition deterrent activities of some plant extracts against tomato leaf miner, *Tuta Absoluta* meyrick (Lepidoptera: Gelechiidae). *J. Bacteriol. Mycol. Open Access* **2019**, *7*, 139–142. [CrossRef]
99. Civelek, H.S.; Çolak, A.M. Effects of some plant extracts and bensultap on *Trichopherus griseus* (Fabricius, 1792) (Coleoptera: Cerambycidae). *World J. Agric. Sci.* **2008**, *4*, 721–725.
100. Jovanović, Z.; Kostić, M.; Popović, Z. Grain-protective properties of herbal extracts against the bean weevil *Acanthoscelides obtectus* Say. *Ind. Crop. Prod.* **2007**, *26*, 100–104. [CrossRef]
101. Islam, A.K.M.M.; Widhalm, J.R. Agricultural uses of juglone: Opportunities and challenges. *Agronomy* **2020**, *10*, 1500. [CrossRef]
102. Khan, A.A.; Fazili, A.B.A.; Bhat, S.A.; Bhat, W.F.; Asghar, M.N.; Khan, M.S.; Bano, B. Purification, characterization and studies of a novel cysteine protease inhibitor from *Juglans regia*: Implications as a potential biopesticide. *J. King Saud Univ. Sci.* **2022**, *34*, 101829. [CrossRef]
103. Terzić, M.; Majkić, T.; Beara, I.; Zengin, G.; Miljić, U.; Đurović, S.; Mollica, A.; Radojković, M. Elderberry (*Sambucus nigra* L.) wine as a novel potential functional food product. *Food Biosci.* **2022**, *50*, 102047. [CrossRef]
104. Ramanauskienė, K.; Inkenienė, A.M.; Puidokaite, E.; Grigonis, A. Quality analysis of semisolid formulations with the liquid extract of elderflower (*Sambucus nigra* L.). *Acta Pol. Pharm.* **2019**, *76*, 1061–1071. [CrossRef]
105. Álvarez, C.A.; Barriga, A.; Albericio, F.; Romero, M.S.; Guzmán, F. Identification of peptides in flowers of *Sambucus nigra* with antimicrobial activity against aquaculture pathogens. *Molecules* **2018**, *23*, 1033. [CrossRef] [PubMed]
106. Antolak, H.; Czyżowska, A.; Kręgiel, D. Antibacterial and antiadhesive activities of extracts from edible plants against soft drink spoilage by *Asaia* spp. *J. Food Prot.* **2017**, *80*, 25–34. [CrossRef] [PubMed]
107. Cioch, M.; Satora, P.; Skotniczny, M.; Semik-Szczurak, D.; Tarko, T. Characterisation of antimicrobial properties of extracts of selected medicinal plants. *Pol. J. Microbiol.* **2017**, *66*, 463–472. [CrossRef] [PubMed]
108. Hearst, C.; McCollum, G.; Nelson, D.; Ballard, L.M.; Millar, B.C.; Goldsmith, C.E.; Rooney, P.J.; Loughrey, A.; Moore, J.E.; Rao, J.R. Antibacterial activity of elder (*Sambucus nigra* L.) flower or berry against hospital pathogens. *J. Med. Plants Res.* **2010**, *4*, 1805–1809. [CrossRef]
109. Salamon, I.; Şimşek Sezer, E.N.; Kryvtsova, M.; Labun, P. Antiproliferative and antimicrobial activity of anthocyanins from berry fruits after their isolation and freeze-drying. *Appl. Sci.* **2021**, *11*, 2096. [CrossRef]
110. Pehlivan Karakaş, F.; Yildirim, A.; Türker, A. Biological screening of various medicinal plant extracts for antibacterial and antitumor activities. *Turk. J. Biol.* **2021**, *36*, 641–652. [CrossRef]
111. Tosun, M.N.; Demirel Zorba, N.N.; Yüceer, Y.K. Anti-quorum sensing and antitumor activity of *Prunella vulgaris*, *Sambucus nigra*, *Calendula officinalis*: Potential use in food industry. *J. Microbiol. Biotechnol. Food Sci.* **2021**, *10*, e2774. [CrossRef]
112. Rodino, S.; Butu, A.; Butu, M.; Cornea, P.C. Comparative studies on antibacterial activity of licorice, elderberry and dandelion. *Dig. J. Nanomater. Biostruct.* **2015**, *10*, 947–955.
113. Mohammadsadeghi, S.; Malekpour, A.; Zahedi, S.; Eskandari, F. The antimicrobial activity of elderberry (*Sambucus nigra* L.) extract against gram positive bacteria, gram negative bacteria and yeast. *Res. J. Appl. Sci.* **2013**, *8*, 240–243.
114. Kačániová, M.; Miklášová, K.; Kunová, S.; Galovičová, L.; Borotová, P.; Váľková, V.; Žiarovská, J.; Terentjeva, M. Antimicrobial and antioxidant activity of black elder, stinging nettle, marigold and ribwort plantain. *Sci. Pap. Anim. Sci. Biotechnol.* **2021**, *54*, 136–142.
115. Allison, B.J.; Allenby, M.C.; Bryant, S.S.; Min, J.E.; Hieromnimon, M.; Joyner, P.M. Antibacterial activity of fractions from three Chumash medicinal plant extracts and in vitro inhibition of the enzyme enoyl reductase by the flavonoid jaceosidin. *Nat. Prod. Res.* **2016**, *31*, 707–712. [CrossRef] [PubMed]

116. Rodino, S.; Negoescu, C.; Butu, M.; Cornea, P.C. Preliminary investigation regarding the antifungal activity of *Sambucus nigra* extracts. *Anim. Sci. Biotechnol.* **2013**, *70*, 391–392.
117. Puia, C.E.; Miclea, R.; Toader, I.; Cornoiu, I. Biological control of potential toxigenic fungi in straw used as bedding material for swine. *Anim. Sci. Biotechnol.* **2012**, *69*, 335–336.
118. Jankowska, B.; Wojciechowicz-Żytka, E. Effect of aqueous extracts of black alder (*Alnus glutinosa* (LINNAEUS, 1753) GAERTNER, 1791) and elder (*Sambucus nigra* LINNAEUS, 1753) on the occurrence of *Brevicoryne brassicae* LINNAEUS, 1758 (Homoptera, Aphidoidea), its parasitoid *Diaeretiella rapae* (M'INTOSH, 1855) (Hymenoptera, Ichneumonidae) and predatory Syrphidae on white cabbage. *Pol. J. Entomol.* **2016**, *85*, 237–246. [\[CrossRef\]](#)
119. Ahmad, M.; Saeed, F.; Mehjabeen; Jahan, N. Evaluation of insecticidal and anti-oxidant activity of selected medicinal plants. *J. Pharmacogn. Phytochem.* **2013**, *2*, 153–158.
120. Ertürk, O. Antifeedant and toxicity effects of some plant extracts on *Thaumetopoea solitaria* Frey. (Lep.: Thaumetopoeidae). *Turk. J. Biol.* **2006**, *30*, 51–57.
121. Moreira, C.D.; Santos, T.B.; Freitas, R.H.C.N.; Pacheco, P.A.F.; da Rocha, D.R. Juglone: A versatile natural platform for obtaining new bioactive compounds. *Curr. Top. Med. Chem.* **2021**, *21*, 2018–2045. [\[CrossRef\]](#)
122. Sharma, N.; Ghosh, P.; Sharma, U.K.; Sood, S.; Sinha, A.K.; Gulati, A. Microwave-assisted efficient extraction and stability of juglone in different solvents from *Juglans regia*: Quantification of six phenolic constituents by validated RP-HPLC and evaluation of antimicrobial activity. *Anal. Lett.* **2009**, *42*, 2592–2609. [\[CrossRef\]](#)
123. Cai, Y.Y.; Zou, G.M.; Xi, M.H.; Hou, Y.J.; Shen, H.Y.; Ao, J.F.; Li, M.; Wang, J.; Luo, A.W. Juglone inhibits *Listeria monocytogenes* ATCC 19115 by targeting cell membrane and protein. *Foods* **2022**, *11*, 2558. [\[CrossRef\]](#) [\[PubMed\]](#)
124. Medina, L.F.C.; Stefani, V.; Brandeli, A. Use of 1,4-naphthoquinones for control of *Erwinia carotovora*. *Can. J. Microbiol.* **2004**, *50*, 951–956. [\[CrossRef\]](#) [\[PubMed\]](#)
125. Faidallah, H.M.; Al-Shaikh, K.M.A.; Sobahi, T.R.; Khan, K.A.; Asiri, A.M. An Efficient approach to the synthesis of highly congested 9,10-dihydrophenanthrene-2,4-dicarbonitriles and their biological evaluation as antimicrobial agents. *Molecules* **2013**, *18*, 15704–15716. [\[CrossRef\]](#) [\[PubMed\]](#)
126. Li, K.; Xing, S.; Wang, M.; Peng, Y.; Dong, Y.; Li, X. Anticomplement and antimicrobial activities of flavonoids from *Entada phaseoloides*. *Nat. Prod. Commun.* **2012**, *7*, 867–871. [\[PubMed\]](#)
127. de Almeida Roger, J.; Magro, M.; Spagnolo, S.; Bonaiuto, E.; Baratella, D.; Fasolato, L.; Vianello, F. Antimicrobial and magnetically removable tannic acid nanocarrier: A processing aid for *Listeria monocytogenes* treatment for food industry applications. *Food Chem.* **2018**, *267*, 430–436. [\[CrossRef\]](#) [\[PubMed\]](#)
128. Guimaraes, I.; Costa, R.; Madureira, S.; Borges, S.; Oliveira, A.L.; Pintado, M.; Baptista-Silva, S. Tannic acid tailored-made microsystems for wound infection. *Int. J. Mol. Sci.* **2023**, *24*, 4826. [\[CrossRef\]](#) [\[PubMed\]](#)
129. Kernou, O.-N.; Azzouz, Z.; Madani, K.; Rijo, P. Application of rosmarinic acid with its derivatives in the treatment of microbial pathogens. *Molecules* **2023**, *28*, 4243. [\[CrossRef\]](#) [\[PubMed\]](#)
130. Pereira, V.V.; Pereira, N.R.; Pereira, R.C.G.; Duarte, L.P.; Takahashi, J.A.; Silva, R.R. Synthesis and antimicrobial activity of ursolic acid ester derivatives. *Chem. Biodivers.* **2022**, *19*, e202100566. [\[CrossRef\]](#)
131. Shukla, Y.N.; Srivastava, A.; Kumar, S.; Kumar, S. Phytotoxic and antimicrobial constituents of *Argyrea speciosa* and *Oenothera biennis*. *J. Ethnopharmacol.* **1999**, *67*, 241–245. [\[CrossRef\]](#)
132. El-Nagar, A.; Elzaawely, A.A.; Taha, N.A.; Nehela, Y. The antifungal activity of gallic acid and its derivatives against *Alternaria solani*, the causal agent of tomato early blight. *Agronomy* **2020**, *10*, 1402. [\[CrossRef\]](#)
133. Li, Z.-J.; Liu, M.; Dawuti, G.; Dou, Q.; Ma, Y.; Liu, H.-G.; Aibai, S. Antifungal activity of gallic acid in vitro and in vivo. *Phyther. Res.* **2017**, *31*, 1039–1045. [\[CrossRef\]](#) [\[PubMed\]](#)
134. Lam, P.-L.; Gambari, R.; Kok, S.H.-L.; Lam, K.-H.; Tang, J.C.-O.; Bian, Z.-X.; Lee, K.K.-H.; Chui, C.-H. Non-toxic agarose/gelatin-based microencapsulation system containing gallic acid for antifungal application. *Int. J. Mol. Med.* **2015**, *35*, 503–510. [\[CrossRef\]](#) [\[PubMed\]](#)
135. Nguyen, D.-M.-C.; Seo, D.-J.; Lee, H.-B.; Kim, I.-S.; Kim, K.-Y.; Park, R.-D.; Jung, W.-J. Antifungal activity of gallic acid purified from *Terminalia nigrovenulosa* bark against *Fusarium solani*. *Microb. Pathog.* **2013**, *56*, 8–15. [\[CrossRef\]](#)
136. Khatkar, A.; Nanda, A.; Kumar, P.; Narasimhan, B. Synthesis, antimicrobial evaluation and QSAR studies of gallic acid derivatives. *Arab. J. Chem.* **2017**, *10*, S2870–S2880. [\[CrossRef\]](#)
137. Lima, V.N.; Oliveira-Tintino, C.D.M.; Santos, E.S.; Morais, L.P.; Tintino, S.R.; Freitas, T.S.; Geraldo, Y.S.; Pereira, R.L.S.; Cruz, R.P.; Menezes, I.R.A.; et al. Antimicrobial and enhancement of the antibiotic activity by phenolic compounds: Gallic acid, caffeic acid and pyrogallol. *Microb. Pathog.* **2016**, *99*, 56–61. [\[CrossRef\]](#) [\[PubMed\]](#)
138. Gauthier, L.; Bonnin-Verdal, M.-N.; Marchegay, G.; Pinson-Gadais, L.; Ducos, C.; Richard-Forget, F.; Atanasova-Penichon, V. Fungal biotransformation of chlorogenic and caffeic acids by *Fusarium graminearum*: New insights in the contribution of phenolic acids to resistance to deoxynivalenol accumulation in cereals. *Int. J. Food Microbiol.* **2016**, *221*, 61–68. [\[CrossRef\]](#)
139. Khan, F.; Bamunuarachchi, N.I.; Tabassum, N.; Kim, Y.-M. Caffeic acid and its derivatives: Antimicrobial drugs towards microbial pathogens. *J. Agric. Food Chem.* **2021**, *69*, 2979–3004. [\[CrossRef\]](#)
140. Gandhi, B.; Juliya, J.; Dileep, V.; Rajeswari Batchu, U.; Misra, S.; Kaki, S.S. Antioxidant and biological activities of novel structured monoacylglycerol derivatives with phenolic acids. *Eur. J. Lipid Sci. Technol.* **2021**, *123*, 2100055. [\[CrossRef\]](#)

141. Liu, S.; Jiang, J.; Ma, Z.; Xiao, M.; Yang, L.; Tian, B.; Yu, Y.; Bi, C.; Fang, A.; Yang, Y. The role of hydroxycinnamic acid amide pathway in plant immunity. *Front. Plant Sci.* **2022**, *13*, 922119. [[CrossRef](#)]
142. Wijayanti, E.D.; Safitri, A.; Siswanto, D.; Triprisila, L.F.; Fatchiyah, F. Antimicrobial activity of ferulic acid in Indonesian purple rice through toll-like receptor signaling. *Makara J. Sci.* **2021**, *25*, 247–257. [[CrossRef](#)]
143. de Morais, M.C.; Perez-Castillo, Y.; Silva, V.R.; Santos, L.S.; Soares, M.B.P.; Bezerra, D.P.; de Castro, R.D.; de Sousa, D.P. Cytotoxic and antifungal amides derived from ferulic acid: Molecular docking and mechanism of action. *Biomed Res. Int.* **2021**, *2021*, 3598000. [[CrossRef](#)] [[PubMed](#)]
144. Speranza, B.; Cibelli, F.; Baiano, A.; Carlucci, A.; Raimondo, M.L.; Campaniello, D.; Viggiani, I.; Bevilacqua, A.; Corbo, M.R. Removal ability and resistance to cinnamic and vanillic acids by fungi. *Microorganisms* **2020**, *8*, 930. [[CrossRef](#)] [[PubMed](#)]
145. Jaisinghani, R.N. Antibacterial properties of quercetin. *Microbiol. Res.* **2017**, *8*, 13–14. [[CrossRef](#)]
146. Sadeghi-Ghadi, Z.; Vaezi, A.; Ahangarkani, F.; Ilkit, M.; Ebrahimnejad, P.; Badali, H. Potent in vitro activity of curcumin and quercetin co-encapsulated in nanovesicles without hyaluronan against *Aspergillus* and *Candida* isolates. *J. Mycol. Med.* **2020**, *30*, 101014. [[CrossRef](#)] [[PubMed](#)]
147. Xi, K.-Y.; Xiong, S.-J.; Li, G.; Guo, C.-Q.; Zhou, J.; Ma, J.-W.; Yin, J.-L.; Liu, Y.-Q.; Zhu, Y.-X. Antifungal activity of ginger rhizome extract against *Fusarium solani*. *Horticulturae* **2022**, *8*, 983. [[CrossRef](#)]
148. Yin, J.A.; Peng, X.D.; Lin, J.; Zhang, Y.; Zhang, J.; Gao, H.; Tian, X.; Zhang, R.; Zhao, G. Quercetin ameliorates *Aspergillus fumigatus* keratitis by inhibiting fungal growth, toll-like receptors and inflammatory cytokines. *Int. Immunopharmacol.* **2021**, *93*, 107435. [[CrossRef](#)] [[PubMed](#)]
149. Ghasemzadeh, F.; Darzi, G.N.; Mohammadi, M. Extraction and purification of ursolic acid from the apple peel and in vitro assessment of the biochemical antibacterial, antioxidant and wound healing characteristics. *Appl. Food Biotechnol.* **2022**, *9*, 17–30. [[CrossRef](#)]
150. Hassan, S.; Hamed, S.; Almuhayawi, M.; Hozzein, W.; Selim, S.; Abdelgawad, H. Bioactivity of ellagic acid and velutin: Two phenolic compounds isolated from marine algae. *Egypt. J. Bot.* **2021**, *61*, 219–231. [[CrossRef](#)]
151. Tavares, W.S.; Martin-Pastor, M.; Tavares, A.G.; Sousa, F.F.O. Biopharmaceutical activities related to ellagic acid, chitosan, and zein and their improvement by association. *J. Food Sci.* **2018**, *83*, 2970–2975. [[CrossRef](#)]
152. Ielciu, I.; Niculae, M.; Pall, E.; Barbalata, C.; Tomuta, I.; Olah, N.-K.; Burtescu, R.F.; Benedec, D.; Oniga, I.; Hanganu, D. Antiproliferative and antimicrobial effects of *Rosmarinus officinalis* L. loaded liposomes. *Molecules* **2022**, *27*, 3988. [[CrossRef](#)]
153. Chanioti, S.; Katsouli, M.; Tzia, C. Chapter 9— $\beta$ -Sitosterol as a functional bioactive. In *A Century of Valuable Plant Bioactives*, 1st ed.; Mushtaq, M., Anwar, F., Eds.; Academic Press: Amsterdam, The Netherlands, 2021; pp. 193–212.

**Disclaimer/Publisher’s Note:** The statements, opinions and data contained in all publications are solely those of the individual author(s) and contributor(s) and not of MDPI and/or the editor(s). MDPI and/or the editor(s) disclaim responsibility for any injury to people or property resulting from any ideas, methods, instructions or products referred to in the content.