



Yifei Sun^{1,2}, Xuexue Xia^{1,2}, Ganjun Yuan^{1,2,*}, Tongke Zhang¹, Beibei Deng², Xinyu Feng² and Qixuan Wang²

- ¹ Biotechnological Engineering Center for Pharmaceutical Research and Development, Jiangxi Agricultural University, Nanchang 330045, China
- ² Laboratory of Natural Medicine and Microbiological Drug, College of Bioscience and Bioengineering, Jiangxi Agricultural University, Nanchang 330045, China
- * Correspondence: gyuan@jxau.edu.cn; Tel.: +86-0791-83813459

Abstract: Four Chinese herbs from the Citrus genus, namely Aurantii Fructus Immaturus (Zhishi), Aurantii Fructus (Zhiqiao), Citri Reticulatae Pericarpium Viride (Qingpi) and Citri Reticulatae Pericarpium (Chenpi), are widely used for treating various cardiovascular and gastrointestinal diseases. Many ingredients have already been identified from these herbs, and their various bioactivities provide some interpretations for the pharmacological functions of these herbs. However, the complex functions of these herbs imply undisclosed cholinergic activity. To discover some ingredients with cholinergic activity and further clarify possible reasons for the complex pharmacological functions presented by these herbs, depending on the extended structure-activity relationships of cholinergic and anti-cholinergic agents, a simple method was established here for quickly discovering possible choline analogs using a specific TLC method, and then stachydrine and choline were first identified from these Citrus herb decoctions based on their NMR and HRMS data. After this, two TLC scanning (TLCS) methods were first established for the quantitative analyses of stachydrine and choline, and the contents of the two ingredients and synephrine in 39 samples were determined using the valid TLCS and HPLC methods, respectively. The results showed that the contents of stachydrine (3.04‰) were 2.4 times greater than those of synephrine (1.25%) in Zhiqiao and about one-third to two-thirds of those of Zhishi, Qingpi and Chenpi. Simultaneously, the contents of stachydrine, choline and synephrine in these herbs present similar decreasing trends with the delay of harvest time; e.g., those of stachydrine decrease from 5.16‰ (Zhishi) to 3.04‰ (Zhike) and from 1.98‰ (Qingpi) to 1.68‰ (Chenpi). Differently, the contents of synephrine decrease the fastest, while those of stachydrine decrease the slowest. Based on these results, compared with the pharmacological activities and pharmacokinetics reported for stachydrine and synephrine, it is indicated that stachydrine can be considered as a bioactive equilibrist for synephrine, especially in the cardio-cerebrovascular protection from these citrus herbs. Additionally, the results confirmed that stachydrine plays an important role in the pharmacological functions of these citrus herbs, especially in dual-directionally regulating the uterus, and in various beneficial effects on the cardio-cerebrovascular system, kidneys and liver.

Keywords: choline; *Citrus*; method; analysis; decoction; Aurantii Fructus Immaturus; Aurantii Fructus; effect; cardiovascular protection; uterus

1. Introduction

The fruits or peels of some citrus genus plants were traditionally used as *Qi*-regulating Chinese herbs [1]. Among them, the herbs Aurantii Fructus Immaturus (*Zhishi*) and Aurantii Fructus (*Zhiqiao*) are collected from the dried young and near-mature fruits, respectively, of *Citrus aurantium* L., *Citrus sinensis* Osbeck or their cultivated varieties, and harvested from May to June and in July, respectively. These herbs are also widely used in prescriptions for treating various gastrointestinal and cardiovascular diseases in clinics, and many ingredients, including alkaloids, flavonoids, essential oil, coumarins, limonoids, etc., were identified and considered to be responsible for their pharmacological functions [2,3].



Citation: Sun, Y.; Xia, X.; Yuan, G.; Zhang, T.; Deng, B.; Feng, X.; Wang, Q. Stachydrine, a Bioactive Equilibrist for Synephrine, Identified from Four *Citrus* Chinese Herbs. *Molecules* **2023**, *28*, 3813. https:// doi.org/10.3390/molecules28093813

Academic Editors: Antonio Tiezzi, Elisa Ovidi and Valentina Laghezza Masci

Received: 23 March 2023 Revised: 26 April 2023 Accepted: 27 April 2023 Published: 29 April 2023



Copyright: © 2023 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (https:// creativecommons.org/licenses/by/ 4.0/). It was reported that two alkaloids, synephrine and *N*-methyltyramine, discovered in these herbs can produce various adrenergic effects including raising blood pressure, constricting peripheral blood vessels, dilating the pupil, stimulating the uterus and relaxing the intestines [3–5]. However, some animal experiments and clinical results indicated that the decoctions or aqueous extracts of *Zhishi* and *Zhiqiao* connoted some pharmacological effects contrary to synephrine and *N*-methyltyramine [6]. For example, (1) transiently or inapparently raising, or sometimes even lowering blood pressure [7,8]; (2) various cardiovascular protections [9,10]; (3) an excitatory effect to the in vitro and in vivo uteruses of pregnant and nonpregnant rabbits [11]; (4) a two-way regulating effect on gastrointestinal smooth muscle, not only exciting the gastrointestinal tract and enhancing its peristalsis, but also reducing the tension of gastrointestinal smooth muscle and relieving spasm [12]. These facts showed that there are probably some other ingredients with cholinergic or anticholinergic activities in the decoctions or aqueous extracts of *Zhishi* and *Zhiqiao*, although some reports indicated that citrus flavonoids and essential oil should play a certain role in these complicated functions [13–15].

Based on the above, it is easy to associate these ingredients with the agonists of cholinergic receptors or the antagonists of adrenergic receptors, the former being more probable. The extended structure-activity relationships indicated whether cholinergic or anticholinergic agents present a similar structural characterization (Figure 1). As shown in the center of Figure 1, a nitrogen atom of a tertiary amine or quaternary ammonium is linked to an oxygen atom by an alkane chain of 2–3 carbons, or by a hydrocarbon chain of 3-4 carbons containing cis-double bonds, with the commensurate space distance. This structural fragment can bind with the cholinergic receptor or choline esterase, and then initiate various cholinergic or anticholinergic activities. This led us to focus on the discovery of choline analogs from these herb decoctions. As choline does not contain any chromophore and has no ultraviolet absorption, it is unable to probe according to our previous structure-oriented thinking using the HPLC method [16]. However, choline has the structural fragment of quaternary ammonium, and belongs to alkaloids. Thus, an efficient and specific chromogenic reagent was first explored here for establishing a sensitive thin-layer chromatography (TLC) method to detect choline analogs from these citrus herbs or other plant resources. Then, probable choline analogs in these herbs were isolated and identified, and the quantitative analyses of them and synephrine were performed for clarifying the substance bases of various pharmacological activities and presenting possible reasons for the complex pharmacological functions presented by these Chinese citrus herbs.



Figure 1. The extended structure-activity relationships of cholinergic and anti-cholinergic agents.

2. Results

2.1. Detection of Choline Analogs in These Chinese Citrus Herbs

For detecting possible choline analogs from botanical resources, choline chloride was used as a positive indicator. Considering that synephrine, γ -aminobutyric acid and their analogs were already discovered in these four Chinese citrus herbs [2,3], γ -aminobutyric acid and synephrine were taken as two controls to exclude the similar primary and secondary amines, respectively, in many botanical samples. Various proportions of two solvent systems I (ethyl acetate–95% ethanol–formic acid) and II (*n*-butanol–glacial acetic acid–water) were tested for botanical samples, and the results indicated that there was no obvious difference in developing effects between these two solvent systems. The better proportions of solvent systems I and II were 10:4:5 and 4:1:5 (lower layer), respectively. Considering that solvent system II developed more slowly on the plate than solvent system I, and that a tiering operation should be simultaneously completed for solvent system II, solvent system I with the proportions of 10:4:5 was selected for the subsequent TLC analyses.

Furthermore, the color reactions (Figure S1) on the thin-layer plates indicated that choline and synephrine presented brown and orange-red strips, respectively, for improved Dragendorff reagents, while no strips were observed from all the lanes of choline, synephrine and γ -aminobutyric acid after being colored with Dragendorff and Wagner reagents. Although Dragendorff and Wagner reagents are two broad-spectrum chromogenic agents for alkaloids, they generally do not work for many compounds with lower molecular weights, such as choline and synephrine. From Figure S1, it is indicated that the improved Dragendorff reagent can effectively detect possible choline analogs, which usually have lower molecular weights. It can also give synephrine, a catecholamine amine with a low molecular weight, an orange-red color. This further indicated that a compound presenting a red or orange-red color, like many alkaloids, is unlikely a choline analog after being colored with the improved Dragendorff reagent.

According to the general procedure of TLC analysis, possible choline analogs in four Chinese citrus herbs were detected with a positive indicator of choline, using the optimized developing solvent and chromogenic reagent. The results (Figure 2a) showed that all four herbs contained choline or a probable choline analog. There are probably more analogs and larger contents in the decoction sample of *Zhishi* than those of *Zhiqiao*, Citri Reticulatae Pericarpium Viride (*Qingpi*) and Citri Reticulatae Pericarpium (*Chenpi*). The TLC analyses further indicated that samples from different producing areas contain different choline analogs with different contents, such as the TLC analyses for ten *Zhishi* samples from different producing areas (Figure 2b).



Figure 2. Detection of choline analogs in four Chinese herbs from the Citrus genus. (a) Chinese citrus herbs; 1, Choline; 2, Chenpi (No. 2010019); 3, Qingpi (No. 2010041); 4, Zhishi (No. 2010001); 5, Zhiqiao

(No. 2010032). (b) Chinese herb Zhishi from different producing areas in China; 1, Choline; 2, No. 2008004 (Zizhong, Sichuan); 3, No. 2010002 (Shanggao, Jiangxi); 4, No. 2010003 (Danleng, Sichuan); 5, No. 2010004 (Baisha, Chongqing); 6, No. 2010005 (Anyue, Sichuan); 7, No. 2010006 (Ziyang, Sichuan); 8, No. 2010007 (Tonglian, Sichuan); 9, No. 2010008 (Jintang, Sichuan); 10, No. 2010009 (Jiangjin, Chongqing); 11, No. 2010010 (Lezhi, Sichuan).

2.2. Isolation and Identification of Choline Analogs in Citrus Chinese Herb Zhishi

Considering that the decoctions of *Zhishi* samples contained more choline analogs and higher contents, ten samples of *Zhishi* from different producing areas (Figure 2b) were mixed and pulverized for obtaining more probable analogs of choline. Using the preparative TLC, probable choline analogs **1**, **2** and **3** were isolated from the sampling solution of *Zhishi* according to the process in Section 4.2.2. Based on their spectroscopic data of HRMS, or ¹H and ¹³C NMR, compounds **1**, **2** and **3** were identified as choline, stachydrine (Figure 3) and synephrine, respectively. This indicated that there is a similar fragment, which is surrounded by a blue coil for stachydrine in Figure 3, in the structure of stachydrine and choline.



Figure 3. Chemical structure of probable choline analogs. **1**, choline; **2**, the mixed sample of Chinese herb *Zhishi*; **3**, stachydrine; spots (**a**–**c**) correspond to compounds **1**, **2** and **3**, and the chemical structures of choline analogs **1** and **2** are shown as (**a**,**b**), respectively, on the right side of the thin-layer plate.

The related data for their identification are as follows.

1: A white amorphous powder; a brown spot colored with the improved Dragendorff reagent on the thin-layer plates; HRESIMS m/z 104.1060 [M]⁺ (calcd. for C₅H₁₄NO, 104.1075); ¹H NMR (MeOH- d_4 , 600 MHz) δ : 3.91 (H-3), 3.56 (H-2) and 3.24 (H-4 to H-6, s); ¹³C NMR (MeOH- d_4 , 150 MHz) δ : 68.9 (C-2), 56.9 (C-3) and 54.5 (C-4, 5 and 6). These NMR data are in accordance with those reported for choline [17].

2: A white amorphous powder; a reddish-brown spot colored with the improved Dragendorff reagent on the thin-layer plates; HRESIMS *m*/*z* 144.1010 [M]⁺ (calcd. for C₇H₁₄NO₂, 144.1025); ¹H NMR (D₂O, 400 MHz) δ : 4.07 (1H, t, *J* = 9.2 Hz, H-2), 3.70 (1H, m, H-5a), 3.53 (1H, m, H-5b), 3.29 (3H, s, H-6), 3.10 (3H, s, H-7), 2.48 (1H, m, H-3a), 2.33~2.24 (1H, m, H-3b) and 2.23~2.11 (2H, m, H-4); ¹³C NMR (D₂O, 100 MHz) δ : 171.3 (C-8), 76.4 (C-2), 67.2 (C-5), 52.1 (C-6), 45.9 (C-7), 25.3 (C-3) and 18.6 (C-4). These NMR data are in accordance with those reported for stachydrine [18].

3: A white amorphous powder; an orange-red spot colored with the improved Dragendorff reagent on the thin-layer plates; HRESIMS m/z 168.1009 [M + H]⁺ (calcd. for C₉H₁₄NO₂, 168.1024), 150.0903 [M + H - H₂O]⁺ (calcd. for C₉H₁₂NO, 150.0919) which presented as base peak ion.

The above indicated that compound **3** is not a choline analog. After spraying with the improved Dragendorff reagent, the spots of compounds **1** and **2**, two choline analogs, presented brown or reddish-brown color, while compound **3** showed an orange-red spot

on the thin-layer plates. These indicated that the compounds represented by the spots are unlikely choline analogs if the color of the spots is red. A detailed discussion is given in Section 3.

2.3. The Contents of Stachydrine, Choline and Synephrine in Four Chinese Citrus Herbs

Along with the identification of stachydrine and choline from the four Chinese citrus herbs Zhishi, Zhiqiao, Qingpi and Chenpi, it is known that three alkaloids, namely stachydrine, choline and synephrine, are extensively distributed in these herbs, and their proportions and contents in these herbs are different from each other. As we mentioned in the introduction, the decoctions or aqueous extracts of Zhishi and Zhiqiao connoted probable cholinergic activities, contrary to the adrenergic effects of synephrine. Simultaneously, these herbs have different pharmacological functions according to the theory of Chinese medicine. Thus, it was worth exploring whether there were some relationships between the complex pharmacological functions of these herbs and the contents and proportions of these three ingredients. To achieve this, the contents of these three ingredients were determined.

2.3.1. Validation of Quantitative Analyses

As there is no chromophore in the structures of stachydrine and choline, it is unsuitable to perform their quantitative analyses using the HPLC-UV method. Based on the detection procedure of choline analogs and their separation effects on thin-layer plates, the TLC scanning (TLCS) method was considered for the quantitative analyses of stachydrine and choline. Meanwhile, the quantitative analysis for synephrine was established using the HPLC-UV method, referring to that described in the Chinese pharmacopeia [1].

The methodology validation showed that the chromatographic peaks of both choline and stachydrine presented good symmetry, and both compounds can be well separated with an identical resolution of 1.26 from their nearest peaks, according to the procedure of TLCS analysis described in Section 4.4.1. The limit of detection (LOD) and limit of quantitation (LOQ) for stachydrine were 1.0 μ g (0.20 mg·mL⁻¹) with a relative standard deviation (RSD) of 0.08% and 2.0 μ g (0.40 mg·mL⁻¹) with an RSD of 0.18%, respectively, and those for choline were 0.5 μ g (0.10 mg·mL⁻¹) with an RSD of 0.08% and 0.8 μ g $(0.15 \text{ mg} \cdot \text{mL}^{-1})$ with an RSD of 0.18%, respectively. The repeatability tests showed that the RSDs for a strip of stachydrine and choline on a thin-layer plate were 0.34% and 0.48%, respectively. The precision tests indicated that the RSDs for six strips of stachydrine and choline on the same thin-layer plate were 1.20% and 3.55%, respectively, and those for the identical solutions of stachydrine and choline on six thin-layer plates were 4.05% and 4.15%, respectively. A good linearity correlation y = 6565.7x + 5522.7 (r = 0.9996) (Figure S2a) between the amounts (x) and peak areas (y) was presented for stachydrine in a range from 2.0 to 14.0 μ g, and an acceptable linearity correlation y = 12302.1x - 909.3 (r = 0.9987) (Figure S2b) was presented for choline in a range from 1.0 to 4.0 μ g. Using a powder sample of Zhishi (No. 2010001), the reproducibility was assessed, and the results showed that the contents of stachydrine and choline in this sample were 0.302% (with an RSD value of 2.39%) and 0.085% (with an RSD value of 2.03%), respectively. Using this Zhishi sample, the recoveries of choline and stachydrine were tested. The results showed that the average recovery of stachydrine was 100.28% with an RSD value of 3.08 (Table S1), and that of choline was 101.04% with an RSD value of 2.31 (Table S2). These results together indicated that the established TLCS methods were valid, and can be used for the quantitative analyses of stachydrine and choline in these herbs.

Similarly, the chromatographic peaks of synephrine presented good symmetry and can be well separated with a resolution of 4.06 from its nearest peaks, according to the procedure of HPLC analysis described in Section 4.5.1. The limit of detection (LOD) and limit of quantitation (LOQ) for synephrine were 0.02 μ g and 0.05 μ g, respectively. The repeatability experiments presented an RSD value of 0.42% for a standard solution of synephrine. A good linearity correlation y = 4435448.7x + 2217.2 (r = 1.0000) (Figure S2c)

between the amounts (x) and peak areas (y) was presented for synephrine in a range from 0.5 to 32.0 μ g. Using a powder sample of Zhishi (No. 2010001), the reproducibility experiment showed that the contents of synephrine in this sample were 1.10% with an RSD value of 1.77%, and the sample solution showed good stability with an RSD value of 1.60% in 24 h. Using this Zhishi sample, the recovery of synephrine was tested, and the results showed that the average recovery of synephrine was 99.89% with an RSD value of 3.19 (Table S3). These results together indicated that the established HPLC method was validated and can be used for the quantitative analyses of synephrine in these herbs.

2.3.2. Contents of Stachydrine, Choline and Synephrine in Four Chinese Citrus Herbs

Using the above validated methods for the quantitative analyses, the contents of stachydrine, choline and synephrine in these citrus herbs were determined, and the results are shown in Table 1.

 Table 1. Contents of stachydrine, choline and synephrine in four Chinese herbs from Citrus genus ^a.

Name of Chinese Herbs	Producing Area	Batch No.	Harvest Dates	Contents in Chinese Herb (mg·g ⁻¹)			Content Ranges (mg·g ⁻¹)		
				Stachydrine (SC)	Choline (CL)	Synephrine (SN)	Stachydrine (SC)	Choline (CL)	Synephrine (SN)
Zhishi	Zizhong, Sichuan Shanggao, Jiangxi	2010001 2010002 2010002	June 2020 June 2020	3.10 6.42	0.87 0.16 0.20	11.03 21.07 2.56			
	Baisha, Chongqing Anyue, Sichuan	2010003 2010004 2010005	June 2020 June 2020 June 2020	3.23 5.31	b 0.42	5.21 19.84	2 948 53	0.000.87	3 56-21 07
	Ziyang, Sichuan Tongliang, Chongqing Jintang, Sichuan	2010006 2010007 2010008	June 2020 June 2020 June 2020	6.46 3.74 8.53	0.47 0.11 0.29	20.10 4.34 12.58	2.94~0.55	0.00-0.07	3.30~21.07
	Jiasi, Chongqing Lezhi, Sichuan	2010009 2010010	June 2020 June 2020	2.94 6.48	0.21 0.31	9.64 10.80			
Zhiqiao	Baisha, Chongqing Tongnan, Chongqing Bazhong, Sichuan	2010031 2010032 2010033	July 2020 July 2020 July 2020	1.43 2.51 5.13	0.10 0.21 0.14	2.42 1.82 0.59			
	Zizhong, Sichuan Jiasi, Chongqing Zhangshu, Jiangxi	2010034 2010035 2010036	July 2020 July 2020 July 2020	3.92 3.11 2.31	0.20 0.12 0.19	1.25 1.77 1.73	1.43~5.13	0.00~0.21	0.00~2.42
	Dazu, Chongqing Dazhu, Sichuan Ji'an Jiangxi	2010037 2010038 2010039	July 2020 July 2020 July 2020	2.66 4.68 1.62	0.19	1.63			
	Quzhou, Zhejiang Zhangshu, Jiangxi	2010041 2010042	July 2020 July 2020 July 2020	2.35 2.63	0.34 0.32	5.16 6.18			
Qingpi	Danleng, Sichuan Fengyuzhen, Sichuan Ziyang, Sichuan	2010043 2010044 2010045	July 2020 July 2020 July 2020	2.22 1.93	0.47 0.32 0.29	6.11 6.37 5.30			
	Huangshui, Sichuan Meishan, Sichaun	2010045 2010046 2010047	July 2020 July 2020 July 2020	1.15 3.51	0.21 0.47	6.29 5.08	0.99~3.51	0.21~0.60	4.39~7.19
	Pingshan, Sichuan Ji'an, Jiangxi Shuangliu, Sichuan	2010048 2010049 2010050	July 2020 July 2020 July 2020	1.64 0.99 2.36	0.60 0.59 0.53	7.19 4.39 5.56			
Chenpi	Jintang, Sichuan Yibin, Sichuan	2010011 2010012	January 2020 January 2020	0.96 1.03		1.98 2.04			
	Nanchong, Sichuan Meishan, Sichuan Neijiang, Sichuan	2010013 2010014 2010015	January 2020 January 2020 January 2020	1.92 2.93 0.86	0.16 - 0.12	2.71 1.86 2.74	0.06 0.06	0.00.0.00	1.0(0.00
	Yiyang, Hunan Anyue, Sichuan Lozhi, Sichuan	2010016 2010017 2010018	January 2020 January 2020 January 2020	3.26 1.33 1.16	0.18	3.80 2.43 2.16	0.86~3.26	0.00~0.20	1.86~3.80
	Dazhou, Sichuan Xinhui, Guangzhou	2010018 2010019 2010020	January 2020 January 2020 January 2020	1.10 1.89 1.43	0.09	2.10 2.52 2.74			

^a: These citrus herbs were commercially available from Chengdu Huichu Technology Co., Ltd., Chengdu, China; herbs Zhishi and Zhiqiao are the dried young and near-mature fruits, respectively, of *Citrus aurantium* L. or its cultivated varieties, and herbs Qingpi and Chenpi are the dried peel from young (or immature) and mature fruits, respectively, of Citrus reticulata Blanco or its cultivated varieties. ^b: –, the content was lower than the LOD and was set as 0.00 for subsequent calculations.

From Table 1, the contents of the three ingredients in the four herbs fluctuate greatly. Overall, the contents of stachydrine and synephrine are obviously higher than those of choline, and the content fluctuations of synephrine are greater than those of stachydrine in these herbs. Simultaneously, the contents of synephrine and stachydrine are obviously higher in Zhishi than in the other three herbs. It is worth noting that the content of synephrine in the herb Zhiqiao is the lowest among these herbs, while that of stachydrine in the herb Zhiqiao is very high and just lower than that in the herb Zhishi.

2.3.3. Statistical Analysis for the Content Data of Three Ingredients

To make the content differences between these three ingredients clearer to explain the differences in the pharmacological functions of these herbs, the data in Table 1 were further analyzed using statistical methods. The results confirmed that the contents of stachydrine and synephrine in the herb Zhishi are larger than in the other three herbs (p < 0.05 or p < 0.01), and those of choline in Qingpi (or/and Zhishi) are larger than in Zhiqiao and Chenpi (p < 0.05 or p < 0.01). Simultaneously, stachydrine is the largest of these three ingredients in Zhiqiao (p < 0.05 or p < 0.01).

Since herbs Zhishi and Zhiqiao are the dried young fruits of *Citrus aurantium* L. (or its cultivated varieties), and Qingpi and Chenpi are the dried peel from young (or immature) fruits of Citrus reticulata Blanco (or its cultivated varieties), it was further concluded that all three ingredients will reduce (p < 0.05) with the prolongation of growth time for Citrus genus plants, except for stachydrine in the herbs Qingpi and Chenpi. Among them, the contents of synephrine decrease the most rapidly, while those of stachydrine decrease the most slowly. Although all their contents reduce with the prolongation of growth time for these Citrus genus plants, the contents of synephrine are significantly higher than those of stachydrine and choline, except those of stachydrine are largest in the herb Zhiqiao. Moreover, since herbs Zhishi (or Zhiqiao) and Qingpi (or Chenpi) are derived from the fruits and the pericarps, the fact that synephrine and stachydrine distribute more in the exocarp than in the mesocarp could be further inferred from their decreasing speeds and degrees with the growth time of these Citrus genus plants, and this inference was also in accordance with a previous report [19]. However, there is possibly no obvious difference in the distribution of choline in exocarp and mesocarp.

2.4. Comprehensive Analyses for the Pharmacological Effects of Stachydrine and Synephrine

These four herbs are derived from the fruits or peels of *Citrus* genus plants. Herbs Zhishi and Zhiqiao originate from the fruits of Citrus aurantium L. or its cultured varieties at different harvest times, and the herbs *Qingpi* and *Chenpi* originate from the peels of *Citrus* reticulata Blanco or its cultured varieties at different harvest times [1]. Their decoctions or water extracts have various pharmacological effects on the digestive system, cardiovascular system, respiratory system and so on [2,3,11]. Many reports indicate that flavonoids, alkaloids, coumarins, essential oil and limonoids are the main components of these herbs [2,3]. It was reported that some flavonoids (such as narirutin, naringin, hesperidin, neohesperidin and nobiletin) and alkaloids (such as synephrine and N-methyl tyramine), with higher content and a wider distribution in these herbs, have various bioactivities in the digestive system, cardiovascular system and respiratory system, and which are mainly responsible for the pharmacological effects of these herbs [11,12,20]. However, some pharmacological activities of these herbs in the human body have few related investigations, such as the excitatory effect of the herbs Zhishi, Zhiqiao and Qingpi on the in vitro and in vivo uteruses of both pregnant and non-pregnant rabbits. Moreover, some related investigations remain insufficiently clear, such as which components are responsible for the two-way regulating effects on gastrointestinal smooth muscle, showed by the decoction of Zhiqiao [12,20].

Considering that the decoctions or aqueous extracts of these herbs conceal some possible cholinergic activities, here, probable choline analogs in these herbs were detected and determined. The results from Table 2 show that stachydrine (or plus choline) and synephrine presented commensurate contents in these herbs and similar changing trends with the increase in growth time. However, the contents of synephrine decrease most rapidly, while those of stachydrine decrease most slowly. Since synephrine is a sympathomimetic amine and has intrinsic sympathomimetic activity, it was inferred that they possibly have different or even contrary pharmacological activities. If this is true, different contents and proportions of the two compounds in these herbs would have important impacts on their pharmacological functions in the human body, which would probably give some reasonable interpretations for the difference in pharmacological functions of these herbs, although other components also have important roles. Moreover, this would also fluctuate the pharmacological effects of these herbs with different producing areas and harvest times. To clarify these, the main bioactivities of stachydrine (plus choline) and synephrine were summarized and are listed in Table 3.

Table 2. Statistical analyses for the contents of stachydrine, choline and synephrine in four citrus herbs ^a.

Norma of Chiraca	Avera	age Content \pm SD (m	Sequencing of the	Sequencing of the		
Name of Chinese Herbs	Stachydrine (SC)	Choline (CL)	Synephrine (SN)	Contents of SC, CL and SN ^c	Contents of SN, and SC Plus SC ^d	
Zhishi	5.16 ± 1.87 **##++	0.32 ± 0.24 *#	11.82 ± 6.61 ##+	SN ^{!!} > $SC > CL $ ^{!!}	$SN > (SC + CL)^{\ddagger}$	
Zhiqiao	3.04 ± 1.29 #+	0.13 ± 0.08 ⁺⁺	1.25 ± 0.86 ##++	$SC^{\S} > SN > CL^{\S}$	(SC + CL) [‡] > SN	
Qingpi	1.98 ± 0.81 *	0.41 ± 0.14 **##	5.76 ± 0.81 **##	$SN $ $^{!} > SC > CL $ $^{!!}$	$SN > (SC + CL)^{\ddagger}$	
Chenpi	1.68 ± 0.83 *	0.08 ± 0.08 ⁺⁺	2.50 ± 0.56 ^++	SN ^{!!} > $SC > CL $ ^{!!}	$SN > (SC + CL)^{\ddagger}$	
Sequencing in herbs	Zhishi > Zhiqiao > Qingpi (Chenpi)	Qingpi (Zhishi) > Zhiqiao (Chenpi)	Zhishi > Qingpi > Chenpi > Zhiqiao			

^a: The data before analysis are shown in Table 1. ^b: *, ⁺ and [#] indicate that the differences are significant (p < 0.05) compared with Zhiqiao, Qingpi and Chenpi, respectively; **, ⁺⁺ and ^{##} indicate that the differences are significant (p < 0.01) compared with Zhiqiao, Qingpi and Chenpi, respectively. ^c: [!] and [§] indicate that the differences are significant (p < 0.05) compared with stachydrine (SC) and synephrine (SN), respectively; ^{!!} and ^{§§} indicate that the differences are significant (p < 0.01) compared with stachydrine (SC) and synephrine (SN), respectively; ^{!!} and ^{§§} indicate that the differences are significant (p < 0.01) compared with stachydrine (SC) or remarkably significant (p < 0.01) compared with synephrine (SN).

From Table 3, synephrine, as a partial agonist of α_1 -adrenoreceptor and an antagonist of α_2 -adrenoreceptor, can constrict peripheral blood vessels, cerebrovascular and aorta, while stachydrine, an ingredient coexisting with synephrine in these citrus herbs, presents various cardio-cerebrovascular protections including rapid vascular relaxation, accelerating blood circulation, increasing coronary and myocardial blood flow, relieving myocardial necrosis, slowing heart rate and decreasing cardiac output, suppressing and ameliorating myocardial fibrosis, ameliorating cardiac hypertrophy and fibrosis. Some results were obtained from animal models induced by adrenergic receptor agonists (marked in bold font in Table 3). Simultaneously, synephrine would increase the level of platelet [37], while stachydrine can inhibit platelet aggregation and ameliorate platelet-mediated thromboinflammation [27,28,38]. Synephrine can contract the uterus (pregnancy) [47], while stachydrine can regulate the uterus, such as through the inhibition of convulsive uterus, the stimulation of uterine contraction [49] and reducing uterine bleeding [52]. Moreover, synephrine can be rapidly absorbed and predominantly metabolized in the liver [72], and this would lead to some possible unfavorable influences on the liver, especially at large doses of citrus herbs or some related juices containing synephrine. However, stachydrine can rapidly relax blood vessels by activating the endothelial nitric oxide synthase in the vascular endothelium [35], and has various helpful effects on the liver, such as anti-inflammatory action, ameliorating hepatic fibrosis [44] and treating non-alcoholic fatty liver [26]. Thus, the unfavorable influence of synephrine on the liver would be theoretically eradicated by stachydrine coexisting in these citrus herbs.

Fable 3. Main bioactivities	of stachydrine	(plus choline)	and synephrine ^a .
------------------------------------	----------------	----------------	-------------------------------

Effected Tissues,	Pharmacological Effects					
Organs or Systems	Synephrine	Stachydrine (Choline)				
Eye	Exciting α_1 -adrenoreceptor and dilating the pupil [21].	/				
cardio- cerebrovascular system	A partial agonist of α_1 -adrenoreceptor and an antagonist of α_2 -adrenoreceptor, and can weakly bind on α_1 - and α_2 -adrenoreceptors. The effects on β_1 - and β_2 -adrenoreceptors are very small and can be ignored [4,5,10,22–24]. (1) Constricting peripheral blood vessels including mesenteric artery, and raising blood pressure; (2) Complex responses of the coronary artery by the excitation of α_1 -adrenoceptor and TAARs [22]; (3) Constricting aorta directly by the excitation of α_1 -adrenoceptor and 5-HT1D [25], not by 5-HT1B and β -receptor [23]; (4) Cerebral vasoconstriction deduced from it acting on the α_1 -adrenoceptor.	Cardiovascular system protection [26]: (1) Accelerating blood circulation, increasing coronary and myocardial blood flow in adrenaline-induced myocardial ischemia [27,28]; (2) Relieving myocardial necrosis, lowering blood viscosity and vascular resistance, improving microcirculation [27,28]; (3) Slowing heart rate and decreasing cardiac output [27,28]; (4) Suppressing and ameliorating myocardial fibrosis [29,30]; (5) Ameliorating isoproterenol-induced cardiac hypertrophy and fibrosis [31]; (6) Inhibiting norepinephrine-induced cardiomyocyte hypertrophy [32–34]; (7) Rapid vascular relaxation mediated by the activation of endothelial nitric oxide synthase in vascular endothelial cells [35]; (8) Ameliorating endothelial dysfunction induced by homocysteine [36].				
Blood	Increasing the level of platelet [37].	Inhibiting platelet aggregation and ameliorating platelet-mediated thrombo-inflammation [27,28,38];				
Neuroprotective effects	/	 Protecting the neuronal injury [39]; Inhibiting inflammatory reactions and improving pathological changes after cerebral ischemia [40]; Inhibition of neuronal apoptosis, improvement of energy metabolism disorder, and microcirculation of brain [41]. 				
Respiratory system	No bronchial constriction [42].	Antitussive effects by reducing citric acid-induced coughing [43].				
Digestive system	A partial agonist of α_1 -adrenoreceptor and an antagonist of α_2 -adrenoreceptor. (1) Relaxing the intestinal smooth muscle and the intestine [3]; (2) A modest reduction in contractions for rabbit duodenum [42]; (3) Both of the above are also supported with it is an antagonist of α_2 -adrenoreceptor [5].	 (1) Treating non-alcoholic fatty liver disease [26]; (2) Ameliorating carbon tetrachloride-induced hepatic fibrosis [44]; (3) For choline, maintaining the function and health of liver [45,46]. 				
Uterus	Uterine contraction (pregnancy), deduced from the fact that synephrine is an agonist α_1 -adrenoreceptor [47].	 Regulation of uterus effect (pregnancy and non-pregnancy) [27,48]: (1) Stimulation of uterine contraction [49,50]; (2) Inhibition of convulsive uterus [51]; (3) Reducing uterine bleeding [52]. 				
Blood sugar	Inhibiting α_1 -adrenoreceptor and α -glycosidase, and presenting a hypoglycemic effect which can be also deduced from it being an antagonist of α_2 -adrenoreceptor [5,53,54].	Ameliorating and protecting high-glucose-induced endothelial cell senescence by upregulation of SIRT1 and downregulation of p16 ^{INK4A} [55].				
Anti-inflammatory effect	/	 (1) Inhibition of TXB2 and IL-10 secretion, and production of NO [56]; (2) Inhibition of NF-κB and AKT signal pathways [57]; (3) Improvement of cellular membrane permeability, and inhibition of inflammatory factors and lipid peroxidation [58]. 				
Antidepressant activity	Anti-depressant activity by modulating noradrenergic neurotransmission and stimulating α_1 -adrenoceptor [59–61].	/				
Anti-obesity	Weight loss, anti-obesity, and regulating fat metabolism, due to that synephrine is a partial agonist β_3 -adrenoreceptor, and can weakly bind on β_3 -adrenoreceptor [62], together with lipolytic and thermogenic effects [63].	/				
Renal protection	/	 Reducing and ameliorating renal interstitial fibrosis [64]; Ameliorating hydrogen peroxide-induced renal tubular epithelial cell injury [65]; Protecting adenine-induced chronic renal failure [66]; Inducing diuresis [27]. 				
Pharmacokinetics	 Pharmacokinetic characteristics [67–69]: (1) Oral ingestion absorption was fast, and the time to peak is approximately ranged from 1 to 2 h after administration; (2) The biological half-life is about 2 h; (3) The bioavailability is approximately 22%; (4) The metabolism is exerted predominantly in the liver, and it can be rapidly removed from the bloodstream by hepatic uptake; (5) Cannot cross the blood–brain barrier 	 Pharmacokinetic characteristics [70,71]: (1) Rapid absorption after oral administration (2) Fast and extensive distribution; (3) The biological half-life is about 4 h; (4) The time to peak is approximately 3 h after administration; (5) The bioavailability is above 90%; (6) Most excreted from urine. 				

^a: /, no related reports obtained.

From the pharmacokinetic characteristics (Table 3) of synephrine and stachydrine, both compounds can be rapidly absorbed after oral administration. The relative bioavailability is approximately 22% for synephrine and 90% for stachydrine, respectively. Considering that the contents of stachydrine in these herbs except *Zhiqiao* are approximately one-third to two-thirds of that of synephrine, some pharmacological contributions from the bioavailable

differences in the two ingredients would be balanced, to a great extent, by their contents in these herbs. Together with most contrary pharmacological activities mentioned above (shown in Table 3), these indicate that both ingredients can be considered, only from their pharmacological activities, as a pair of antagonists in these citrus herbs. However, they both present many contrary bioactivities and also show some different or synergetic bioactivities, such as neuroprotective, hepatoprotective, renal protective, antitussive and anti-inflammatory effects from stachydrine, and gastrointestinal relaxation, antidepressant and anti-obesity activities from synephrine, along with their synergistic activities in uterine contraction and anti-diabetes. Thus, when they are used for other medicinal purposes, their contrary bioactivities, usually acting as their individual adverse effects, can be partly canceled out by each other. From this view, they can also be considered as a pair of synergists or associates from their contributions to the pharmacological functions and safety of these herbs.

It is noteworthy that there are no related reports for another compound with some of the pharmacological activities that are presented in Table 3 for synephrine or stachydrine. However, this does not mean that it has no similar, different or even contrary bioactivities to the identical organ or tissue. As described above, stachydrine was eventually identified from these herbs, using the detection method of choline analogs. Simultaneously, from the pharmacological activities, especially various cardio-cerebrovascular effects, stachydrine presents various contrary effects compared to synephrine and other adrenergic receptor agonists in Table 3 (marked in bold font for items (1), (5), (6) and (8) of the row "cardio-cerebrovascular system"). Thus, it is indicated that stachydrine seems to have some cholinergic activities, and is more like an agonist of M-type cholinergic receptor. Considering that their contents changed with the harvest time of these herbs simultaneously and similarly, stachydrine and synephrine can be also considered as a pair of bioactive equilibrists in the *Citrus* genus, like a pair of sympathetic and parasympathetic neurotransmitters in the human body.

Based on the above, many convoluted pharmacological functions for the aqueous extract or the decoction of *Zhishi*, *Zhiqiao*, *Qingpi* or/and *Chenpi* can be scientifically and rationally interpreted, including an excitatory effect on the in vitro and in vivo uteruses of both pregnant and non-pregnant rabbits, reducing cerebrovascular resistance and increasing cerebral blood flow, constricting gallbladder and a two-way regulating effect on gastrointestinal smooth muscle. Moreover, some explanations for the four Chinese herbs harvested from the different parts and growth times of the *Citrus* genus having different functions are presented in Section 3.

3. Discussion

Inspired by the traditional and modern pharmacodynamics of four Chinese citrus herbs, here, possible choline analogs were discovered from these herbs using the TLC method with a specific chromogenic reagent, which led to the identification of stachydrine and choline based on their NMR and HRMS data. After this, a TLCS method was first established for the quantitative analyses of stachydrine and choline, and the contents of both ingredients and synephrine in 39 samples were determined. Based on this, the statistical analyses of the contents of these three ingredients were performed, and then the pharmacological effects and pharmacokinetics reported for stachydrine and synephrine were comprehensively compared and analyzed. The results showed that stachydrine and synephrine can be considered as a pair of bioactive equilibrists, especially in the cardio-cerebrovascular protection from these citrus herbs, and which can, to a great extent, present some reasonable interpretation for the complex pharmacological functions of these herbs. Moreover, some important and relevant aspects are further discussed and developed as follows.

3.1. A Simple Method Detecting Choline Analogs from Plant Resource

Based on the extended structure–activity relationships of cholinergic and anti-cholinergic agents (Figure 1), choline analogs were speculated to be agonists or antagonists of cholinergic receptors, and have various bioactivities in the cardio-cerebrovascular, digestive and nervous systems, and the eyes, among others. Simultaneously, some of them also belong to betaine analogs, and these compounds have various bioactivities including anti-ulcer, regulating gastrointestinal function and treating liver diseases, and have multiple effects on homocysteine metabolism, which is very helpful for the protection of the cardio-cerebrovascular system and the kidneys [73]. For example, stachydrine was recently reported to have very extensive bioactivities (Table 3), and it was also isolated from another Chinese herb *Yimucao* (the overground part of *Leonurus japonicus* Houtt.) [26]. Based on the above TLC procedure for discovering stachydrine and choline from these citrus herbs, a simple and rapid method was established for quickly discovering possible choline analogs from botanical resource, and schemed as Figure 4.



Figure 4. A simple procedure for quickly discovering choline analogs from botanical resources. The specific chromogenic reagent (improved Dragendorff's reagent) was prepared by adding a mixture of bismuth nitrite (0.82 g), potassium iodide (11.06 g) and 50% (v/v) phosphoric acid (90 mL) into a 100 mL volumetric flask to make a constant volume using water.

The procedure (Figure 4) can be used for the discovery of choline analogs, including some betaine analogs. There are two key factors for discovering choline analogs: one is the specific chromogenic reagent (improved Dragendorff's reagent), and the other is the color of the spots. Generally, two chromogenic reagents, Dragendorff's and Wagner, are used for the color reaction of most alkaloids, but they present poor effects for some alkaloids with a low molecular weight. Considering that an acidic environment should be provided for the color reaction, the acetic acid was replaced with phosphoric acid in Dragendorff's reagent for improving the chromogenic sensitivity, referring to what Zhang N, et al. reported [74]. Moreover, if the color of the spots is red, such as orange-red or brownish-red, the compounds represented by the spots are likely not choline analogs but rather alkaloids, possibly some multi-methoxy flavonoids, or multi-methoxybenzenes (such as α - and β -asarones), since the potassium bismuth iodide reagent colors these compounds to orange-red or brownish-red. This was also supported by the fact that compound **3** was found not to be a choline analog, and colored an orange-red spot **c** (Figure 3). Furthermore, the brown color can be more accurately defined, as the absorption wavelengths ranged

from 550 to 580 nm when the thin-layer plate was scanned in 20 to 50 min after being taken from the chromogenic reagent.

Moreover, possible choline analogs can be quickly identified by the combinational method of preparative TLC and ¹H NMR, since these compounds would present one to three single peaks with some specific ¹H chemical shifts ranging from 3.20 to 3.60 ppm (4.45 to 4.75 ppm for methylpyridine-type choline analogs, such as trigonelline), assigned to three to nine hydrogens (one to three methyl groups). It is noteworthy that these analyses should eliminate the signal peaks from the deuterated methanol used for the NMR experiments or the possible residual methanol in the process of sample preparation. Thus, the deuterated solvents MeOH- d_4 should be avoided to use for the NMR experiments as much as possible. Considering that these compounds contain a structural fragment of the quaternary ammonium, D₂O is considered the preferred solvent.

According to the LODs of stachydrine and choline, the detectable content of choline analogs in botanical resources is approximately 1.0 to 1.4 μ M per gram of sample powder, and equal to contents of 0.01% to 0.05% in dried botanical resources, assuming the molecular weights of choline analogs are less than 500. If a highly efficient thin-layer plate is used for the method, the detection sensitivity would be increased. Using this detection procedure of choline analogs, choline and stachydrine were also detected in samples from the herbs Xiangyuan (dried fruits of Citrus wilsonii Tanaka) and Foushou (dried fruits of Citrus medica L. var. sarcodactylis Swingle) (Figure S3a), and this was also in accordance with previous publications [43,75]. These indicated that choline and stachydrine are widely distributed in Chinese herbs of the *Citrus* genus. Moreover, the results showed that stachydrine and/or choline were also discovered in the leaves of *Citrus* genus plants, with mostly higher contents than in their individual fruits (Figure S3b). Moreover, some possible choline analogs can be also detected in other botanical resources, including some Chinese herbs such as Huangliang (Coptidis Rhizoma), Juhua (dried capitulum of Chrysanthemum morifolium Ramat.) and *Chuanxiong* (dried rhizome of *Ligusticum chuanxiong* Hort.) (Figure S4). This indicates that the method can effectively detect choline or its analogs in various botanical resources.

3.2. The Contents of Stachydrine, Choline and Synephrine in These Citrus Herbs

According to the theory of Chinese medicine, these four citrus herbs have different pharmacological functions [1]. To clarify their substance bases and the reasons for their different functions, many studies on their active ingredients such as flavonoids, alkaloids and essential oil were reported [2,3,76]. Citrus flavonoids and alkaloids were generally considered the two main components responsible for the pharmacological functions of these herbs [76]. However, it remains a convoluted fact that for all the pharmacological activities of the ingredients reported in these herbs, it is difficult to clarify the pharmacological functions of their decoctions. Among them, synephrine acts as an agonist of adrenore-ceptor, and is widely distributed in these herbs. Here, stachydrine and choline were also discovered from these herbs and were reported to have various bioactivities. Considering that stachydrine and synephrine can be considered as a pair of bioactive equilibrists, it was expected to give some reasonable explanations for the convoluted pharmacological functions presented by these herbs. Thus, the contents of these three ingredients in these herbs were further determined.

As there is no chromophore in the structures of stachydrine and choline, it is unsuitable to perform their quantitative analyses using the HPLC-UV method, although this method was used for the quantitative analyses of stachydrine in Yimucao and choline in various plants. According to the detection procedure of choline analogs, stachydrine and choline can be perfectly isolated from their adjacent spots in these herbs (Figure 1). Thus, here, the TLCS method was selected for the quantitative analyses of stachydrine and choline, although the HPLC-MS/MS method can also be used for both components [71].

The quantitative analyses indicated these three ingredients have similar changing trends with the increase in growth time. The contents of all three compounds decrease from

Zhishi (harvested in June) to *Zhiqiao* (harvested in July), and from *Qingpi* (harvested in July) to *Chenpi* (harvested in January of next year). However, the decreasing speed of stachydrine is slower than that of synephrine, which was also supported by the depth analyses for the ratio values of stachydrine and synephrine, comparing *Zhishi* (0.44 ± 0.16) with *Zhiqiao* (2.43 ± 1.03) (p < 0.05) and *Qingpi* (0.34 ± 0.14) with *Chenpi* (0.67 ± 0.33) (p < 0.05). Although they were collected from different plants, the changing trends of synephrine, choline and stachydrine in 39 batches of samples are certain according to statistical analyses (Table 2), and among them, those of synephrine in various citrus herbs were also supported by many reports [76].

It is noteworthy that some multi-methoxy flavonoids having various bioactivities in the cardiovascular system, such as nobiletin and tangeretin [77], are reported to be widely distributed in the fruits or peels of many Citrus genus plants, including these herbs [78,79]. Simultaneously, their contents in the peels are much higher than those in other tissues (such as sarcocarp and seed) of these herbs [78,80], in accordance with the distribution of stachydrine in the fruits and peels of these herbs. Moreover, their contents in the fruits or peels of Citrus genus plants also present a decreasing trend with the delay of harvest time [81,82], which is similar to the decreasing trend of stachydrine and choline. It was reported that stachydrine is a proline betaine, and choline is also a precursor of glycine betaine [73,83,84]. All the bio-syntheses of stachydrine, choline and multi-methoxy flavonoids were catalyzed by S-adenosyl-methionine (SAM)-dependent methyltransferases with a universal methyl donor SAM [85–87]. Differently, the subclassified N-methyltransferases are responsible for the bio-syntheses of stachydrine and choline, while the sub-classified O-methyltransferases are responsible for that of multimethoxy flavonoids [87]. The similar changing trends of stachydrine/choline and multimethoxy flavonoids in the same tissue of the *Citrus* genus plants indicate there is a kind of intrinsic mechanism for simultaneously regulating both sub-classified enzymes with a similar effect. More probably, there are some physiological needs regulating the whole biosynthesis pathway involving SAM-dependent methyltransferases down in the fruits and peels of these citrus genus plants. Thus, it is worth further studying the physiological regulations of these Citrus genus plants to these components in their fruits and peels, which would be very helpful for clarifying the different pharmacological functions of these herbs and the regulation relationship of the two sub-classified enzymes in *Citrus* genus plants.

3.3. Communication between Active Ingredients and Pharmacological Effects of These Herbs

It was reported that these citrus herbs mainly contain flavonoids, alkaloids, essential oils and coumarins [2,15,88]. Many experiments have indicated that Citrus flavonoids exert multiple beneficial effects on cardiovascular and metabolic health through antioxidant, antidiabetic and anti-inflammatory activities, and by modulating lipid metabolism and adipocyte differentiation, etc. [89,90]. Simultaneously, essential oils have extensive pharmacological activities in the central nervous system, such as sedation, hypnosis, antianxiety and anti-depression, and also in the digestive system, including gastro- and hepatoprotective effects [91]. Considering that the decoctions of these herbs were usually used for treating some diseases, the contributions of these ingredients to the pharmacological effects from the application of these herbs would be lower than anticipated because of the weak hydrophilicity of many flavonoids, essential oil and coumarins in these herbs. Here, two water-soluble components, stachydrine and choline, present extensive biological activities, including beneficial effects on the cardio-cerebrovascular and nervous systems and the kidney, liver, and blood, and obviously regulating uterus effects (pregnancy and non-pregnancy), among others. (Table 3). Taken together with the contents of stachydrine and synephrine in these herbs, and the balance from their pharmacokinetics after oral administration, it was inferred that this pair of equilibrists plays an unignorable role in the pharmacological effects of these herb decoctions.

Some reports indicated that aqueous extracts or the decoctions of these herbs can contract the in vitro and in vivo uteruses of both pregnant and non-pregnant rabbits, and relax the in vitro uterus of pregnant rats or mice [92–94]. Ahangarpour et al. confirmed that the aqueous extract of *C. aurantium* flowers can reduce spontaneous motility and decrease the uterus contractions of pregnant rats, related to voltage-dependent calcium channels and without involving β -adrenoceptors and opioid receptors [95]. From Table 3, many reports have indicated that stachydrine can not only stimulate uterine contraction but also inhibit uterine spasms in both pregnant and non-pregnant rabbits [27,48,49,51]. Thus, the discovery of stachydrine with good water solubility and considerable content in these herbs can provide some reasonable interpretations for the heterogeneous effects of these herb decoctions on the uterus, depending on the different contents of stachydrine and other related components in these herbs and the physiological states of the uterus. Moreover, choline has similar effects on the uterus as stachydrine [51], while its contents in these herbs are only about 1/5 to 1/25 of those of stachydrine (Table 2).

Traditionally, these herbs are used for the treatment of some cardiovascular disease, and have various beneficial effects on cardiovascular health, including antioxidant and anti-inflammatory, hypolipidemic, anti-thrombosis and anti-atherosclerosis effects, and cardio-cerebrovascular protection, etc. [3,96,97]. Recently, Mahmoud et al. summarized that *Citrus* flavonoids confer cardiovascular protection via their antioxidant, antidiabetic, anti-inflammatory, anti-atherosclerosis and other biological activities [89,90], and it was reported that some citrus flavonoids such as a multi-methoxy flavonoid nobiletin have antihypertensive activity [6]. Simultaneously, Pontifex et al. also pointed out that Citrus fruits should be encouraged in the diet for their potential neurological benefits [98]. However, they also recommended that further studies and clinical trials should be performed for evaluating their efficacy. Simultaneously, it is noteworthy that the poor bioavailability of many citrus flavonoids dampens their systemic effects after oral administration, due to the combination of their degradation by intestinal bacterial enzymes, the poor hydrophobic nature of aglycone, the efflux of intestinal P-glycoprotein and the metabolism of cytochrome P450 [99–101]. Moreover, the concentrated distributions of some citrus flavonoids such as tangeretin and naringenin in the kidney, lung and liver will also influence the actual effects on the cerebrovascular system [102,103]. Thus, these factors would reduce the actual effects of citrus flavonoids on the cardio-cerebrovascular system, although their beneficial effects are confident. Based on these findings, stachydrine with high bioavailability (above 90%) and good tissue distribution would play an important role in cardiovascular and cerebral protection, since it has extensive beneficial effects on the cardio-cerebrovascular system (Table 3). Moreover, the rapid vascular relaxation, slowing heart rate and decreasing cardiac output, and the increasing coronary and myocardial blood thanks to stachydrine can also offset the possible adverse effects of synephrine and N-methyltyramine on the heart for non-drug purposes, which is probably one of the reasons why blood pressure sometimes presented an inapparent or transient raise and even decline after the oral administration of these herb decoctions to experimental animals [7,8]. Furthermore, Liu et al. reported that choline also has anti-hypertensive and cardiovascular protective effects [104].

It was reported that the concentrated solutions (1.5 g dried herbs per milliliter) of these citrus herb decoctions have obvious in vitro antiplatelet aggregation activities in human platelets [105]. Synephrine would increase the level of platelets [37]. Conversely, stachydrine can not only inhibit platelet aggregation but also ameliorate platelet-mediated thrombo-inflammation [27,28,38]. Thus, considering that stachydrine has good water solubility and bioavailability, and its considerable contents in these herbs, it can be confirmed that stachydrine is an important component in the anti-platelet aggregation of these herb decoctions, although some flavonoids, such as hesperidin and naringin [106,107], were also reported to have inhibitory activity against platelet aggregation [89,90].

Furthermore, choline is already used for the treatment of nonalcoholic fatty liver disease [46,108]. Thus, it can be deduced that choline in the decoctions of these herbs, especially *Qingpi*, would play an important role in maintaining the function and health of the liver [45,46]. Moreover, choline is also beneficial for cardiovascular and atherosclerosis diseases, and possibly neurological disorders [109,110]. Besides the protective effects of

Citrus flavonoids on the cardiovascular system mentioned above, they also have beneficial effects on gastrointestinal health and function [13,14], and renal, hepatic and nervous system protections [111–113]. Nevertheless, the two-way regulating effects of these herb decoctions on gastrointestinal smooth muscle should result from the combinational effects of flavonoids, essential oil and synephrine [13,15,92]. Simultaneously, citrus flavonoids, coumarins, stachydrine and choline should play main roles in the neuroprotective effects of these citrus herbs [2,88,113], while essential oil, γ -aminobutyric acid and synephrine should be responsible for various functions of the central nervous system, such as sedation, hypnosis, anxiolytic, anticonvulsant and anti-depression [11]. Moreover, citrus flavonoids, essential oil and stachydrine in these herbs could have antitussive and expectorant effects on the respiratory system.

Taken together, stachydrine should play an important role in the pharmacological functions of these citrus herbs, especially it can dual-directionally regulate the uterus and has various beneficial effects on the cardio-cerebrovascular system, blood, kidney and liver. Simultaneously, as a pair of bioactive equilibrists, the cardio-cerebrovascular protection of stachydrine can counteract the possible cardiovascular risk brought out from synephrine, which is very beneficial for the safe use of these citrus herbs. Moreover, together with the pharmacological activities of alkaloids (choline, synephrine, *N*-methyl tyramine and γ -aminobutyric acid) and *Citrus* flavonoids, essential oil and coumarins, these can more scientifically and reasonably interpret the substance bases for the various pharmacological effects of these citrus herb decoctions. Conversely, the differences in the formation, content, water solubility, extractability and pharmacokinetic characteristics of these components would lead to efficient differences among these four Chinese citrus herbs.

4. Materials and Methods

4.1. Materials, Chemicals and Reagents

Ten dried samples (Table 1) of Zhishi, Qingpi and Chenpi, together with nine samples (Table 1) of Zhiqiao, from different places of production were purchased from Chengdu Huichu Technology Co., Ltd. (Chengdu, China). Moreover, eight leaf and fruit samples of Citrus aurantium L. and its cultivated varieties, Citrus junos Siebold ex Tanaka, and Citrus reticulata Blanco 'Zhangtouhong' were collected from different habitats and growth ages and were identified by senior agronomist Yao Nie, and some fresh leaves and fruits of Xiangyuan were collected in Xinyu, China. The following Chinese herbs were purchased from Yifeng Pharmacy (Xialuo Branch, Nanchang, China): Foshou, the dried fruits of Citrus medica L. var. sarcodactylis Swingle (Guangxi, China); Xiangyuan, the dried fruits of Citrus wilsonii Tanaka; Huanglian, the dried rhizome of Coptis chinensis Franch (Sichuan, China); *Chuanxiong*, the dried rhizome of *Ligusticum chuanxiong* Hort. (Sichuan, China); *Dafupi*, the dried peel of Areca catechu L. (Yunnan, China); Banxia, the processed products according to the legal process for the dried tuber *Pinellia ternata* (Thunb.) Breit; *Juhua*, the dried capitulum of Chrysanthemum morifolium Ramat.; Mahuang, or Ephedrae Herba; Duzhong, the dried bark of Eucommia ulmoides Oliv.; Kushen, the dried root of Sophora flavescens Ait.; and Gancao, or Glycyrrhizae Radix et Rhizoma.

Reference standard choline chloride (No. C12799084) with a purity of 98% was purchased from Shanghai Macklin Biochemical Co., Ltd. (Shanghai, China), and stachydrine hydrochloride (No. PS012344) and synephrine (No. PS000966), each with purity of more than 98%, were purchased from Chengdu Push Bio-Technology Co., Ltd. (Chengdu, China).

TLC silica gel plates were purchased from Qingdao Ocean Chemical Co., Ltd. (Qingdao, China), and HPTLC Silica gel 60 F254 was purchased from Merck KGaA (Darmstadt, Germany). Bismuth subnitrate used for preparing the chromogenic reagent of TLC analyses was purchased from Aladdin Chemistry Co., Ltd. (Shanghai, China). Chromatographicgrade methanol (Anhui Tedia High Purity Solvents Co., Ltd., Anqing, China) and sodium dodecyl sulfonate (Shanghai Baihe Chemical Plant, Shanghai, China) were used for the HPLC analyses of synephrine. The ultra-pure water was prepared by the TST-UPB-10 ultra-pure water machine (Shijiazhuang TST Equipment Co., Ltd., Shijiazhuang, China). Other chemicals (analytical purity) were purchased from China National Pharmaceutical Group Co., Ltd. (Beijing, China).

4.2. Detection of Choline Analogs

4.2.1. Controls and Chromogenic Reagents

Choline chloride was used as a positive control, and γ -aminobutyric acid and synephrine were taken as excluded controls. Two chromogenic reagents (Dragendorff's and Wagner) and an improved Dragendorff reagent for alkaloids were selected for color reactions on the thin-layer plate. The stock solutions were prepared by the equal mixture of solution I (0.85 g of bismuth nitrite dissolved in 10.0 mL of acetic acid with analytical purity and 40.0 mL of water) and solution II (8.00 g of potassium iodide dissolved in 20.0 mL of water), and 1.0 mL of the stock solution, 2.0 mL of acetic acid (analytical reagent) and 10.0 mL of water were mixed as Dragendorff's reagent before use. Then, 1 g of iodine and 10.0 g of potassium iodide were dissolved in 50 mL of water, and the Wagner reagent was prepared by transferring this solution into a 100 mL volumetric flask, subsequently supplementing it with 2.0 mL of acetic acid and the required amount of water to make a constant volume. The improved chromogenic reagent was prepared by adding a mixture of bismuth nitrite (0.82 g), potassium iodide (11.06 g) and 50% (v/v) phosphoric acid (90 mL) into a 100 mL volumetric flask to make a constant volume using water.

4.2.2. Reference and Sample Solutions

A total of 25.0 mg each of choline chloride, stachydrine hydrochloride and synephrine was transferred into a volumetric flask, and then a concentration of $1.0 \text{ mg} \cdot \text{mL}^{-1}$ for three reference solutions was prepared by dissolving and supplementing with 80% (v/v) ethanol to a constant volume of 25 mL.

For Chinese herb slices, 20.0 g of herb slices was placed into a decocting jar, and 250 mL of purified water was added to it, soaking for 30 min. Next, the mixture was decocted for 30 min on MSD-1-12 ceramic decocting equipment produced by Meisidi Craft Products Factory (Caozhou, China), and then the extracted liquid was poured out of the jar to obtain decoction I. Subsequently, another 250 mL of purified water was added to the decocting jar and the herb residue was decocted for 20 min, and the extracted liquid was poured out to obtain decoction II. Finally, decoctions I and II were put together to obtain the decoction of each herb. For the detection of choline analogs, 100 mL of the decoction was concentrated into a small amount of mixture under vacuum, which was then transferred to a 10 mL volumetric flask to make a constant volume with purified water. Then, 1000 μ L of the suspension was transferred to a centrifuge tube with a pipette, diluting with equal volume of 80% ethanol (v/v), and centrifuged to obtain the supernatant. After filtering with a 0.45 μ m microporous membrane, the sample solution for the decoction of each herb was obtained.

Fresh samples were cut into thin slices and then dried under 60 °C ambient temperature in a blast drying oven. All dried samples with the amount of 20 to 30 g were crushed into coarse powder and then passed through a 65-mesh sieve to obtain their corresponding powders. Then, 2.0 g of each powder sample was placed into a 50 mL Erlenmeyer flask with a stopper, to which 25 mL of 80% (v/v) ethanol was added. After sonicating twice for 10 min each time in a DK-410T water bath sonicator with a frequency of 40 kHz, the mixture was filtered, and the residue was washed twice with 80% (v/v) ethanol. The filtrate was concentrated under vacuum, and the residual suspension was transferred into a volumetric flask to prepare 2 mL of the sample solution with 80% (v/v) ethanol.

4.2.3. TLC Analysis for Choline Analogs in Chinese Herbs

According to the general procedure of TLC analysis, the reference solutions of choline, γ -aminobutyric acid and synephrine were each sampled on a thin-layer plate, and then developed with solvent systems I (ethyl acetate–95% ethanol–formic acid) and II (*n*-butanol–

glacial acetic acid–water), respectively. After drying, the thin-layer plates were visualized with Dragendorff, Wagner and improved Dragendorff reagents.

According to the optimized procedure, the TLC analyses for sample solutions from Chinese herbs *Zhishi*, *Zhiqiao*, *Qingpi* and *Chenpi* were performed for discovering probable choline analogs. After this, other herbs from the *Citrus* genus, including *Foushou* and *Xiangyuan*, and some samples of the leaves from the plants of the *Citrus* genus were also detected for possible choline analogs. Moreover, to further verify the efficiency of the method for detecting choline analogs, the TLC analyses for sample solutions from some other Chinese herbs, including *Huanglian*, *Juhua*, *Mahuang*, *Chuanxiong*, *Dafupi*, *Banxia*, *Duzhong*, *Kushen* and *Gancao*, were also determined. Among these herbs, it was reported that herbs *Chuanxiong* and *Huanglian* contain choline or its analogs.

4.3. Isolation and Identification of Choline Analogs in Chinese Herbs Zhishi

According to the sample solution process in Section 4.2.2, a solution of the *Zhishi* mixed sample from 10 different producing areas was prepared. Using preparative TLC, probable choline analogs in the Chinese herb *Zhishi* were isolated. Solvent I (ethyl acetate–95% ethanol–formic acid, 10:4:5) was used as the developing agent, and probable choline analogs were located by visualization of the slices of thin-layer plates immersed in the improved Dragendorff's reagent.

Compounds **1** and **2** were identified based on the spectral analyses of their NMR and MS (Figure S5), combining their physicochemical analyses and the comparison of reported data [17,18]. The NMR data were recorded on a Bruker AV-600 or AV-400 MHz NMR spectrometer, and MeOH- d_4 and D₂O were used as the solvents dissolving compounds **1** and **2**, respectively. Compound **3** was identified based on the spectral analyses for its HRMS, comparing the TLC profiles of synephrine, compound **3** and their mixture. All HRMS data were obtained with ESI ion source (positive ion mode) from a TripleTOF 5600⁺ hybrid quadrupole time-of-flight mass spectrometer system (AB Sciex, Framingham, MA, USA).

4.4. *Quantitative Analyses of Chlorine and Stachydrine with TLCS Analysis* 4.4.1. Procedure of TLCS Analysis

Reference and sample solutions were prepared according to the procedure described in Section 4.2.2, and all of them were filtered by a 0.45 μ m microporous membrane before use. Certain volumes of sample solutions and reference solutions of choline chloride or stachydrine hydrochloride were sampled on an analytical and glass-based silica gel TLC plate (Qingdao Ocean Chemical Co., Ltd., Qingdao, China) placed on a YOKO-TD electric strip spotter (Whyoko New Technology Development Co., Ltd., Wuhan, China), and the plate was placed in a developing chamber. After being saturated with the solvent of ethyl acetate–95% ethanol–formic acid (10:4:5, v/v/v) for 15 min, the spots were developed for a span length of 60 mm. Subsequently, the plate was taken from the chamber and dried, and then was immersed in the improved Dragendorff reagent for 2 to 4 s. After taking them from the stained jar for 20 min, the TLCS analyses were performed with the reflection absorption method for 50 min on a KH-3000 Plus TLC Scanner (Shanghai Keze Biochemical Technology Co., Ltd., Shanghai, China), and the measured/reference wavelengths for choline and stachydrine were 568/820 nm and 574/820 nm, respectively. The contents of choline or stachydrine in Chinese herbs from the Citrus genus were calculated from the mean peak areas of three bands of choline or stachydrine from reference and sample solutions.

In the above procedure, the measured/reference wavelengths and the chromogenic stability were simultaneously determined from the 10 min interval determination in 70 min for the absorption curves of choline and stachydrine bands on the TLC plate after being visualized, and the sampling volume was 5.0 μ L. Depending on the typical chromatographic profile of TLC analyses for the sample solutions of Chinese citrus herbs, the resolution between choline (or stachydrine) and its nearest strip was calculated.

4.4.2. Methodology Validation

According to the general procedure of methodology validation, the limit of detection (LOD), the limit of quantitation (LOQ), precision, repeatability, linearity and range were evaluated using the reference standard of choline or stachydrine. The reproducibility was assessed using the powder of Zhishi. The recovery was tested using the powder of Zhishi with a known content of choline or stachydrine, and the reference standard of choline or stachydrine.

In detail, the LOD was defined as the lowest concentration of choline or stachydrine at which the signal-to-noise ratio was from 3 to 4, and the LOQ was defined as the lowest concentration of choline or stachydrine at which the signal-to-noise ratio was greater than or equal to 10 with a precision below 5%. According to the procedure of TLCS analysis in Section 4.4.1, six scans of a strip of choline or stachydrine on a plate were performed for evaluating the repeatability, using a choline or stachydrine solution $(1.0 \text{ mg} \cdot \text{mL}^{-1})$, and the sampling volume was 10.0 μ L. Simultaneously, intra- and inter-plate precision were assessed by detecting choline or stachydrine solutions, respectively, on a thin-layer plate with six replicates and on six thin-layer plates. The linear correlation was established using seven sampling volumes (three replicates for each) at a concentration of $1.0 \text{ mg} \cdot \text{mL}^{-1}$ for choline or stachydrine solution. The reproducibility was evaluated by the quantitative determination of choline or stachydrine in Zhishi powder (No. 20101001) with six replicates, and the sampling volume was 2.0 µL. The recovery was tested from 9 samples prepared by adding an amount of choline or stachydrine into the powders of Zhishi (No. 20101001) with a known content of choline or stachydrine, including three different amounts for choline or stachydrine with three replicates for each.

4.4.3. Quantitative Analyses for Samples

According to the validated procedure of TLC analyses, the quantitative analyses of choline or stachydrine in the 39 purchased samples, including 10 of Zhishi, 10 of Qingpi, 10 of Chenpi and 9 of Zhiqiao, were performed with the external standard method in triplicate for each sample. Three spots were sampled on the plate for each sample or standard solution. The sampling volume of each spot for the sample solutions of Zhishi was 2.0 μ L, and that for the sample solutions of Zhiqiao, Qingpi or Chenpi was 3.0 μ L. Simultaneously, the sampling volumes were 1.0 and 3.0 μ L for the reference solutions of choline and stachydrine, respectively. After being visualized, each lane on the plates was scanned to obtain the peak areas of choline and stachydrine in samples and corresponding standards, and then the contents of choline and stachydrine in various samples were calculated.

4.5. Quantitative Analyses of Synephrine with HPLC

4.5.1. Procedure of HPLC Analysis

A 5.0 mg·mL⁻¹ stock solution for synephrine was prepared by dissolving 10.0 mg of synephrine into 80% (v/v) ethanol, and transferred into a 2 mL volumetric flask which was further supplemented with 80% (v/v) ethanol to a constant volume. From this stock, a concentration of 0.5 mg·mL⁻¹ for standard solution was prepared by 10-fold dilution.

The concentrations of synephrine in sample solutions were determined with the external standard method, referring to the procedure of the Chinese pharmacopoeia [1]. Briefly, the quantitative analyses of synephrine were performed using a Waters e2695 separation system consisting of a model 2998 ultraviolet detector (Milford, CT, USA), and the detection wavelength was set at 275 nm. A SinoChrom ODS2 (4.6 mm × 250 mm, 5.0 μ m) (Elite, Dalian, China) was used as the chromatographic column, and the temperature was kept at 30 °C. A methanol and phosphate buffer with a ratio of 67:33 (v/v) was used as the mobile phase of the isocratic elution, and the flow rate was set at 1.0 mL/min. The phosphate buffer was prepared by dissolving 0.60 g of KH₂PO₄, 1.00 g of sodium dodecyl sulfonate and 1.0 mL of glacial acetic acid into 700 mL of water, which was then transferred into a 1000 mL volumetric flask and subsequently supplemented with the

required amount of water to make a constant volume. Moreover, the injection volume for all sample solutions and standard solutions was 10.0 μ L. Depending on the typical chromatographic profile of HPLC analyses for sample solutions of Chinese citrus herbs, the resolution between synephrine and its nearest chromatographic peak was automatically calculated by the HPLC system.

4.5.2. Methodology Validation

Similar to the validation of TLC analysis, the LOD was defined as the lowest concentration of synephrine at which the signal-to-noise ratio was from 3 to 4, and the LOQ was defined as the lowest concentration of synephrine at which the signal-to-noise ratio was greater than or equal to 10 with precision below 5%. The linear correlation was established using seven concentrations (0.05, 0.10, 0.20, 0.40, 0.80, 1.60 and 3.20 mg/mL) of synephrine solution with three replicates for each. Six injections for a standard solution with a concentration of 0.50 mg·mL⁻¹ were performed to evaluate the repeatability. Using a powder sample of Zhishi (No. 2010001), the stability of a sample solution in 24 h was evaluated, and the reproducibility was also assessed by the quantitative determination of synephrine in the powder sample with six replicates. The recovery was tested from nine samples prepared by adding an amount of synephrine into the powders of Zhishi (No. 20101001), including three different amounts of synephrine with three replicates for each.

4.5.3. Quantitative Analyses for Samples

All sample solutions used for the quantitative analyses of chlorine and stachydrine in Section 4.4 were simultaneously used for the quantitative analyses of synephrine according to the valid procedure, and the synephrine concentration of the reference solution was $0.5 \text{ mg} \cdot \text{mL}^{-1}$.

4.6. Statistical Analysis for the Contents of Three Ingredients in These Four Chinese Citrus Herbs

All statistical analyses were performed using the Excel program of Microsoft Office 2016, and a bilateral *t*-test was used for the comparison of two groups. A paired *t*-test was selected when the number of data was the same (both n = 10), while two-sample equivariance was selected for the *t*-test when the number of data was different (n = 10 and 9). A *p* value less than 0.05 shows that the data difference between two groups is significant, and that less than 0.01 indicates that the data difference between two groups is very significant.

4.7. Comprehensive Analyses for Pharmacological Effects of Four Chinese Citrus Herbs

Studies published in the past 10 years on the bioactivities of synephrine, chlorine and stachydrine were unsystematically searched using the Google Scholar search engine and the databases Medline, CNKI and RSC, using the keywords "pharmacological" or "activity" or "review", and "synephrine" or "chlorine" or "stachydrine". Furthermore, the relevant references in the obtained literature were also checked. Based on the literature results and the contents of synephrine, chlorine and stachydrine in these four Chinese citrus herbs, the pharmacological effects of these compounds were comprehensively analyzed, together with their contributions to the pharmacological effects of the four Chinese citrus herbs. Some reasonable explanations for the differences in the pharmacological activities of these four Chinese herbs have been presented here.

5. Conclusions

To discover some ingredients with cholinergic activity and further clarify possible reasons for the complex functions presented by four Chinese citrus herbs, a simple and specific method was first established for quickly discovering possible choline analogs, and stachydrine and choline were discovered in these citrus herbs. Then, a TLCS method was first established for the quantitative analyses of stachydrine and choline, and the contents of both ingredients and synephrine in 39 herb samples were determined. The

results showed that stachydrine and synephrine have commensurate contents in these herbs, and the contents of the two ingredients and choline in these herbs present similar decreasing trends with the delay of harvest time. However, the contents of synephrine decrease the fastest, while those of stachydrine decrease the slowest. Based on these findings, the pharmacological activities and pharmacokinetics reported for stachydrine and synephrine were compared, and the results indicated that stachydrine and synephrine can be considered as a pair of bioactive equilibrists in these citrus herbs, especially for their effects on the cardio-cerebrovascular system. Additionally, the results confirmed that stachydrine plays an important role in the pharmacological functions of these citrus herbs, especially in dual-directionally regulating the uterus, and in various beneficial effects on the cardio-cerebrovascular system, kidney and liver. Considering these findings, some more scientifical and reasonable interpretations would be concluded with the help of network pharmacology analyses performed for the main ingredients and the various pharmacological functions of these citrus herb decoctions, and which would indicate that the component differences in the formation, content, water solubility, extractability and pharmacokinetic characteristics can lead to the differences in the pharmacological functions of these citrus herbs. Furthermore, the quality markers of these herbs should be reevaluated based on this study, the results of which would present a good reference for the quality control of these herbs.

Supplementary Materials: The following supporting information can be downloaded at: https://www.mdpi.com/article/10.3390/molecules28093813/s1, Figure S1: Color development reactions of choline (1), synephrine (2) and γ -aminobutyric acid (3) on thin-layer plates; Figure S2: The linearity correlations between the amounts (*x*) and peak areas (*y*) of stachydrine (**a**), choline (**b**) and synephrine (**c**); Figure S3: Detection of choline analogs in samples originated from *Citrus* genus plants on thin-layer plates; Figure S4: Detection of choline analogs in samples originated from other plants on thin-layer plates; Figure S5: ¹H (up), ¹³C (middle) NMR and MS (down) spectra of compound **2**; Table S1: The recovery of stachydrine (*n* = 9); Table S2: The recovery of choline (*n* = 9).

Author Contributions: Conceptualization, G.Y.; methodology, G.Y. and Y.S.; software, G.Y., Y.S. and Q.W.; validation, G.Y. and Y.S.; formal analysis, G.Y., Y.S. and X.X.; investigation, Y.S., G.Y., X.X., T.Z., B.D., X.F. and Q.W.; resources, G.Y.; data curation, G.Y., Y.S., X.X., X.F. and B.D.; writing—original draft preparation, G.Y. and Y.S.; writing—review and editing, G.Y.; visualization, G.Y., T.Z. and B.D.; supervision, G.Y.; project administration, G.Y.; funding acquisition, G.Y. All authors have read and agreed to the published version of the manuscript.

Funding: This research was funded by the National Natural Science Foundation of China, grant numbers 82073745 and 81960636.

Institutional Review Board Statement: Not applicable.

Informed Consent Statement: Not applicable.

Data Availability Statement: Not applicable.

Acknowledgments: The authors are thankful to senior agronomist Yao Nie, who worked in Jiangxi Shunfutang Traditional Chinese Medicine Slices Co., Ltd., for his helps in the collections and identifications of various leaf and fruit samples of *Citrus aurantium* L. and its cultivated varieties, *Citrus junos* Siebold ex Tanaka, and *Citrus reticulata* Blanco 'Zhangtouhong'.

Conflicts of Interest: The authors declare no conflict of interest.

Sample Availability: Samples of the compounds 1, 2 and 3 are available from the authors.

References

- Chinese Pharmacopoeia Commission. *Pharmacopoeia of People's Republic of China*; Part 1; China Medicinal Science and Technology Press: Beijing, China, 2020; pp. 257–258, 199–200, 205–206.
- Lv, X.; Zhao, S.; Ning, Z.; Zeng, H.; Shu, Y.; Tao, O.; Xiao, C.; Lu, C.; Liu, Y. Citrus fruits as a treasure trove of active natural metabolites that potentially provide benefits for human health. *Chem. Cent. J.* 2015, *9*, 68. [CrossRef] [PubMed]

- 3. Xu, S.; Xu, J.; Zhang, X.; Zhang, T.; Liu, C. Research progress on Citri Reticulatae Pericarpium, Aurantii Fructus Immaturus, and Aurantii Fructus and Q-marker predictive analysis. *Chin. Tradit. Herb. Drugs* **2018**, *49*, 35–44.
- Jordan, R.; Midgley, J.M.; Thonoor, C.M.; Williams, C.M. Beta-adrenergic activities of octopamine and synephrine stereoisomers on guinea-pig atria and trachea. *J. Pharm. Pharmacol.* 1987, 39, 752–754. [CrossRef] [PubMed]
- Ma, G.; Bavadeka, S.A.; Schaneberg, B.T.; Khan, I.A.; Feller, D.R. Effects of synephrine and beta-phenylephrine on human alpha-adrenoreceptor subtypes. *Planta Med.* 2010, 76, 981–986. [CrossRef] [PubMed]
- 6. Kim, J.J.; Kim, K.; Jung, Y.R.; Bian, Y.; Ngo, T.; Bae, O.N.; Lim, K.M.; Chung, J.H. Co-existence of hypertensive and antihypertensive constituents, synephrine, and nobiletin in *Citrus unshiu* peel. *Molecules* **2019**, *24*, 1197. [CrossRef]
- 7. Cui, H.; Zhou, Y.; Lv, S.; Feng, S.; Huang, Y.; Song, Z.; Wang, C.; Liu, Z.; Sun, M. The different effects of 2 species of Fructus Aurantii Immaturus on cardiovascular and respiratory system of rats. *Chin. J. Inform. Tradit. Chin. Med.* **2010**, *17*, 41–43.
- 8. Chen, X.; Huang, Q.; Zhou, T. Studies of *Citrus aurantium* and its hypertensive ingredients on the cardiac functions and hemodynamic in comparison with dopamine and dobutamine. *Acta Pharm. Sin.* **1980**, *15*, 71–77.
- Parmar, H.S.; Kar, A. Antiperoxidative, antihypoidal, antihyperglycemic and cardioprotective role of *Citrus sinensis* peel extract in male mice. *Phytother. Res.* 2008, 22, 791–795. [CrossRef]
- Parmar, H.S.; Kar, A. Medicinal values of fruit peels from *Citrus sisensis*, *Punica granatum*, and *Musa paradisiacal* with respect to alterations in tissue lipid peroxidation and serum concentration of glucose, insulin, and thyroid hormones. *J. Med. Food* 2008, 11, 376–381. [CrossRef]
- 11. Suntar, I.; Khan, H.; Patel, S.; Celano, R.; Rastrelli, L. An overview on *Citrus aurantium* L.: Its functions as food ingredient and therapeutic agent. *Oxid. Med. Cell. Longev.* **2018**, 2018, 7864269. [CrossRef]
- 12. Wang, H.; Zhong, G.; Zhang, S.; He, J.; Zeng, J. Research progress on chemical constituents and pharmacological effects of *Zhiqiao* (Aurantii Fructus) and predictive analysis on quality markers. *Chin. Arch. Tradit. Chin. Med.* **2022**, 40, 184–192.
- Wang, S.; Bao, Y.; Li, T.; Yu, T.; Chang, X.; Yang, G.; Meng, X. Mechanism of Fructus Aurantii flavonoids promoting gastrointestinal motility: From organic and inorganic endogenous substances combination point of view. *Pharmacogn. Mag.* 2017, 13, 372–377. [CrossRef] [PubMed]
- 14. Stevens, Y.; Van Rymenant, E.; Grootaert, C.; Van Camp, J.; Possemiers, S.; Masclee, A.; Jonkers, D. The intestinal fate of citrus flavanones and their effects on gastrointestinal health. *Nutrients* **2019**, *11*, 1464. [CrossRef] [PubMed]
- González-Mas, M.C.; Rambla, J.L.; López-Gresa, M.P.; Blázquez, M.A.; Granell, A. Volatile compounds in citrus essential oils: A comprehensive review. *Front. Plant. Sci.* 2019, 10, 12. [CrossRef] [PubMed]
- 16. Cao, S.; Du, X.; Li, P.; Yuan, G.; Chen, S.; Chen, W.; Song, X. A chemical screening method for menaquinone-producing strains based on HPLC-UV technology. *J. Microbiol. Meth.* **2020**, *172*, 105907. [CrossRef]
- 17. Takeda, J.; Iwao, Y.; Karashima, M.; Yamamoto, K.; Ikeda, Y. Structural evaluation of the choline and geranic acid/water complex by SAXS and NMR Analyses. *ACS Biomater. Sci. Eng.* **2021**, *7*, 595–604. [CrossRef]
- 18. Hudec, J.; Mojzis, J.; Habanova, M.; Saraiva, J.A.; Hradil, P.; Liptaj, T.; Kobida, L.; Haban, M.; Holovicova, M.; Zvercova, D. In vitro cytotoxic effects of secondary metabolites present in *Sarcopoterium Spinosum* L. *Appl. Sci.* **2021**, *11*, 5300. [CrossRef]
- 19. Mercolini, L.; Mandrioli, R.; Trerè, T.; Bugamelli, F.; Ferranti, A.; Raggi, M.A. Fast CE analysis of adrenergic amines in different parts of *Citrus aurantium* fruit and dietary supplements. *J. Sep. Sci.* 2010, *33*, 2520–2527. [CrossRef]
- 20. Gong, B.; Li, Q.; Hu, X.; Xiao, Q.; Huang, L. Advances in chemical constituents and pharmacological activities of Fructus Aurantii. *South Chin. For. Sci.* **2019**, 47, 40–45.
- Porter, J.; Yoon, G.; Lozano, D.; Wolfing, J.; Tumbar, R.; Macrae, S.; Cox, I.G.; Williams, D.R. Aberrations induced in wavefrontguided laser refractive surgery due to shifts between natural and dilated pupil center locations. *J. Cataract. Refract. Surg.* 2006, 32, 21–32. [CrossRef]
- 22. Koh, A.H.W.; Chess-Williams, R.; Lohning, A.E. Differential mechanisms of action of the trace amines octopamine, synephrine and tyramine on the porcine coronary and mesenteric artery. *Sci. Rep.* **2019**, *9*, 10925. [CrossRef]
- Hibino, T.; Yuzurihara, M.; Kase, Y.; Takeda, A. Synephrine, a component of Evodiae Fructus, constricts isolated rat aorta via adrenergic and serotonergic receptors. J. Pharmacol. Sci. 2009, 111, 73–81. [CrossRef]
- Brown, C.M.; McGrath, J.C.; Midgley, J.M.; Muir, A.G.; O'Brien, J.W.; Thonoor, C.M.; Williams, C.M.; Wilson, V.G. Activities of octopamine and synephrine stereoisomers on alpha-adrenoceptors. *Br. J. Pharmacol.* 1988, 93, 417–429. [CrossRef]
- 25. Ruffolo, R.R., Jr.; Waddell, J.E. Aromatic and benzylic hydroxyl substitution of imidazolines and phenethylamines: Differences in activity at alpha-1 and alpha-2 adrenergic receptors. *J. Pharmacol. Exp. Ther.* **1983**, 224, 559–566.
- 26. Cheng, F.; Zhou, Y.; Wang, M.; Guo, C.; Cao, Z.; Zhang, R.; Peng, C. A review of pharmacological and pharmacokinetic properties of stachydrine. *Pharmacol. Res.* 2020, 155, 104755. [CrossRef]
- 27. Liu, X.; Yan, D.; Deng, X.; Zhao, B.; Xue, X.; Wang, S.; Zhang, Y.; Meng, J. Quality assessment of crude and processed Leonuri Fructus by chemical and color analysis combined with chemometric method. *Chin. Herb. Med.* **2018**, *10*, 388–395. [CrossRef]
- 28. Jiang, J.; Xiao, Q. Handbook of Effective Ingredients of Botanicals, 1st ed.; People's Health Publishing House: Beijing, China, 1986; pp. 991–992.
- 29. Chen, H.H.; Zhao, P.; Zhao, W.X.; Tian, J.; Guo, W.; Xu, M.; Zhang, C.; Lu, R. Stachydrine ameliorates pressure overload-induced diastolic heart failure by suppressing myocardial fibrosis. *Am. J. Transl. Res.* **2017**, *9*, 4250–4260.
- Liu, X.; Shan, X.; Chen, H.; Li, Z.; Zhao, P.; Zhang, C.; Guo, W.; Xu, M.; Lu, R. Stachydrine ameliorates cardiac fibrosis through inhibition of angiotensin II/transformation growth factor β₁ fibrogenic axis. *Front. Pharmacol.* 2019, 10, 538. [CrossRef]

- Zhao, L.; Wu, D.; Sang, M.; Xu, Y.; Liu, Z.; Wu, Q. Stachydrine ameliorates isoproterenol-induced cardiac hypertrophy and fibrosis by suppressing inflammation and oxidative stress through inhibiting NF-κB and JAK/STAT signaling pathways in rats. *Int. Immunopharmacol.* 2017, 48, 102–109. [CrossRef]
- 32. Zhang, C.; Shan, X.L.; Liao, Y.L.; Zhao, P.; Guo, W.; Wei, H.C.; Lu, R. Effects of stachydrine on norepinephrine-induced neonatal rat cardiac myocytes hypertrophy and intracellular calcium transients. *BMC Complement. Altern. Med.* **2014**, *14*, 474. [CrossRef]
- 33. Shan, X.; Zhang, C.; Liao, Y.; Wei, H.; Lu, R. Inhibitory effects of Stachydrine of Leonurus on cardiaomyocyte hypertrophy induced by norepinephrine. *Shanghai J. Tradit. Chin. Med.* **2013**, *47*, 70–72.
- Sun, Z.; Li, H.; Lv, R.; Zhao, P. Effects on calcium uptake capacity and activity of SERCA in rat sarcoplasmic reticulum of myocardial hypertrophy cell of stachydrine. *Chin. J. Exp. Trad. Med. Form.* 2010, 16, 118–122.
- Xie, X.; Yang, C.; Cui, Q.; Ma, W.; Liu, J.; Yao, Q.; Zhang, Z.; Xiao, L.; Wang, N. Stachydrine mediates rapid vascular relaxation: Activation of endothelial nitric oxide synthase involving amp-activated protein kinase and Akt phosphorylation in vascular endothelial cells. J. Agric. Food. Chem. 2019, 67, 9805–9811. [CrossRef] [PubMed]
- 36. Xie, X.; Zhang, Z.; Wang, X.; Luo, Z.; Lai, B.; Xiao, L.; Wang, N. Stachydrine protects eNOS uncoupling and ameliorates endothelial dysfunction induced by homocysteine. *Mol. Med.* **2018**, *24*, 10. [CrossRef] [PubMed]
- 37. D'Andrea, G.; Granella, F.; Leone, M.; Perini, F.; Farruggio, A.; Bussone, G. Abnormal platelet trace amine profiles in migraine with and without aura. *Cephalalgia* **2006**, *26*, 968–972. [CrossRef]
- Sun, X.; Zhou, M.; Pu, J.; Wang, T. Stachydrine exhibits a novel antiplatelet property and ameliorates platelet-mediated thromboinflammation. *Biomed. Pharmacother.* 2022, 152, 113184. [CrossRef]
- Yu, N.; Hu, S.; Hao, Z. Benificial effect of stachydrine on the traumatic brain injury induced neurodegeneration by attenuating the expressions of Akt/mTOR/PI3K and TLR4/NFκ-B pathway. *Transl. Neurosci.* 2018, 9, 175–182. [CrossRef]
- 40. Miao, M.; Wang, T.; Lou, X.; Bai, M.; Xi, P.; Liu, B.; Chang, B. The influence of stachydrine hydrochloride on the reperfusion model of mice with repetitive cerebral ischemia. *Saudi. J. Biol. Sci.* **2017**, *24*, 658–663. [CrossRef]
- 41. Liu, B.; Bai, M.; Peng, M.; Li, R.; Liu, T.; Miao, M. Protective effects and its mechanism of stachydrine on focal cerebral ischemia reperfusion injury in mice. *Chin. J. Clin. Pharmacol.* **2018**, *34*, 2295–2298.
- Tainter, M.L.; Seidenfeld, M.A. Comparative actions of sympathomimetic compounds: Synephrine-isomers and -ketone. J. Pharmacol. Exp. Ther. 1930, 40, 23–42.
- 43. Shi, Q.; Liu, Z.; Yang, Y.; Geng, P.; Zhu, Y.Y.; Zhang, Q.; Bai, F.; Bai, G. Identification of anti-asthmatic compounds in Pericarpium citri reticulatae and evaluation of their synergistic effects. *Acta Pharmacol. Sin.* **2009**, *30*, 567–575. [CrossRef]
- Zhang, J.; Yang, A.; Wu, Y.; Guan, W.; Xiong, B.; Peng, X.; Wei, X.; Chen, C.; Liu, Z. Stachydrine ameliorates carbon tetrachlorideinduced hepatic fibrosis by inhibiting inflammation, oxidative stress and regulating MMPs/TIMPs system in rats. *Biomed. Pharmacother.* 2018, 97, 1586–1594. [CrossRef]
- 45. Mehedint, M.G.; Zeisel, S.H. Choline's role in maintaining liver function: New evidence for epigenetic mechanisms. *Curr. Opin. Clin. Nutr. Metab. Care* **2013**, *16*, 339–345. [CrossRef]
- 46. Sherriff, J.L.; O'Sullivan, T.A.; Properzi, C.; Oddo, J.L.; Adams, L.A. Choline, its potential role in nonalcoholic fatty liver disease, and the case for human and bacterial genes. *Adv. Nutr.* **2016**, *7*, 5–13. [CrossRef]
- 47. Ko, H.C.; Chen, K.T.; Chen, C.F.; Su, J.P.; Chen, C.M.; Wang, G.J. Chemical and biological comparisons on evodia with two related species of different locations and conditions. *J. Ethnopharmacol.* **2006**, *108*, 257–263. [CrossRef]
- He, C.; Peng, C.; Dai, O.; Yan, L.; Liu, J.; Guo, L.; Xiong, L.; Liu, S. Chemical constituents from *Leonurus japonicus* injection. *Chin. Tradit. Herb. Drugs* 2014, 45, 3048–3052.
- 49. Dai, L.; Xie, X.; Sun, C.; Ao, H.; Dong, Y.; Peng, C. Effect of alkaloid monomer of *Yimucao* injection on isolated smooth muscle of uterus. *Nat. Prod. Res. Dev.* **2016**, *28*, 1633–1638.
- Zhou, F.; Liu, F.; Liu, J.; He, Y.L.; Zhou, Q.M.; Guo, L.; Peng, C.; Xiong, L. Stachydrine promotes angiogenesis by regulating the VEGFR2/MEK/ERK and mitochondrial-mediated apoptosis signaling pathways in human umbilical vein endothelial cells. *Biomed. Pharmacother.* 2020, 131, 110724. [CrossRef]
- 51. Zhen, J.; Wang, Y. The regulatory effects of stachydrine, choline and their combination on the contraction of isolated uterus. *Chin. J. Reprod. Health* **2017**, *28*, 127–130+137.
- Li, X.; Wang, B.; Li, Y.; Wang, L.; Zhao, X.; Zhou, X.; Guo, Y.; Jiang, G.; Yao, C. The Th1/Th2/Th17/Treg paradigm induced by stachydrine hydrochloride reduces uterine bleeding in RU486-induced abortion mice. *J. Ethnopharmacol.* 2013, 145, 241–253. [CrossRef]
- 53. Vatsavai, L.K.; Kilari, E.K. Interaction of *p*-synephrine on the pharmacodynamics and pharmacokinetics of gliclazide in animal models. *J. Ayurveda Integr. Med.* **2018**, *9*, 183–189. [CrossRef] [PubMed]
- 54. Taslimi, P.; Akıncıoglu, H.; Gülçin, İ. Synephrine and phenylephrine act as *α*-amylase, *α*-glycosidase, acetylcholinesterase, butyrylcholinesterase, and carbonic anhydrase enzymes inhibitors. *J. Biochem. Mol. Toxicol.* **2017**, *31*, e21973. [CrossRef]
- Servillo, L.; D'Onofrio, N.; Longobardi, L.; Sirangelo, I.; Giovane, A.; Cautela, D.; Castaldo, D.; Giordano, A.; Balestrieri, M.L. Stachydrine ameliorates high-glucose induced endothelial cell senescence and SIRT1 downregulation. *J. Cell Biochem.* 2013, 114, 2522–2530. [CrossRef] [PubMed]
- 56. Hu, Y.; Mao, A.; Yu, Z.; He, K. Anti-endotoxin and anti-inflammatory effects of Chinese herbal medicinal alkaloid ingredients in vivo. *Microb. Pathog.* 2016, 99, 51–55. [CrossRef] [PubMed]

- 57. Meng, J.; Zhou, C.; Zhang, W.; Wang, W.; He, B.; Hu, B.; Jiang, G.; Wang, Y.; Hong, J.; Li, S.; et al. Stachydrine prevents LPS-induced bone loss by inhibiting osteoclastogenesis via NF-κB and Akt signalling. *J. Cell Mol. Med.* **2019**, *23*, 6730–6743. [CrossRef]
- 58. Wang, F.; Wang, C. Study on the anti-inflammatory activity of stachydrine. Chin. Pharm. 2012, 23, 212–214.
- 59. Kim, K.W.; Kim, H.D.; Jung, J.S.; Woo, R.S.; Kim, H.S.; Suh, H.W.; Kim, Y.H.; Song, D.K. Characterization of antidepressant-like effects of *p*-synephrine stereoisomers. Naunyn-Schmiedeb. *Arch. Pharmacol.* **2001**, *364*, 21–26. [CrossRef]
- Song, D.K.; Suh, H.W.; Jung, J.S.; Wie, M.B.; Son, K.H.; Kim, Y.H. Antidepressant-like effects of *p*-synephrine in mouse models of immobility tests. *Neurosci. Lett.* 1996, 214, 107–110. [CrossRef]
- Malik, H.; Javaid, S.; Rasool, M.F.; Samad, N.; Ahamad, S.R.; Alqahtani, F.; Imran, I. Amelioration of scopolamine-induced amnesic, anxiolytic and antidepressant effects of *Ficus benghalensis* in behavioral experimental models. *Medicina* 2020, 56, 144. [CrossRef]
- Carpéné, C.; Galitzky, J.; Fontana, E.; Atgié, C.; Lafontan, M.; Berlan, M. Selective activation of β₃-adrenoceptors by octopamine: Comparative studies in mammalian fat cells. *Naunyn-Schmiedeb. Arch. Pharmacol.* **1999**, 359, 310–321. [CrossRef]
- 63. Mercader, J.; Wanecq, E.; Chen, J.; Carpéné, C. Isopropylnorsynephrine is a stronger lipolytic agent in human adipocytes than synephrine and other amines present in *Citrus aurantium*. *J. Physiol. Biochem.* **2011**, *67*, 443–452. [CrossRef]
- 64. Zhang, C.; Lu, Y.; Zhou, Y.; Tong, Q.; Wang, S.; Liang, W. Effect of stachydrine on expression of PERK of endoplasmic reticulum in renal tissue of rats with unilateral ureteral obstruction. *Chin. Tradit. Herb. Drugs* **2014**, *45*, 1591–1596.
- Liu, H.; Wang, R.; Shi, M.; Luo, Y.; Qiu, C.; Jia, R. Effect of stachydrine chloride on apoptosis induced by oxidative stress in renal tubular epithelial cells. *Chin. J. Int. Trad. West. Nephrol.* 2008, 9, 760–763.
- 66. Chen, M.; Chen, L.; Lian, Y.; Xie, L. Expression of connexin 40 and connexin 45 in renal tissue of rats with chronic renal failure and effect of treatment with Astragalus polysaccharide and stachydrine combination. *Chin. J. Pathophysiol.* **2014**, *30*, 494–502.
- 67. Hengstmann, J.H.; Aulepp, H. Pharmacokinetics and metabolism of ³H-synephrine. Arzneimittelforschung **1978**, 28, 2326–2331.
- Suzuki, O.; Matsumoto, T.; Oya, M.; Katsumata, Y. Oxidation of synephrine by type A and type B monoamine oxidase. *Experientia* 1979, 35, 1283–1284. [CrossRef]
- 69. da Silva-Pereira, J.F.; Bubna, G.A.; Gonçalves Gde, A.; Bracht, F.; Peralta, R.M.; Bracht, A. Fast hepatic biotransformation of p-synephrine and *p*-octopamine and implications for their oral intake. *Food Funct.* **2016**, *7*, 1483–1491. [CrossRef]
- 70. Li, Y.; Xia, L.; Wang, X. The pharmacokinetics of stachydrine in rats. J. Anhui Trad. Chin. Med. Col. 2007, 26, 48–50.
- Wen, Y.Q.; Gong, L.Y.; Wang, L.; Zhao, N.; Sun, Q.; Kamara, M.O.; Ma, H.Y.; Meng, F.H. Comparative pharmacokinetics study of leonurine and stachydrine in normal rats and rats with cold-stagnation and blood-stasis primary dysmenorrhoea after the administration of *Leonurus japonicus* Houtt electuary. J. Sep. Sci. 2019, 42, 1725–1732. [CrossRef]
- 72. Peixoto, J.S.; Comar, J.F.; Moreira, C.T.; Soares, A.A.; De Oliveira, A.L.; Bracht, A.; Peralta, R.M. Effects of *Citrus aurantium* (bitter orange) fruit extracts and *p*-synephrine on metabolic fluxes in the rat liver. *Molecules* **2012**, *17*, 5854–5869. [CrossRef]
- Arumugam, M.K.; Paal, M.C.; Donohue, T.M., Jr.; Ganesan, M.; Osna, N.A.; Kharbanda, K.K. Beneficial effects of betaine: A comprehensive review. *Biology* 2021, 10, 456. [CrossRef] [PubMed]
- Zhang, N.; Wang, M.; Li, Y.; Zhou, M.; Wu, T.; Cheng, Z. TLC-MS identification of alkaloids in Leonuri Herba and Leonuri Fructus aided by a newly developed universal derivatisation reagent optimised by the response surface method. *Phytochem. Anal.* 2021, 32, 242–251. [CrossRef] [PubMed]
- 75. Fu, M.; Zou, B.; An, K.; Yu, Y.; Tang, D.; Wu, J.; Xu, Y.; Ti, H. Anti-asthmatic activity of alkaloid compounds from Pericarpium Citri Reticulata (*Citrus reticulata* 'Chachi'). *Food Funct.* **2019**, *10*, 903–911. [CrossRef] [PubMed]
- Liu, C.; Zhang, T. Theory and Practice of Q-Marker of Traditional Chinese Medicine, 1st ed.; Science Press: Beijing, China, 2019; pp. 561–658.
- Cao, J.; Zhou, S.; Qiu, F.; Kong, W.; Wan, L.; Yang, M. A simple and fast method for the simultaneous quantification of six flavonoids in Fructus aurantii by UPLC–PDA and confirmation by UPLC/ESI-Q-TOF-MS. Anal. *Methods* 2012, 4, 4121–4128. [CrossRef]
- Nogata, Y.; Sakamoto, K.; Shiratsuchi, H.; Ishii, T.; Yano, M.; Ohta, H. Flavonoid composition of fruit tissues of citrus species. Biosci. Biotechnol. Biochem. 2006, 70, 178–192. [CrossRef]
- 79. Duan, L.; Dou, L.L.; Yu, K.Y.; Guo, L.; Bai-Zhong, C.; Li, P.; Liu, E.H. Polymethoxyflavones in peel of *Citrus reticulata* 'Chachi' and their biological activities. *Food Chem.* **2017**, *234*, 254–261. [CrossRef]
- 80. Zhao, X.; Zhu, D.; Ye, X.; Jiang, S.; Xi, Y. The study progress of the citrus flavonoids. Nat. Prod. Res. Dev. 2002, 14, 89–92.
- Li, P.; Zeng, S.L.; Duan, L.; Ma, X.D.; Dou, L.L.; Wang, L.J.; Li, P.; Bi, Z.M.; Liu, E.H. Comparison of Aurantii Fructus Immaturus and Aurantii Fructus based on multiple chromatographic analysis and chemometrics methods. *J. Chromatogr. A* 2016, 1469, 96–107. [CrossRef]
- Zeng, S.L.; Li, S.Z.; Lai, C.J.; Wei, M.Y.; Chen, B.Z.; Li, P.; Zheng, G.D.; Liu, E.H. Evaluation of anti-lipase activity and bioactive flavonoids in the Citri Reticulatae Pericarpium from different harvest time. *Phytomedicine* 2018, 43, 103–109. [CrossRef]
- 83. Day, C.R.; Kempson, S.A. Betaine chemistry, roles, and potential use in liver disease. *Biochim. Biophys. Acta* 2016, 1860, 1098–1106. [CrossRef]
- 84. Lever, M.; Sizeland, P.C.; Bason, L.M.; Hayman, C.M.; Chambers, S.T. Glycine betaine and proline betaine in human blood and urine. *Biochim. Biophys. Acta* **1994**, *1200*, 259–264. [CrossRef]

- Lashley, A.; Miller, R.; Provenzano, S.; Jarecki, S.-A.; Erba, P.; Salim, V. Functional diversification and structural origins of plant natural product methyltransferases. *Molecules* 2023, 28, 43. [CrossRef]
- Liscombe, D.K.; Louie, G.V.; Noel, J.P. Architectures, mechanisms and molecular evolution of natural product methyltransferases. Nat. Prod. Rep. 2012, 29, 1238–1250. [CrossRef]
- Liu, Y.; Fernie, A.R.; Tohge, T. Diversification of chemical structures of methoxylated flavonoids and genes encoding flavonoid-Omethyltransferases. *Plants* 2022, 11, 564. [CrossRef] [PubMed]
- Arigò, A.; Rigano, F.; Russo, M.; Trovato, E.; Dugo, P.; Mondello, L. Dietary intake of coumarins and furocoumarins through citrus beverages: A detailed estimation by a HPLC-MS/MS method combined with the linear retention index system. *Foods* 2021, 10, 1533. [CrossRef]
- 89. Mahmoud, A.M.; Hernández Bautista, R.J.; Sandhu, M.A.; Hussein, O.E. Beneficial effects of citrus flavonoids on cardiovascular and metabolic health. *Oxid. Med. Cell Longev.* **2019**, 2019, 5484138. [CrossRef]
- 90. Testai, L.; Calderone, V. Nutraceutical value of citrus flavanones and their implications in cardiovascular disease. *Nutrients* **2017**, *9*, 502. [CrossRef]
- 91. Dosoky, N.S.; Setzer, W.N. Biological activities and safety of Citrus spp. essential oils. Int. J. Mol. Sci. 2018, 19, 1966. [CrossRef]
- 92. Yang, Y.; Zhang, L.; Guo, Q.; Yang, J.; Duan, Y.; Li, F. Bioactive components of fructus Aurantii Immaturus and Fructus Aurantii and their application. *Food Drug* **2021**, *23*, 476–484.
- Zhang, H.; Zhang, R.; Pang, J.; Wu, M.; Huang, C.; Xia, Y.; Wei, E.; Luo, L. Study on the effect on rabbit isolated vaginal smooth muscle contract *in vitro* with the challenges of Fructus Aurantii Immaturus. *Acta Med. Sin.* 2007, 20, 8–9.
- 94. Yang, Y. The effects of Zhishi and Qingpi on the motivity of smooth muscle. J. Northwest Norm. Univ. Nat. Sci. 2002, 38, 114–117.
- 95. Ahangarpour, A.; Oroojan, A.A.; Amirzargar, A.; Ghanavati, M. Antispasmodic effects of *Citrus aurantium* flowers aqueous extract on uterus of non-pregnant rats. *Iran J. Reprod. Med.* **2011**, *9*, 289–294. [PubMed]
- 96. Yu, J.; Su, J.; Lv, G. Research progress in anti-cardiovascular and cerebrovascular disease activity of Citri Reticulatae Pericarpium. *Chin. Tradit. Herb. Drugs* **2016**, *47*, 3127–3132.
- 97. Zhao, J.; Hou, Y.; Wang, X.; Mo, X.; Tang, Q.; Zhao, Z.; Mao, J. Research progress on the pharmacological effects of Chinese herb Fructus Aurantii treating cardiovascular diseases. *Chin. J. Integr. Med. Cardio-/Cerebrovasc. Dis.* **2019**, *17*, 1162–1165.
- Pontifex, M.G.; Malik, M.M.A.H.; Connell, E.; Müller, M.; Vauzour, D. Citrus polyphenols in brain health and disease: Current perspectives. *Front. Neurosci.* 2021, 15, 640648. [CrossRef]
- Liu, Y.; Yang, F.; Zhao, X.; Wang, S.; Yang, Q.; Zhang, X. Crystal structure, solubility, and pharmacokinetic study on a hesperetin cocrystal with piperine as coformer. *Pharmaceutics* 2022, 14, 94. [CrossRef]
- 100. Bhia, M.; Motallebi, M.; Abadi, B.; Zarepour, A.; Pereira-Silva, M.; Saremnejad, F.; Santos, A.C.; Zarrabi, A.; Melero, A.; Jafari, S.M.; et al. Naringenin nano-delivery systems and their therapeutic applications. *Pharmaceutics* **2021**, *13*, 291. [CrossRef]
- Barreca, D.; Mandalari, G.; Calderaro, A.; Smeriglio, A.; Trombetta, D.; Felice, M.R.; Gattuso, G. Citrus flavones: An update on sources, biological functions, and health promoting properties. *Plants* 2020, *9*, 288. [CrossRef]
- Hung, W.L.; Chang, W.S.; Lu, W.C.; Wei, G.J.; Wang, Y.; Ho, C.T.; Hwang, L.S. Pharmacokinetics, bioavailability, tissue distribution and excretion of tangeretin in rat. J. Food Drug Anal. 2018, 26, 849–857. [CrossRef]
- Peng, H.W.; Cheng, F.C.; Huang, Y.T.; Chen, C.F.; Tsai, T.H. Determination of naringenin and its glucuronide conjugate in rat plasma and brain tissue by high-performance liquid chromatography. J. Chromatogr. B Biomed. Sci. Appl. 1998, 714, 369–374. [CrossRef]
- 104. Liu, L.; Lu, Y.; Bi, X.; Xu, M.; Yu, X.; Xue, R.; He, X.; Zang, W. Choline ameliorates cardiovascular damage by improving vagal activity and inhibiting the inflammatory response in spontaneously hypertensive rats. *Sci. Rep.* **2017**, *7*, 42553. [CrossRef]
- 105. Ji, Z.; Song, L.; Niu, Q. Studies on effects of 15 Chinese herbs for regulating *Qi* on in vitro aggregation of human platelet. *Chin. Tradit. Herb. Drugs* **2001**, *32*, 428–430.
- 106. Xiao, Y.; Li, L.L.; Wang, Y.Y.; Guo, J.J.; Xu, W.P.; Wang, Y.Y.; Wang, Y. Naringin administration inhibits platelet aggregation and release by reducing blood cholesterol levels and the cytosolic free calcium concentration in hyperlipidemic rabbits. *Exp. Ther. Med.* 2014, *8*, 968–972. [CrossRef]
- 107. Jin, Y.R.; Han, X.H.; Zhang, Y.H.; Lee, J.J.; Lim, Y.; Chung, J.H.; Yun, Y.P. Antiplatelet activity of hesperetin, a bioflavonoid, is mainly mediated by inhibition of PLC-gamma2 phosphorylation and cyclooxygenase-1 activity. *Atherosclerosis* 2007, 194, 144–152. [CrossRef]
- 108. Corbin, K.D.; Zeisel, S.H. Choline metabolism provides novel insights into nonalcoholic fatty liver disease and its progression. *Curr. Opin. Gastroenterol.* 2012, 28, 159–165. [CrossRef]
- 109. Wortmann, S.B.; Mayr, J.A. Choline-related-inherited metabolic diseases-a mini review. *J. Inherit. Metab. Dis.* **2019**, *42*, 237–242. [CrossRef]
- Zeisel, S.H.; da Costa, K.A. Choline: An essential nutrient for public health. Nutr. Rev. 2009, 67, 615–623, Erratum in J. Inherit. Metab. Dis. 2020, 43, 156. [CrossRef]
- Evans, J.A.; Mendonca, P.; Soliman, K.F.A. Neuroprotective effects and therapeutic potential of the citrus flavonoid hesperetin in neurodegenerative diseases. *Nutrients* 2022, 14, 2228. [CrossRef]

- 112. Nakajima, A.; Ohizumi, Y. Potential benefits of nobiletin, a citrus flavonoid, against Alzheimer's disease and Parkinson's disease. *Int. J. Mol. Sci.* **2019**, *20*, 3380. [CrossRef]
- 113. Furukawa, Y.; Okuyama, S.; Amakura, Y.; Sawamoto, A.; Nakajima, M.; Yoshimura, M.; Igase, M.; Fukuda, N.; Tamai, T.; Yoshida, T. Isolation and characterization of neuroprotective components from citrus peel and their application as functional food. *Chem. Pharm. Bull.* 2021, 69, 2–10. [CrossRef]

Disclaimer/Publisher's Note: The statements, opinions and data contained in all publications are solely those of the individual author(s) and contributor(s) and not of MDPI and/or the editor(s). MDPI and/or the editor(s) disclaim responsibility for any injury to people or property resulting from any ideas, methods, instructions or products referred to in the content.