



# **Structural Elucidation and Cytotoxic Activity of New Monoterpenoid Indoles from** *Gelsemium elegans*

Da Song <sup>1,2,†</sup>, Jia-Jun Liang <sup>1,†</sup>, Shi-Biao Pu <sup>3,†</sup>, Pan-Pan Zhang <sup>1</sup>, Yun-Lin Peng <sup>1</sup>, Xia Liu <sup>1</sup>, Ting-Ting Feng <sup>1</sup>, Xiang Pu <sup>1</sup>, Ying Zhou <sup>1</sup>, Xiong-Wei Liu <sup>1,\*</sup> and Xin Wei <sup>1,\*</sup>

- <sup>1</sup> School of Pharmacy, Guizhou University of Traditional Chinese Medicine, Guiyang 550025, China
- <sup>2</sup> School of Humanities and Management, Guizhou University of Traditional Chinese Medicine, Guiyang 550025, China
- <sup>3</sup> College of Chinese Materia Medica, Yunnan University of Chinese Medicine, Kunming 650500, China
- \* Correspondence: ashevy0819@163.com (X.-W.L.); sfweixin@163.com (X.W.)
- + These authors contributed equally to this work.

**Abstract:** Two new monoterpenoid indole alkaloids, gelselegandines F (1) and G (2), were isolated from the aerial parts of *Gelsemium elegans*. Their structures were elucidated by means of spectroscopic techniques and quantum chemical calculations. The ECD calculations were conducted at the B3LYP/6-311G(d,p) level and NMR calculations were carried out using the Gauge-Including Atomic Orbitals (GIAO) method. Structurally, the two new compounds possessed rare, cage-like, monoterpenoid indole skeletons. All isolated compounds and the total alkaloids extract were tested for cytotoxicity against four different tumor cell lines. The total alkaloids extract of *G. elegans* exhibited significant antitumor activity with IC<sub>50</sub> values ranging from 32.63 to 82.24 ug/mL. In order to discover anticancer leads from the active extraction, both new indole compounds (1–2) were then screened for cytotoxicity. Interestingly, compound **2** showed moderate cytotoxicity against K562 leukemia cells with an IC<sub>50</sub> value of 57.02 uM.

Keywords: gelselegandines F and G; cytotoxicity; leukemic cells; Gelsemium elegans

# 1. Introduction

Natural monoterpenoid indole alkaloids (MIAs) are widely distributed in many plants from the families Apocynaceae, Loganiaceae, and Rubiaceae [1–3]. Previous intensive studies of antitumor indole alkaloids have resulted in considerable groundbreaking discoveries in the past decades [4,5]. Especially due to star natural antitumor indoles entering the clinical forefront, such as vinblastine, vincristine, and vindesine, MIAs with remarkable biological activities and fascinating structures have attracted more and more attention from the pharmaceutical industry [6,7]. However, it is difficult to obtain clear NMR signals or fine single crystals from a large number of compounds, which limits the rapid determination of the stereoconfiguration of MIAs with multiple chiral centers. With the advancement of ECD and nuclear magnetic resonance (NMR) calculation methods, bioactive compound identification based on quantum chemical calculations has become the mainstream approach for the discovery of leading structures [8–11].

*Gelsemium elegans*, also known as "Gou Wen" or "Duan Chang Cao," is a famous medicine in traditional Chinese medicine (TCM) [12]. *G. elegans* was historically used as a treatment for cancer, nervous pain, and skin ulcers by the folk people of China [12,13]. As part of our continuing research on MIAs [4,14,15], two new monoterpenoid indole alkaloids, gelselegandines F (1) and G (2), were isolated from the aerial parts of *G. elegans* (Figure 1). Both isolated compounds possessed the rare, cage-like, monoterpenoid indole skeleton. For the identification of their absolute configuration, spectroscopic techniques as well as ECD at the B3LYP/6-311G(d,p) level and NMR calculations using the Gauge-Including Atomic Orbitals (GIAO) method were carried out. In addition, the total alkaloids



Citation: Song, D.; Liang, J.-J.; Pu, S.-B.; Zhang, P.-P.; Peng, Y.-L.; Liu, X.; Feng, T.-T.; Pu, X.; Zhou, Y.; Liu, X.-W.; et al. Structural Elucidation and Cytotoxic Activity of New Monoterpenoid Indoles from *Gelsemium elegans. Molecules* **2023**, *28*, 2531. https://doi.org/10.3390/ molecules28062531

Academic Editors: Cesar M. Compadre and Toshio Morikawa

Received: 3 February 2023 Revised: 5 March 2023 Accepted: 8 March 2023 Published: 10 March 2023



**Copyright:** © 2023 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (https:// creativecommons.org/licenses/by/ 4.0/). extract of *G. elegans* exhibited significant antitumor activity with  $IC_{50}$  values ranging from 32.63 to 82.24 ug/mL. Based on the antitumor effects of the total alkaloids extracts, new compounds **1** and **2** were then screened for cytotoxicity. Compound **2** showed moderate cytotoxicity against K562 leukemia cells with an  $IC_{50}$  value of 57.02 uM. This work provides a systematic approach to obtain an active compound from the total alkaloids of *G. elegans*, thereby supporting its traditional anti-cancer application.



Figure 1. Structures of compounds 1-2.

#### 2. Results and Discussion

#### 2.1. Structure Elucidation

Compound 1 displayed a positive reaction to Dragendorff's reagent. Based on its HR-ESI-MS spectra, the quasi-molecular ion peak at m/z 403.1785 [M]<sup>+</sup> (calcd for  $C_{22}H_{28}N_2O_3Cl^+$  403.1783) assigned the molecular formula as  $C_{22}H_{28}N_2O_3Cl$ . The <sup>13</sup>C and <sup>1</sup>H NMR spectroscopic data of compound 1 (Table 1) displayed 22 carbon resonances for humantenine-type alkaloids, including three methyl groups ( $\delta_C$  13.4, 48.6, and 64.3), four methylenes ( $\delta_C$  27.8, 29.4, 65.6, and 54.7), nine methines ( $\delta_C$  33.1, 34.5, 71.0, 74.1, 108.7, 124.8, 126.8, 130.0, and 130.8), and five quaternary carbons ( $\delta_C$  57.0, 130.0, 131.2, 139.5, and 174.9). The above spectroscopic data were similar to those of the compound humantenine [16], except for the remaining CH<sub>2</sub> ( $\delta_C$  69.4) and chlorine groups. Furthermore, the HMBC correlations (Figure 2) from  $\delta_H$  3.27 ( $N_4$  -CH<sub>3</sub>) and  $\delta_H$  4.40 (H-21) to  $\delta_C$  69.4 supported the linkage between the remaining -CH<sub>2</sub>Cl unit and  $N_4$ .



**Figure 2.** Selective and key HMBC ( ), <sup>1</sup>H-<sup>1</sup>H COSY ( ) and ROESY ( ) correlations of compound **1**.

The relative configuration of compound **1** was confirmed by NOE correlations. The *Z*-configured double bond of C-19/20 was established by the NOE correlation of  $\delta_{\rm H}$  2.92 (H-15) with  $\delta_{\rm H}$  6.03 (H-19) in its ROESY spectrum. In a molecular model, the rigid skeleton of a humantenine-type alkaloid required  $\beta$ -orientation for H-3 and H-15 to form the cage-like polycyclic system, which was consistent with the known analogue compound humantenine [16]. The NOE correlations of H-5/H-16/H-22 suggested the same orientation (*a*-orientation) of these protons. Meanwhile, the NOE correlation of H-9 with H-17 supported the relative configuration of C-7. Considering the solved relative configuration of compound **1**, its stereoconfiguration was assigned as two mutually enantiomers, 3*R*, 4*S*, 5*S*, 7*S*, 15*R*, 16*S* and 3*S*, 4*R*, 5*R*, 7*R*, 15*S*, 16*R*.

NO.	Compound 1 <sup>a</sup>		Compound 2 <sup>a</sup>	
	$\delta_{ m H}$	$\delta_{\rm C}$	$\delta_{ m H}$	$\delta_{\rm C}$
2		174.9		131.2
3	3.65 (d, J = 7.0, 1H)	74.1	5.29 (d, J = 10.3, 1H)	61.6
5	4.56 (m, 1H)	71.0	3.93 (m, 1H)	67.4
6	2.78 (dd, <i>J</i> = 17.3, 4.3, 1H) 2.64 (dd, <i>J</i> = 17.3, 8.2, 1H)	29.4	3.26 (dd, <i>J</i> = 18.3, 4.8, 1H) 3.91 (d, <i>J</i> = 4.6, 1H)	21.1
7		57.0		103.9
8		131.2		126.7
9	7.54 (d, <i>J</i> = 7.5, 1H)	126.8	7.51 (d, <i>J</i> = 7.9, 1H)	119.5
10	7.20 (td, <i>J</i> = 7.6, 0.9, 1H)	124.8	7.11 (m, 1H)	121.2
11	7.41 (td, <i>J</i> = 7.7, 0.9, 1H)	130.0	7.21 (m, 1H)	124.3
12	7.11 (d, <i>J</i> = 7.7, 1H)	108.7	7.40 (d, <i>J</i> = 8.2, 1H)	112.9
13		139.5		139.1
14	2.41 (ddd, <i>J</i> = 15.7, 11.5, 7.3, 1H) 2.28 (dd, <i>J</i> = 15.7, 55, 1H)	27.8	2.53 (m, 1H) 3.07 (dd, J = 13.3, 4.4, 1H)	31.3
15	2.20 (ad, $j = 10.7, 50.0, 111)2.92 (m. 1H)$	34.5	313(d I = 411H)	35.9
16	2.86 (dd, I = 10.1, 5.1, 1H)	33.1	0.10 (0,) 1.1, 111,	53.3
17	4.33 (d, J = 11.5, 1H)4.19 (dd, J = 11.5, 4.8, 1H)	65.6	3.85 (m, 2H)	67.5
18	1.86 (d, J = 6.9, 3H)	13.4	1.73 (d, <i>J</i> = 7.0, 3H)	13.0
19	6.03 (q, J = 6.9, 1H)	130.8	5.73 (qd, J = 7.0,4.5, 1H)	122.9
20	-	130.0	-	128.4
21	5.48 (d, <i>J</i> = 11.1, 1H) 4.40 (d, <i>J</i> = 14.5, 1H)	54.7	4.77 (d, <i>J</i> = 16.4, 1H), 4.60 (d, <i>J</i> = 16.2, 1H)	60.2
22	5.46 (s, 2H)	69.4	5.52 (d, <i>J</i> = 10.4, 1H) 5.40 (d, <i>J</i> = 10.4, 1H)	67.8
$N_1$ -OCH <sub>3</sub>	4.03 (s, 3H)	64.3		
$N_4$ -CH <sub>3</sub>	3.27 (s, 3H)	48.6		
COOCH <sub>3</sub> COOCH <sub>3</sub>			3.01 (s, 3H)	52.7 171.7

**Table 1.** <sup>1</sup> H (400 MHz) and <sup>13</sup> C NMR (100 MHz) data for compounds 1–2 ( $\delta$  in ppm, *J* in Hz).

<sup>a</sup> Recorded in CD<sub>3</sub>OD.

The absolute configuration of compound **1** was finally solved by quantum chemical calculation (Supporting Information). As shown in Figure 3A, the calculated ECD spectrum for compound **1** (3*R*, 4*S*, 5*S*, 7*S*, 15*R*, 16*S*) matched well with the experimental spectrum. In addition, the <sup>13</sup>C NMR calculation was further used to support its absolute configuration [17]. The results showed that the correlation coefficient (R<sup>2</sup>) from linear regression analysis between its calculated and experimental <sup>13</sup>C NMR data was 0.9980 and the corrected mean absolute deviation (CMAD) was 1.55 (Figure 3B, Table S3). Based on the above findings, compound **1** was identified as shown in Figure 1 and named gelselegandine F.

The molecular formula of compound **2** was suggested as  $C_{22}H_{26}N_2O_3Cl$  by the quasimolecular ion at m/z 401.1625 [M]<sup>+</sup> (calcd for 401.1626). In the <sup>13</sup>C and <sup>1</sup>H NMR spectra of compound **2** (Table 1), a total of two methyl groups, five methylenes, eight methines, and seven quaternary carbon resonances were discovered. Such spectral data could be specifically assigned to sarpagine-type alkaloid N(b)-methylaknammidine [18], except the  $N_4$ -methyl in N(b)-methylaknammidine was replaced by a CH<sub>2</sub> ( $\delta_C$  67.8, C-22) moiety in compound **2**. Given the remaining chlorine group in the HR-ESI-MS analysis, the other end residue of C-22 was speculated to connect with chlorine. In addition, the HMBC correlations from  $\delta_H$  5.52 (H-22) to  $\delta_C$  61.6 (C-3) and  $\delta_C$  60.2 (C-21) further supported the previous reasoning and established the planar structure of compound **2** (Figure 4).



**Figure 3.** Quantum chemical calculations of compound **1**. (**A**) Calculated and experimental ECD spectra of compound **1**; (**B**) Regression analysis of experimental versus calculated <sup>13</sup>C NMR chemical shifts of compound **1**.



**Figure 4.** Selective and key HMBC ( ), <sup>1</sup>H-<sup>1</sup>H COSY ( ) and ROESY ( ) correlations of compound **2**.

The relative configuration of compound **2** was confirmed by NOE correlations. In the ROESY spectrum of compound **2** (Figure 4), the NOE correlation of  $\delta_{\rm H}$  5.73 (H-19) with  $\delta_{\rm H}$  3.13 (H-15) suggested the *Z*-configured double bond of C-19/20. Meanwhile, NOE correlations of  $\delta_{\rm H}$  3.07 (Ha-14) with  $\delta_{\rm H}$  5.29 (H-3) and  $\delta_{\rm H}$  2.53 (Hb-14) with  $\delta_{\rm H}$  3.13 (H-15) established the  $\alpha$ -orientations of H-3 and H-15, respectively. The relative configuration of quaternary C-16 was determined by the NOE correlation of H-9 with the hydrogen signal of a methoxy group. The NOE correlation between H-5 and H-22 supported the co-face of these protons (*a*-orientation). Thus, the relative configuration of compound **2** was identified and its absolute configuration was limited to two possibilities, 3*S*, 4*S*, 5*S*, 15*S*, 16*S* and 3*R*, 4*R*, 5*R*, 15*R*, 16*R*, which were mutual enantiomers.

Finally, the absolute configuration of compound **2** was identified by means of quantum chemical calculations [17] (Supporting Information). As shown in Figure 5A, the calculated ECD spectrum for compound **2** (3*S*, 4*S*, 5*S*, 15*S*, 16*S*) matched well with the experimental spectrum. In addition, the <sup>13</sup>C NMR calculation showed that the correlation coefficient (R<sup>2</sup>) from linear regression analysis between its calculated and experimental <sup>13</sup>C NMR data was 0.9982 and the corrected mean absolute deviation (CMAD) was 1.46 (Figure 5B, Table S3). Therefore, the structure of compound **2** was established and the new compound was named gelselegandine G.



**Figure 5.** Quantum chemical calculations of compound **2**. (**A**) Calculated and experimental ECD spectra of compound **2**; (**B**) Regression analysis of experimental versus calculated <sup>13</sup>C NMR chemical shifts of compound **2**.

# 2.2. The Cytotoxicity of Total Alkaloids and Compound 2

The inhibitory activity for the total alkaloids of *G. elegans* against four tumor cell lines (A549, Hela, K562, and PC-3) was preliminarily evaluated at a concentration of 160 ug/mL. Based on the results of MTT assay, the total alkaloids exhibited significant cytotoxicity towards the all Hela, K562, PC-3, and A549 cell lines (Figure 6). Under the treated concentration, the cell viability means of the four cell lines were lower than 30% (Figure 6). Especially for Hela cervical cancer cells and K562 leukemia cells, the total alkaloids of *G. elegans* showed the best inhibitory activity among of the four tested cell lines. Furthermore, the IC<sub>50</sub> values of the total alkaloids were determined using MTT method. As shown in Figure 7, the IC<sub>50</sub> values of the total alkaloids against K562, A549, Hela, and PC-3 cell lines were 49.07, 63.98, 32.63, and 82.24 ug/mL, respectively.

Based on the above antitumor clues of the total alkaloids, both new indole compounds **1** and **2** were then screened for cytotoxicity against A549, Hela, K562, and PC-3 cell lines. However, only compound **2** showed moderate cytotoxicity against K562 leukemia cells with an IC<sub>50</sub> value of 57.02 uM (Figure 7), which was consistent with the better cytotoxicity of the total alkaloids toward Hela and K562 cells. Meaningful cytotoxicity against the other cell lines was not detected (IC<sub>50</sub> > 100 uM). This study reports antitumor activity tracking from the total alkaloids extract to new monomer compounds, which might provide a new type of lead for the inhibition of K562 leukemia cells.



**Figure 6.** The cell viability of A549, Hela, K562, and PC-3 cells treated with the total alkaloids of *G. elegans* at a concentration of 160 ug/mL.



Figure 7. The IC<sub>50</sub> values of the total alkaloids, compound 2, and positive control drug cisplatin.

## 3. Experimental Section

3.1. General Experimental Procedures

Optical rotations were measured using an Autopol VI (serial #91058). IR spectra were determined using a Bruker Vertex 70 instrument with KBr pellets. HR-ESIMS data were obtained using a SHIMADZU UPLC-IT-TOF. UV spectra were measured using a SHIMADZU UV-2401PC. The 1D and 2D NMR spectra were measured on a Bruker Avance NEO (400 MHz). TLC analyses were carried out on precoated silica gel GF-254 plates and column chromatography was performed on 200–300 mesh silica gel (Qingdao Marine Chemical Plant, Qingdao, China), MCI-gel (Mitsubishi Chemical Co., Ltd., Tokyo, Japan), and ODS-gel (50  $\mu$ m, YMC, Kyoto, Japan). HPLC was carried out SEP LC-52 using an MWD UV detector (Separation Technology Co., Ltd., Beijing, China) and semi-preparative C18 columns (250  $\times$  10 mm).

#### 3.2. Plant Material

The aerial parts of *G. elegans* were purchased from Kunming Zhifen Biotechnology Co., Ltd. (Kunming, China) in April 2021, and identified by An-Rui Lou from Kunming Zhifen Biotechnology Co., Ltd. A voucher specimen (No. WX\_20210401) was deposited in Guizhou University of Traditional Chinese Medicine.

#### 3.3. Extraction and Isolation

The aerial parts of *G. elegans* (20 kg) were extracted with 95% ethanol (30 L × 3) under reflux conditions at 70 °C, for two hours each time. After removal of the organic solvent under reduced pressure, the crude extract (3893 g) was obtained. The ethanol extract was dissolved in H<sub>2</sub>O and acidified with dilute acid water to pH 2, then basified with NH<sub>3</sub>.H<sub>2</sub>O to pH 10 and extracted with CH<sub>2</sub>Cl<sub>2</sub> to obtain the total alkaloids (221 g). The total alkaloids were separated by flash silica gel column chromatography using a MeOH/CH<sub>2</sub>Cl<sub>2</sub> (1:100–1:0, *v:v*) gradient to give 25 fractions (L1–L25). Then, L-9 and L-10 were merged together and separated by ODS column chromatography using a MeOH/H<sub>2</sub>O (2:3–1:0, *v:v*) gradient to give 15 sub-fractions. Sub-fraction 15 was successively separated by MCI column chromatography using a MeOH/H<sub>2</sub>O (1:1–1:0, *v:v*) gradient and by HPLC preparative chromatographic isocratic elution with MeOH/H<sub>2</sub>O (3:7, *v:v*) to give compounds **1** (20.0 mg) and **2** (47.0 mg).

# 3.3.1. Gelselegandine F (1)

Amorphous solid;  $[\alpha]^{18}_{D}$  -102.1 (*c* 0.06, MeOH); UV (MeOH)  $\lambda_{max}$  207, 254, 279 nm; IR (KBr)  $\nu_{max}$  3432, 2926, 1718, 1619, 1384, and 1077 cm<sup>-1</sup>; HRESIMS *m*/*z* 403.1785 [M]<sup>+</sup> (calcd for C<sub>22</sub>H<sub>28</sub>N<sub>2</sub>O<sub>3</sub>Cl<sup>+</sup> 403.1783); <sup>1</sup>H and <sup>13</sup>C NMR data, see Table 1.

#### 3.3.2. Gelselegandine G (2)

Amorphous solid;  $[\alpha]^{18}_{D}$  -18.1 (*c* 0.07, MeOH); UV (MeOH)  $\lambda_{max}$  219, 269, 278, 285 nm; IR (KBr)  $\nu_{max}$  3430, 2926, 1729, 1623, 1457, 1234, and 1074 cm<sup>-1</sup>; HRESIMS *m*/*z* 401.1625 [M]<sup>+</sup> (calcd for C<sub>22</sub>H<sub>26</sub>N<sub>2</sub>O<sub>3</sub>Cl<sup>+</sup> 401.1626); <sup>1</sup>H and <sup>13</sup>C NMR data, see Table 1.

#### 3.4. ECD Calculation

The ECD calculations of compounds **1** and **2** were carried out using Gaussian 09 (Supporting Information). At first, all conformers were optimized at PM6. Room temperature equilibrium populations were calculated according to the Boltzmann distribution law (Supporting Information), based on which dominant conformers of the population (over 1%) were kept. The chosen conformers were further optimized at B3LYP/6-31G(d,p) in gas phase. Vibrational frequency analysis confirmed the stable structures. ECD calculations were conducted at the B3LYP/6-311G(d,p) level in methanol with the IEFPCM model using time-dependent density functional theory (TD-DFT). The ECD spectrum was simulated using the ECD/UV analysis tool by overlapping Gaussian functions for each transition (Supporting Information). The spectra of the enantiomers were produced directly by mirror inversion about the horizontal axis [17].

# 3.5. <sup>13</sup>C NMR Calculation

The conformers of compounds **1** and **2** were directly derived from the previous ECD calculations. NMR calculations were carried out using the Gauge-Including Atomic Orbitals (GIAO) method at the mPW1PW91/6-311+G(2d,p) level in methanol simulated by the IEFPCM model (Supporting Information). The TMS-corrected NMR chemical shift values were averaged according to Boltzmann distribution and fitted to the experimental values by linear regression. The calculated <sup>13</sup>C-NMR chemical shift value of TMS in methanol was 187.37 ppm [17].

## 3.6. Cell Lines and Cell Culture

All cell lines were purchased from the Chinese Academy of Sciences, Kunming Cell Bank. All cells were cultured in RPMI-1640 or DMEM media (Gibco, Beijing, China) supplemented with 10% fetal bovine serum, 1% glutamine, 100 U/mL penicillin, and 100 mg/mL streptomycin in a humidified atmosphere with 5% CO<sub>2</sub> at 37 °C. The compounds were stored at -20 °C after being dissolved in DMSO. Cisplatin was purchased from Aladdin Company (Shanghai, China).

## 3.7. Cell Viability Assay

Cells in the logarithmic growth phase were washed with PBS and the cell density was regulated as  $5 \times 10^3$  cells/mL. The cells (80 µL/well) were inoculated in 96-well plates and incubated at 37 °C under 5% CO<sub>2</sub>. The cells were cultured with total alkaloids (1.25, 2.5, 5, 10, 20, 40, 80, 160, 320, and 640 µg/mL), compounds **1** and **2**, and positive control cisplatin (100, 50, 25, 12.5, and 6.25 µM, respectively, for monomer compounds) in medium for 48 h. Each group had equal five wells and five blank wells without cells were set at the same time. Then, 10 µL of MTT (Solarbio, Beijing Solarbio Science & Technology Co., Ltd., Beijing, China) was added to each well and the cells were incubated for 4 h at 37 °C under 5% CO<sub>2</sub>. The absorbance of each well was measured at 490 nm using an automatic microplate reader (Thermo Fisher Scientific 3020, Waltham, MA, USA). Cell viability % = (OD value of detection well – background OD value)/(OD value of blank control group – background OD value) × 100%.

# 4. Conclusions

In conclusion, as part of our continuing study on MIAs from traditional Chinese medicine, two new monoterpenoid indole alkaloids, gelselegandines F (1) and G (2), were isolated from the aerial parts of *G. elegans*. For their absolute configuration identification, ECD and NMR calculations were used together with spectroscopic techniques. The cell viability assay results exhibited significant and broad antitumor effects of the total alkaloids from *G. elegans*, and the IC<sub>50</sub> values of the total alkaloids against K562, A549, Hela, and PC-3 cell lines were 49.07, 63.98, 32.63, and 82.24 ug/mL, respectively. Based on the antitumor clues provided by the total alkaloids, both new indole compounds 1 and 2 were then screened for cytotoxicity against the above four cell lines. Compound 2 showed moderate cytotoxic activity tracking from a total alkaloids extract to new monomer compounds, thus shedding light on the antitumor ingredients of *G. elegans*.

**Supplementary Materials:** The following supporting information can be downloaded at: https:// www.mdpi.com/article/10.3390/molecules28062531/s1. Parts of calculations and experiments, 1D and 2D NMR spectra, HRESIMS, and UV and ECD spectra of compounds 1 and 2 are available as Supporting Information. The Figures S1–S27 and Tables S1–S6 were listed in Supporting Information [19,20].

**Author Contributions:** X.W. and X.-W.L. designed the project and critically revised the manuscript. P.-P.Z., Y.-L.P. and J.-J.L. performed the isolation and structure elucidation. D.S., S.-B.P., T.-T.F. and X.W. drafted the manuscript. D.S., X.L. and X.-W.L. carried out the biological activity assays and statistical analysis. D.S. and X.W. recorded and tested the quantum chemical calculations; X.W., Y.Z. and X.P. reviewed and edited the manuscript. All authors have read and agreed to the published version of the manuscript.

**Funding:** The authors are grateful to the National Natural Science Foundation of China (Grant Nos. 32000276 and 82260874), the Science and Technology Plan Project of Guizhou Province (Qian Ke He Ji Chu-ZK [2023]Yi Ban 430), and Doctoral Initiation Funding of Guizhou University of Traditional Chinese Medicine (2019-017) for financial support.

Data Availability Statement: The research data were available in Supporting Information.

**Acknowledgments:** The authors sincerely thank the Yinfo Cloud Computing Platform for help with the quantum chemical calculations.

Conflicts of Interest: The authors declare no conflict of interest.

Sample Availability: Samples of the compounds are not available from the authors.

#### References

- Zhan, G.; Miao, R.; Zhang, F.; Hao, X.; Zheng, X.; Zhang, H.; Zhang, X.; Guo, Z. Monoterpene indole alkaloids with diverse skeletons from the stems of *rauvolfia vomitoria* and their acetylcholinesterase inhibitory activities. *Phytochemistry* 2020, 177, 112450. [CrossRef] [PubMed]
- Li, F.R.; Liu, L.; Liu, Y.P.; Wang, J.T.; Yang, M.L.; Khan, A.; Qin, X.J.; Wang, Y.D.; Cheng, G.G. HRESIMS-guided isolation of aspidosperma-scandine type bisindole alkaloids from *Melodinus cochinchinensis* and their anti-inflammatory and cytotoxic activities. *Phytochemistry* 2021, 184, 112–673. [CrossRef] [PubMed]
- Thamm, A.M.K.; Qu, Y.; De Luca, V. Discovery and metabolic engineering of ir-idoid/secoiridoid and monoterpenoid indole alkaloid biosynthesis. *Phytochem. Rev.* 2016, 15, 339–361. [CrossRef]
- Wei, X.; Dai, Z.; Yang, J.; Khan, A.; Yu, H.F.; Zhao, Y.L.; Wang, Y.F.; Liu, Y.P.; Yang, Z.F.; Huang, W.Y.; et al. Unprecedented sugar bridged bisindoles selective inhibiting glioma stem cells. *Bioorg. Med. Chem.* 2018, 76, 1776–1783. [CrossRef] [PubMed]
- Liu, Y.P.; Zhao, Y.L.; Feng, T.; Cheng, G.G.; Zhang, B.H.; Li, Y.; Cai, X.H.; Luo, X.D. Melosuavines A-H, Cytotoxic Bisindole Alkaloid Derivatives from *Melodinus suaveolens. J. Nat. Prod.* 2013, 76, 2322–2329. [CrossRef] [PubMed]
- 6. Beni, Z.; Hada, V.; Dubrovay, Z.; Szantay, C.J. Structure elucidation of indole-indoline type alkaloids: A retrospective account from the point of view of current NMR and MS technology. *J. Pharm. Biomed. Anal.* **2012**, *69*, 106–124. [CrossRef] [PubMed]
- 7. Wei, X.; Yang, J.; Dai, Z.; Yu, H.F.; Ding, C.F.; Khan, A.; Zhao, Y.L.; Liu, Y.P.; Luo, X.D. Antitumor pyridine alkaloids hybrid with diverse units from *Alangium chinense*. *Tetrahedron Lett.* **2020**, *61*, 151502. [CrossRef]

- Li, Q.; Yang, S.; Teng, H.; Li, X.; Xie, W.; Wu, Z.; Yang, G.; Xu, J.; Chen, Y. Structural elucidation of two intricate polycyclic polyprenylated acylphloroglucinols using quantum chemical calculations and their hypoglycemic activities. *Arab. J. Chem.* 2022, 15, 104137. [CrossRef]
- 9. Lodewyk, M.W.; Siebert, M.R.; Tantillo, D.J. Computational prediction of <sup>1</sup>H and <sup>13</sup>C chemical shifts: A useful tool for natural product, mechanistic, and synthetic organic chemistry. *Chem. Rev.* **2012**, *112*, 1839–1862. [CrossRef] [PubMed]
- Tsui, K.Y.; Tombari, R.J.; Olson, D.E.; Tantillo, D.J. Reconsidering the structure of serlyticin-A. J. Nat. Prod. 2019, 82, 3464–3468. [CrossRef] [PubMed]
- Unzueta, P.A.; Greenwell, C.S.; Beran, G.J.O. Predicting density functional theory-quality nuclear magnetic resonance chemical shifts via Δ-machine learning. J. Chem. Theory Comput. 2021, 17, 826–840. [CrossRef] [PubMed]
- Jin, G.L.; Su, Y.P.; Liu, M.; Xu, Y.; Yang, J.; Liao, K.J.; Yu, C.X. Medicinal plants of the genus *Gelsemium* (*Gelsemiaceae, Gentianales*)—A review of their phytochemistry, pharmacology, toxicology and traditional use. J. Ethnopharmacol. 2014, 152, 33–52. [CrossRef] [PubMed]
- 13. Zhang, W.; Zhang, S.Y.; Wang, G.Y.; Li, N.P.; Chen, M.F.; Gu, J.H.; Zhang, D.M.; Wang, L.; Ye, W.C. Five new koumine-type alkaloids from the roots of *Gelsemium elegans*. *Fitoterapia* **2017**, *118*, 112–117. [CrossRef] [PubMed]
- Wei, X.; Yang, J.; Ma, H.X.; Ding, C.F.; Yu, H.F.; Zhao, Y.L.; Liu, Y.P.; Khan, A.; Wang, Y.F.; Yang, Z.F.; et al. Antimicrobial indole alkaloids with adductive C9 aromatic unit from *Gelsemium elegans*. *Tetrahedron Lett.* 2018, 59, 2066–2070. [CrossRef]
- 15. Song, D.; Hu, X.Y.; Liang, J.J.; Liu, X.; Pu, X.; Zhang, L.Y.; Zhou, Y.; Wei, X. Chemical constituents with osteoclasts inhibitory activity from *Gelsemium elegans*. Chem. Nat. Compd. 2022, 58, 962–966. [CrossRef]
- Ouyang, S.; Wang, L.; Zhang, Q.W.; Wang, G.C.; Wang, Y.; Huang, X.J.; Zhang, X.Q.; Jiang, R.W.; Yao, X.S.; Che, C.T.; et al. Six new monoterpenoid indole alkaloids from the aerial part of *Gelsemium elegans*. *Tetrahedron* 2011, 67, 4807–4813. [CrossRef]
- 17. Wei, X.; Guo, R.; Wang, X.; Liang, J.J.; Yu, H.F.; Ding, C.F.; Feng, T.T.; Zhang, L.Y.; Liu, X.; Hu, X.Y.; et al. New monoterpenoid indoles with osteoclast activities from *Gelsemium elegans*. *Molecules* **2021**, *26*, 7457. [CrossRef] [PubMed]
- 18. Hu, W.L.; Zhu, J.P.; Hesse, M. Indole alkaloids from Aistonia angustifolia. Planta Med. 1989, 55, 463–466. [CrossRef] [PubMed]
- 19. O'Boyle, N.M.; Veremeersch, T.; Flynn, C.J.; Maguire, A.R.; Hutchison, G.R. Confab—Systematic generation of diverse low-energy conformers. *J. Cheminformatics* **2011**, *3*, 3–8. [CrossRef]
- 20. Frisch, M.J.; Trucks, G.W.; Schlegel, H.B.; Scuseria, G.E.; Robb, M.A.; Cheeseman, J.R.; Scalmani, G.; Barone, V.; Mennucci, B.; Petersson, G.A.; et al. *Gaussian 09 Revision D.01*; Gaussian Inc.: Wallingford, UK, 2009.

**Disclaimer/Publisher's Note:** The statements, opinions and data contained in all publications are solely those of the individual author(s) and contributor(s) and not of MDPI and/or the editor(s). MDPI and/or the editor(s) disclaim responsibility for any injury to people or property resulting from any ideas, methods, instructions or products referred to in the content.