

SUPPORTING INFORMATION

Total Synthesis of Hemerocallisamine I Paved by Gram Scale Synthesis of (2*S*,4*S*)-4-Hydroxyglutamic Acid Lactone

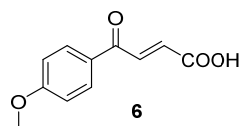
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1. Materials and methods

Unless otherwise noted, all chemicals were purchased from commercial sources and used without further purifications. Column chromatography was carried out using silica 60 A, Davisil, purchased from Fisher Chemicals. Reactions were monitored by thin layer chromatography (TLC), using Macherey-Nagel's pre-coated TLC sheets POLYGRAM SIL G/UV254, which were visualized under UV light (254 nm) or by staining with aqueous basic potassium permanganate or cerium molybdate solutions as appropriate. HPLC analyses were performed on a Varian system using a Macherey-Nagel EC 250/4 Nucleodur Phenyl-Hexyl 5 μ m, CHIRAL ART, Amylose-SA, 250 x 4.6 mm, 5 μ m and Astec CHIROBIOTIC ®T , 250 x 4.6 mm, 5 μ m column. All ^1H and ^{13}C NMR spectra were recorded using Bruker Avance NEO 400 MHz and/or Varian 400 MR spectrometers. Chemical shifts (δ) are given in parts per million (ppm). The ^1H NMR chemical shift scale is referenced to the TMS internal standard (δ = 0 ppm) or solvent residual peak (δ = 2.50 ppm for $\text{DMSO-}d_6$ and δ = 7.26 ppm for CDCl_3). The ^{13}C NMR chemical shift scale is referenced to the solvent residual peak (δ = 39.52 ppm for $\text{DMSO-}d_6$ and δ = 77.16 ppm for CDCl_3). Coupling constants (J) are given in hertz (Hz). The multiplicity of ^1H NMR signals is reported as follows: s = singlet, d = doublet, t = triplet, q = quartet, m = multiplet, bs = broad singlet, "t" for dd with two identical or similar coupling constants, "dt" or "td" for ddd with two identical or similar coupling constants and "q" for ddd with three identical or similar coupling constants. High-resolution mass spectra were measured using Thermo Scientific mass spectrometer with Orbitrap analyzer and HESI ionization.

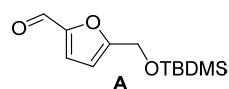
2. Synthesis and characterization of compounds

(*E*)-4-(4-methoxyphenyl)-4-oxobut-2-enoic acid (**6**)



Anisole (20.0 g, 0.185 mol) was dissolved in CH_2Cl_2 (111 ml), and then maleic anhydride (27.2 g, 0.277 mol, 1.5 equiv) and AlCl_3 (37.0 g, 0.277 mol, 1.5 equiv) were added. The reaction mixture was stirred for 20 h at rt. After completion, the reaction mixture was quenched with H_2O (100 ml) and CH_2Cl_2 was evaporated under reduced pressure. The residue was diluted with H_2O (150 ml) and extracted with EtOAc (3 x 150 ml). The collected organic layers were washed with saturated brine (200 ml), dried over anhydrous Na_2SO_4 , and concentrated *in vacuo*, yielding a crude product that was crystallized from EtOAc (120 ml). Acid **6** (21.0 g, 0.102 mol, 55%) was isolated as a yellow solid. The ^1H NMR spectrum of **6** was consistent with the published data.¹

6-(((*tert*-Butyldimethylsilyl)oxy)methyl)-6-hydroxy-2H-pyran-3(6H)-one (**4**)

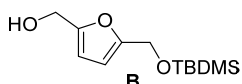


Step 1: Imidazole (5.94 g, 87.2 mmol, 1.1 equiv) and 5-hydroxymethylfurfural (10.0 g, 79.3 mmol) were stirred in CH_2Cl_2 (198 ml) for 15 minutes at rt. TBDMSCl (12.5 g, 83.3 mmol, 1.05 equiv) was added afterwards and the reaction mixture was stirred at rt, under argon, while being monitored by TLC. After 16 h H_2O (200 ml) was added, the organic phase was separated, and the aqueous phase was extracted with CH_2Cl_2 (2 x 150 ml). The combined organic solutions were dried over anhydrous Na_2SO_4 and concentrated *in vacuo*, providing carbaldehyde **A** (18.3 g, 76.1 mmol, 96%) as an orange oil.

Rf 0.77 (EtOAc:Hex 1:5)

^1H NMR (300 MHz, CDCl_3) : δ 9.59 (s, 1H), 7.20 (d, J = 3.6 Hz, 1H), 6.47 (dt, J = 3.5, 0.8 Hz, 1H), 4.73 (d, J = 0.5 Hz, 2H), 0.92 (s, 9H), 0.10 (s, 6H).

^1H -NMR spectrum of **A** was consistent with the published data.²

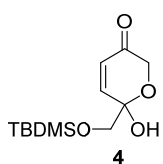


Step 2: NaBH_4 (5.76 g, 152 mmol, 2.0 equiv) was added portionwise to a solution of carbaldehyde **A** (18.3 g, 76.1 mmol) in MeOH (400 ml) at 0 °C, and the reaction mixture was stirred at this temperature for 45 min. Then H_2O (200 ml) was added and MeOH was removed *in vacuo*. The residue was diluted with H_2O (200 ml) and extracted with EtOAc (250 and 2 x 200ml). The combined organic layers were dried over anhydrous Na_2SO_4 and concentrated under reduced pressure. The crude product was purified by flash column chromatography (EtOAc:Hex, 1:1 to 3:1) to give **B** (15.0 g, 61.9 mmol, 81%) as a pale-yellow oil.

Rf 0.44 (EtOAc:Hex 1:4)

^1H NMR (300 MHz, CDCl_3) : δ = 6.24 – 6.20 (m, 1H), 6.17 (d, J = 3.1 Hz, 1H), 4.62 (s, 2H), 4.59 (s, 2H), 1.78 (s, 1H), 0.90 (s, 9H), 0.08 (s, 6H).

^1H -NMR spectrum of **B** was consistent with the published data.³



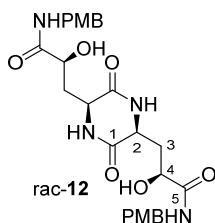
Step 3: *m*-CPBA (18.0 g, 80.5 mmol 1.3 equiv) was added to a solution of **B** (15 g, 61.8 mmol) in CH_2Cl_2 (495 ml) at 0 °C and the mixture was allowed to stir at this temperature for 0.5 h. Afterwards the mixture was let to reach rt and was stirred for additional 4 h while being monitored by TLC. The reaction was quenched with a saturated aqueous Na_2SO_3 solution (150 ml), followed by neutralisation to pH 7–8 with 1M aqueous NaOH. The mixture was extracted with CH_2Cl_2 (2 x 80 mL), the combined organic extracts were washed with brine and dried over anhydrous Na_2SO_4 to give dihydropyranone **4** (13.4 g, 51.9 mmol, 83%) as an off-white solid.

Mp 83 – 84 °C; Rf 0.55 (EtOAc:Hex 1:5)

^1H NMR (400 MHz, CDCl_3) : δ = 6.79 (d, J = 10.4 Hz, 1H), 6.15 (d, J = 10.4 Hz, 1H), 4.59 (d, J = 16.9 Hz, 1H), 4.14 (d, J = 17.0 Hz, 1H), 3.77 (d, J = 10.2 Hz, 1H), 3.68 (d, J = 10.2 Hz, 1H), 0.93 (s, 9H), 0.12 (s, 6H).

^1H -NMR spectrum of **4** was consistent with the published data.³

Isolation of the diketopiperazine byproduct rac-12



Amine rac-**11** (153.0 mg, 0.579 mmol) was dissolved in dry CH_2Cl_2 (2.9 ml) and dihydropyranone **4** (149.6 mg, 0.579 mmol, 1.0 equiv) was added. The reaction mixture was stirred at rt under argon. After 14 h, the reaction mixture was concentrated *in vacuo* and the crude product was treated with a mixture of MeOH and Et_2O (1:1, 2 ml). The precipitate was filtered off and washed with a small amount of MeOH and Et_2O , yielding rac-**12** as a white solid (13.0 mg, 0.025 mmol, 9% yield with respect to the reaction stoichiometry rac-**11**/rac-**12** 2:1).

^1H NMR (400 MHz, $\text{DMSO}-d_6$) : δ = 8.32 – 8.25 (m, 2H, 2xPMB-NH), 8.23 (m, 2H, 2xNH), 7.18 (d, J = 8.7 Hz, 4H, 2xPMB-Ar), 6.86 (d, J = 8.5 Hz, 4H, 2xPMB-Ar), 5.84 (d, J = 5.5 Hz, 1H, OH), 5.79 (d, J = 5.5 Hz, 1H, OH),

4.22 (d, $J = 6.2$ Hz, 4H, 2xPMB-CH₂), 4.19 – 4.12 (m, 2H, 2xH-4), 4.01 – 3.93 (m, 2H, 2xH-2), 3.72 (s, 6H, 2xPMB-Me), 2.05 – 1.83 (m, 4H, 2xH-3)

¹H NMR was assigned on the basis of 2D NMR, the numbering is specified in the figure.

¹³C NMR (101 MHz, DMSO-*d*₆): $\delta = 173.28$ (C-5), 168.51 (C-1), 158.16 (PMB-Ar), 131.54 (PMB-Ar), 128.53 (PMB-Ar), 113.63 (PMB-Ar), 68.00 (C-4), 55.03 (PMB-Me), 51.58 (C-2), 41.23 (PMB-CH₂), 37.56 (C-3).

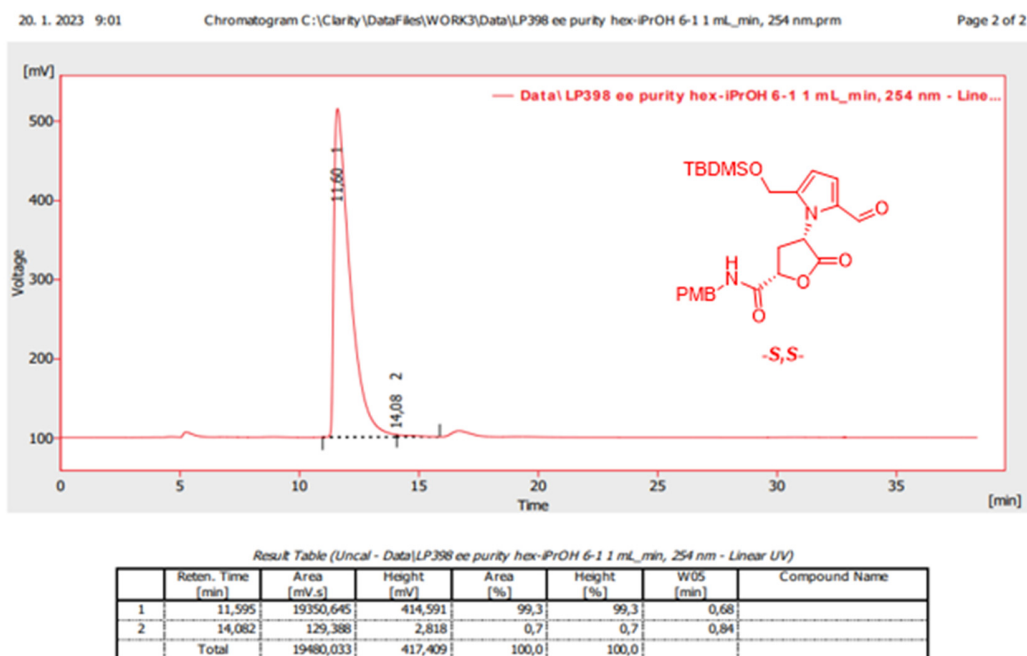
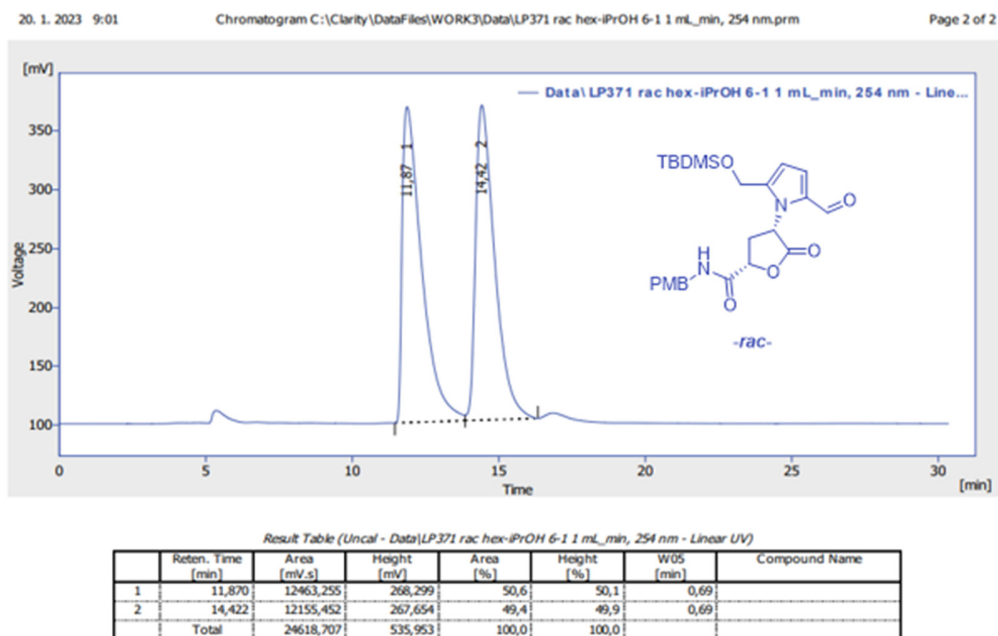
HRMS–HESI (m/z): calcd for C₂₆H₃₂N₄O₈Na [M+Na]⁺, 551.21124, found 551.21137.

3. HPLC data for compounds (S,S)-8 and (S,S)-1

HPLC conditions: CHIRAL ART, Amylose-SA, 250 x 4.6 mm, 5 µm.

Mobile Phase: Hexane:Propan-2-ol 6:1, λ = 254 nm, flow 1 ml/min

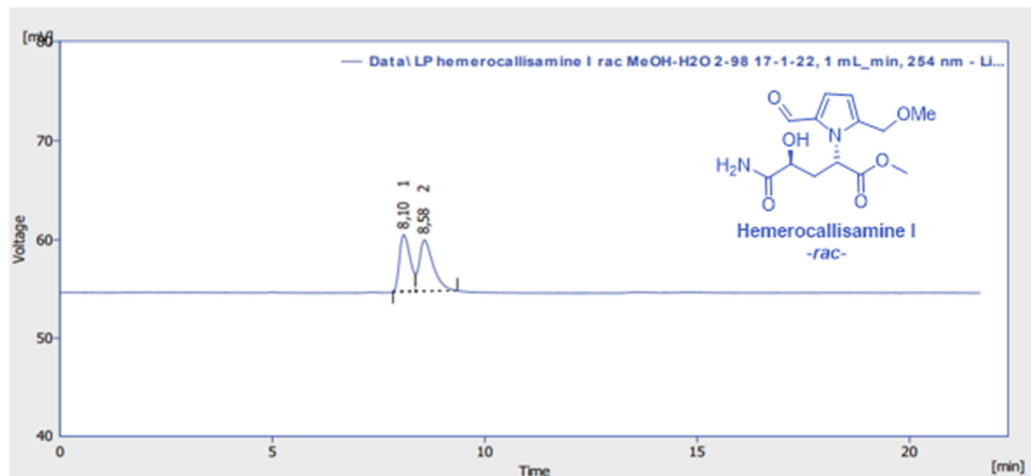
er > 99:1



HPLC conditions: Astec CHIROBIOTIC ®T, 250 x 4.6 mm, 5 µm.
 Mobile Phase: Methanol:Water 2:98, λ = 254 nm, flow 1 ml/min
 er > 99:1

20. 1. 2023 8:55 Chromatogram C:\Clarity\DataFiles\WORK3\Data\LP hemerocallisamine I rac MeOH-H2O 2-98 17-1-22, 1 mL_min, 254 nm.prm

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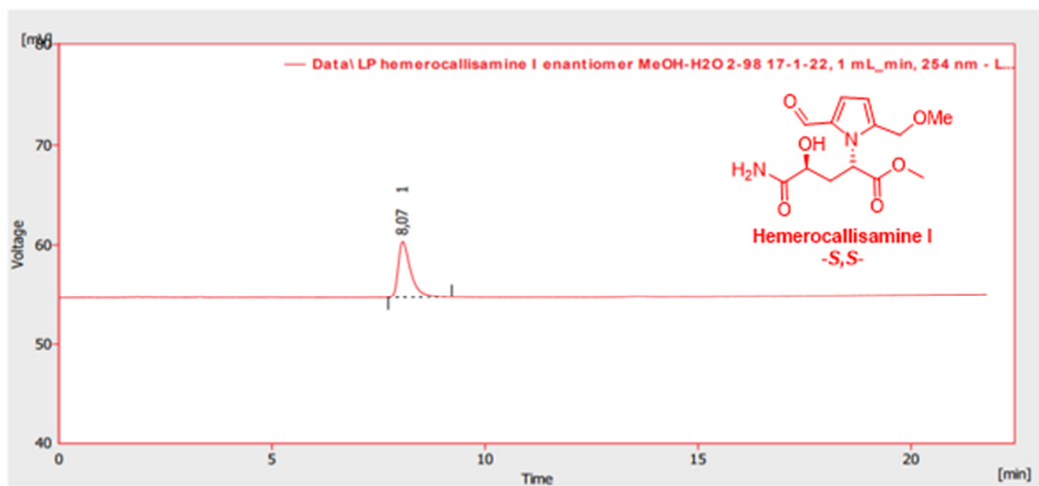


Result Table (Uncal - Data\LP hemerocallisamine I rac MeOH-H2O 2-98 17-1-22, 1 mL_min, 254 nm - Linear UV)

	Reten. Time [min]	Area [mV.s]	Height [mV]	Area [%]	Height [%]	W05 [min]	Compound Name
1	8,095	99,625	5,712	45,5	52,6	0,29	
2	8,578	119,320	5,149	54,5	47,4	0,35	
Total		218,945	10,861	100,0	100,0		

20. 1. 2023 8:55 Chromatogram C:\Clarity\DataFiles\WORK3\Data\LP hemerocallisamine I enantiomer MeOH-H2O 2-98 17-1-22, 1 mL_min, 254 nm.prm

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Result Table (Uncal - Data\LP hemerocallisamine I enantiomer MeOH-H2O 2-98 17-1-22, 1 mL_min, 254 nm - Linear UV)

	Reten. Time [min]	Area [mV.s]	Height [mV]	Area [%]	Height [%]	W05 [min]	Compound Name
1	8,070	114,480	5,611	100,0	100,0	0,30	
Total		114,480	5,611	100,0	100,0		

4. Optimization of the Maillard reaction



General procedure for ^1H -NMR experiment:

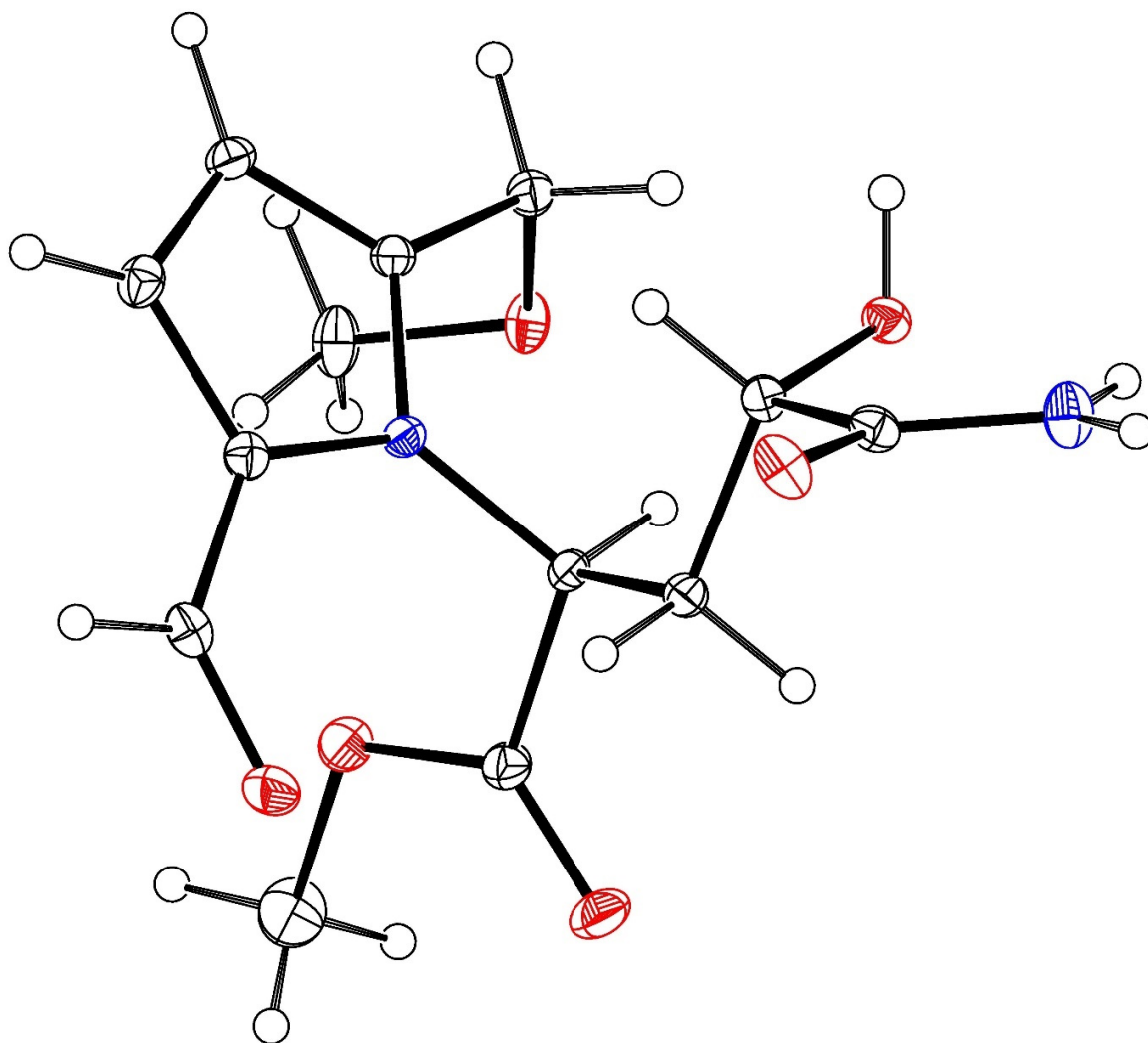
Reagents **rac-11** (9.0 mg, 0.034 mmol) and **4** (8.8 mg, 0.034 mmol, 1.0 equiv) were dissolved in the specified solvents (see the Table). The resulting reaction mixtures were stirred under the specified conditions for 14 h. After completion, the solvent was evaporated *in vacuo*. The reactions yields were determined by ^1H -NMR with the internal standard 1,1,2-trichloroethane (4.5 mg, 0.034 mmol, 3 μl , 1.0 equiv), in CDCl_3 . In selected experiments (A, Q, R, Y, Z), yields of the by-product **rac-12** were determined by ^1H -NMR in $\text{DMSO-}d_6$, with 1,1,2-trichloroethane as the internal standard.

Experiment	A equiv	B equiv	Reaction conditions	P H-NMR – Yield (CDCl ₃)	BP H-NMR – Yield*(DMSO- <i>d</i> ₆)
A	1	1	dry CH ₂ Cl ₂ (0.2M), Ar, rt, 14 h	33%	16%
B	1.1	1	dry CH ₂ Cl ₂ (0.2M), Ar, rt, 14 h	37%	-
C	1	1	dry CH ₂ Cl ₂ (0.2M), Ar, 35 °C, 14 h	41%	-
D	1	1	CH ₂ Cl ₂ (0.2M), rt, 14 h	36%	-
E	1	1	TEA (1 equiv), dry CH ₂ Cl ₂ (0.2M), Ar, rt, 14 h	0%	-
F	1	1	dry CH ₂ Cl ₂ (0.2M), MS (3A), Ar, rt, 14 h	36%	-
G	1	1	dry CH ₂ Cl ₂ (0.2M), Na ₂ SO ₄ , Ar, rt, 14 h	35%	-
H	1	1	DCE (0.2M), Ar, 55 °C, 14 h	43%	-
I	1	1	THF/H ₂ O (0.2M), 55 °C, 14 h	20%	-
J	1	1	CH ₂ Cl ₂ (0.2M), AcOH (cat.), rt, 14 h	32%	-
K	1	1	THF/H ₂ O (0.2M), rt, 14 h	21%	-
L	1	1	THF/H ₂ O (0.2M), AcOH (cat.), rt, 14 h	19%	-
M	1	1	dry MeCN (0.2 M), Ar, rt, 14 h	23%	-
N	1	1	dry MeCN (0.2 M), Ar, 60 °C, 14 h	30%	-
O	1	1	dry THF (0.2 M), Ar, rt, 14 h	17%	-
P	1	1	dry THF (0.2 M), Ar, 60 °C, 14 h	22%	-
Q	1	1	dry toluene (0.2M), Ar, rt, 14 h	36%	25%
R	1	1	dry toluene (0.2M), Ar, 70 °C, 14 h	47%	23%
S	1	1	dry toluene (0.2M), Ar, 90 °C, 14 h	47%	-
T	1	1	dry toluene (0.2M), Ar, 105 °C, 14 h	37%	-
U	1	1	DCE (0.2M), pTsOH (0.88 eq), Ar, rt, 14 h	37%	-
V	1	1	DCE (0.2M), pTsOH (0.75 eq), 55°C, 14 h	23%	-
X	1 (gradual addition over 30 min)	1	dry CH ₂ Cl ₂ (0.08M), Ar, 35°C, 14 h	42%	-
Y	1	1	dry CH ₂ Cl ₂ (0.02 M), rt, 14 h	30%	35%
Z	1	2	dry CH ₂ Cl ₂ (0.02 M), rt, 14 h	37%	30%

* yields with respect to the reaction stoichiometry rac-**11**/rac-**12** 2:1

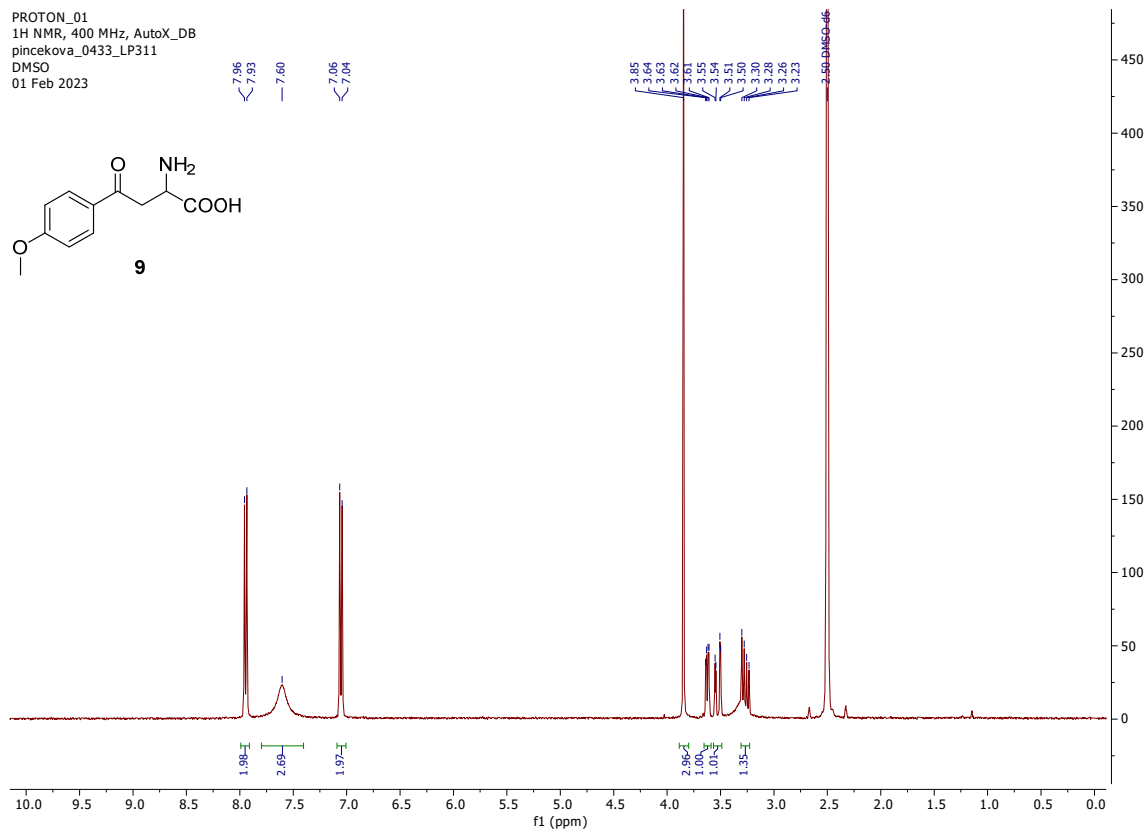
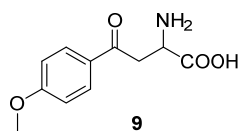
5. X-ray analysis of compounds (*S,S*)-1

Single-crystal diffraction data for (*S,S*)-1 were collected on a Bruker D8 VENTURE Kappa Duo diffractometer equipped with a PHOTON III detector and two I μ S microfocus sealed tubes (Cu, Mo). Data were collected at 120K using monochromated CuK α ($\lambda = 1.54178$ Å) primary radiation. Data reduction was carried out using the diffractometer software. The phase problem was solved by intrinsic phasing (SHELXT)⁴ and the structural model was refined by full-matrix least-squares against F^2 (SHELXL).⁵ Non-hydrogen atoms were refined anisotropically and with no constraints imposed. Hydrogen atoms were refined isotropically. The two hydrogen atoms attached to N1 were refined freely, while all other hydrogen atoms were put into idealized positions and were refined using the riding model.

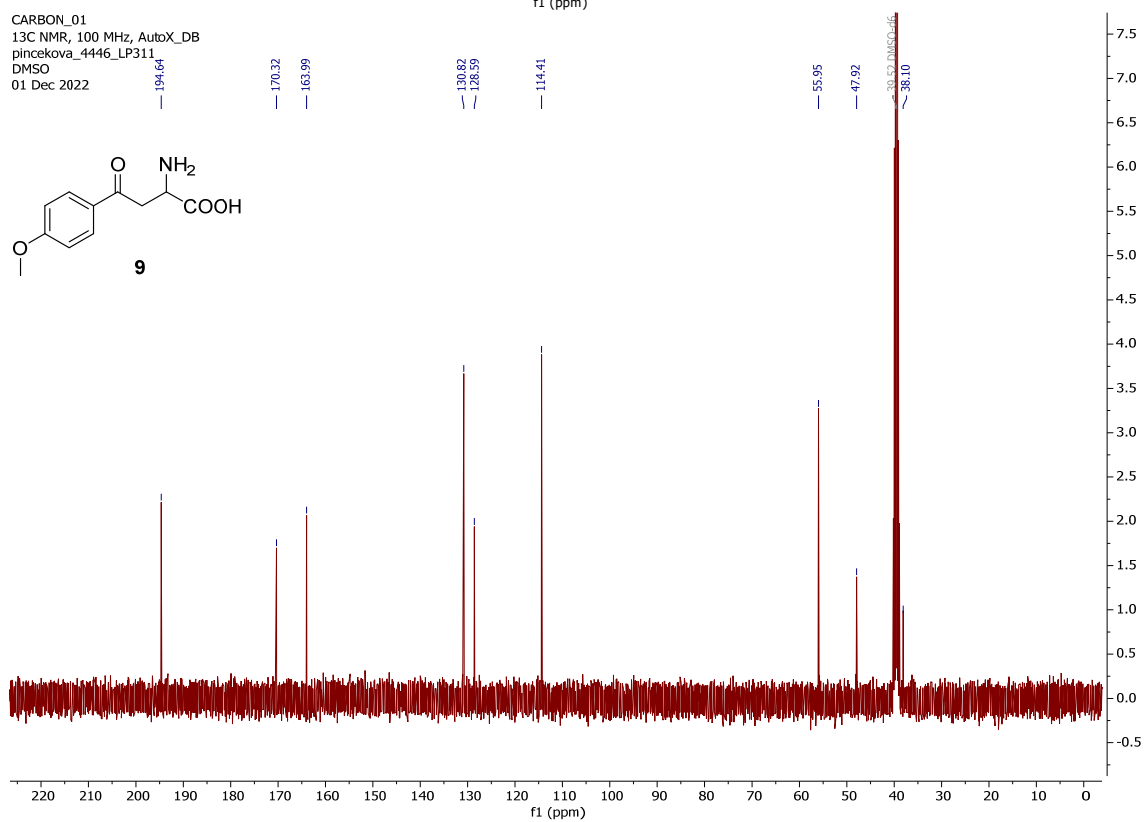
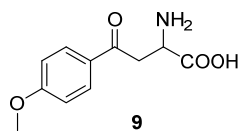


The molecular structure of (*S,S*)-1 in solid state. Non-hydrogen atoms are displayed by thermal ellipsoids on 30% probability level.

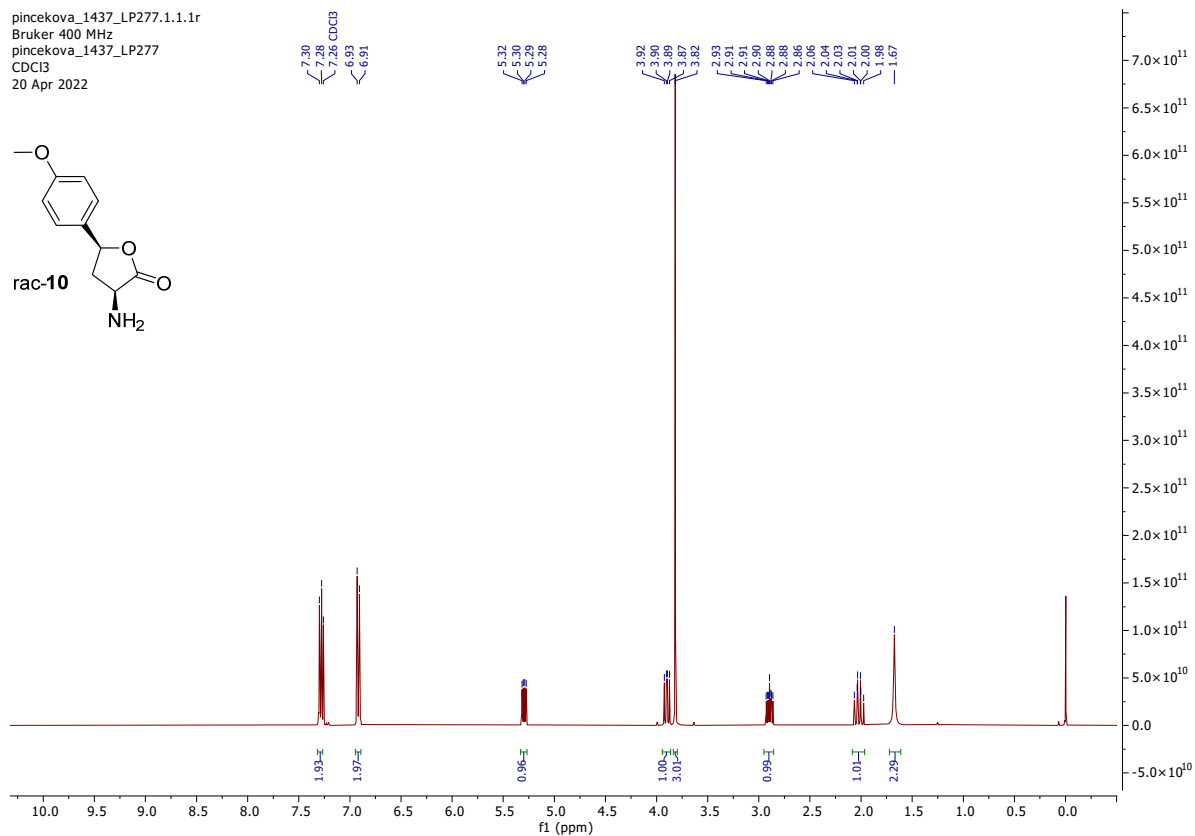
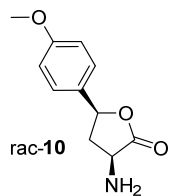
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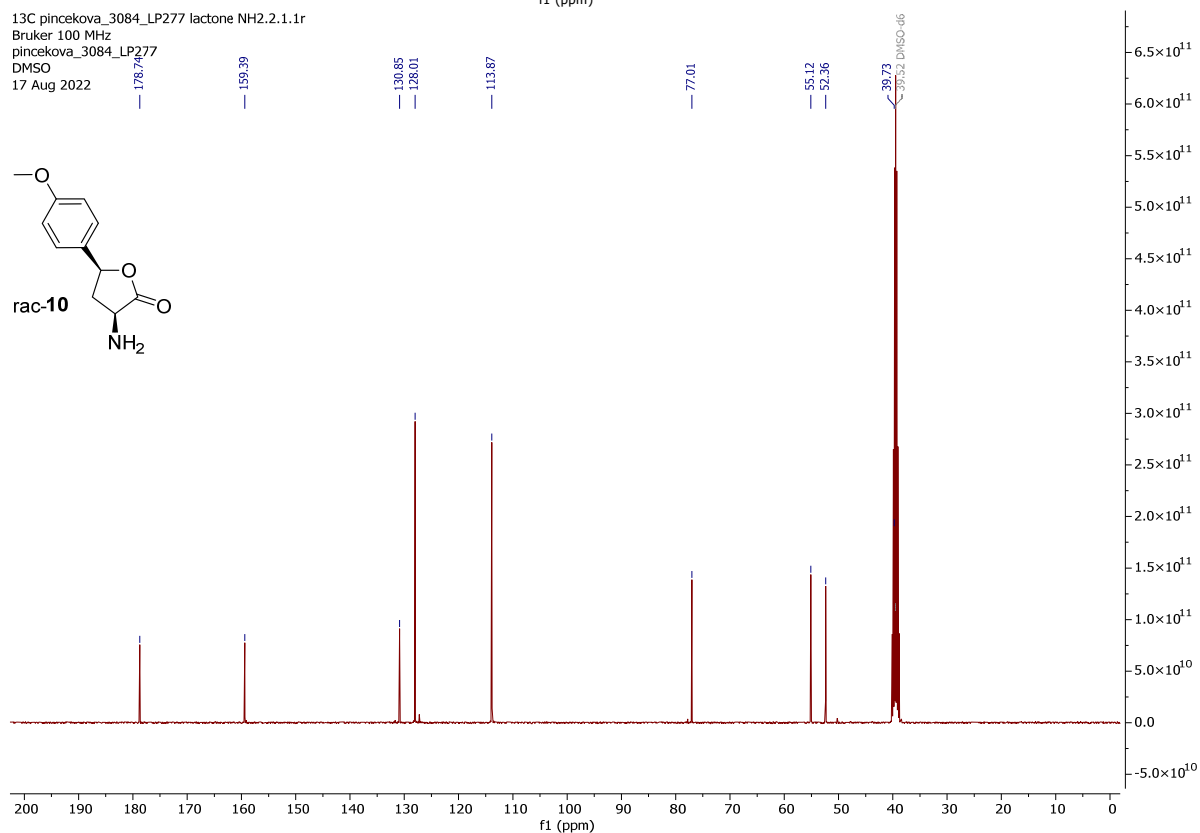
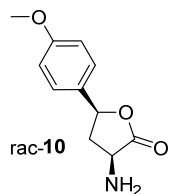
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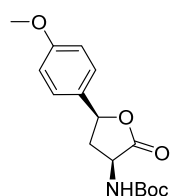
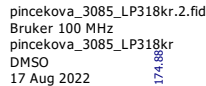


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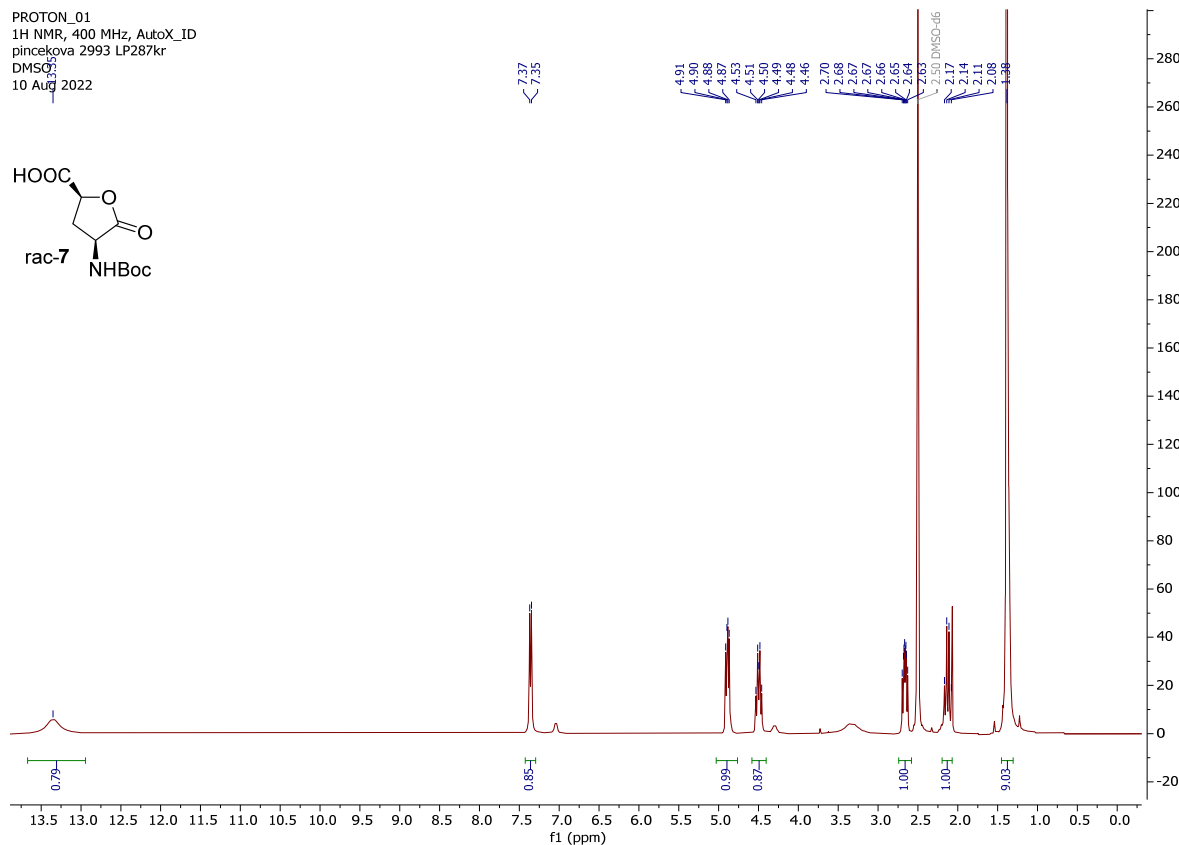
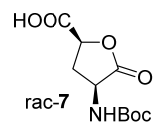


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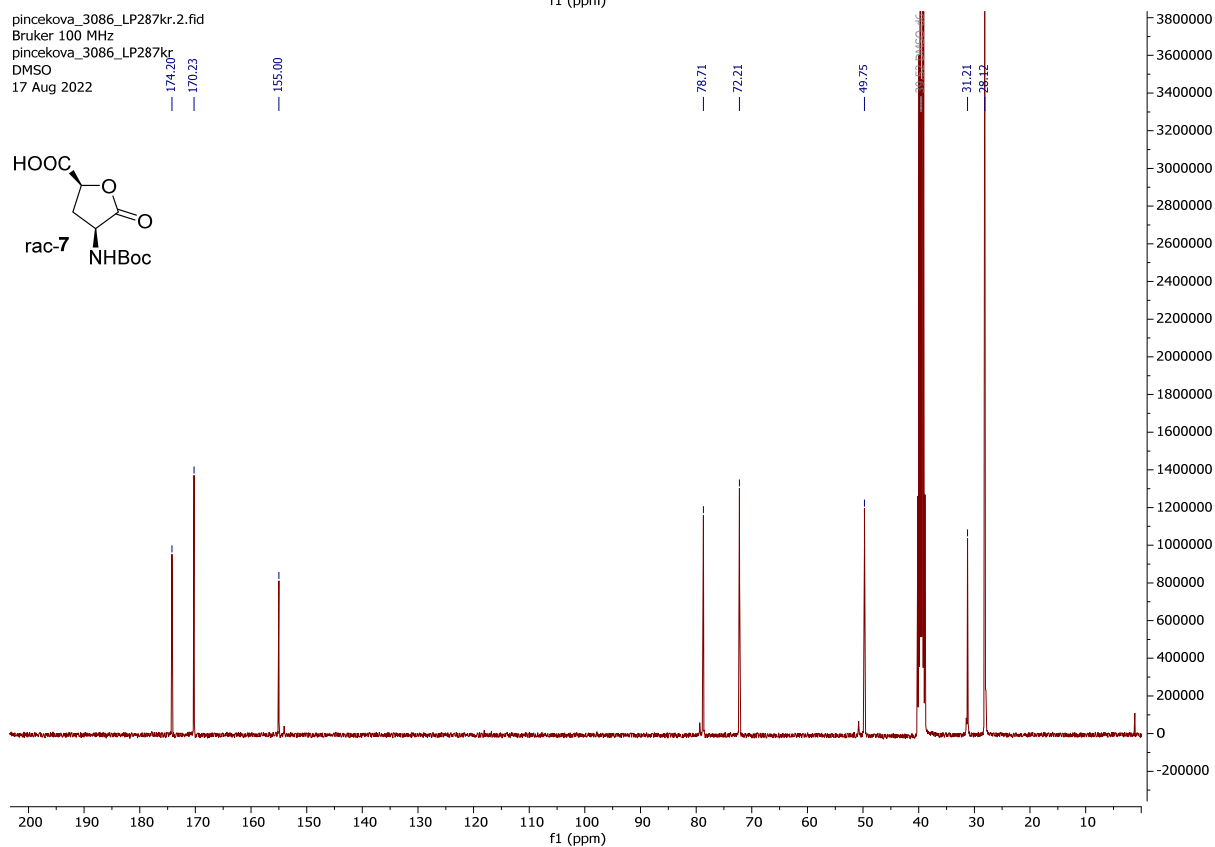
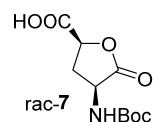


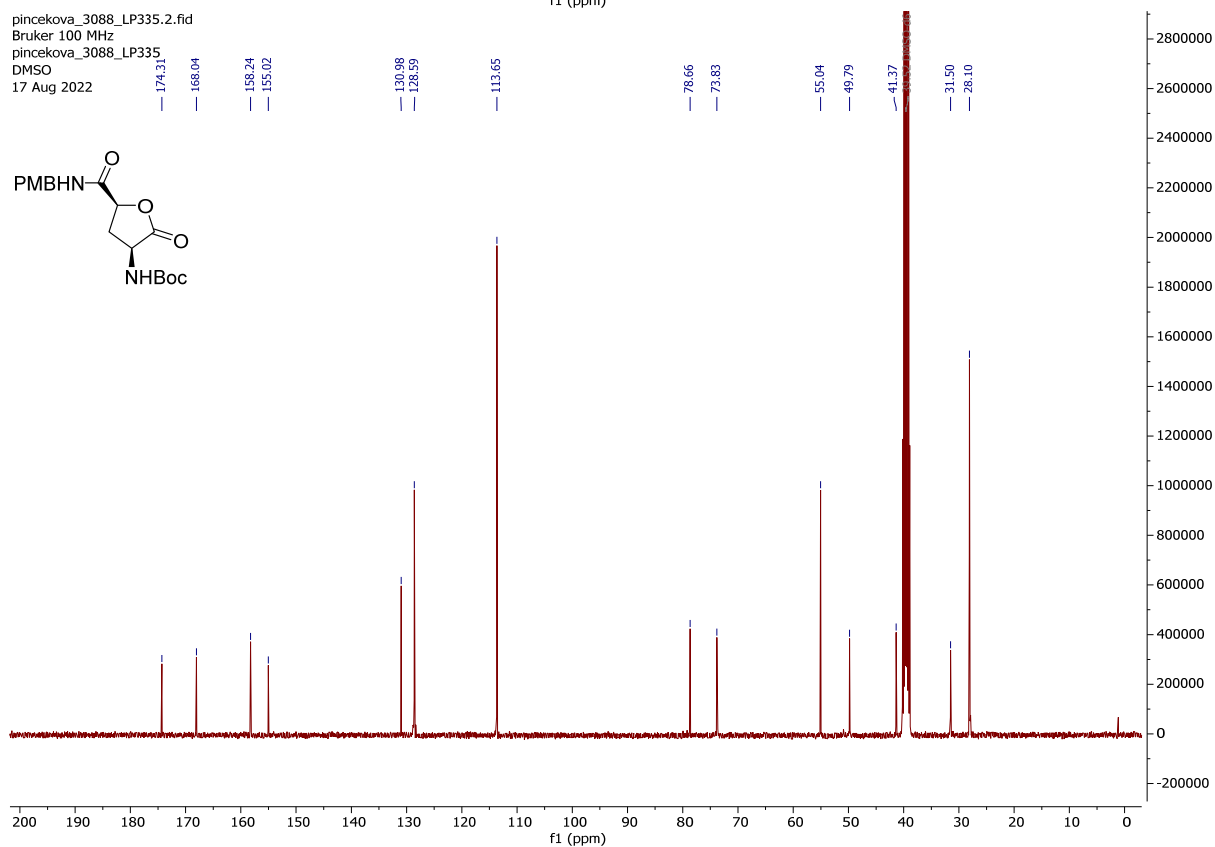
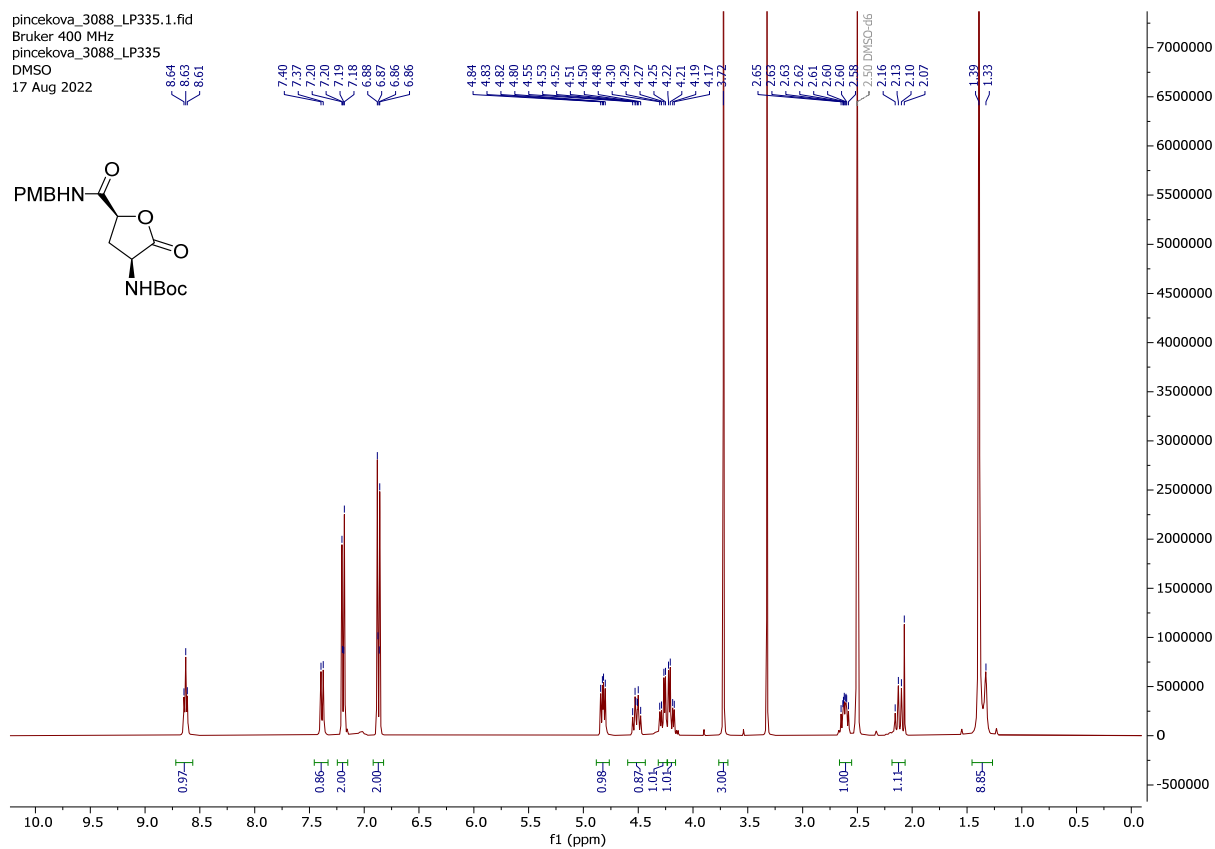
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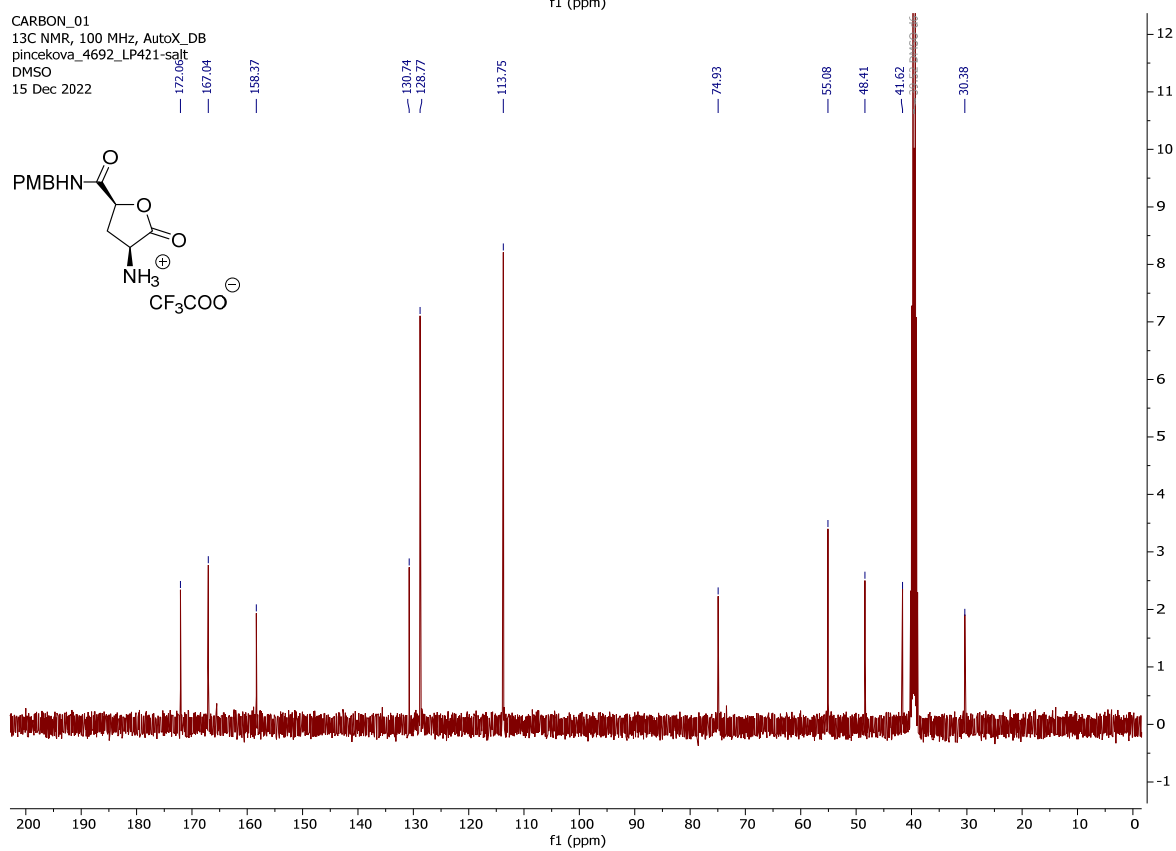
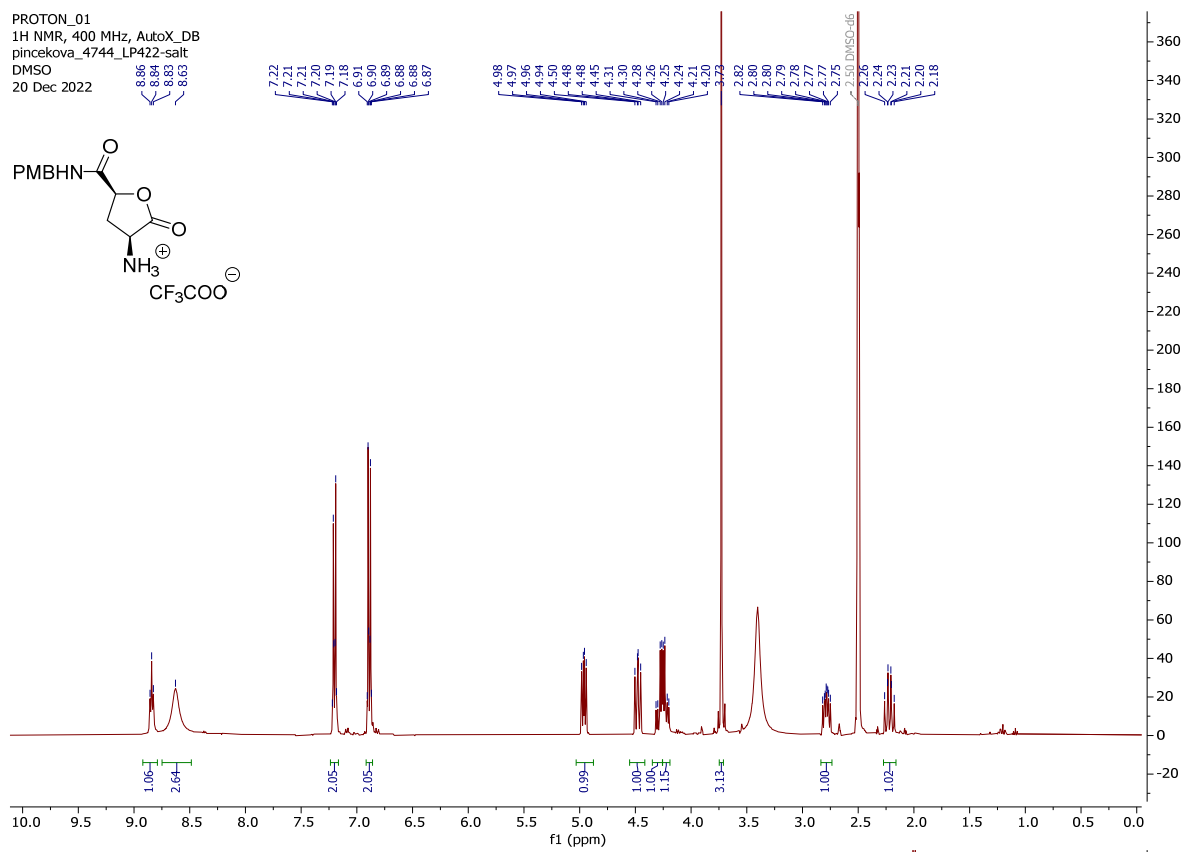
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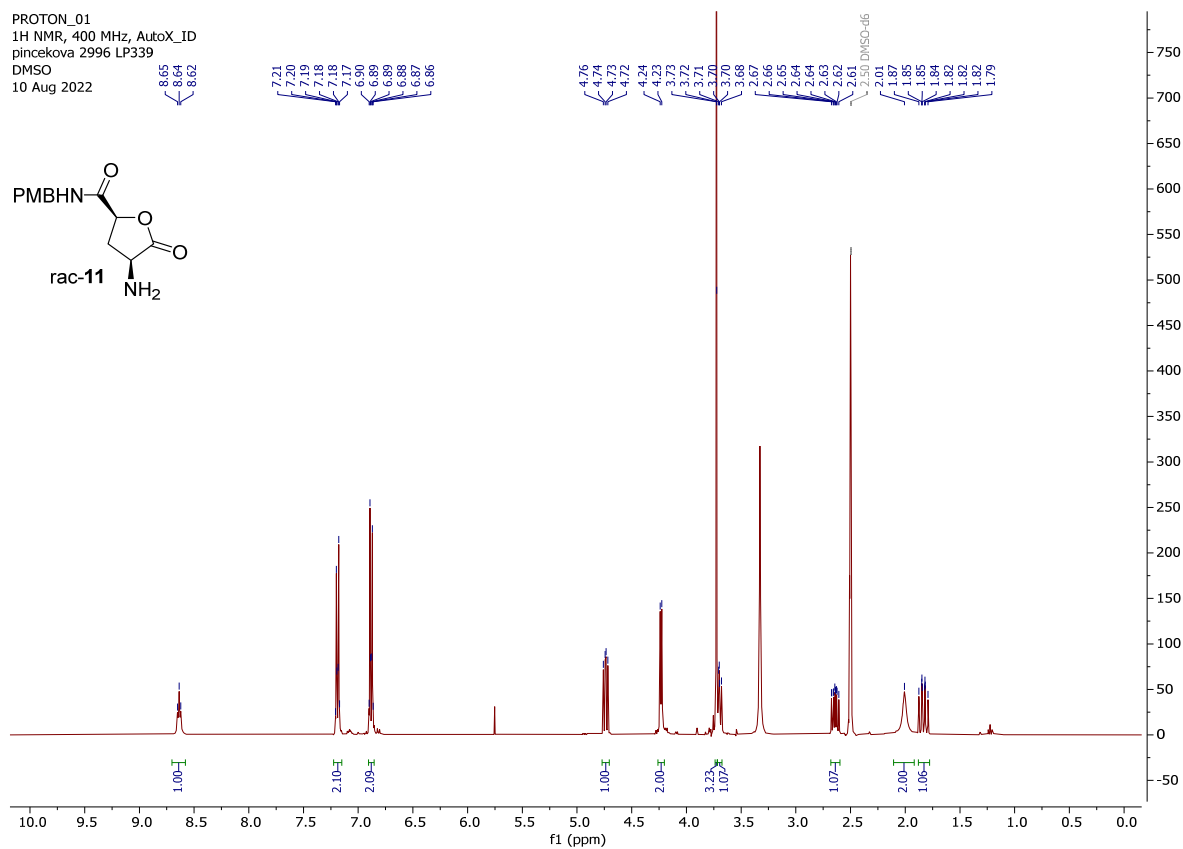
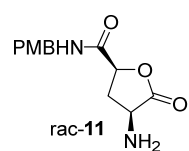
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 17 Aug 2022



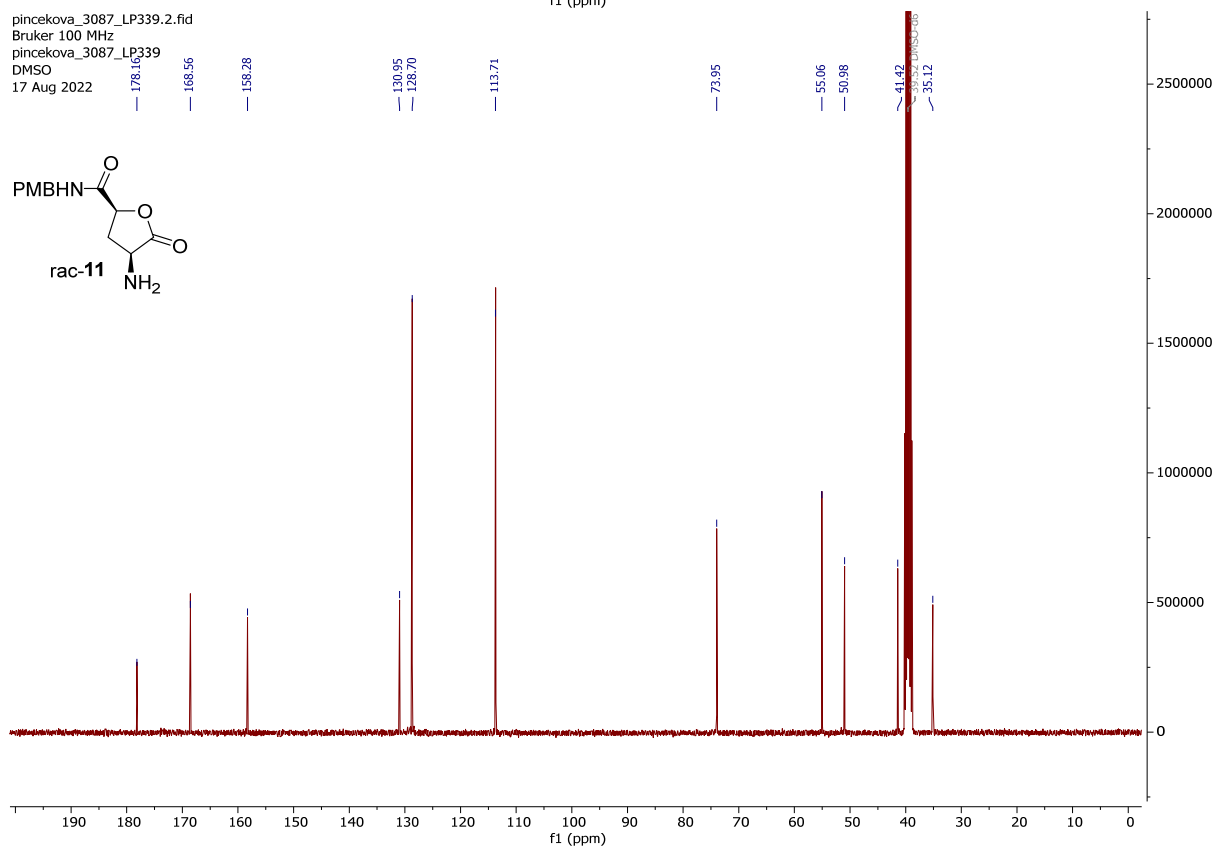
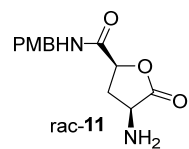


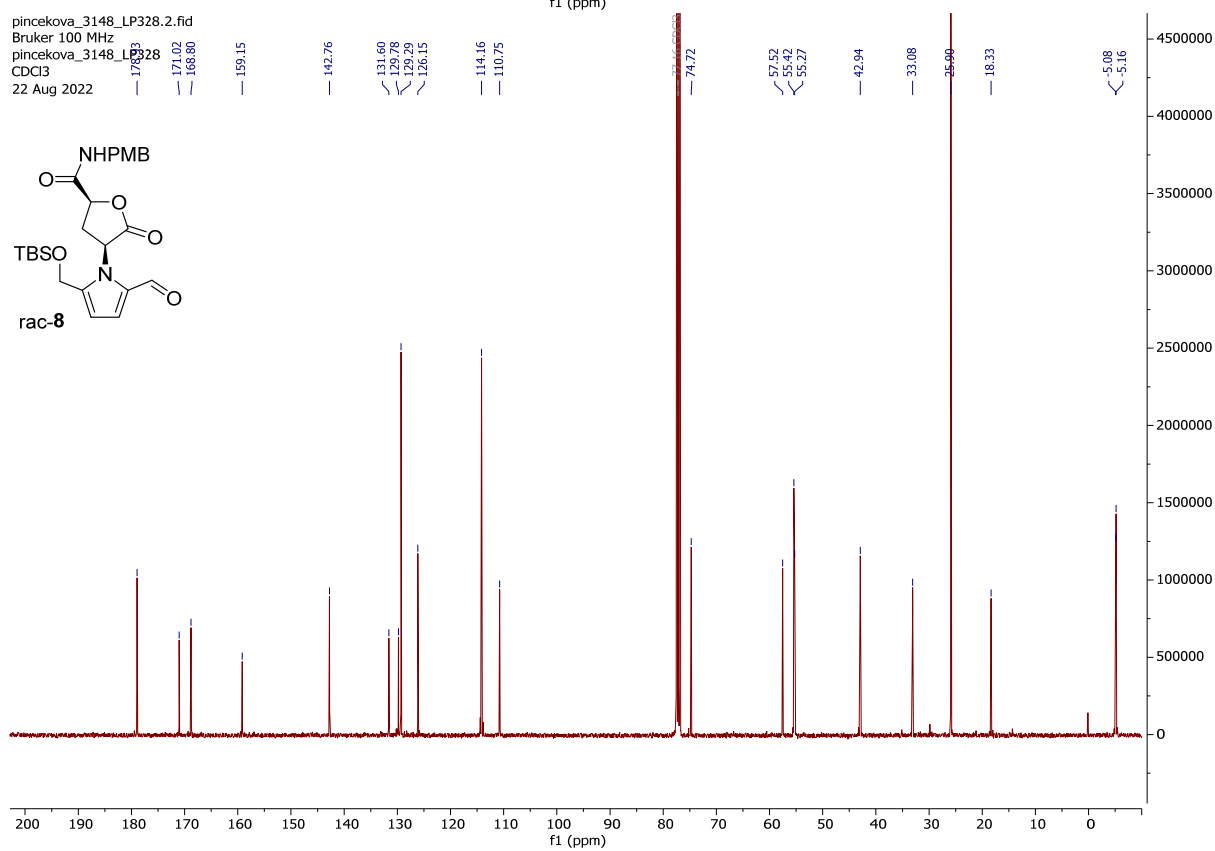
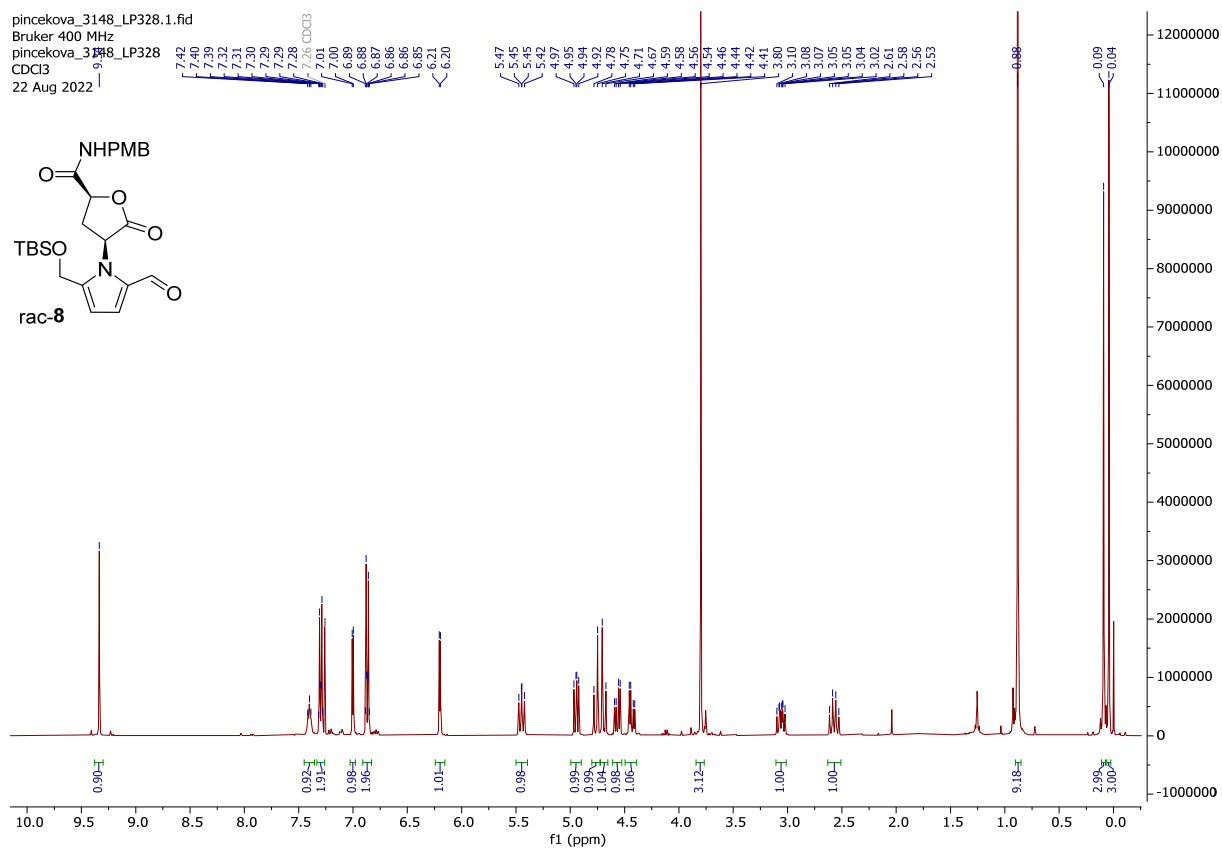


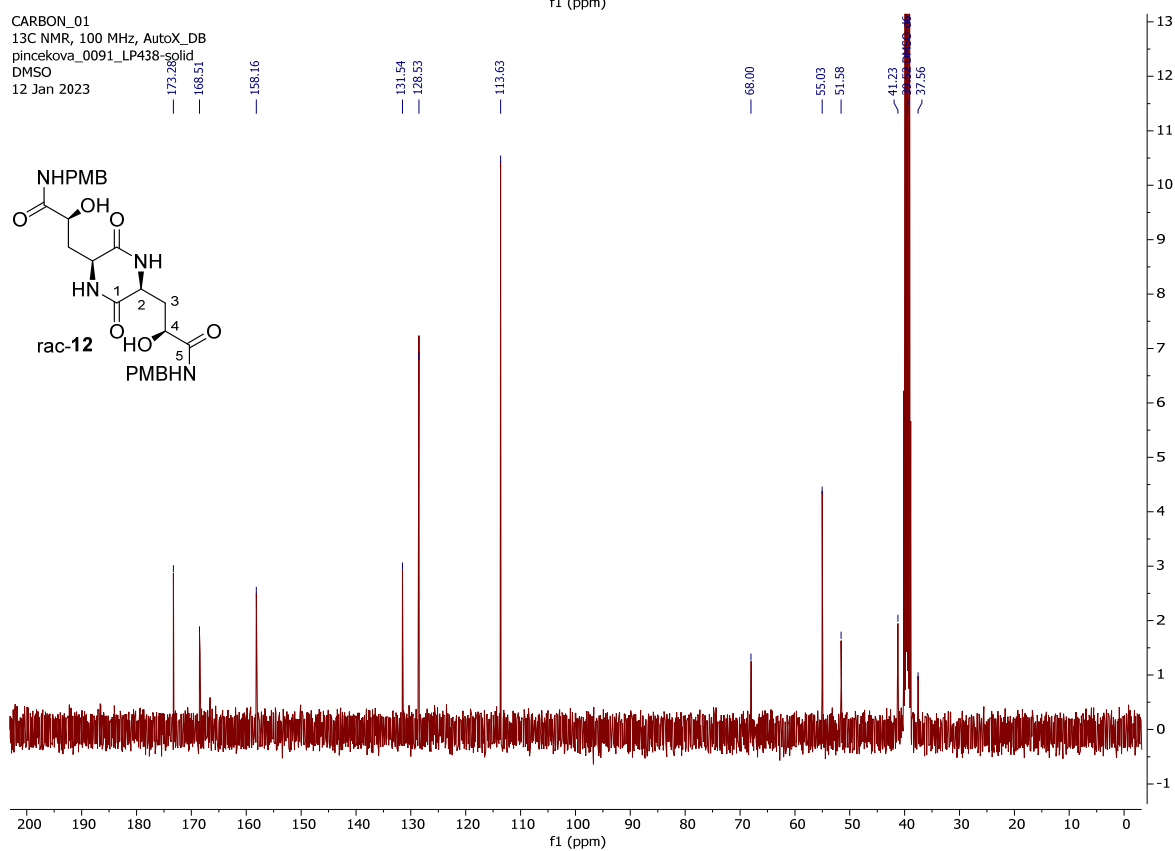
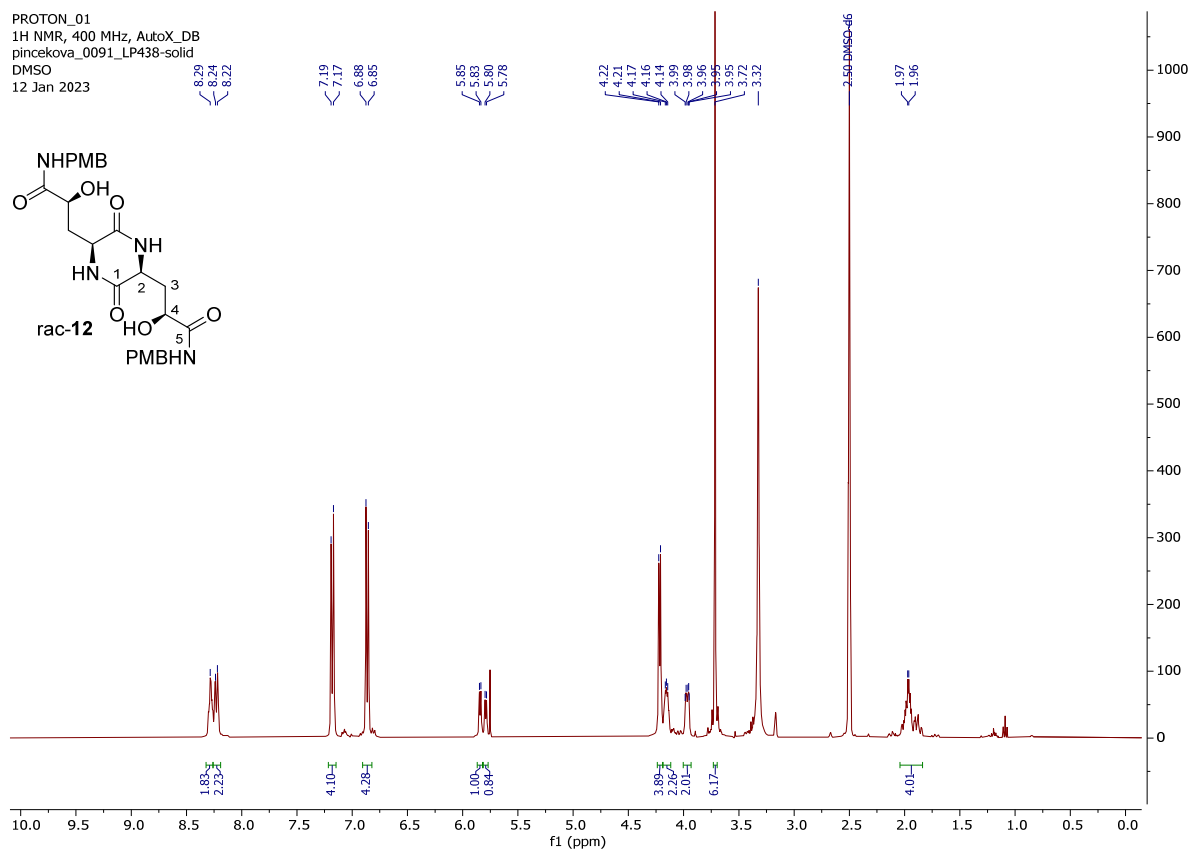
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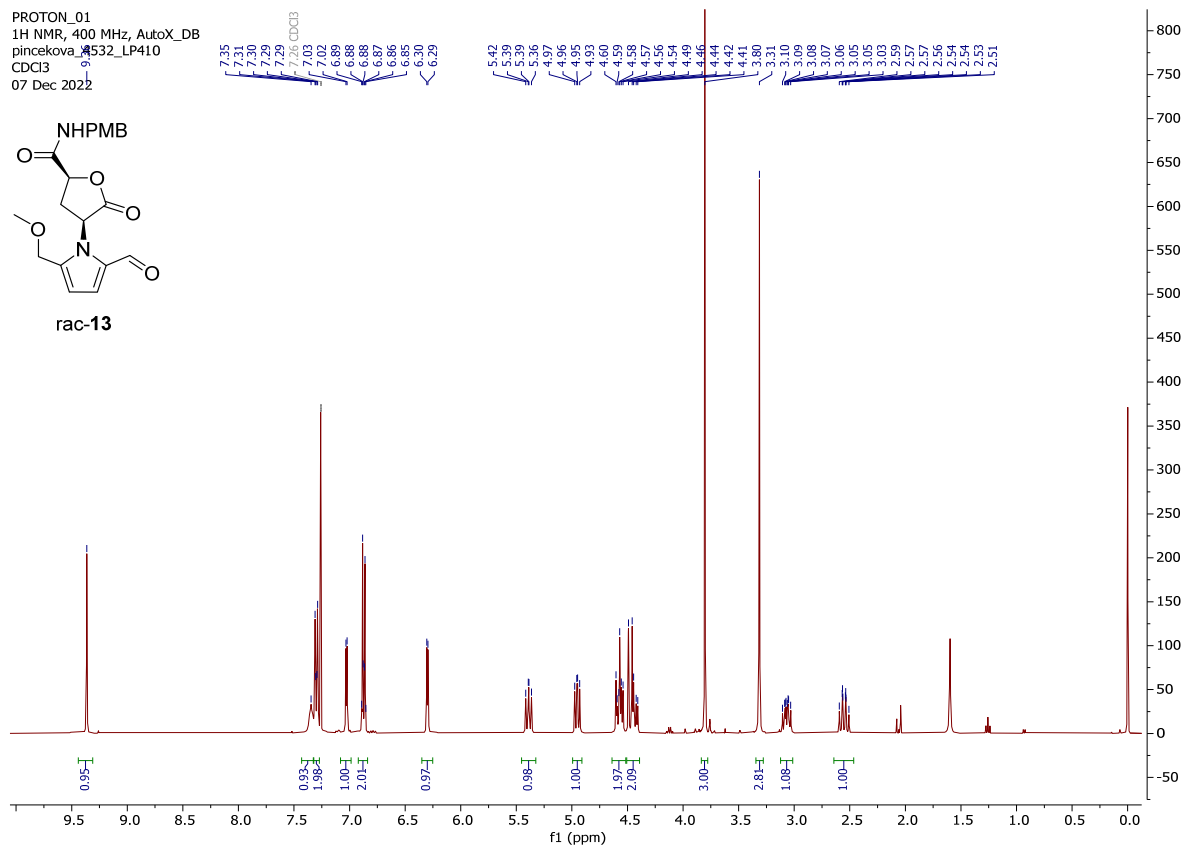
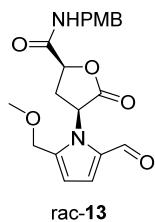
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 17 Aug 2022



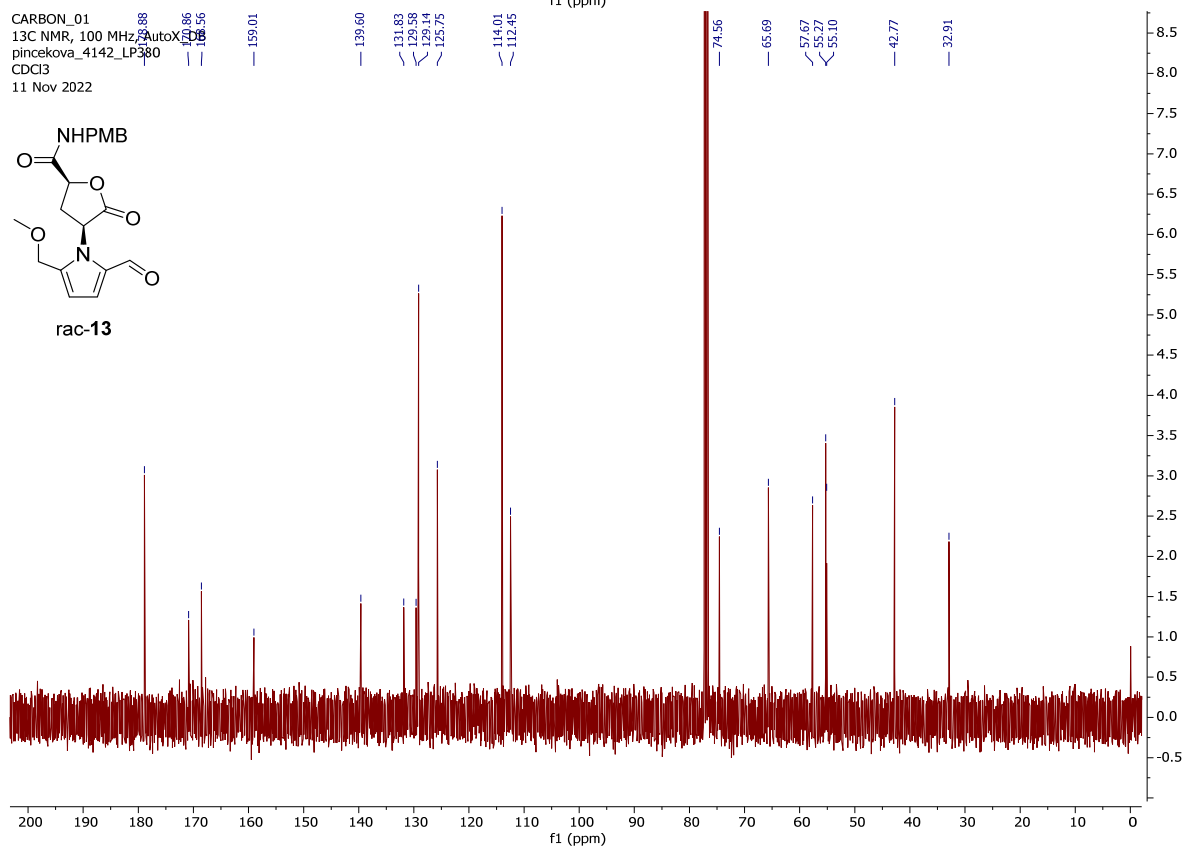
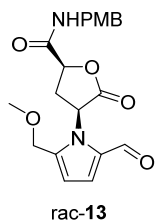




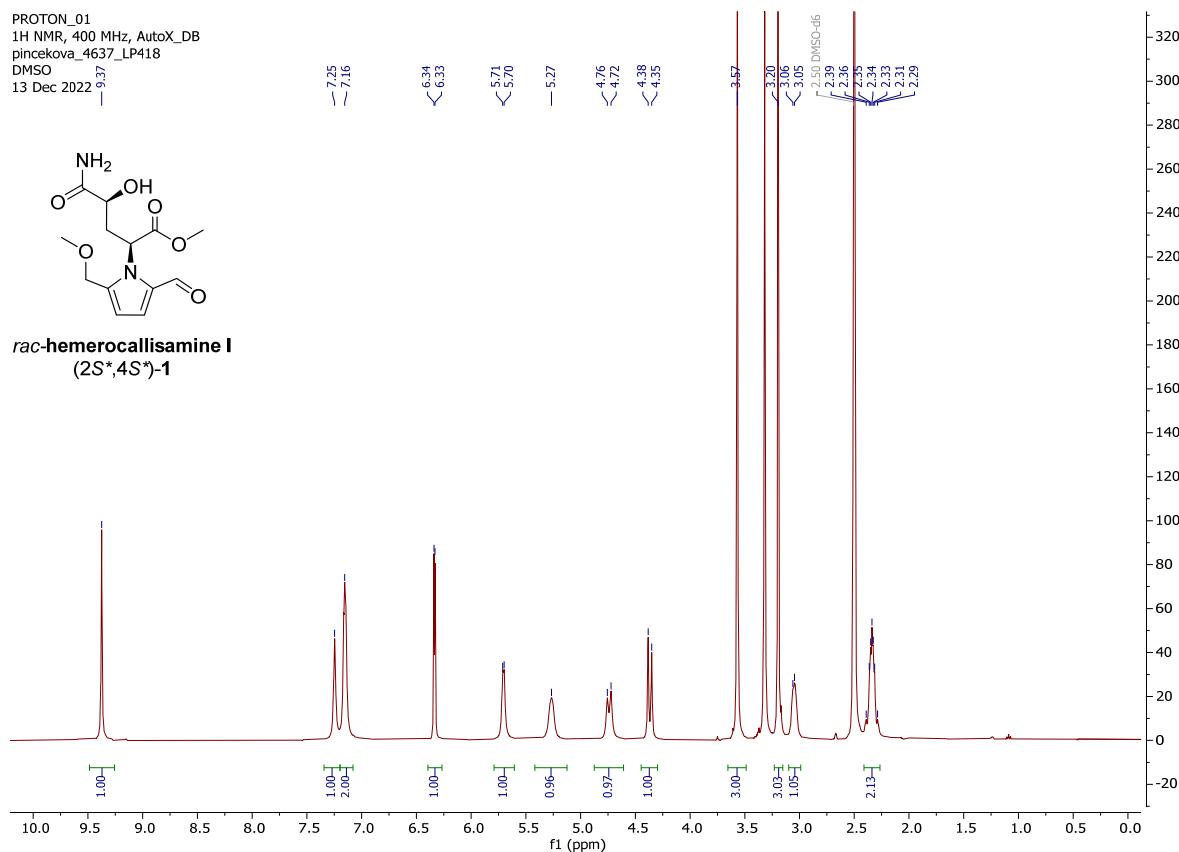
PROTON_01
 1H NMR, 400 MHz, AutoX_DB
 pinckova_4532_LP410
 CDCl3
 07 Dec 2022



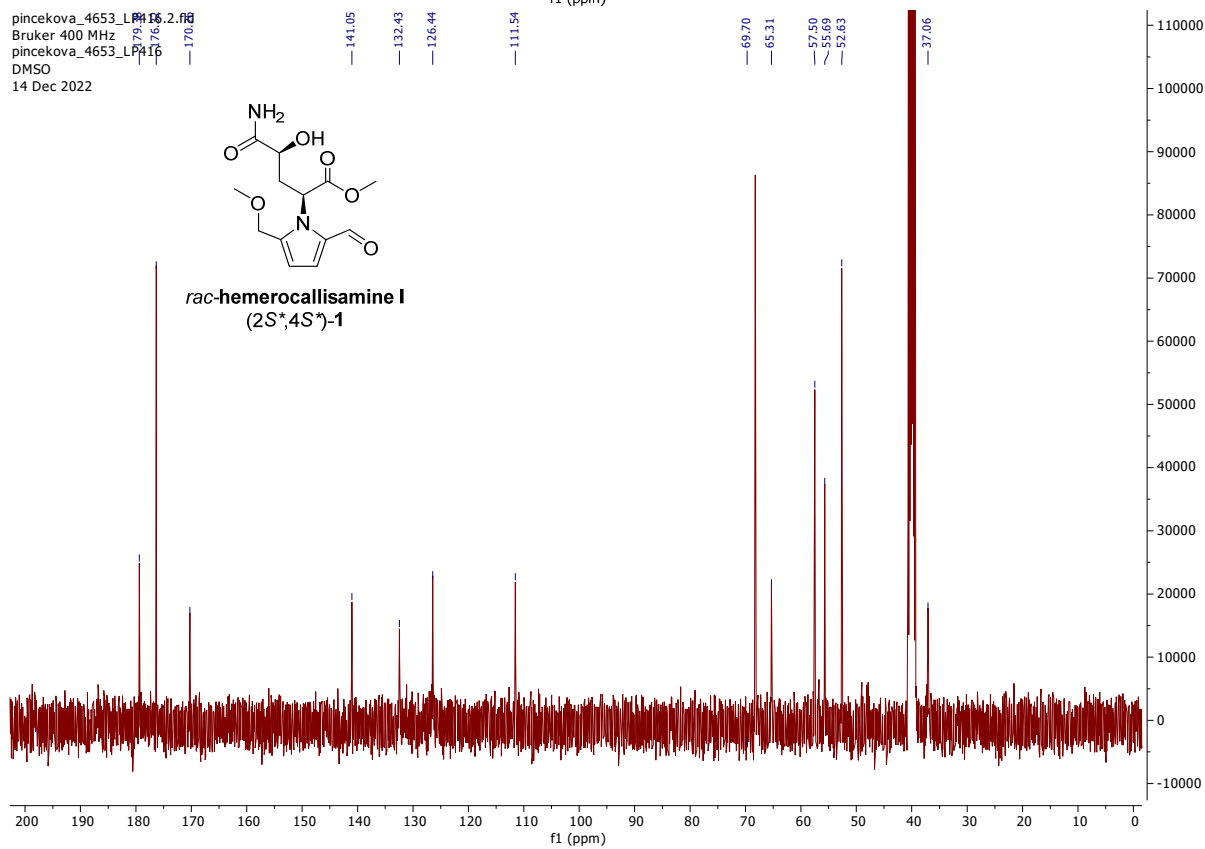
CARBON_01
 13C NMR, 100 MHz, AutoX_DB
 pinckova_4142_LP380
 CDCl3
 11 Nov 2022

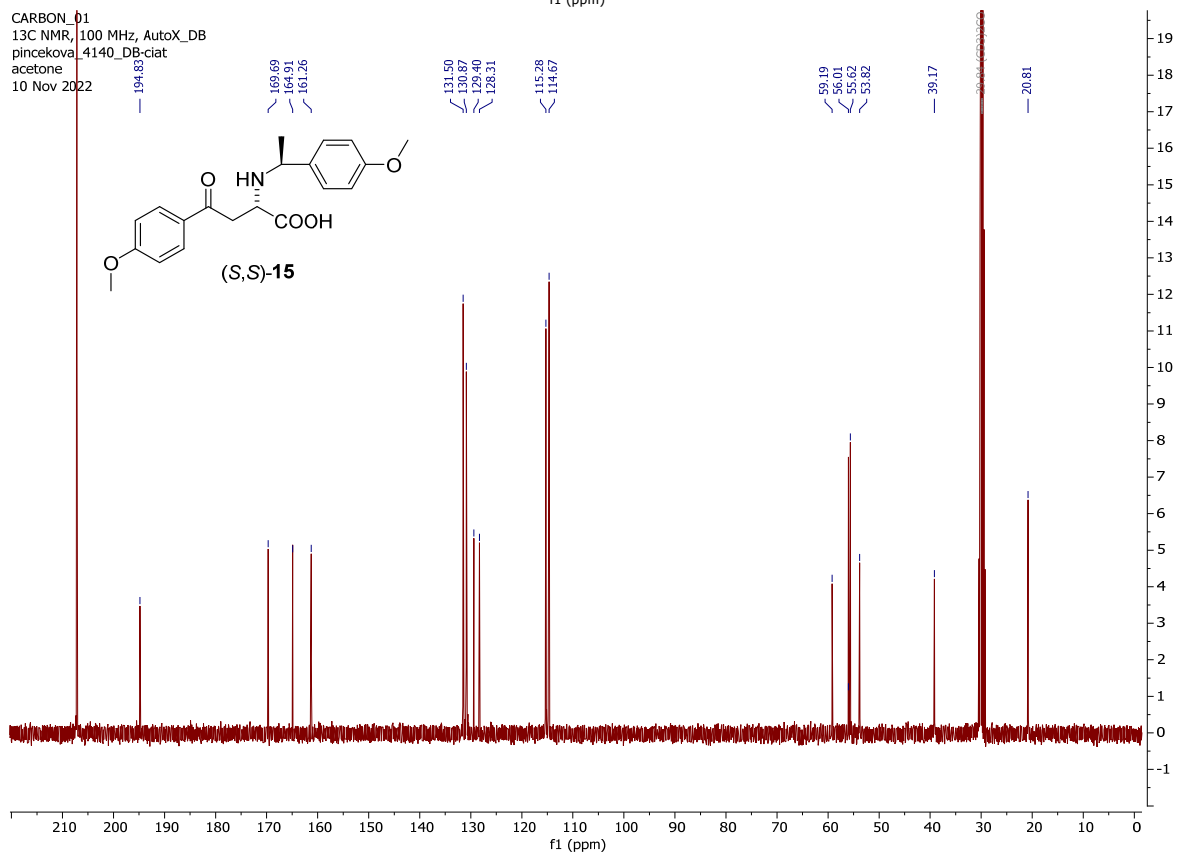
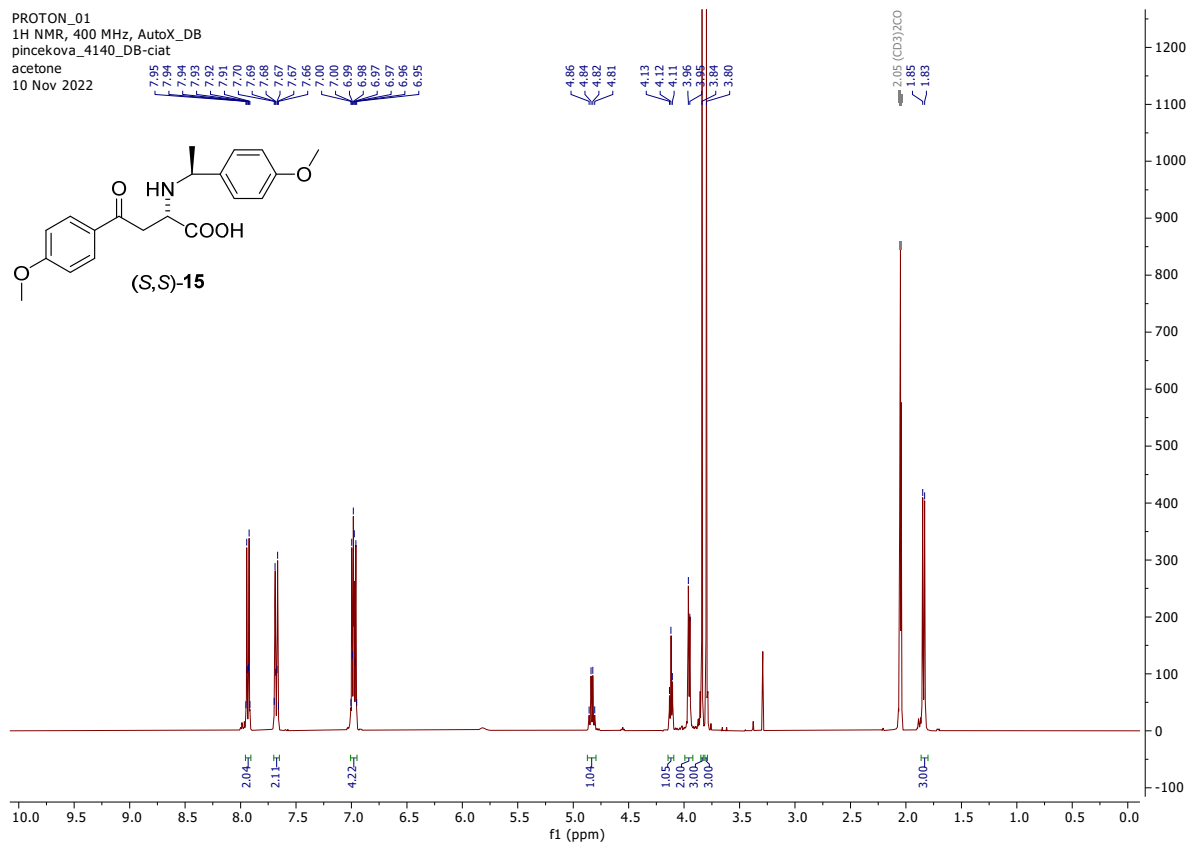


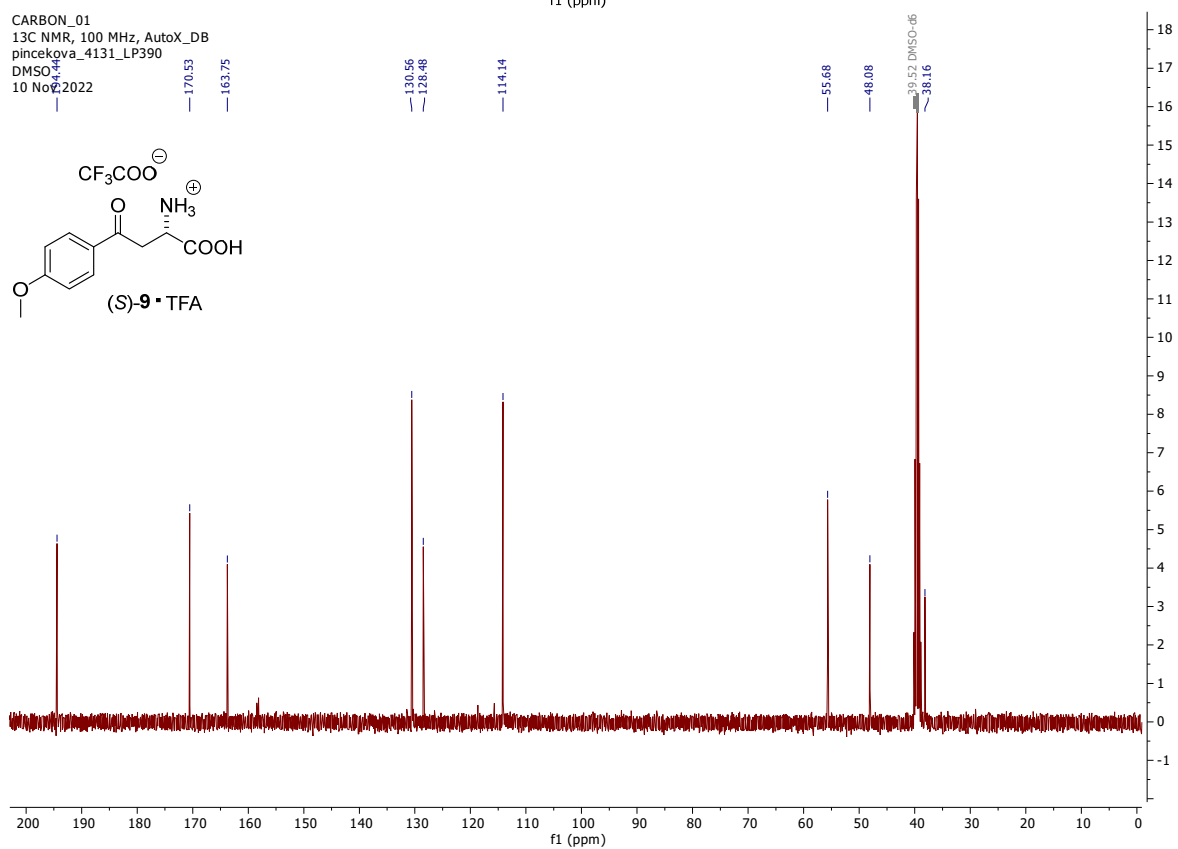
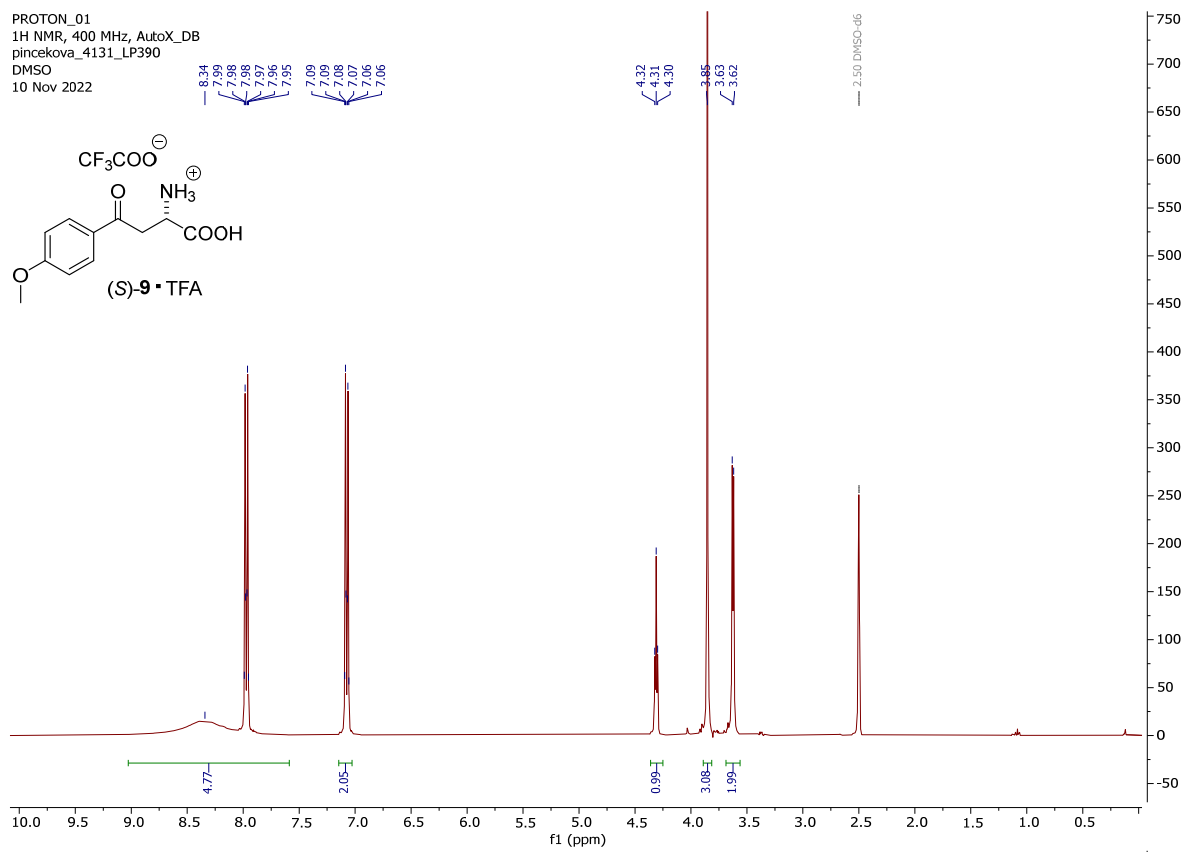
PROTON_01
 1H NMR, 400 MHz, AutoX_DB
 pincekova_4637_LP418
 DMSO
 13 Dec 2022



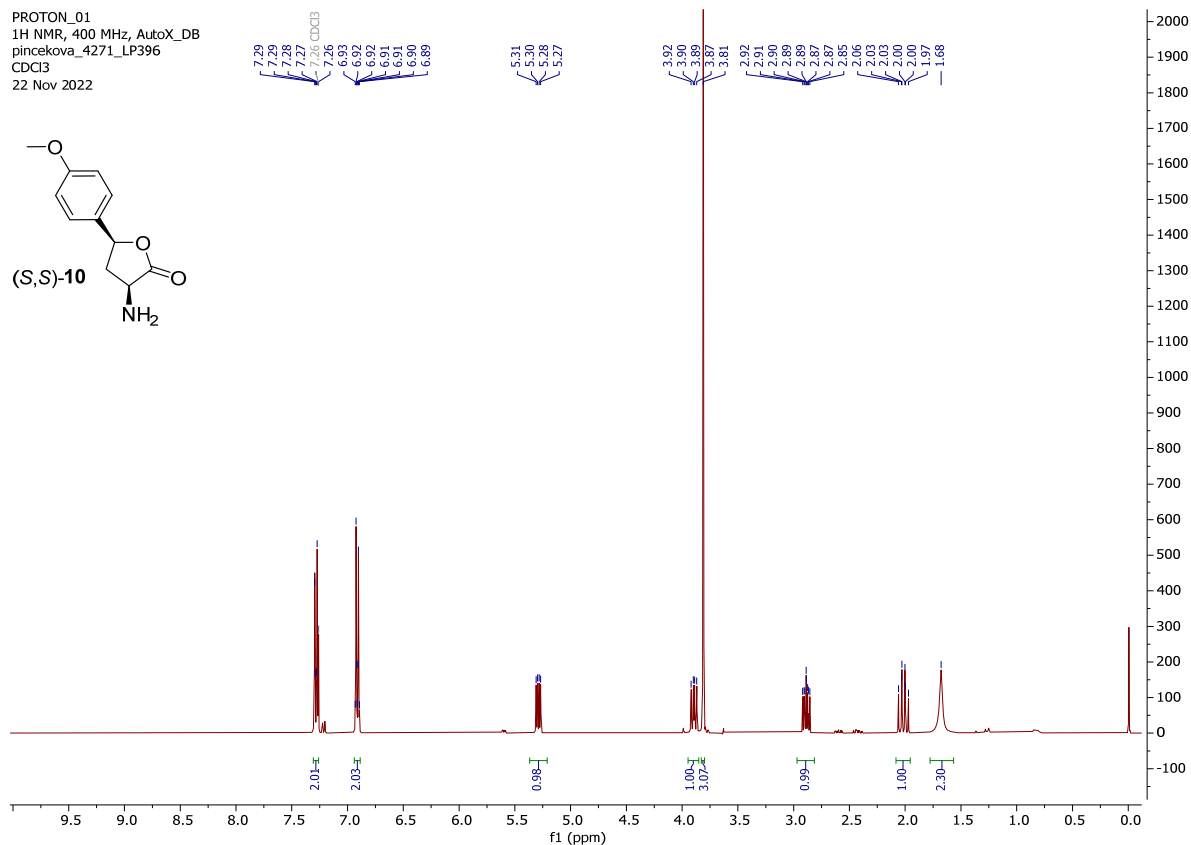
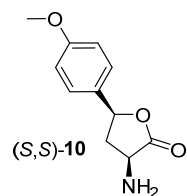
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 Bruker 400 MHz
 pincekova_4653_LP416
 DMSO
 14 Dec 2022



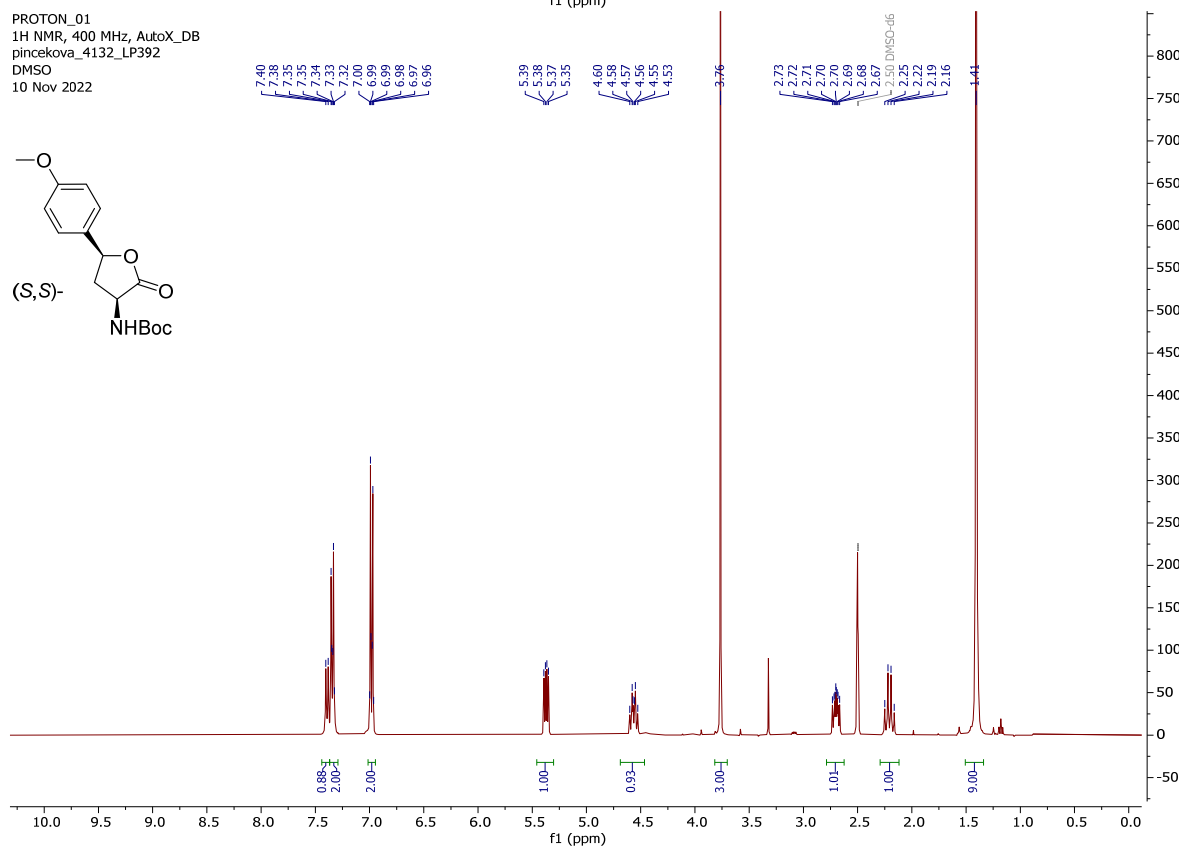
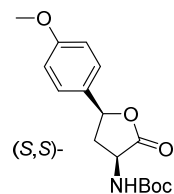




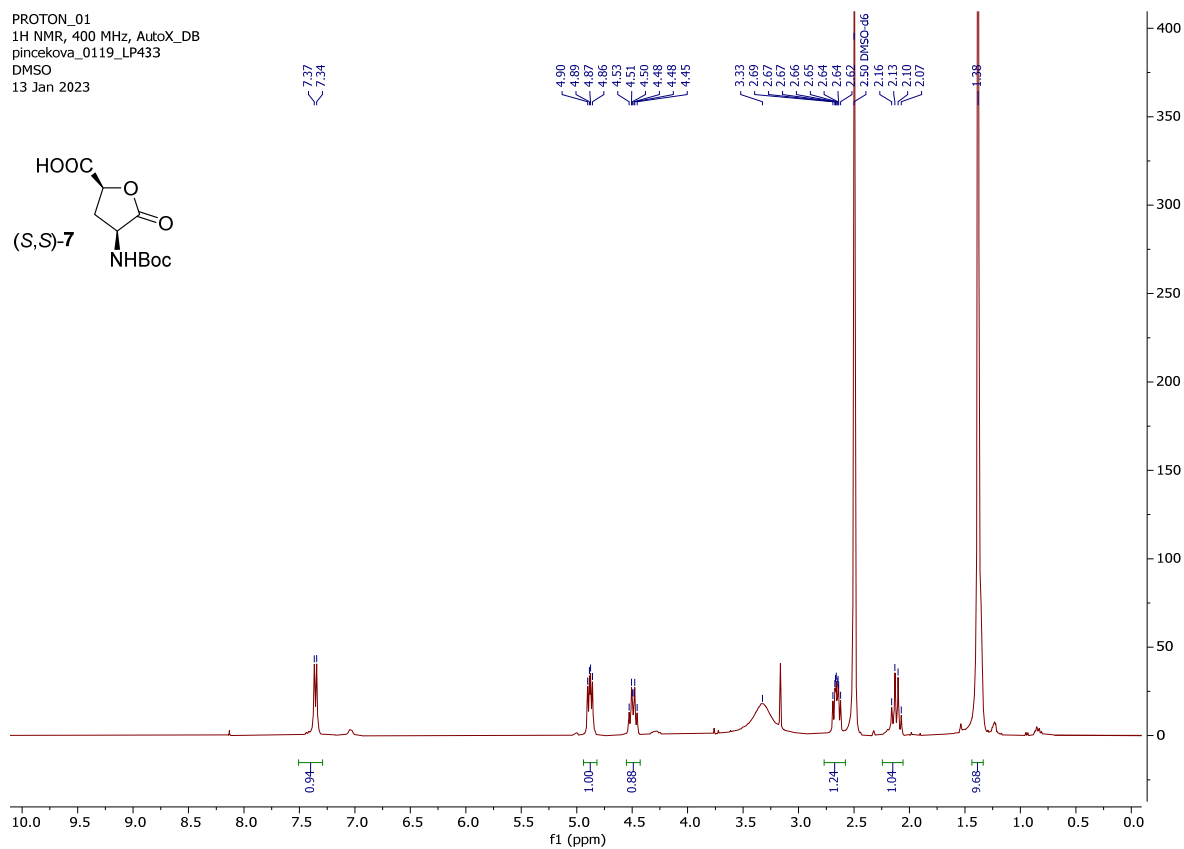
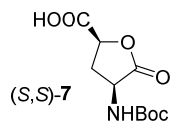
PROTON_01
 1H NMR, 400 MHz, AutoX_DB
 pincekova_4271_LP396
 CDCl3
 22 Nov 2022



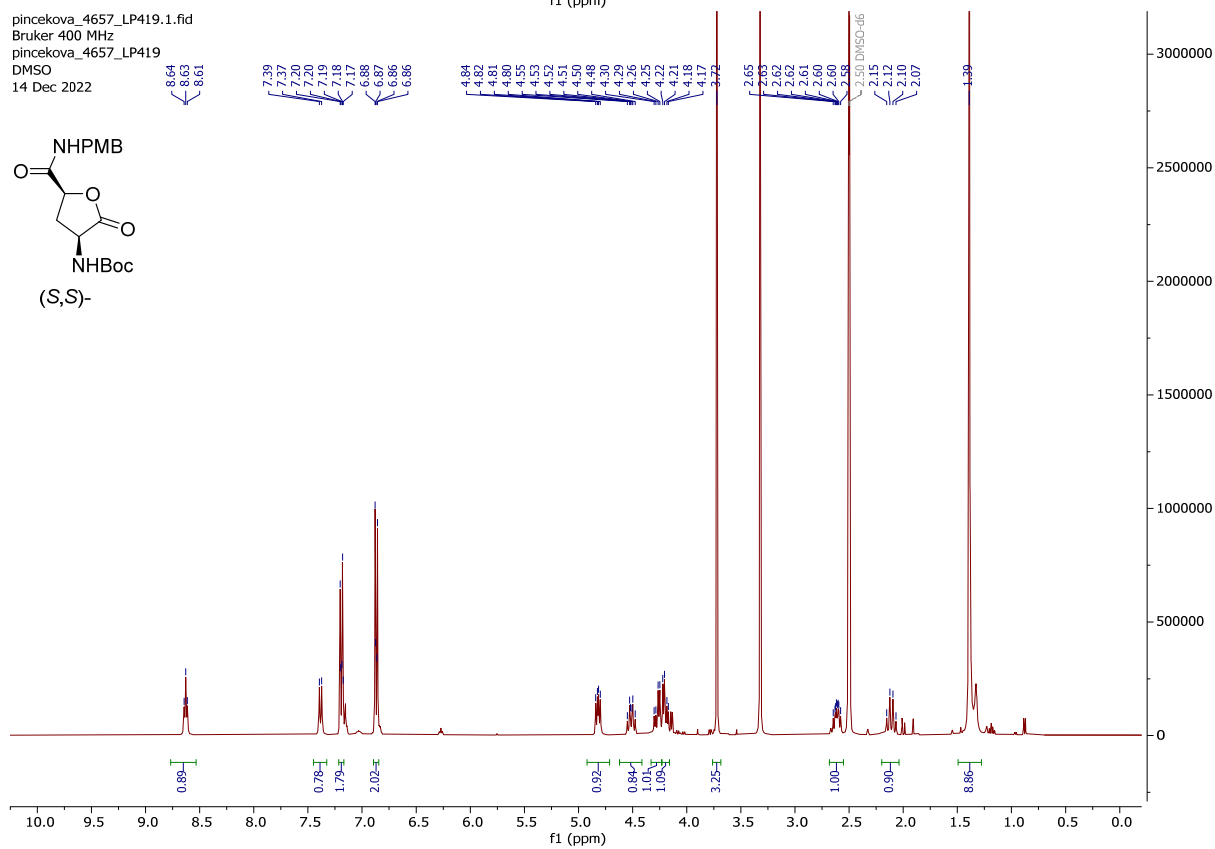
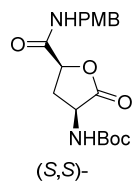
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 1H NMR, 400 MHz, AutoX_DB
 pincekova_4132_LP392
 DMSO
 10 Nov 2022



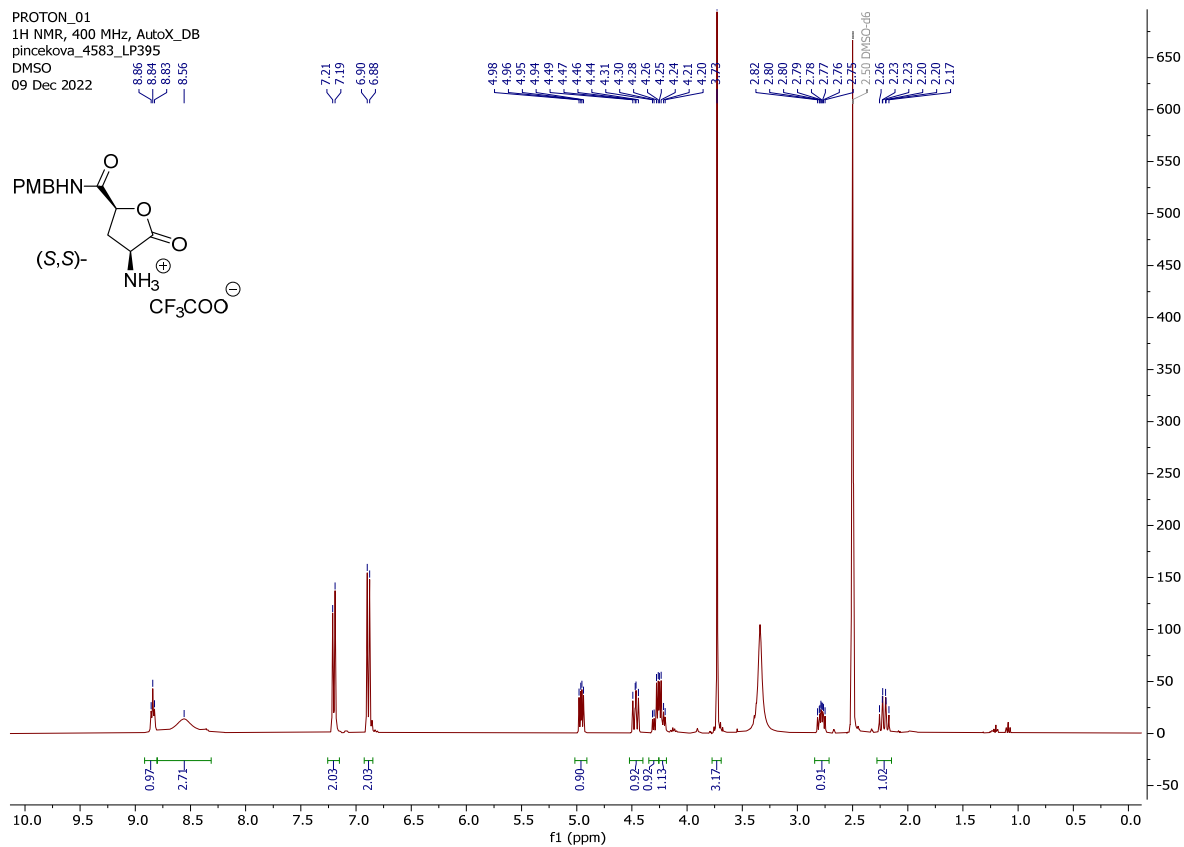
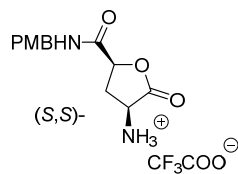
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 1H NMR, 400 MHz, AutoX_DB
 pincekova_0119_LP433
 DMSO
 13 Jan 2023



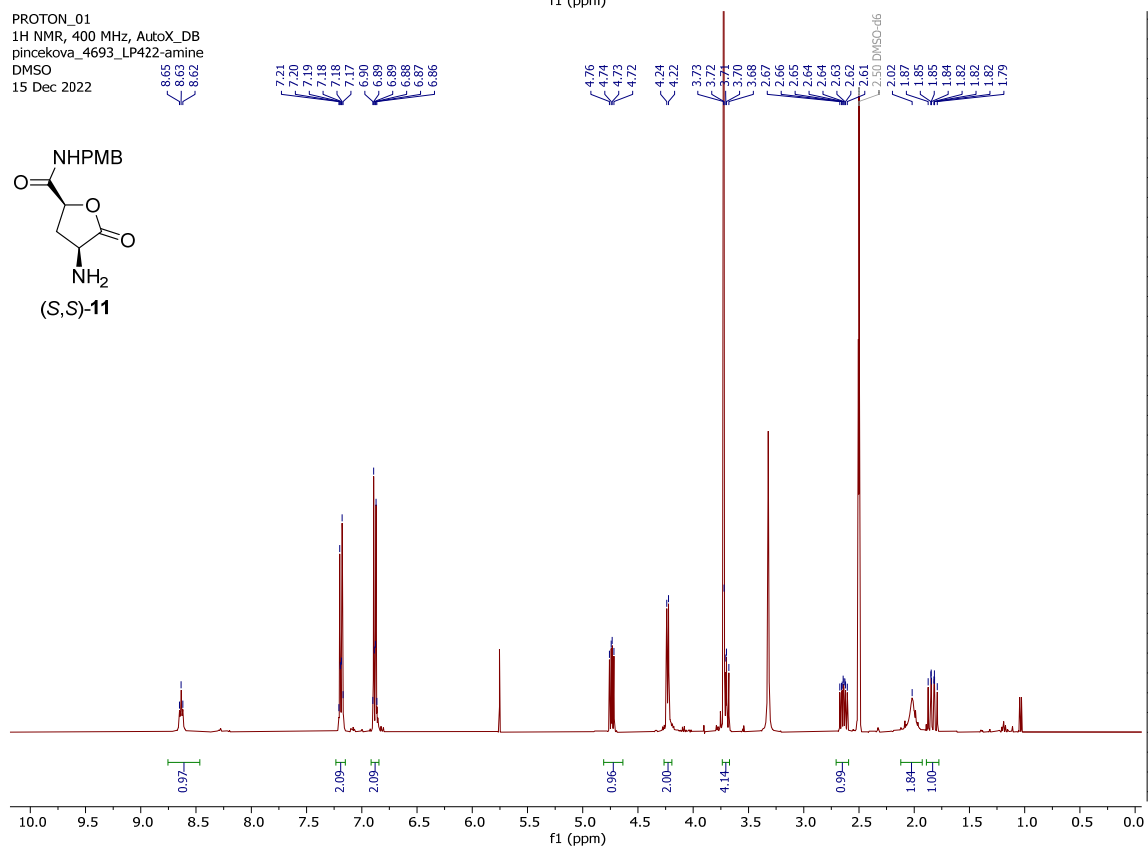
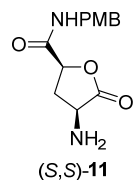
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 Bruker 400 MHz
 pincekova_4657_LP419
 DMSO
 14 Dec 2022



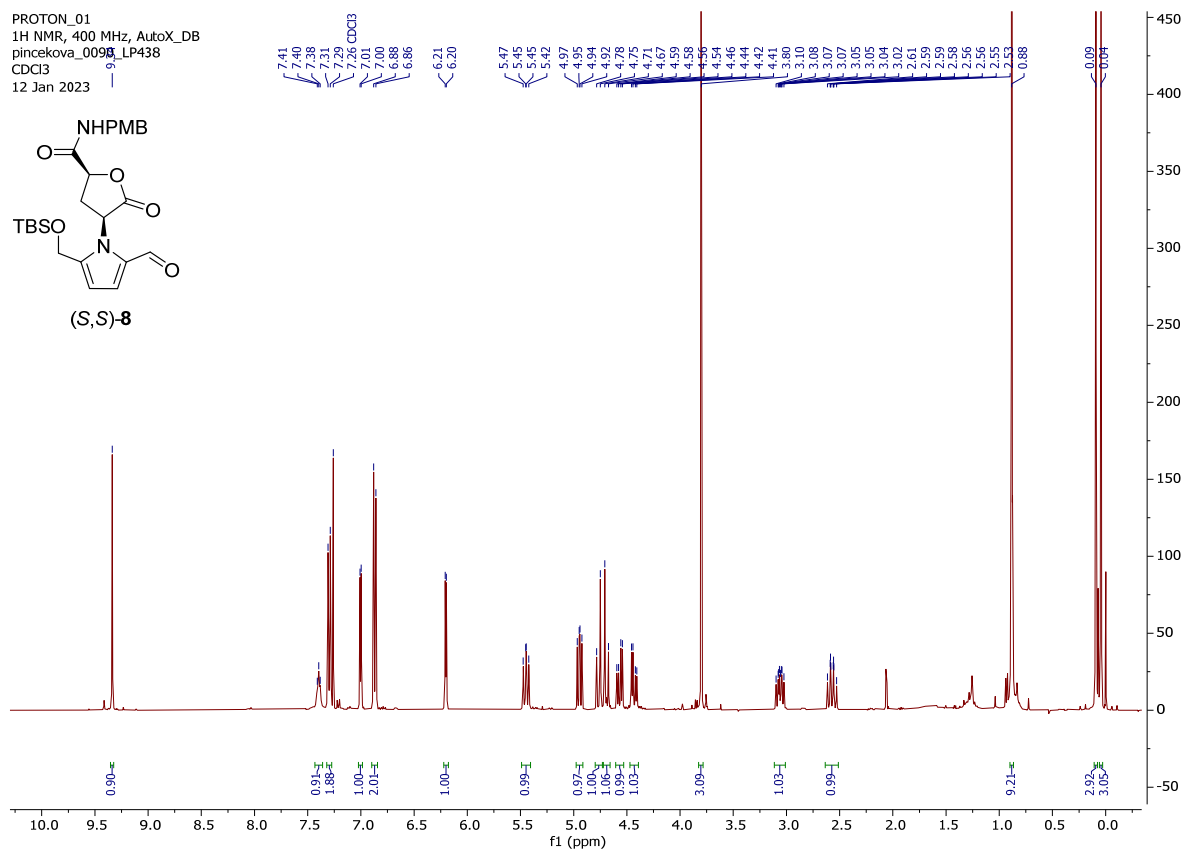
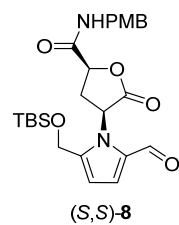
PROTON_01
 1H NMR, 400 MHz, AutoX_DB
 pincekova_4583_LP395
 DMSO
 09 Dec 2022



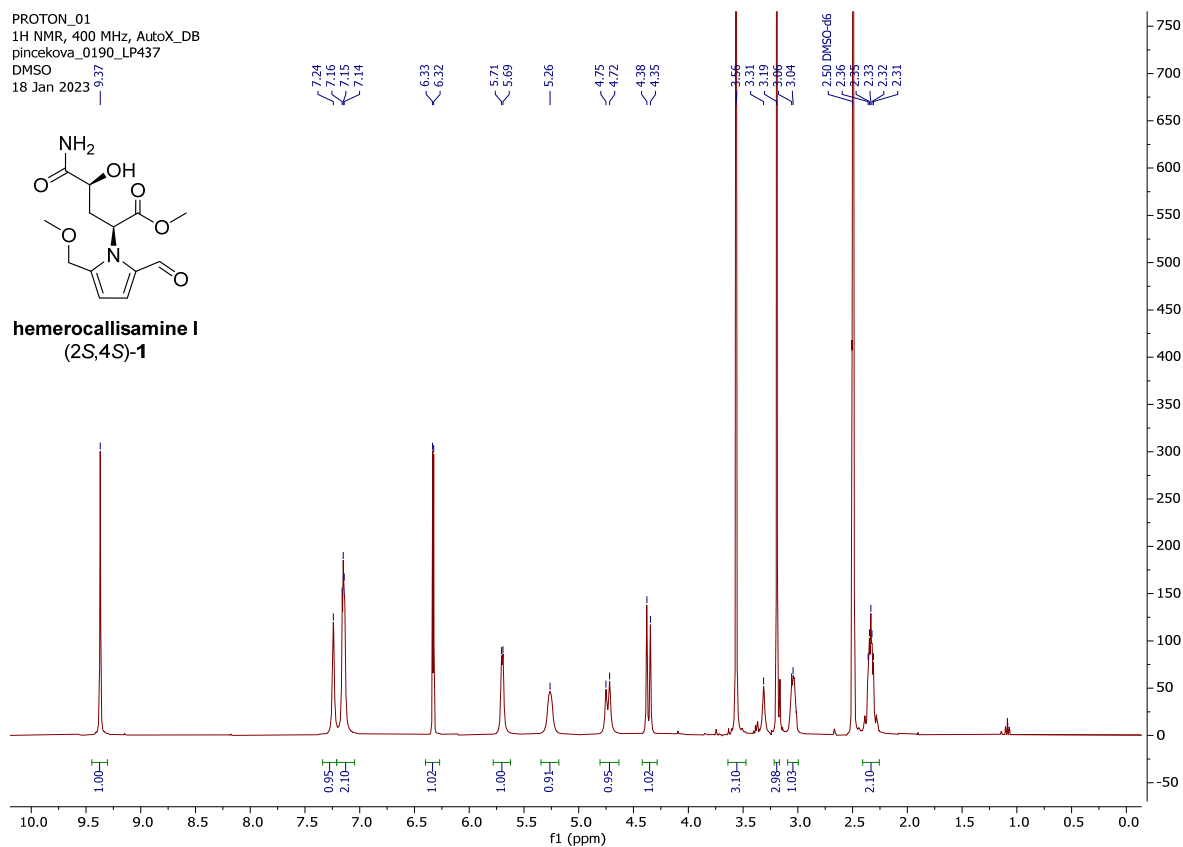
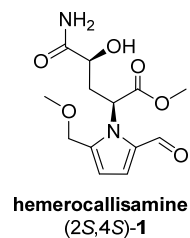
PROTON_01
 1H NMR, 400 MHz, AutoX_DB
 pincekova_4693_LP422-amine
 DMSO
 15 Dec 2022



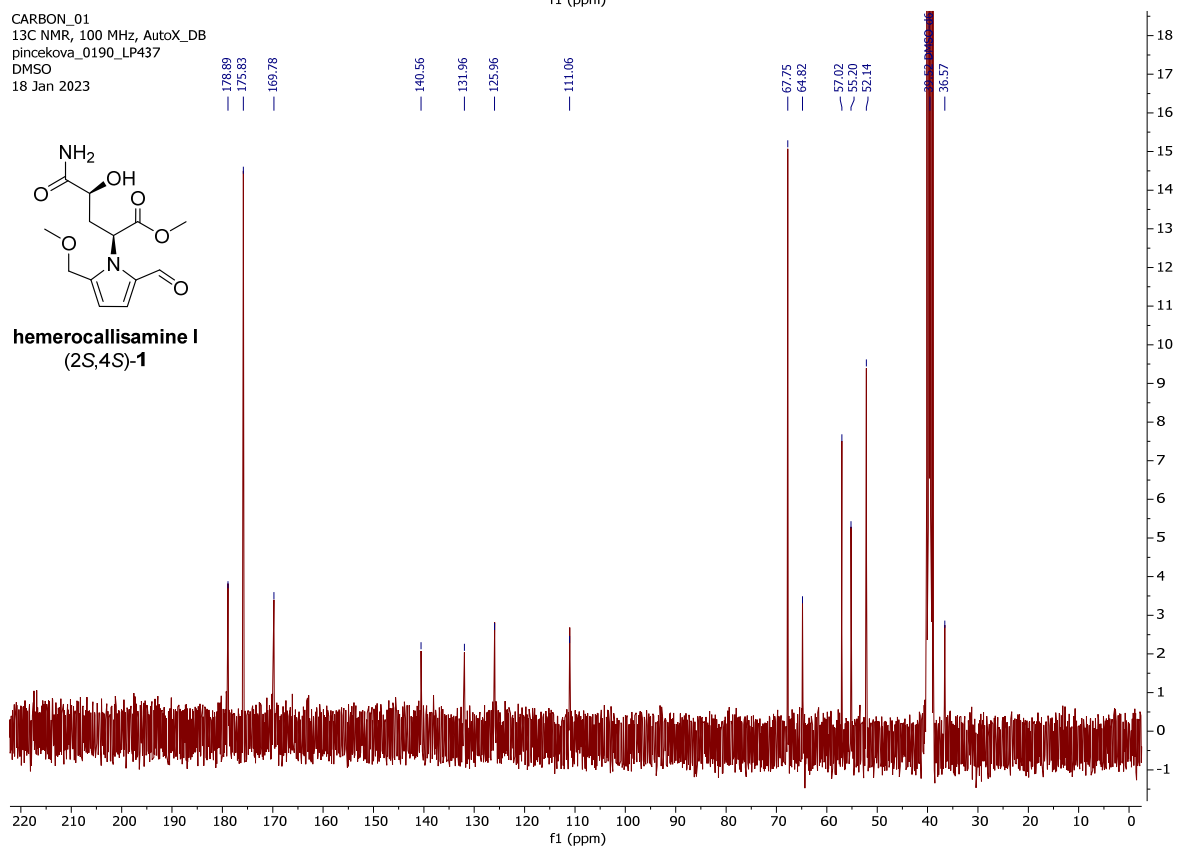
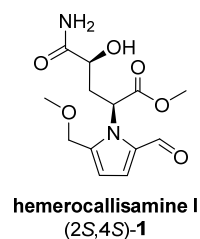
PROTON_01
 1H NMR, 400 MHz, AutoX_DB
 pincekova_0096_LP438
 CDCl3
 12 Jan 2023



PROTON_01
 1H NMR, 400 MHz, AutoX_DB
 pincekova_0190_LP437
 DMSO
 18 Jan 2023



CARBON_01
 13C NMR, 100 MHz, AutoX_DB
 pincekova_0190_LP437
 DMSO
 18 Jan 2023



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