

Supporting Information

BF₂-Azadipyrromethene Fluorophores for Intraoperative Vital Structure Identification

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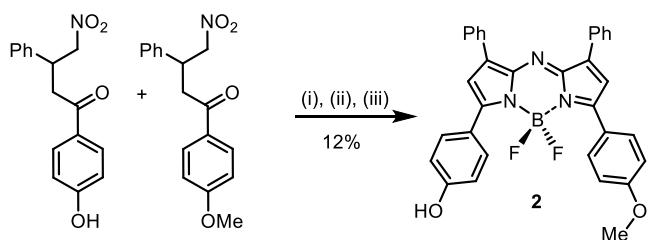


Figure S1. Alternate synthetic route to **2**. (i) NH₄OAc, EtOH, reflux, 16 h; (ii) BF₃.OEt₂, DIPEA, CH₂Cl₂ (anhydrous), rt, 16 h, (iii) silica gel purification, 12% over 2 steps.

Synthesis of BF₂ chelate of (Z)-4-(2-((5-methoxyphenyl)-3-phenyl-1*H*-pyrrol-2-yl)imino)-3-phenyl-2*H*-pyrrol-2*H*-pyrrol-5-yl)phenol (**2**)

1-(4-Hydroxyphenyl)-4-nitro-3-phenylbutan-1-one¹ (1.50 g, 5.26 mmol) and 1-(4-methoxyphenyl)-4-nitro-3-phenylbutan-1-one² (1.57 g, 5.26 mmol) were dissolved in EtOH (37 mL) and ammonium acetate (15.0 g, 194.50 mmol) was added. The reaction was heated for 16 h under reflux with stirring. The reaction was cooled to rt and was further cooled to 0 °C in an ice bath. The resulting precipitate was vacuum filtered, washed with cold EtOH (3 x 20 mL), sat. NaHCO₃ (3 x 20 mL) and H₂O (3 x 20 mL), and vacuum dried overnight to give a crude mixture of azadipyrromethene products (1.24 g) as a blue-black solid. The crude mixture (1.22 g, ~2.46 mmol) was dissolved in dry CH₂Cl₂ (120 mL) and diisopropylethylamine (4.29 mL, 24.62 mmol, 10.0 eq.) was added under N₂. BF₃-diethyletherate (4.25 mL, 34.47 mmol, 14.0 eq.) was added dropwise and the reaction stirred at rt for 16 h. The solvent was removed by rotary evaporation, the crude residue dissolved in EtOAc (120 mL) and washed with sat. NaHCO₃ (2 x 120 mL), H₂O (2 x 120 mL) and brine (2 x 120 mL). The combined organic layers were dried over Na₂SO₄, filtered and concentrated by rotary evaporation yielding a crude residue. The crude was dry loaded (CH₂Cl₂ and silica) onto a silica column and purified by silica gel chromatography eluting with a gradient of CH₂Cl₂ (100 %) going to CH₂Cl₂ / EtOAc (9:1) yielding the product **2** as a red metallic solid (340.1 g, 12 %), mp 145–147 °C. ¹H NMR (400 MHz, DMSO-d⁶) δ: 10.57 (s, 1H), 8.19 – 8.10 (m, 8H), 7.64 (s, 1H), 7.59 – 7.43 (m, 7H), 7.15 (d, J = 8.5 Hz, 2H), 6.96 (d, J = 8.4 Hz, 2H), 3.89 (s, 3H) ppm; ¹³C NMR (101 MHz, DMSO-d⁶) δ: 161.7, 161.4, 158.5, 156.1, 144.9, 144.0, 142.3, 141.2, 132.4, 132.0, 131.8, 131.6, 129.6, 129.3, 129.1, 129.0, 128.7, 123.5, 121.4, 120.1, 119.2, 116.0, 114.4, 55.6 ppm; ¹⁹F NMR (376 MHz, DMSO-d⁶) δ: -130.44 (q, 32.5 Hz) ppm. HRMS (ES): m/z calcd for C₃₃H₂₃BF₂N₃O₂ [M-H]⁻ 542.1863; found 542.1864.

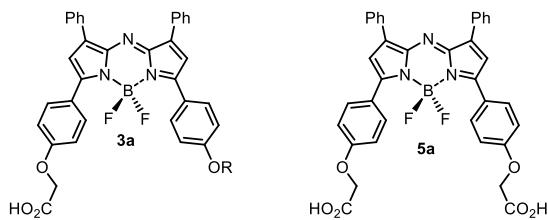
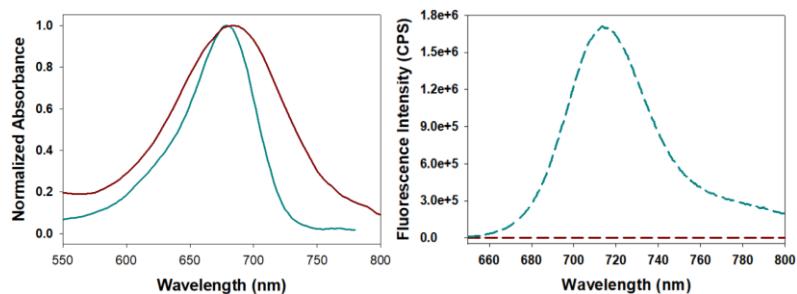
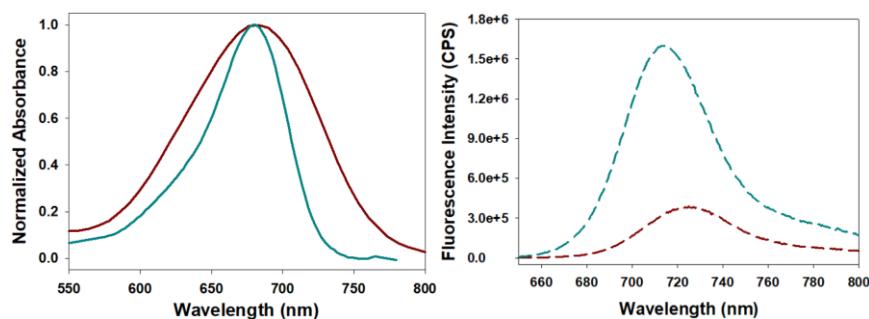


Figure S2. Structures of mono and bis-carboxylic acid functionalized fluorophores **3a** and **5a** employed in the synthesis of mono- and bis- activated ester fluorochromes **4** and **6**.

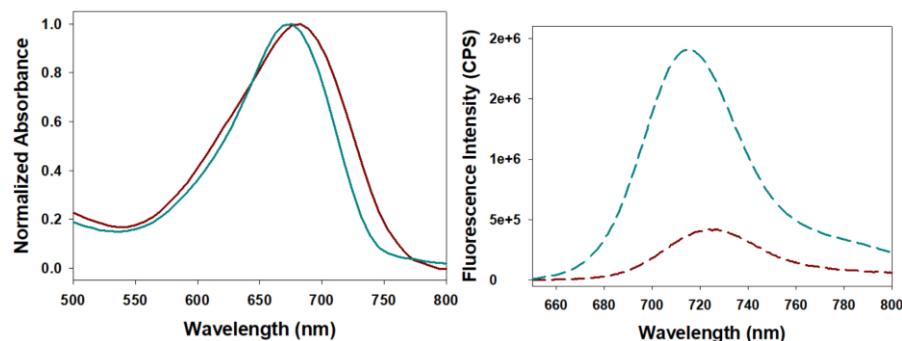
A



B



C



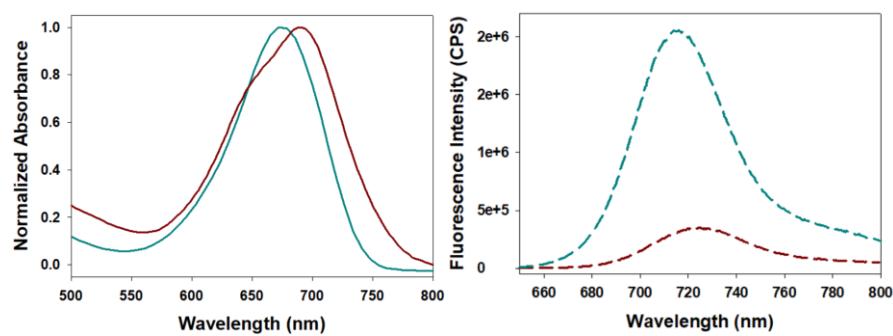
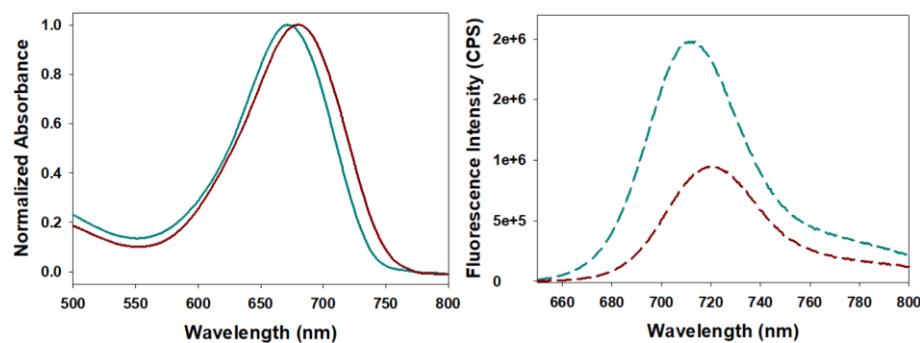
D**E**

Figure S3. Absorbance (left, normalized) and emission (right) spectra in CH₃CN (cyan) and H₂O (red) of (A) 7 ; (B) 8 ; (C) 9 ; D 10 ; (E) 11. Concentration 2 μ M with 2.5 nm slit widths used for fluorescence spectra.

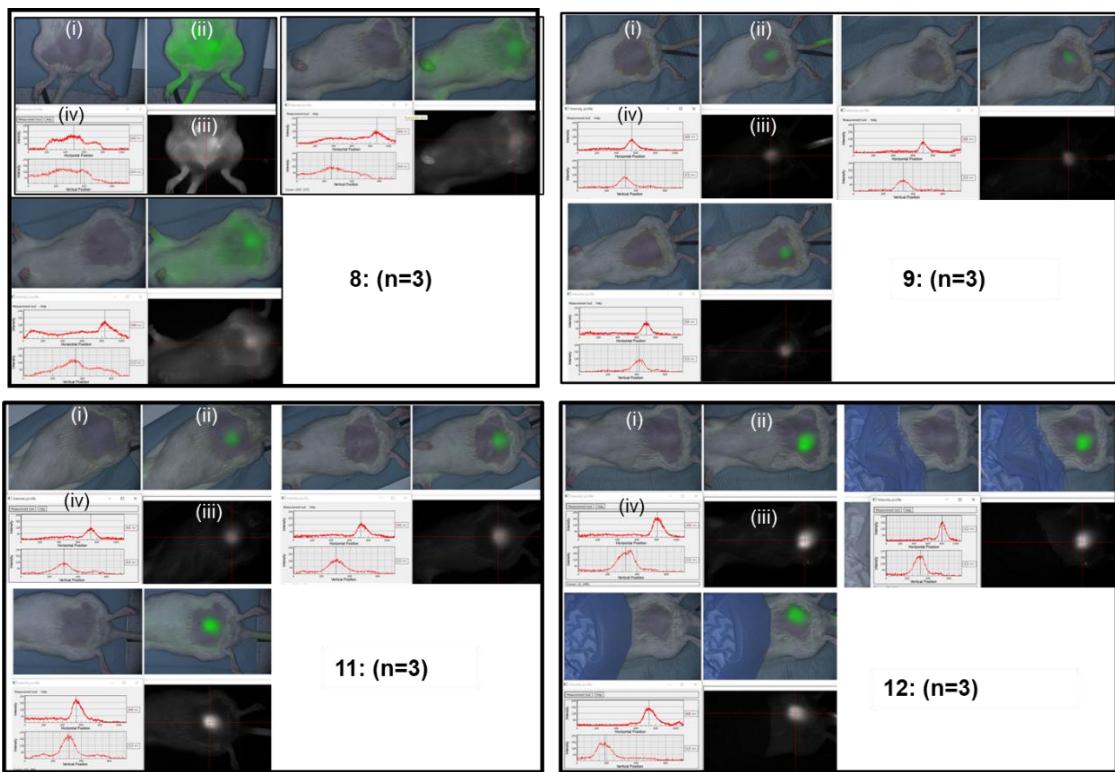


Figure S4. Representative images of the preliminary screening of fluorophores **8**, **9**, **11** and **12** ($n=3$, 1 mg/Kg). Fluorescence intensity from the bladder measured 60 min post administration (i) white light image; (ii) fluorescence (green) overlaid on white light image; (iii) fluorescence shown as black and white for clarity; (iv) averaged pixel analysis of fluorescence intensity through the bladder region of interest for **8** (128 au), **9** (98 au), **11** (155 au), **12** (186 au).

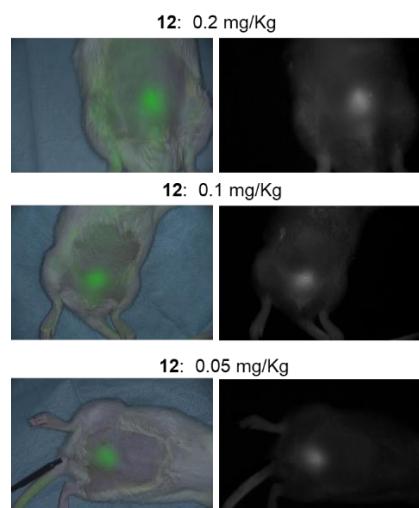


Figure S5. Representative images showing fluorescence from bladder for dose de-escalation study in rodents for **12**.

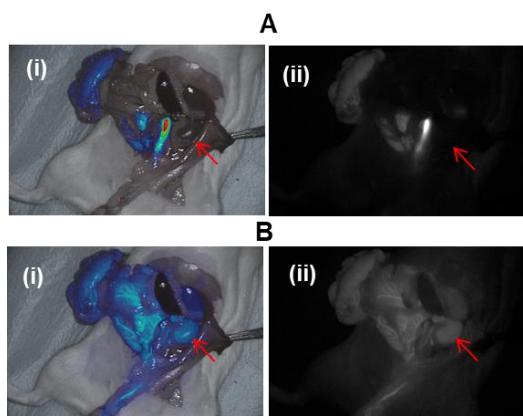


Figure S6. Fluorescence observed immediately upon tail vein administration of **12**. (A) at injection (B) 40 sec later. (i) fluorescence overlaid on white light image; (ii) fluorescence shown as black and white for clarity. Red arrow indicates a kidney.

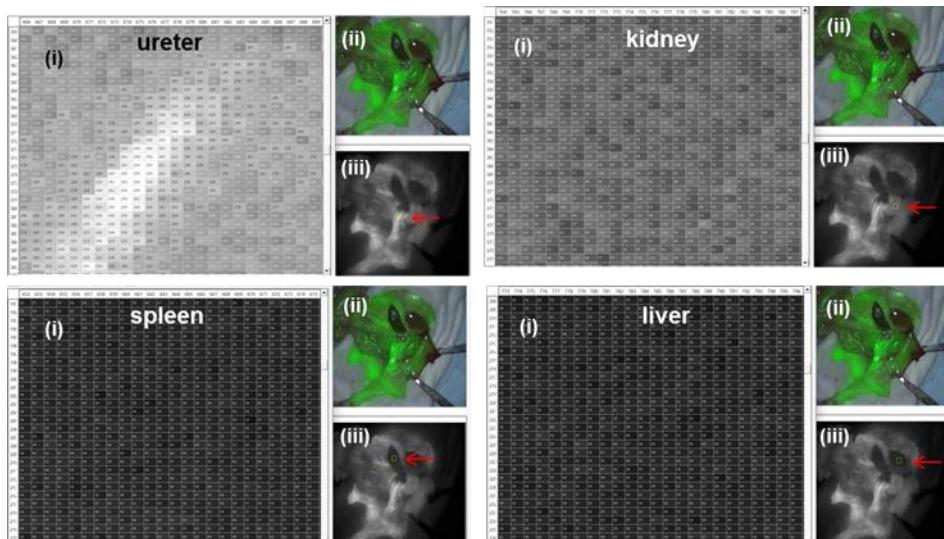


Figure S7. Representative pixel intensity analysis of four regions of interest taken 50 min post administration of **12**. Measured ureter (averaged intensity 167.7 au), kidney (99.3 au), spleen (35.1 au), liver (38.3 au) intensity values. (i) Pixel fluorescence intensity values for measured FOV in yellow box. (ii) fluorescence (green) overlaid on white light image; (iii) fluorescence shown as black and white for clarity. Red arrows indicates yellow box areas from which intensity values were recorded.

Table S1. Rodent urine study confirming renal excretion of **12**. Percentage collection of **12** is calculated from mmol of **12** injected versus mmol of **12** collected 90 min post injection.

entry	mg/kg of 12 injected	mmol of 12 injected	mmol of 12 in urine sample ^a	% of 12 collected
1	2.0	3.77×10^{-5}	1.44×10^{-5}	30
2	0.2	3.77×10^{-6}	1.24×10^{-6}	33
3	0.1	1.88×10^{-6}	5.66×10^{-7}	30
4	0.05	9.42×10^{-7}	2.4×10^{-7}	26

^a Calculated using measured urine volume and standard abs / emission curves of **12**.

Table S2. Percentage body weight contributions of brain, spleen kidneys and liver of male Sprague-Dawley rats on day 8 following total dose intravenous administrations of **12** at 14 (2 x 7 daily administrations) and 10 mg/kg for each dose versus control, n = 3.

Group	Dosage (mg/Kg)	Brain (%)	Spleen (%)	Kidney (%)	Liver (%)
Control (PSB)	0	0.77	0.21	0.86	3.17
12	14 (2 x 7)	0.73	0.27	0.87	3.36
12	10	0.69	0.24	0.95	3.38

Table S3. Selected blood chemistry data for male Sprague-Dawley rats following intravenous administration of **12** at 14 mg/Kg (2 mg/Kg per day for 7 days) and a single dose at 10 mg/kg, n = 3.

Group	AST IU/L	ALT IU/L	Creatinine μ mol/L	Urea mmol/L	Glucose mmol/L
Control (vehicle)	79 (17.1)	50 (7.9)	26.3 (2.3)	4.4 (0.2)	6.49 (0.6)
12 (14 mg/Kg)	69 (3.5)	43 (1.2)	26.0 (1.2)	5.5 (0.4)	6.3 (0.8)
12 (10 mg/Kg)	69 (0.6)	46 (3.6)	25.0 (1.3)	4.6 (0.9)	6.42 (0.7)

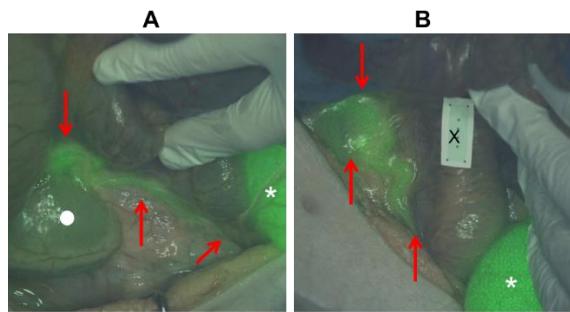
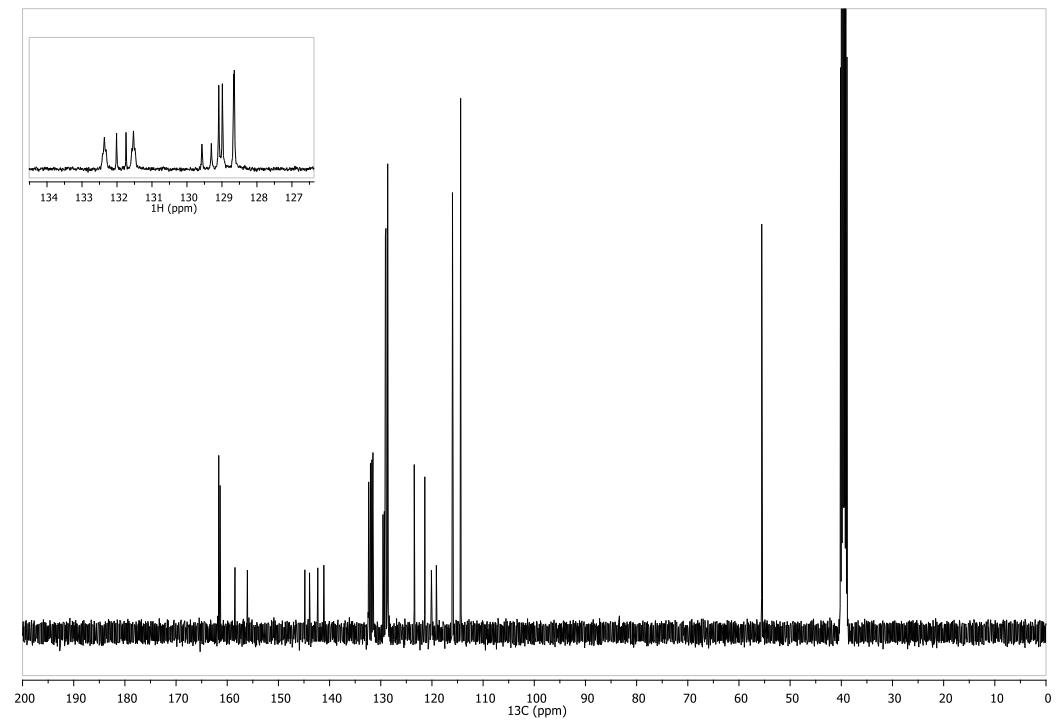
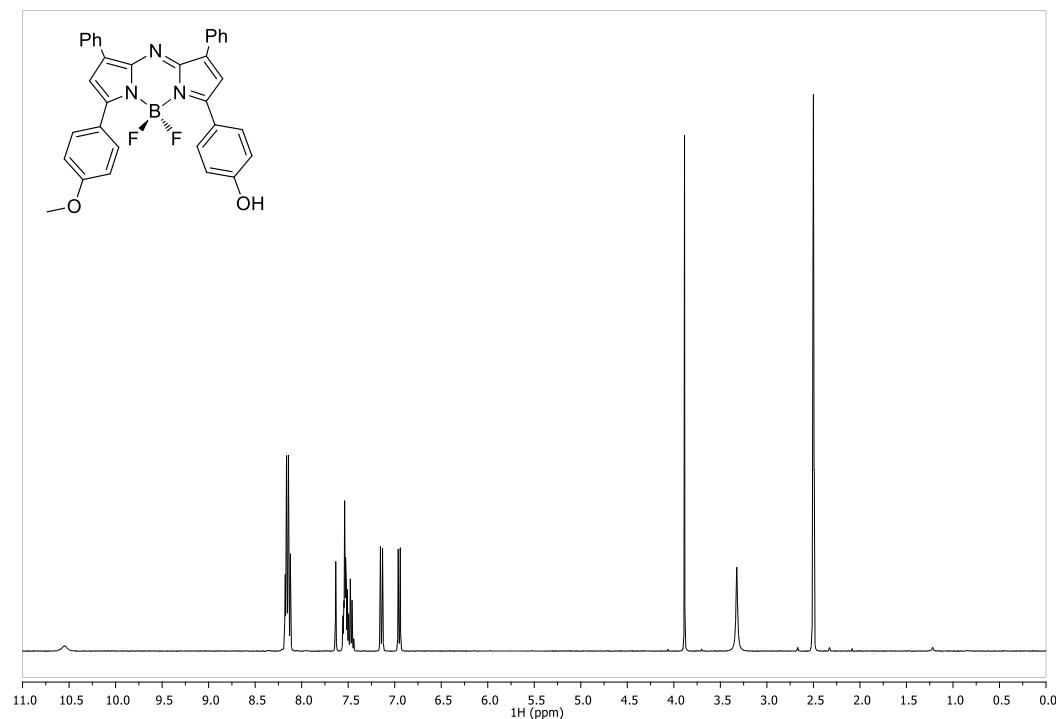
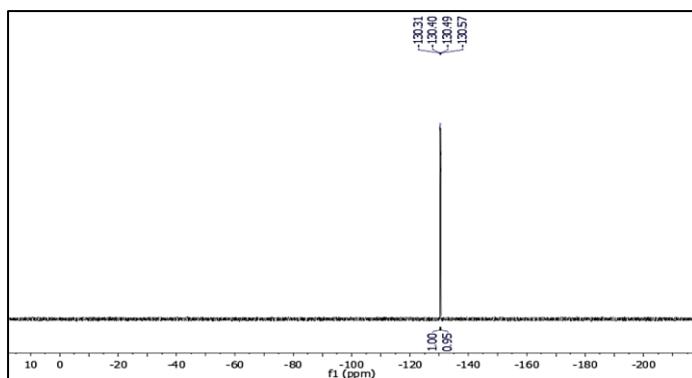


Figure S8. Additional representative images showing ureter identification in porcine model following treatment with **12** (0.5 mg/kg). (A) 25 min post administration (B) 120 min post administration. Red arrows indicate ureter, circle indicates kidney.

Figure S9: Analytical spectra for **2**.



¹⁹F NMR (376 MHz, DMSO-*d*⁶) of **2**



HRMS (ESI⁻) of **2**

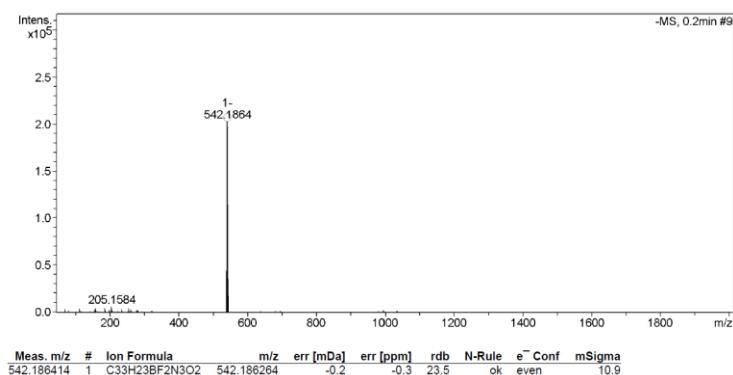
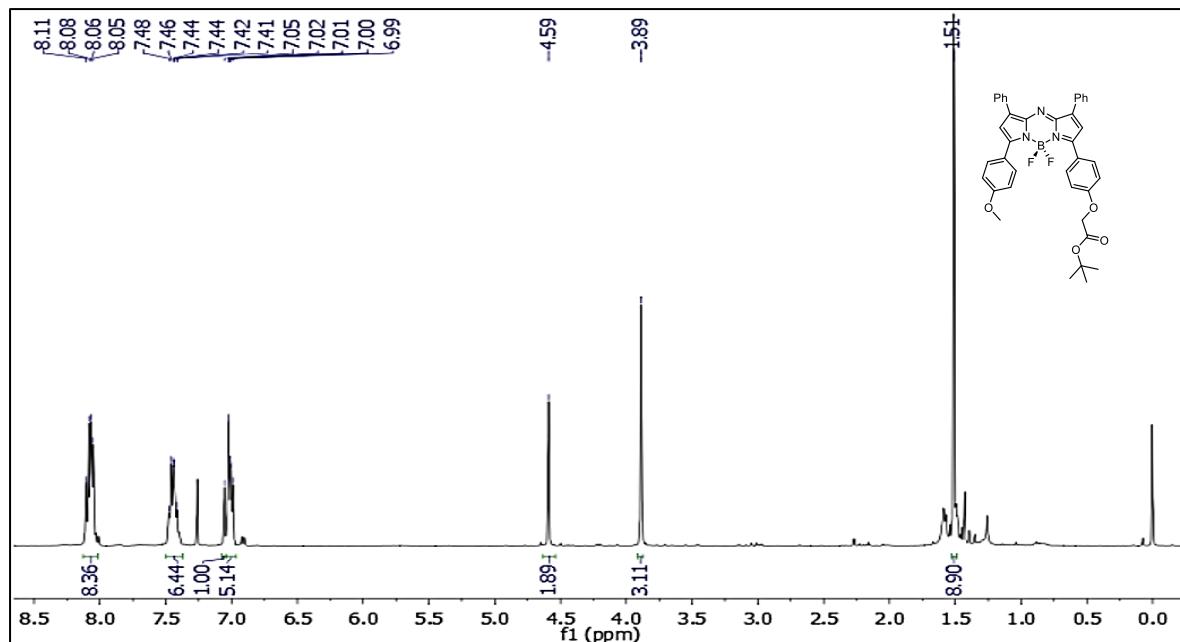
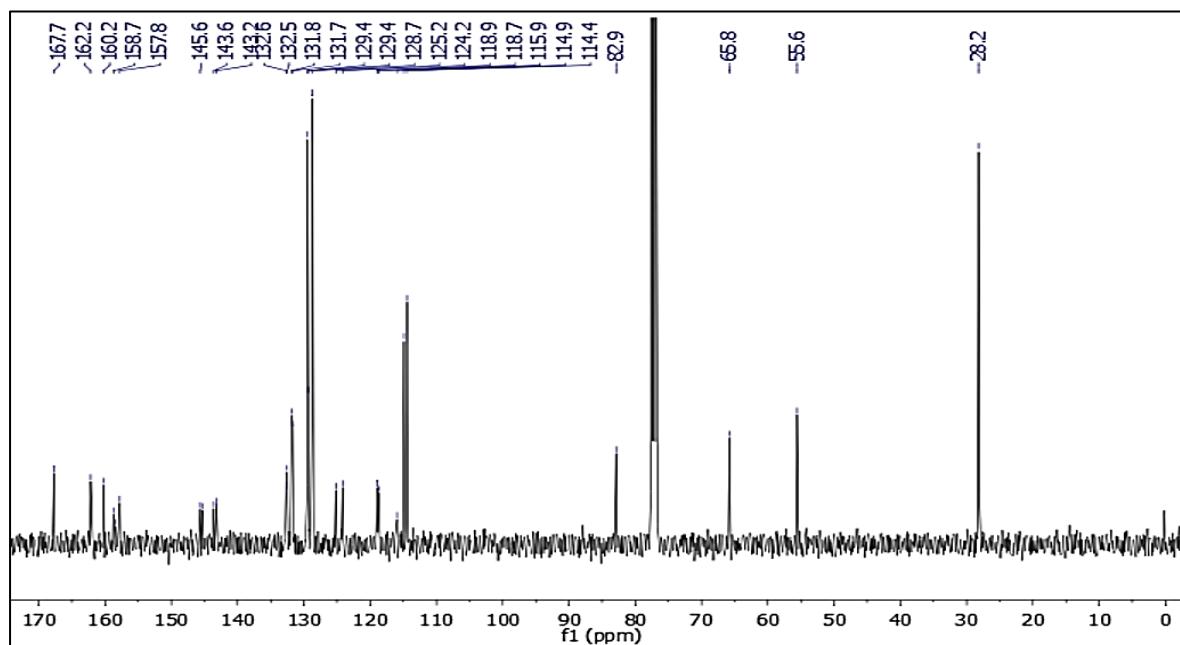


Figure S10: Analytical spectra for **3**

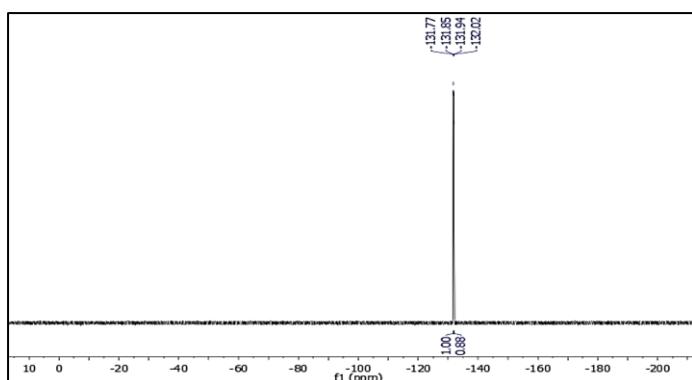
¹H NMR (400 MHz, CDCl₃)



¹³C NMR (101 MHz, CDCl₃)



¹⁹F NMR (376 MHz, CDCl₃)



HRMS (ESI⁺) of **3**

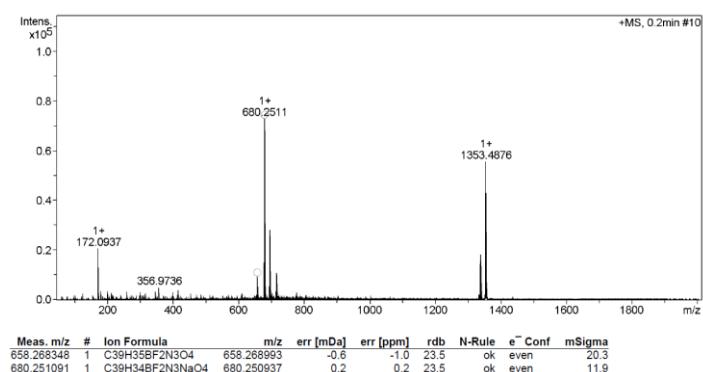
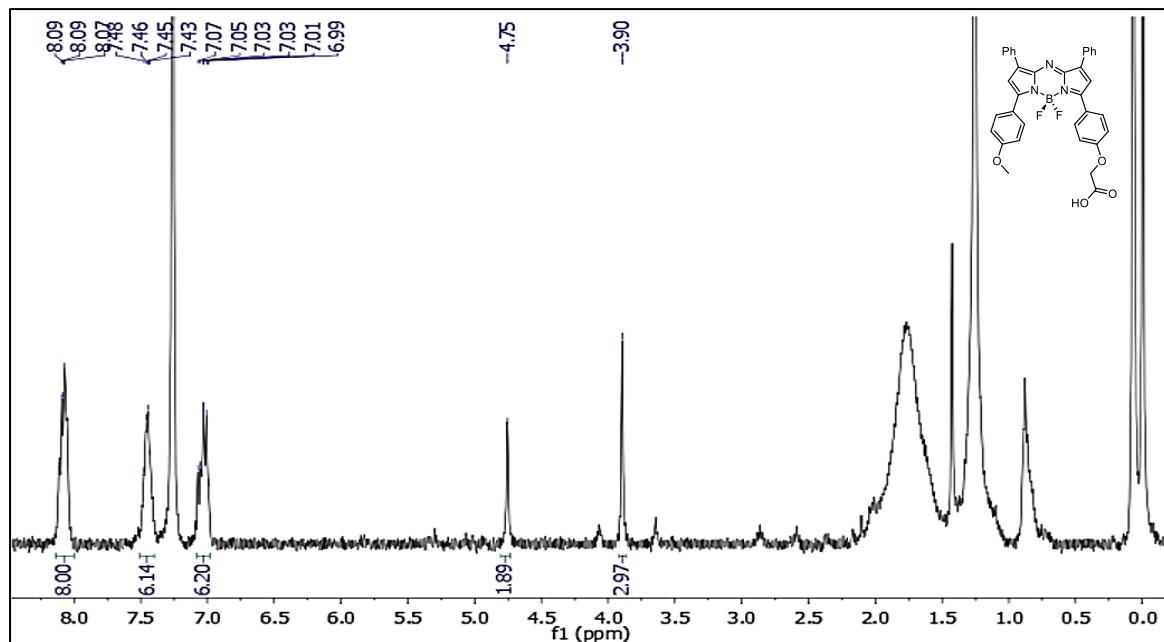
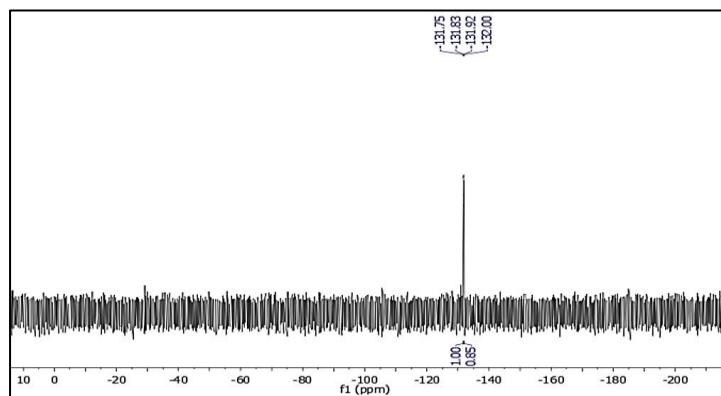


Figure S11: Analytical spectra for **3a**

¹H NMR (400 MHz, DMSO-*d*⁶)



¹⁹F NMR (376 MHz, CDCl₃)



HRMS (ESI⁻) for **3a**

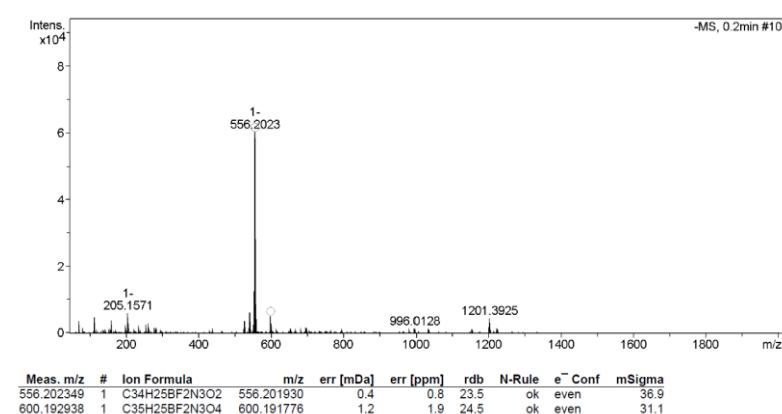
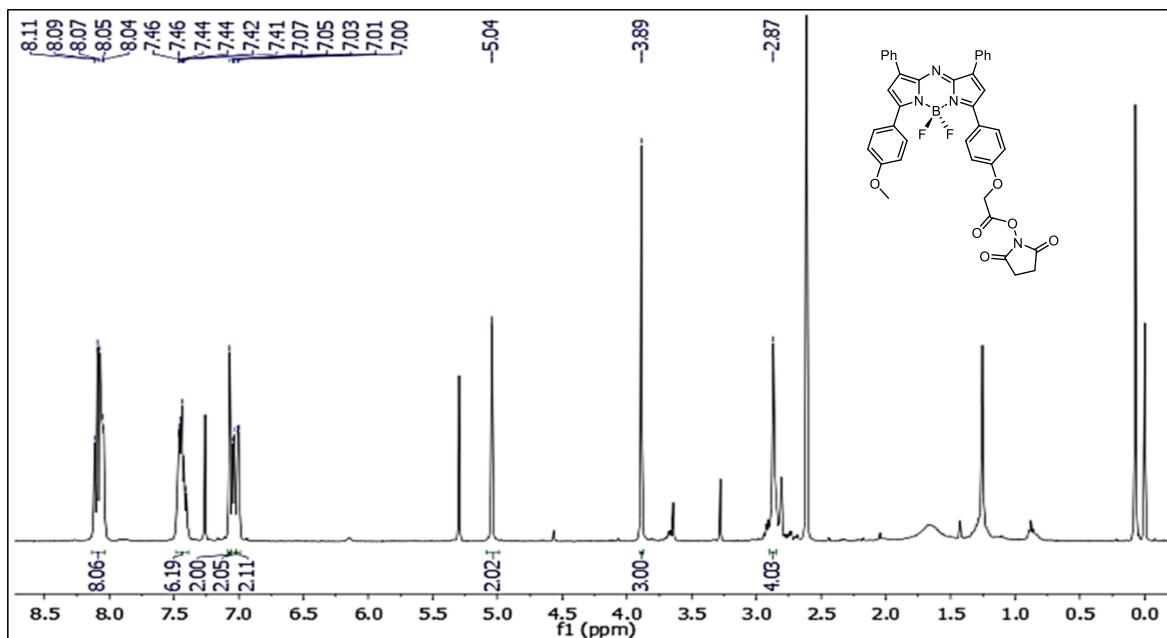
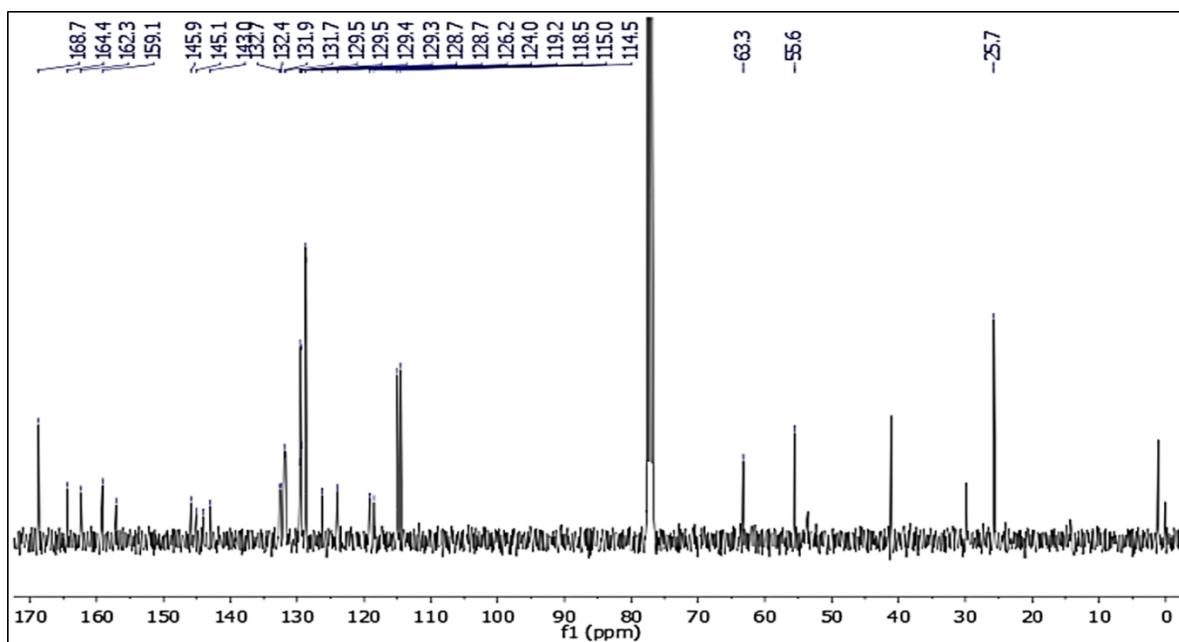


Figure S12: Analytical spectra for **4**

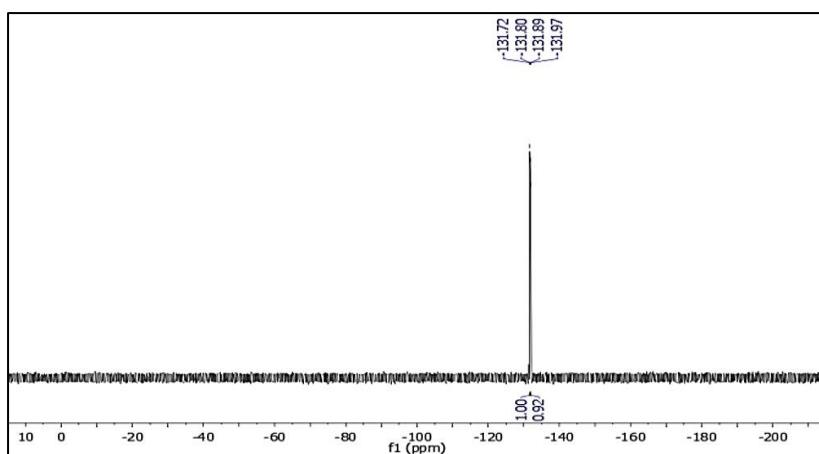
¹H NMR (400 MHz, CDCl₃)



¹³C NMR (101 MHz, CDCl₃)



¹⁹F NMR (376 MHz, CDCl₃)



HRMS (ESI⁺) for **4**

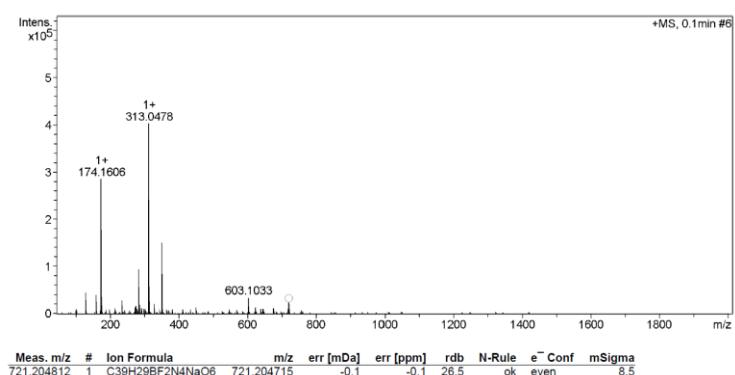
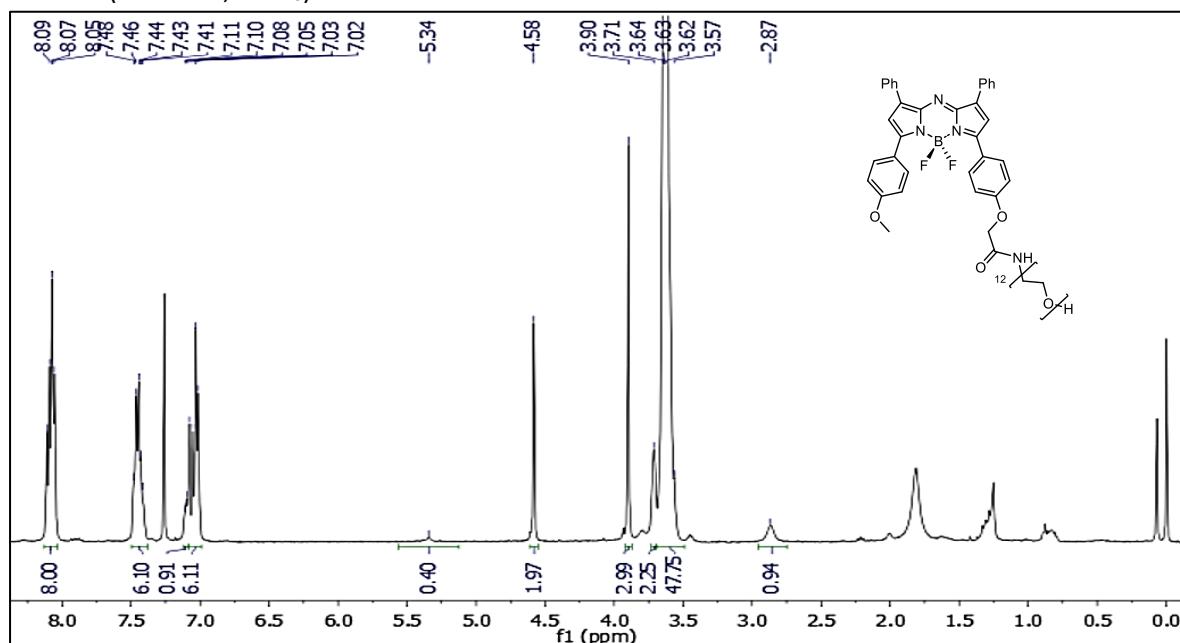
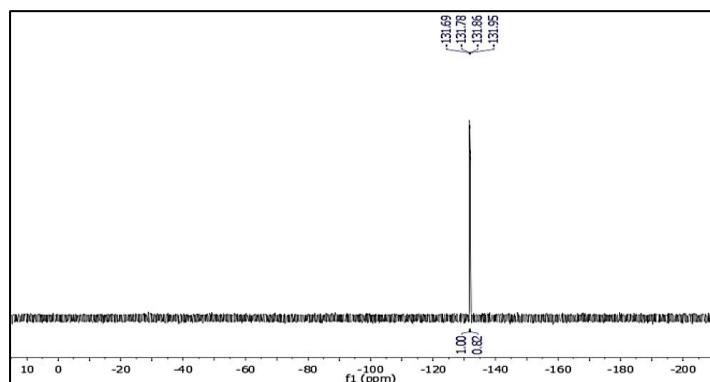


Figure S13: Analytical spectra for **7**

^1H NMR (400 MHz, CDCl_3)



^{19}F NMR (376 MHz, CDCl_3)



MALDI (Q-TOF) for **7**

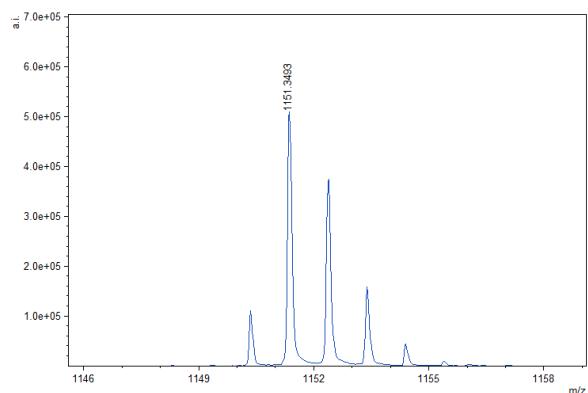
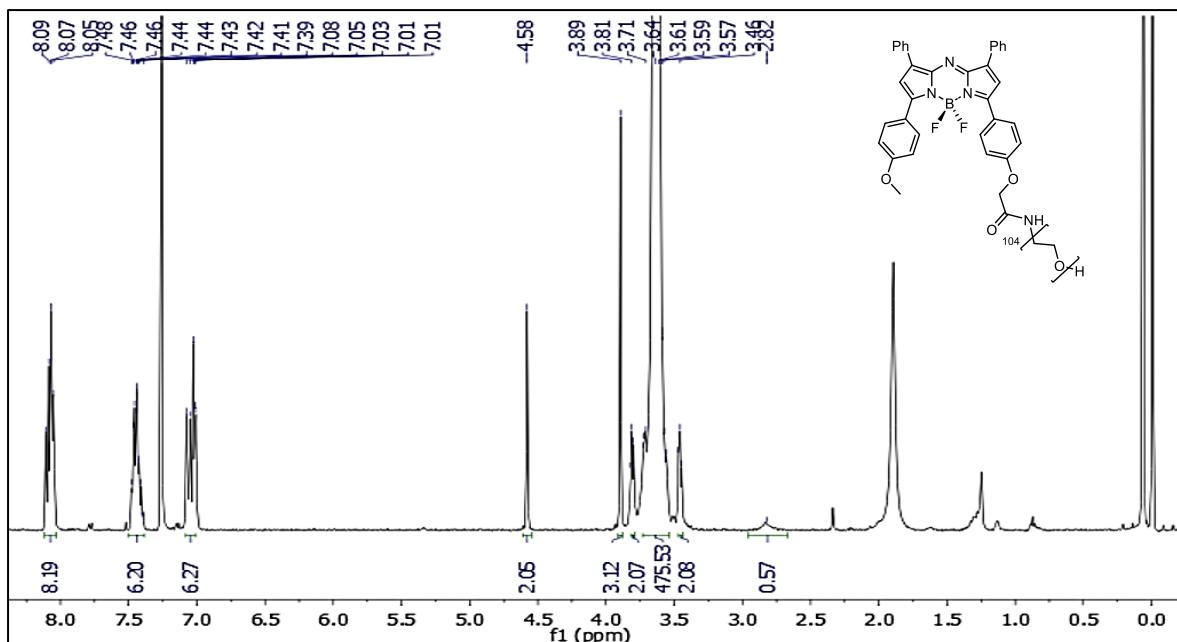
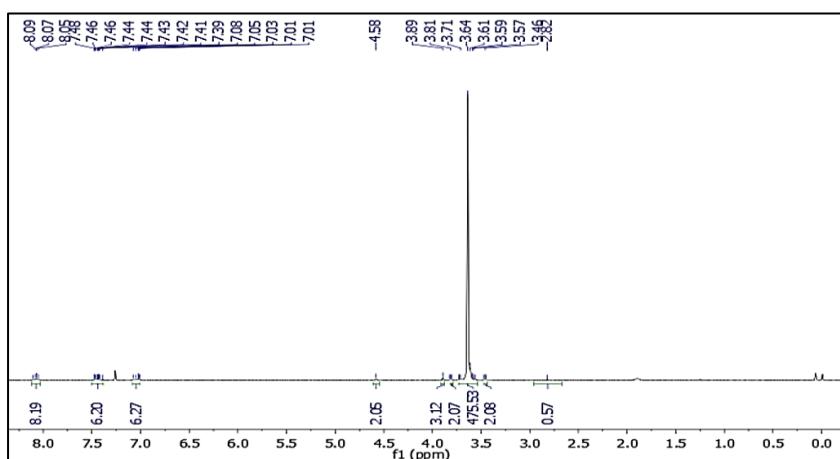


Figure S14: Analytical spectra for **8**

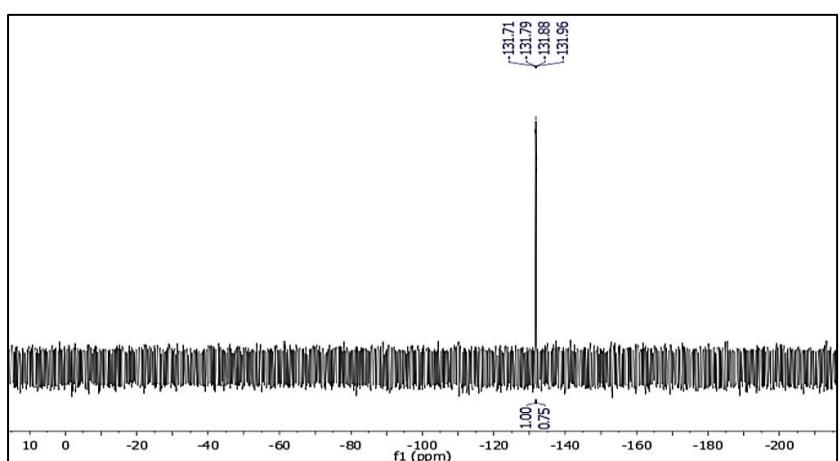
¹H NMR (400 MHz, CDCl₃)



¹H NMR (400 MHz, CDCl₃)



¹⁹F NMR (376 MHz, CDCl₃)



MALDI (Q-TOF) for **8**

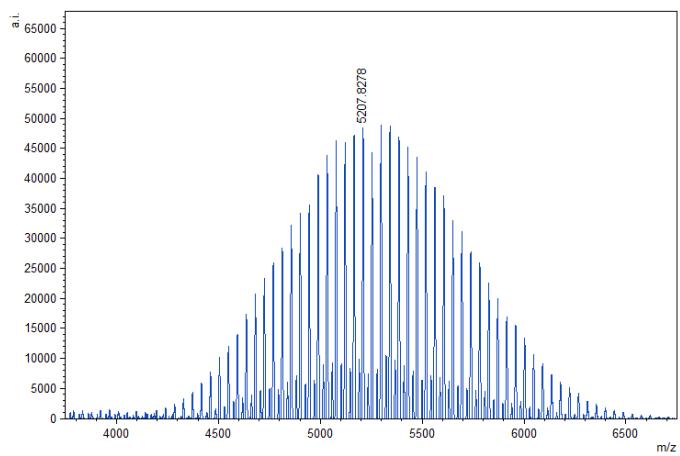
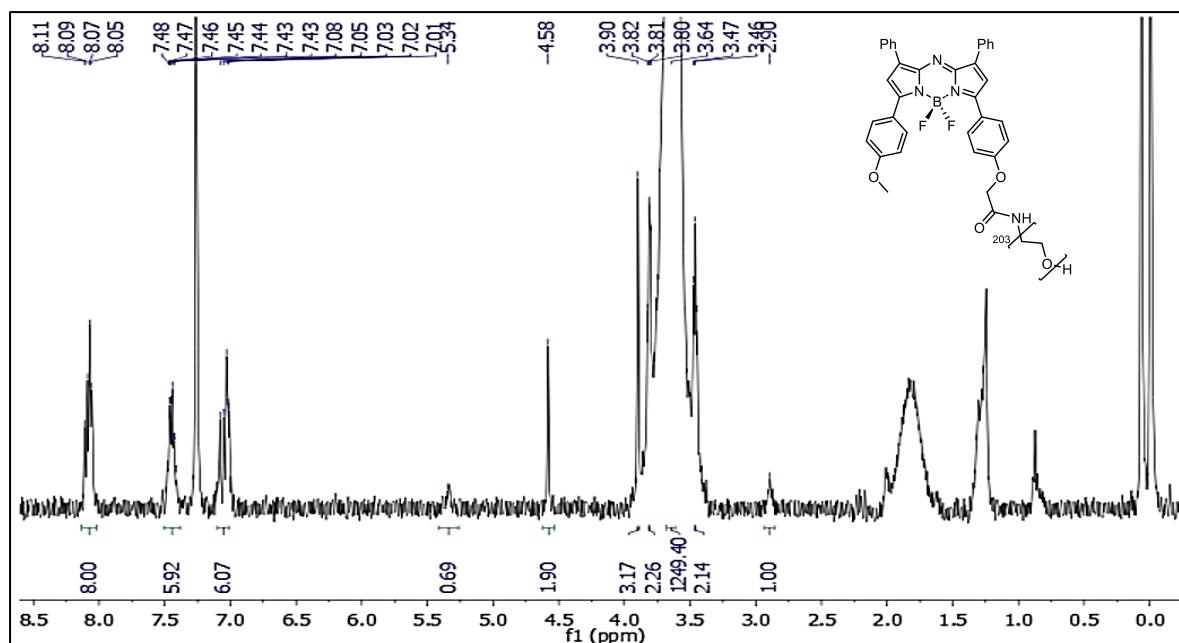
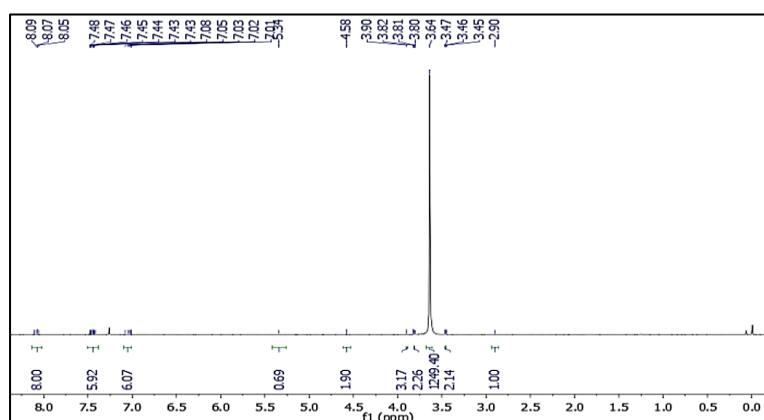


Figure S15: Analytical spectra for 9

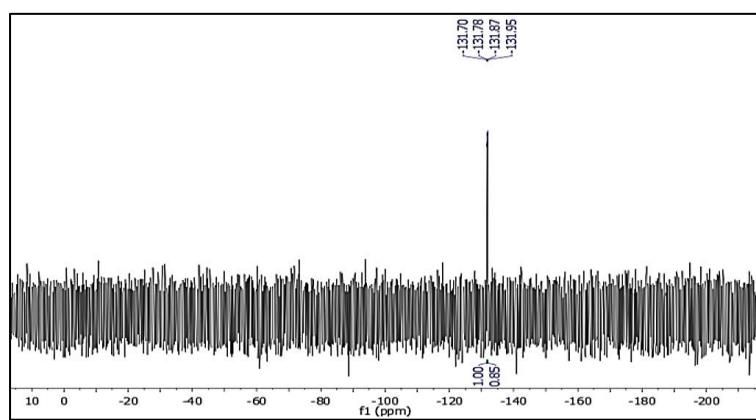
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¹H NMR (400 MHz, CDCl₃)



¹⁹F NMR (376 MHz, CDCl₃)



MALDI (Q-TOF) for **9**

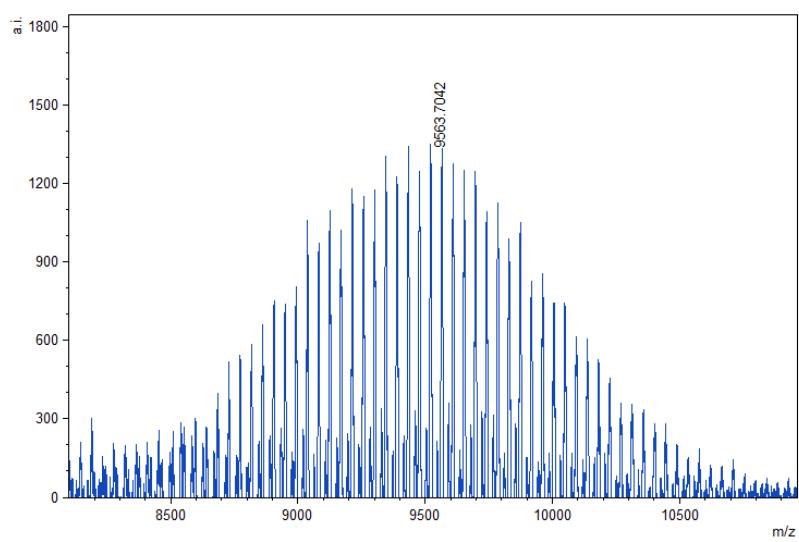
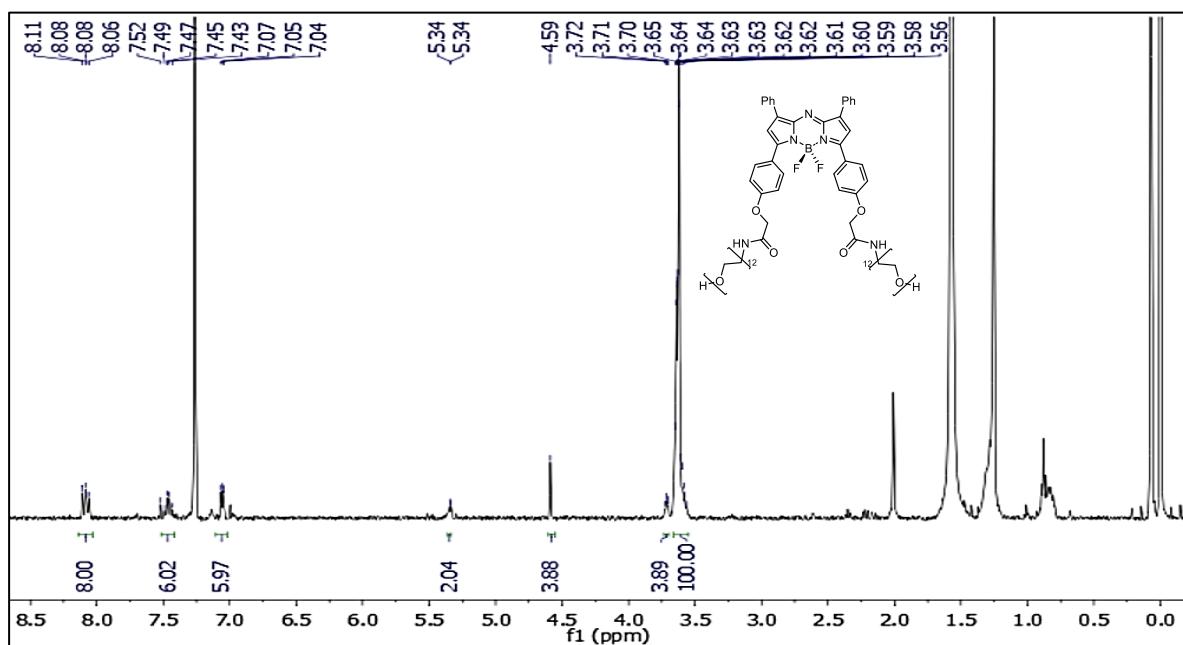
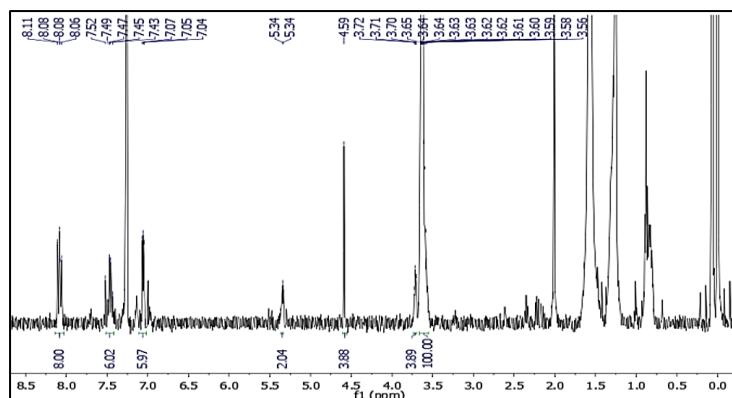


Figure S16: Analytical spectra for **10**

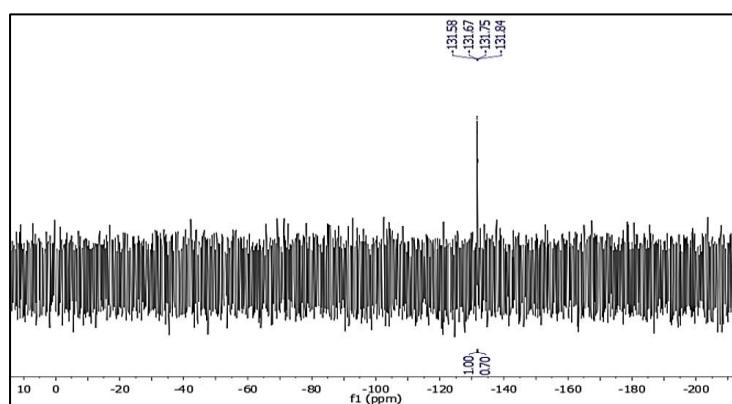
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¹H NMR (400 MHz, CDCl₃)



¹⁹F NMR (376 MHz, CDCl₃)



MALDI (Q-TOF) for **10**

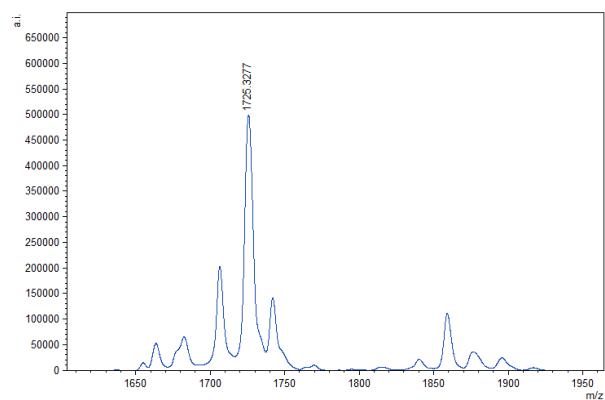
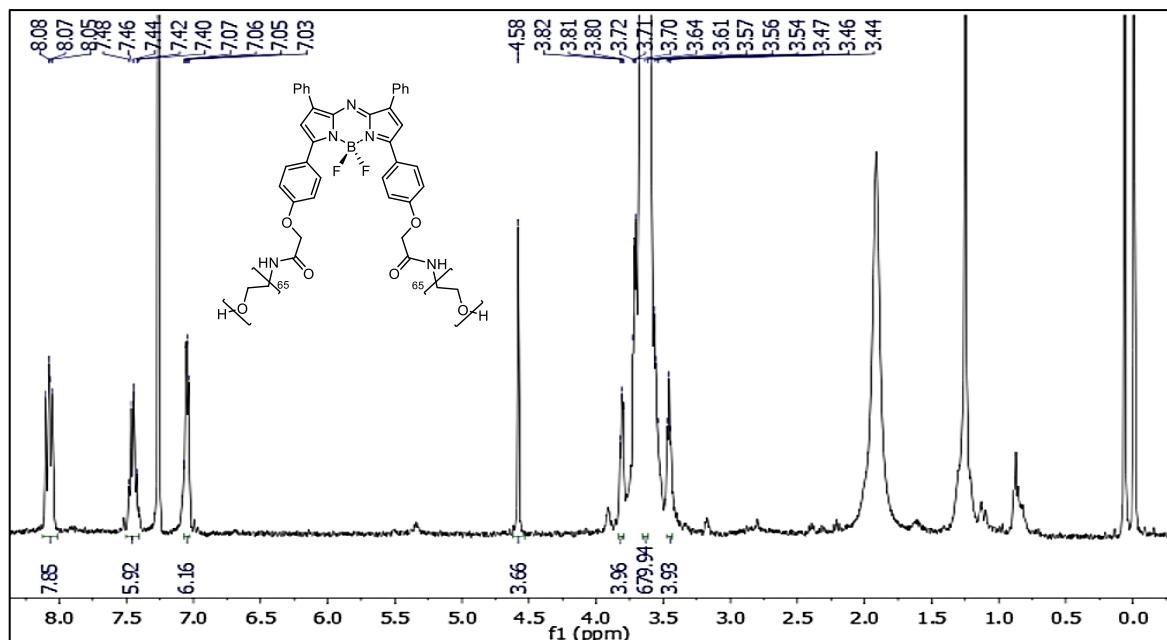
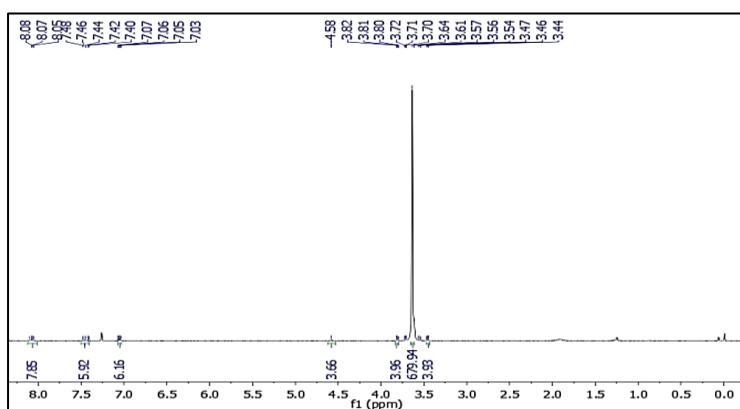


Figure S17: Analytical spectra for **11**

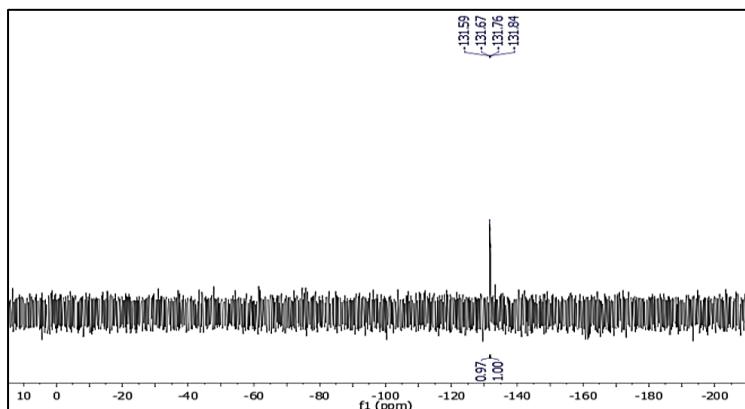
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¹H NMR (400 MHz, CDCl₃)



¹⁹F NMR (376 MHz, CDCl₃)



MALDI (Q-TOF) for **11**

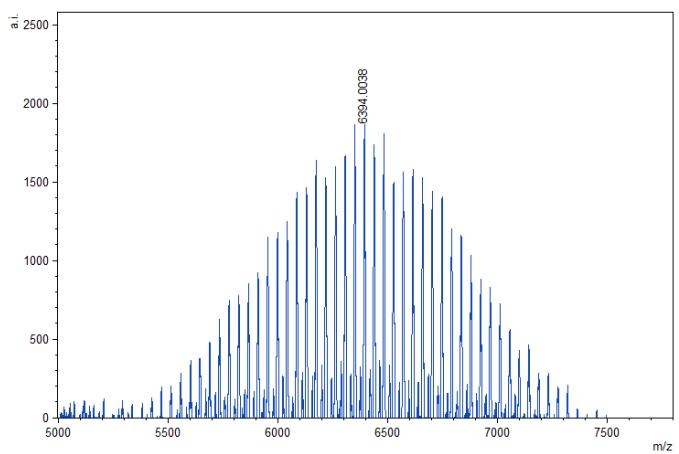
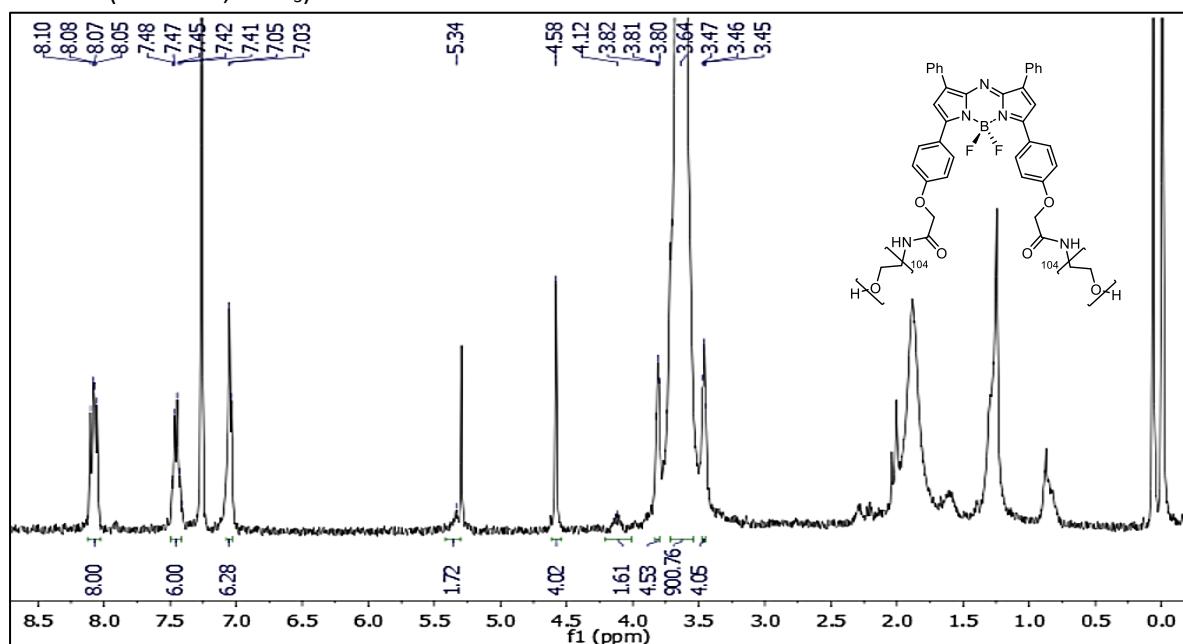
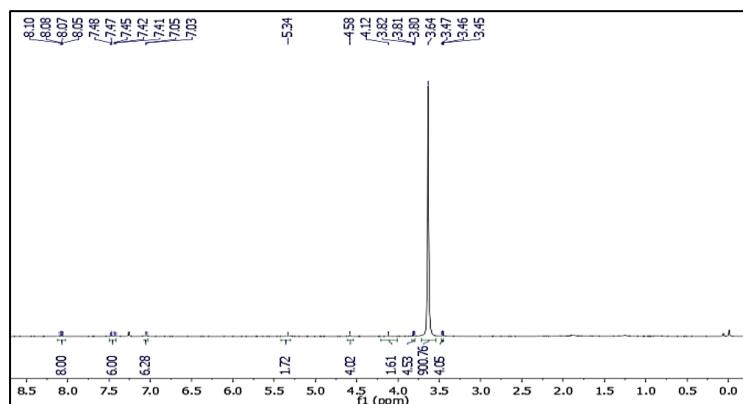


Figure S18: Analytical spectra for **12**

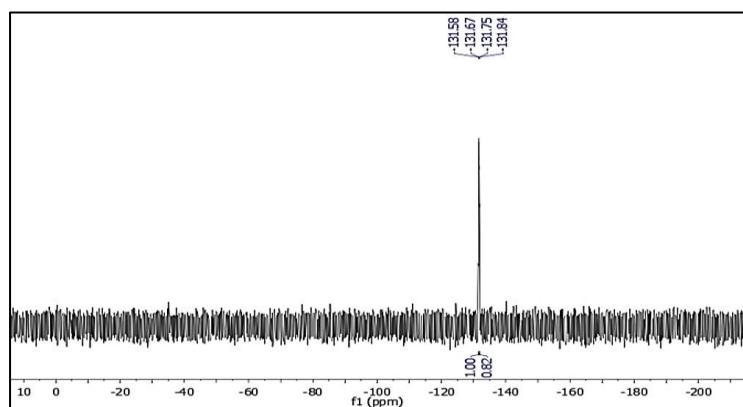
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¹H NMR (400 MHz, CDCl₃)



¹⁹F NMR (376 MHz, CDCl₃)



MALDI (Q-TOF) of **12**

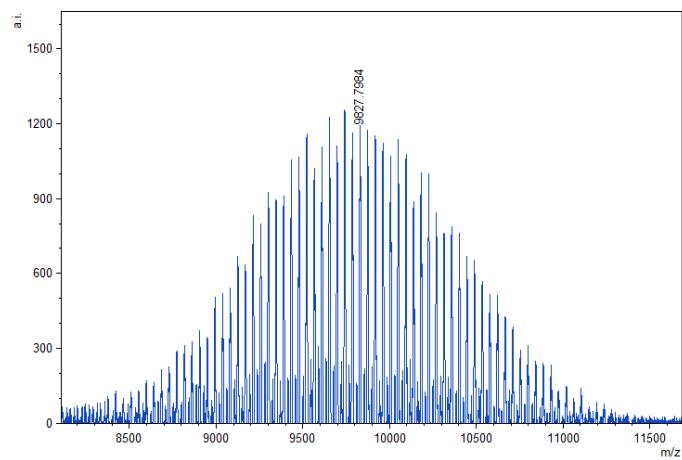
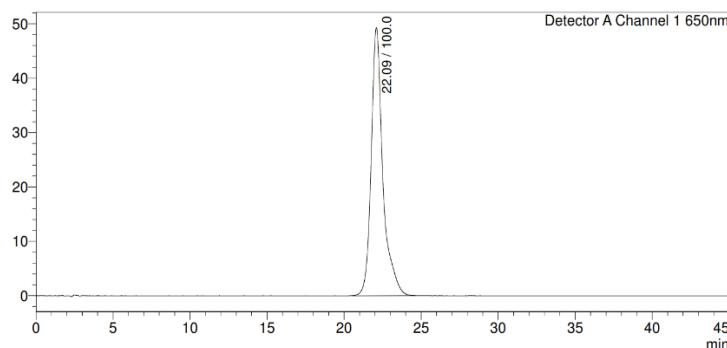
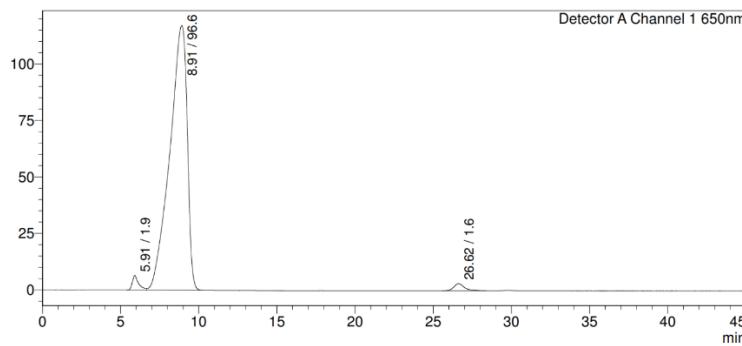


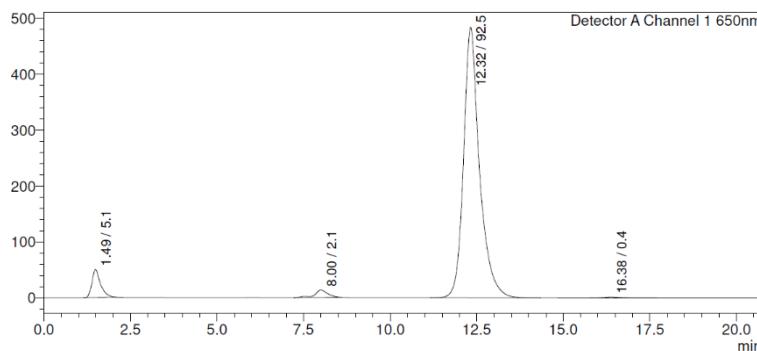
Figure S19: Analytical HPLC traces of **3**, **3a**, **4** and **7 – 12**.



HPLC Trace of **3**. Conditions: Reverse phase-HPLC with YMC triart phenyl column and size: 150 × 4.6 mm I.D., particle size: S-5 μ m, 12 nm hole, detection method: UV-Vis and wavelength for detection: 650 nm and 600 nm. Eluent ACN:H₂O = 70:30 with a flow rate at 1 mL/min.

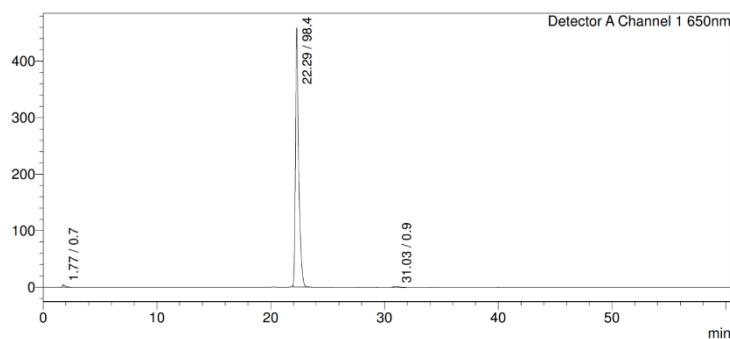


HPLC Trace of **3a**. Conditions: As above for **3**. Elution gradient of ACN:H₂O (40:60 going to 70:30) with a flow rate at 1 mL/min.

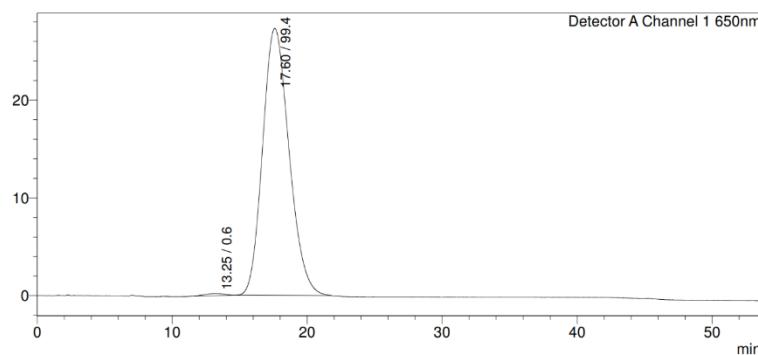


HPLC Trace of **4**. Conditions: As above for **3**. Eluent ACN:H₂O = 70:30 with a flow rate at 1 mL/min.

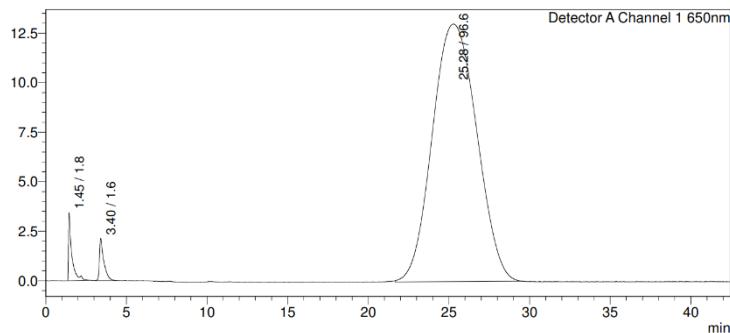
Mono-O-PEG(1.1kDa) 7



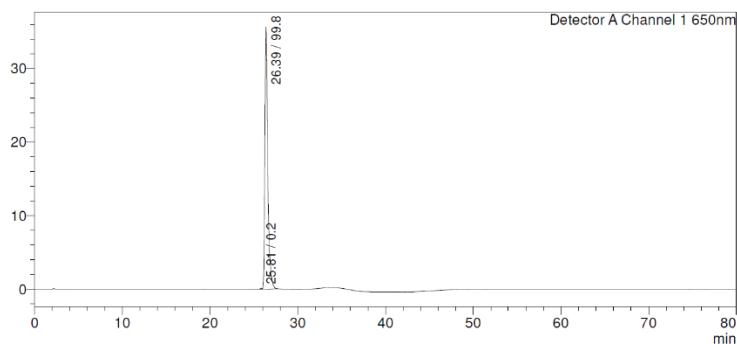
HPLC Trace of **7**. Conditions: As above for **3**. Elution gradient of ACN:H₂O (50:50 going to 70:30) with a flow rate at 1 mL/min.



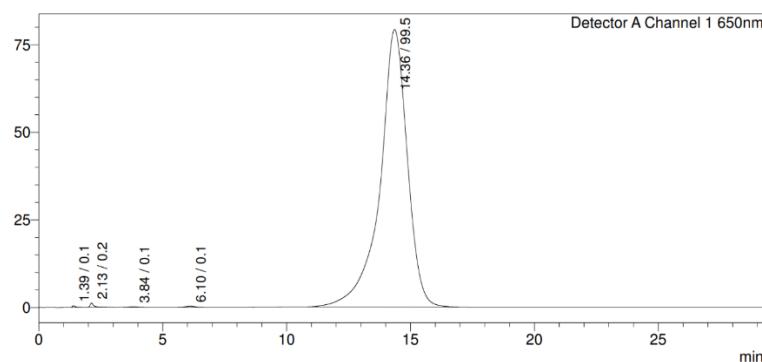
HPLC Trace of **8**. Conditions: As above for **3**. Elution gradient of ACN:H₂O (47.5:52.5 going to 70:30) with a flow rate at 1 mL/min.



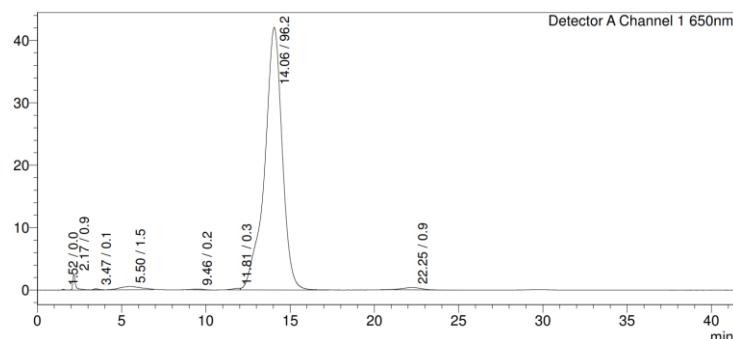
HPLC Trace of **9**. Conditions: As above for **3**. Elution gradient of ACN:H₂O (40:60 going to 70:30) with a flow rate at 1 mL/min.



HPLC Trace of 10. Conditions: As above for **3**. Elution gradient of ACN:H₂O (42.5:57.5 going to 70:30) with a flow rate at 1 mL/min.



HPLC Trace of 11. Conditions: As for **3** above. Elution gradient of ACN:H₂O (45:55 going to 70:30) with a flow rate at 1 mL/min.



HPLC Trace of 12. Conditions: As above for **3**. Elution gradient of ACN:H₂O (45:55 going to 70:30) with a flow rate at 1 mL/min.

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