

Supplementary material

For

First-row Transition Metal Complexes Incorporating the 2-(2'-pyridyl)quinoxaline Ligand (pqx), as Potent Inflammatory Mediator Inhibitors: Cytotoxic Properties and Biological Activities against the Platelet-Activating Factor (PAF) and Thrombin.

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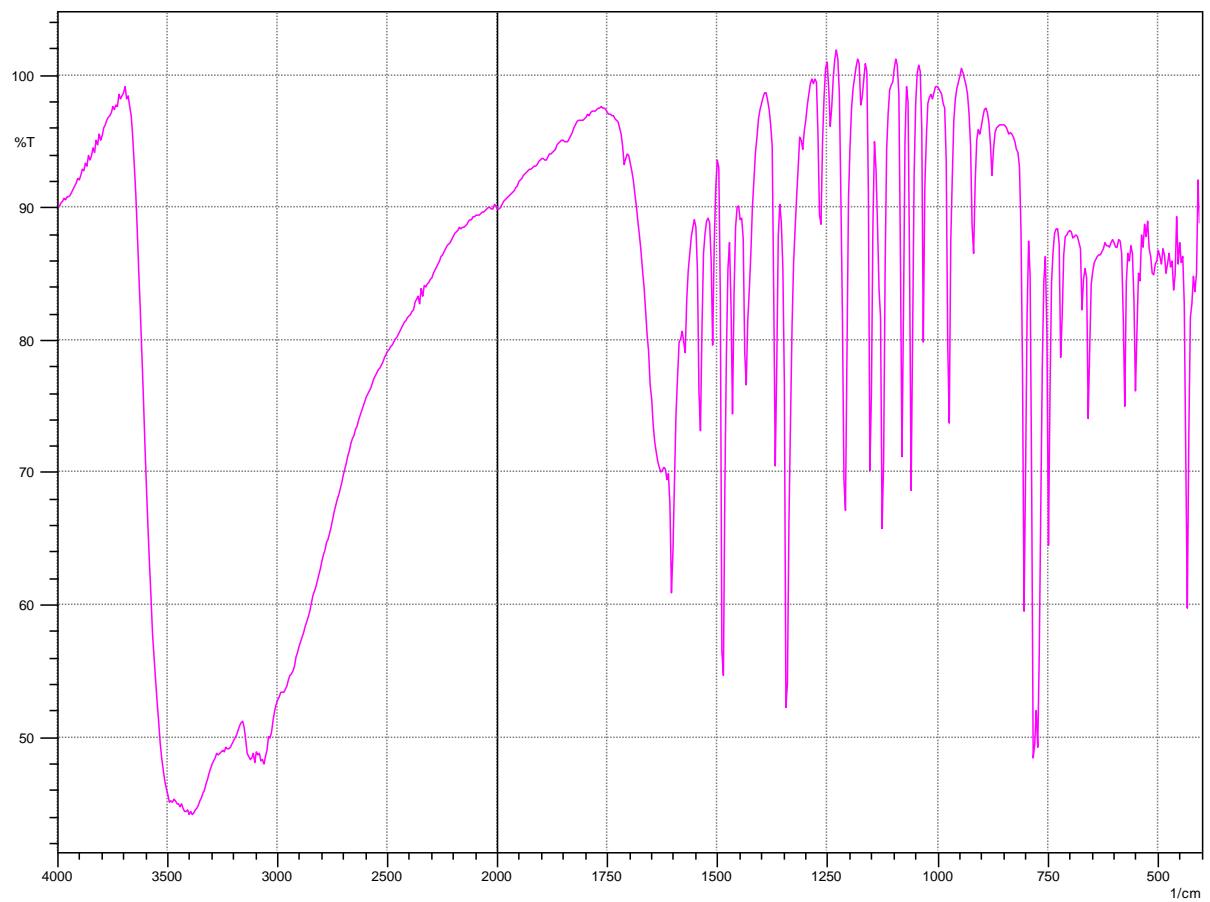
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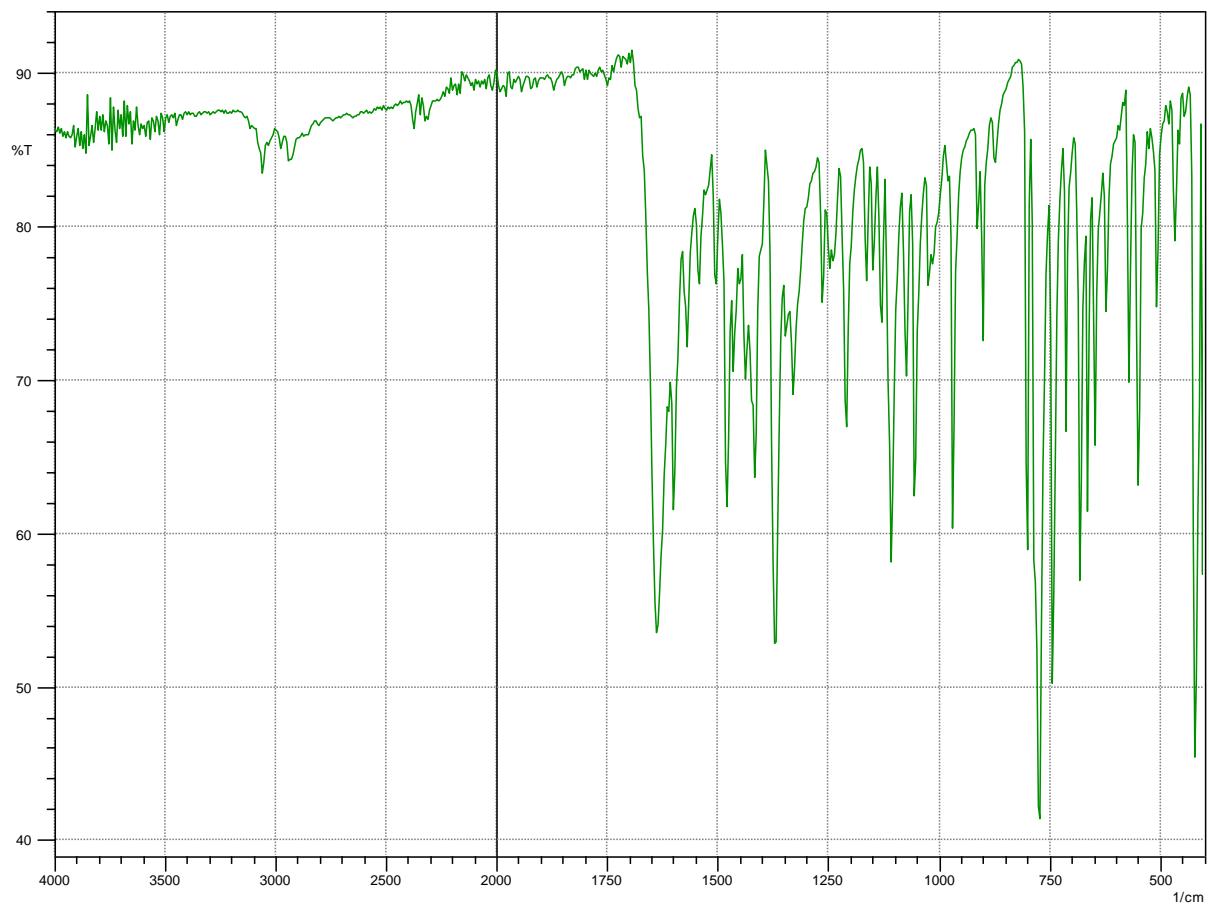
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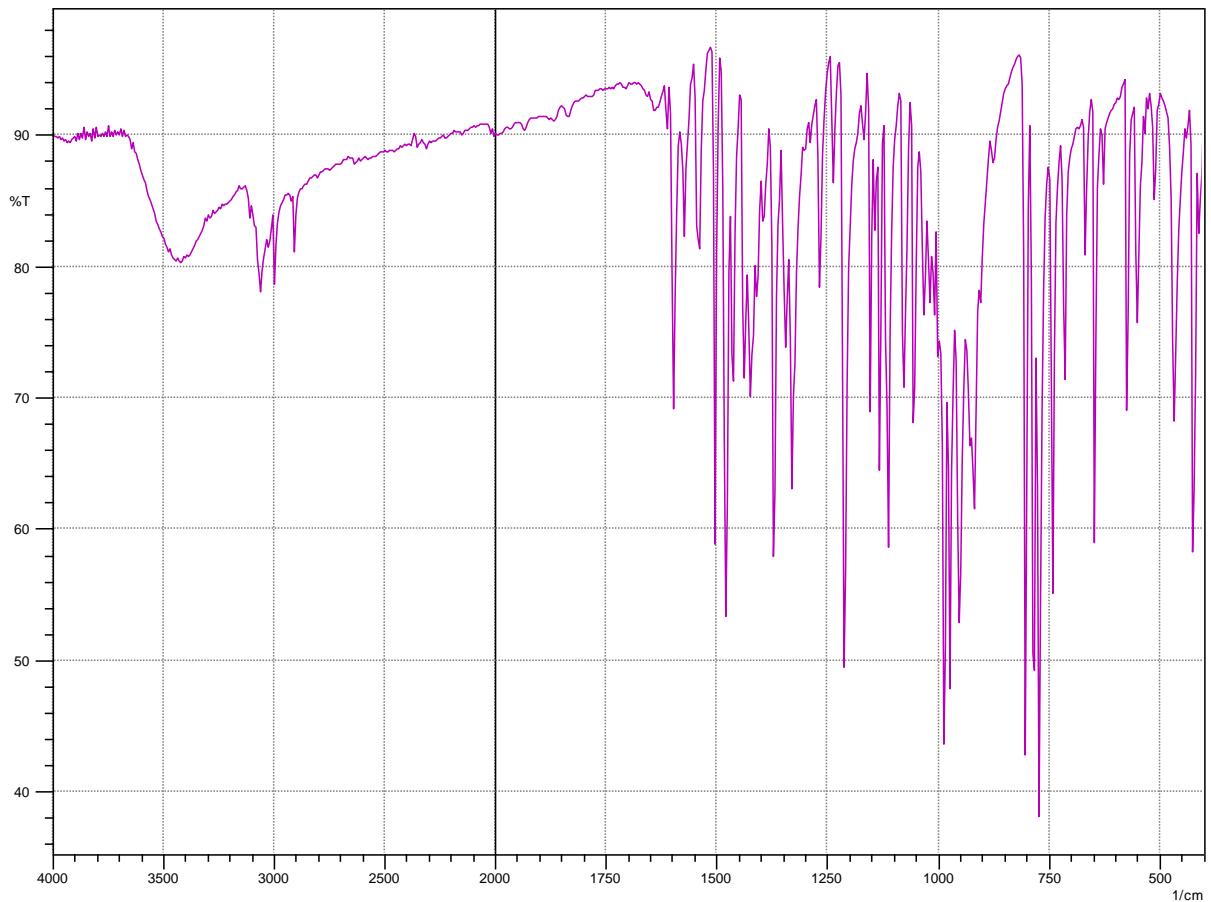
1) **Fig. S1.** FTIR spectra of Cr-pqx (**1**) in KBr



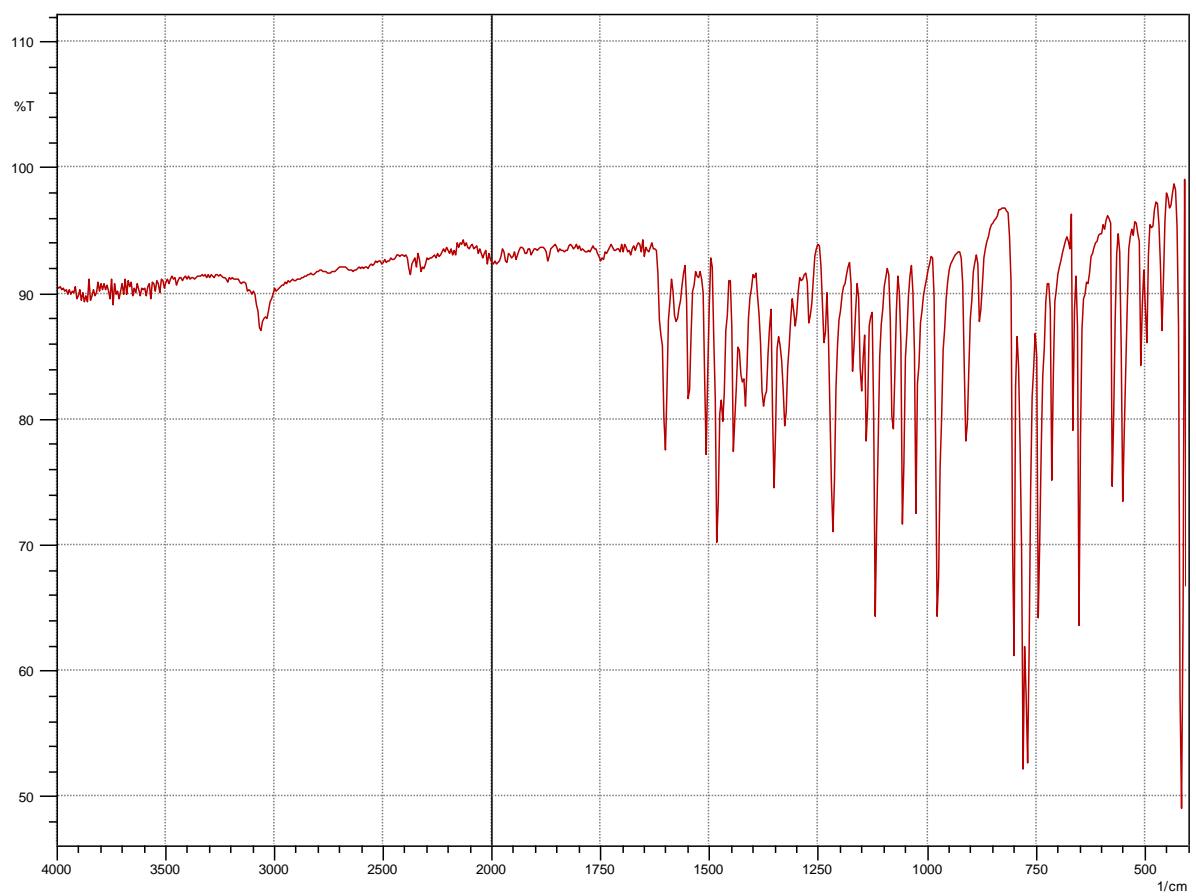
2) **Fig. S2.** ATR-FTIR spectra of Co-pqx (**2**)



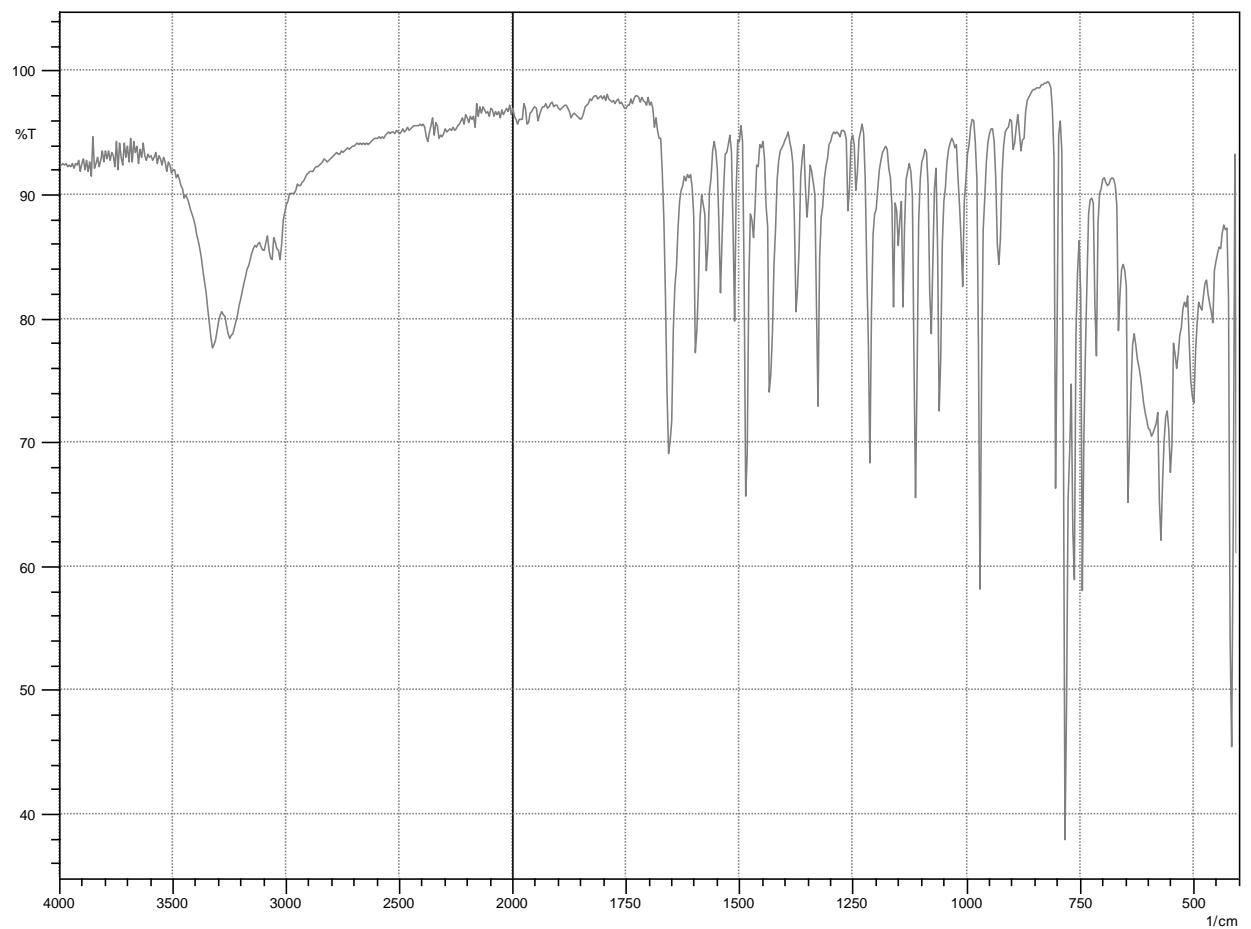
3) **Fig. S3** FTIR spectra of CuLCl₂(DMSO) (**3**) in KBr



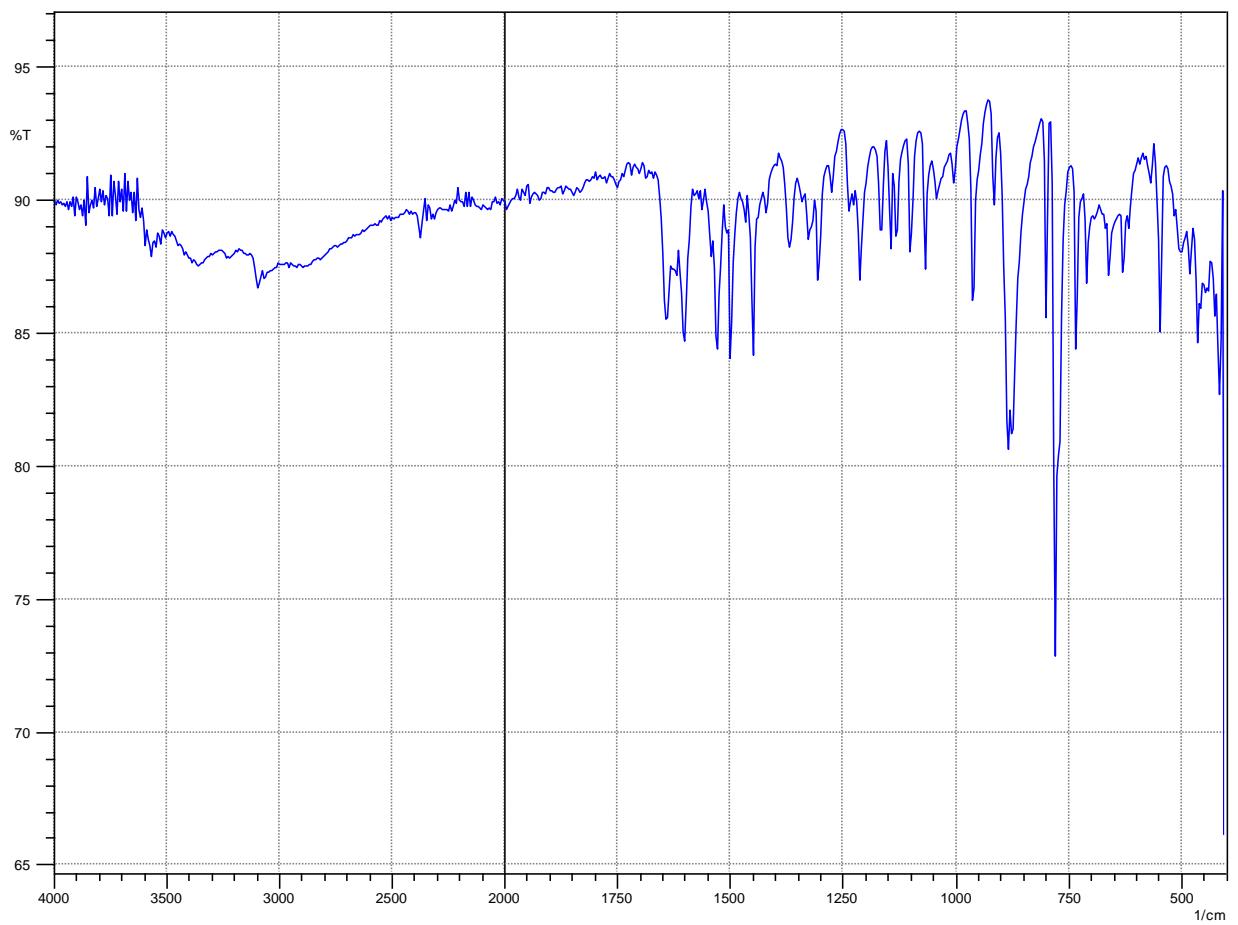
4) **Fig. S4** ATR-FTIR spectra of Zn-pqx (**4**)



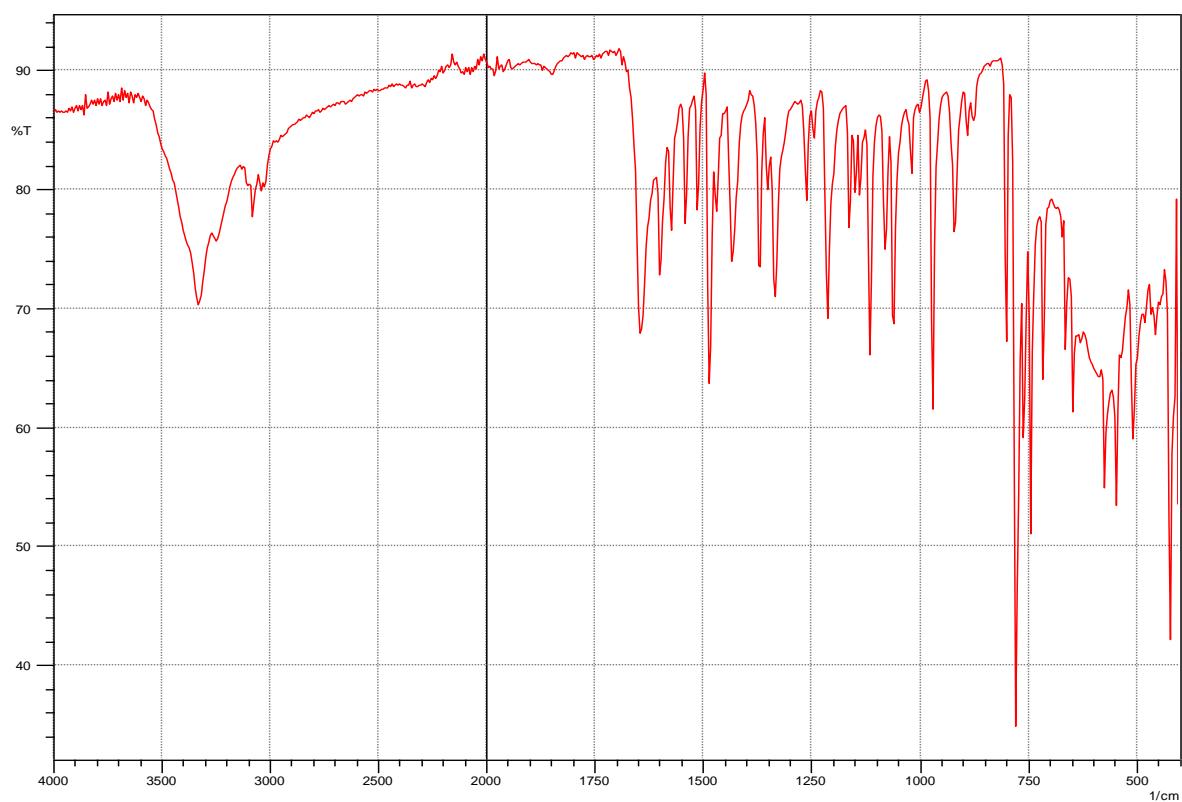
5) **Fig. S5** ATR-FTIR spectra of Mn-pqx (**5**)



6) **Fig. S6** ATR-FTIR spectra of Fe-pqx (**6**)



7) **Fig. S7** ATR-FTIR spectra of Ni-pqx (**7**)



8) **Table S1.** Bond lengths (Å) with estimated standard deviations for **3**

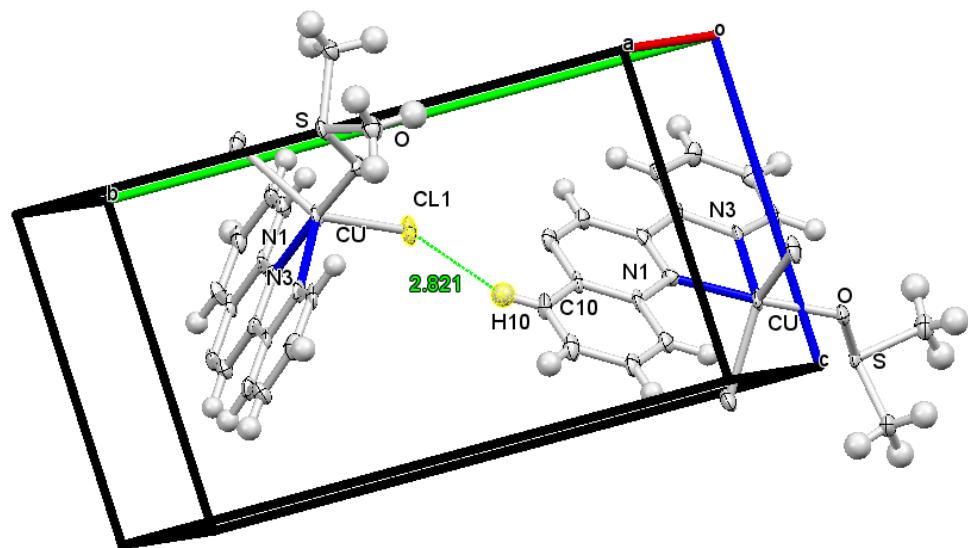
Atom	Atom	Length/ Å	Atom	Atom	Length/ Å
Cu	Cl1	2.3240(15)	N3	C9	1.333(7)
Cu	Cl2	2.3267(16)	C1	C2	1.436(8)
Cu	O	1.970(4)	C1	C5	1.460(8)
Cu	N1	2.020(5)	C3	C4	1.415(8)
Cu	N3	2.128(5)	C3	C10	1.417(8)
S	O	1.527(4)	C4	C13	1.399(9)
S	C14	1.784(6)	C5	C6	1.384(8)
S	C15	1.780(7)	C6	C7	1.382(9)
N1	C1	1.328(8)	C7	C8	1.392(8)
N1	C4	1.374(8)	C8	C9	1.375(8)
N2	C2	1.303(8)	C10	C11	1.358(9)
N2	C3	1.357(8)	C11	C12	1.405(8)
N3	C5	1.361(7)	C12	C13	1.366(9)

9) **Table S2.** Bond angles ($^{\circ}$) with estimated standard deviations for 3

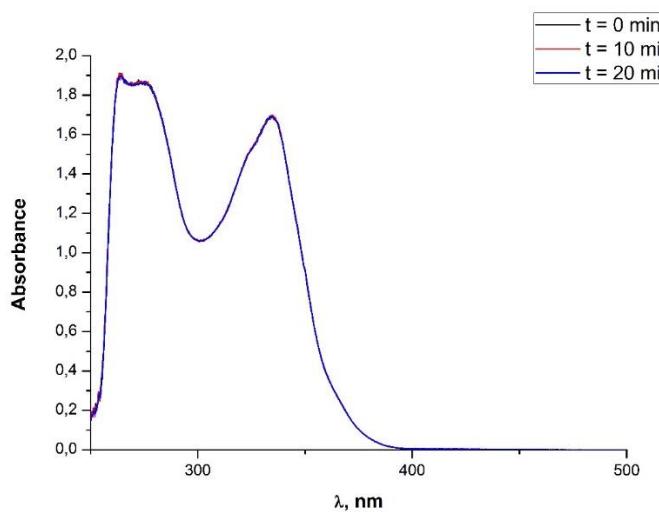
Bond angles ($^{\circ}$) Bond angles ($^{\circ}$)

N1 Cu N3	78.88(19)	N1 C4 C3	119.0(5)
N3 Cu Cl1	109.70(14)	N1 C4 C13	120.6(5)
N3 Cu Cl2	115.37(13)	C13 C4 C3	120.4(5)
O S C14	103.9(3)	N3 C5 C1	114.2(5)
O S C15	105.4(3)	N3 C5 C6	121.2(5)
C15 S C14	98.5(3)	C6 C5 C1	124.6(5)
S O Cu	121.5(2)	C7 C6 C5	119.9(6)
C1 N1 Cu	116.1(4)	C6 C7 C8	118.3(6)
C1 N1 C4	119.1(5)	C9 C8 C7	119.2(6)
C4 N1 Cu	124.8(4)	N3 C9 C8	122.8(6)
C2 N2 C3	117.4(5)	C11 C10 C3	119.8(6)
C5 N3 Cu	113.0(4)	C10 C11 C12	120.6(6)
C9 N3 Cu	128.2(4)	C13 C12 C11	121.3(6)
C9 N3 C5	118.7(5)	C13 C12 C4	119.0(6)

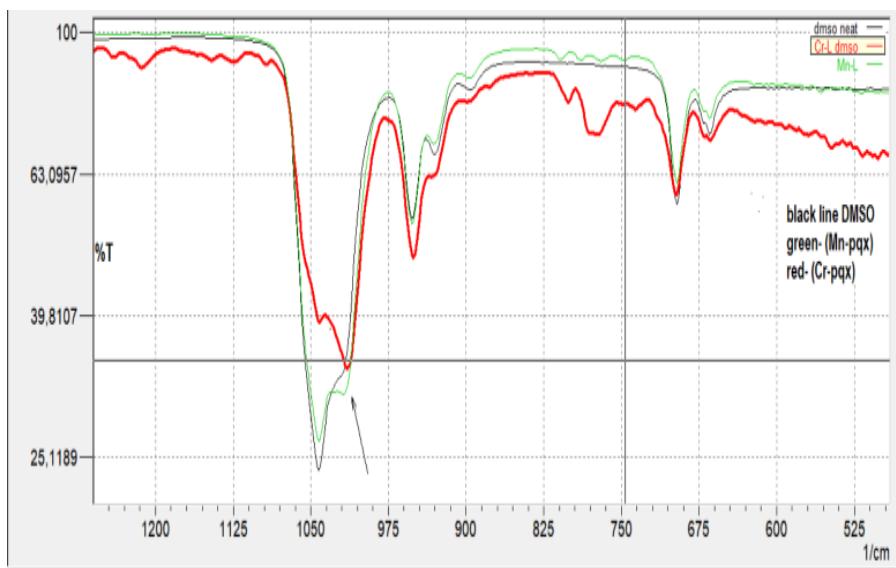
10) **Figure S8.** Hydrogen bonding interactions in complex 3



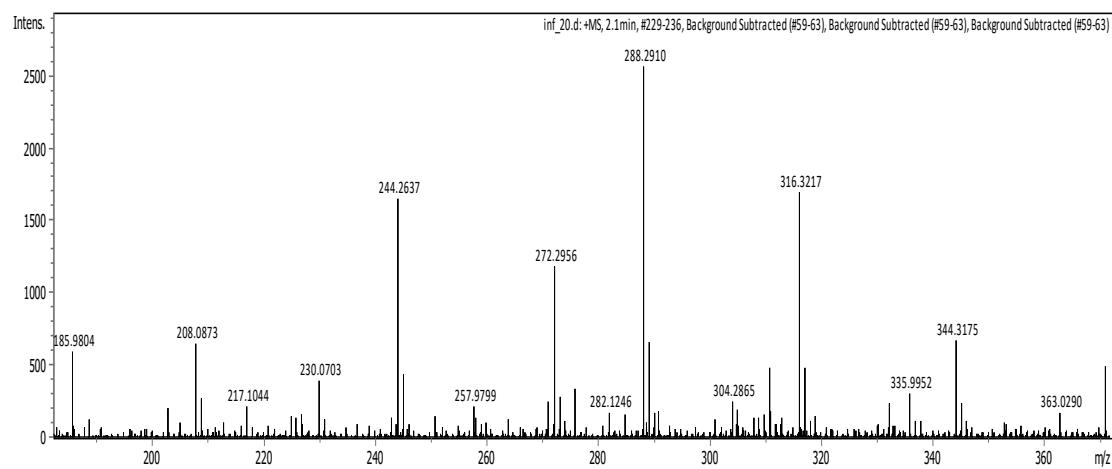
11) Figure S9. UV-Vis of complex 7



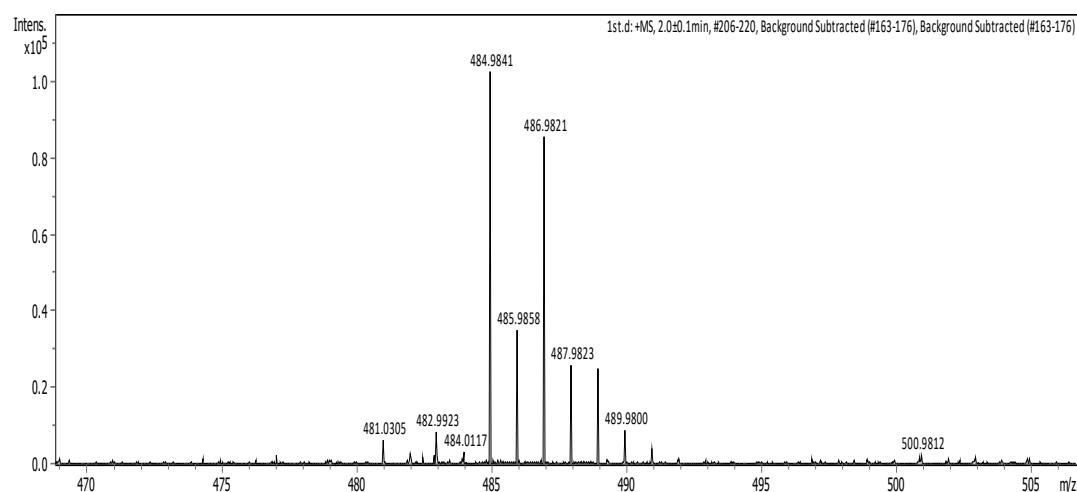
12) Figure S10. FT-IR spectra of **1** and **5** in DMSO



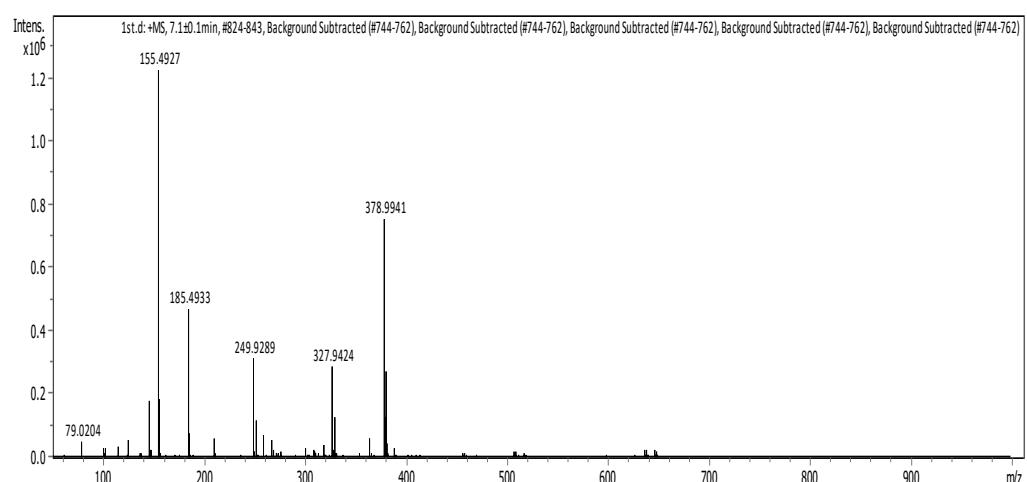
13) Figure S11. ESI-HRMS spectra of pqx in DMSO/methanol



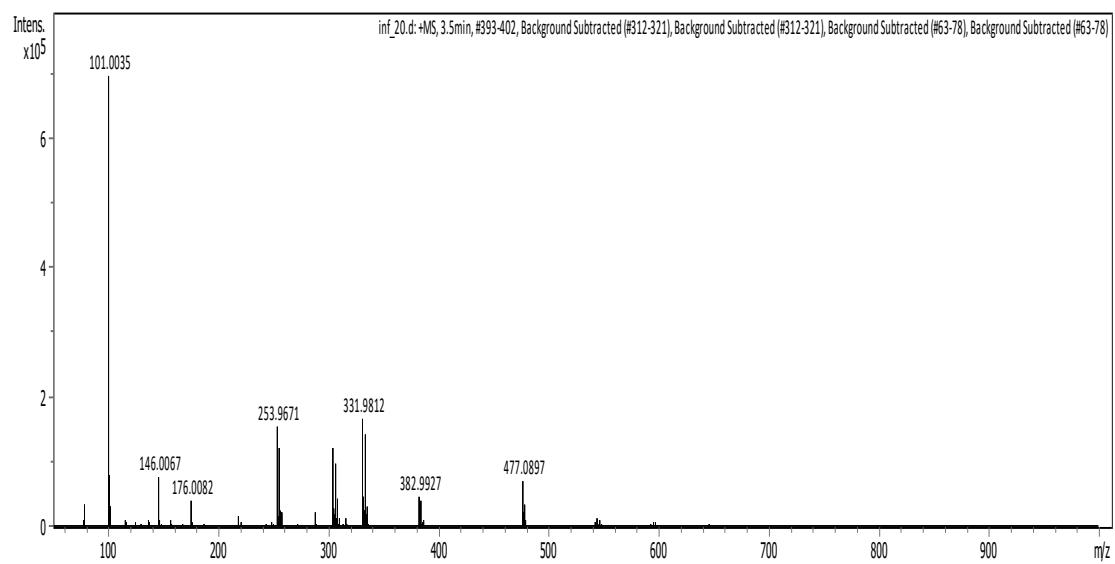
14) Figure S12. ESI-HRMS spectra of **1** in DMSO/methanol



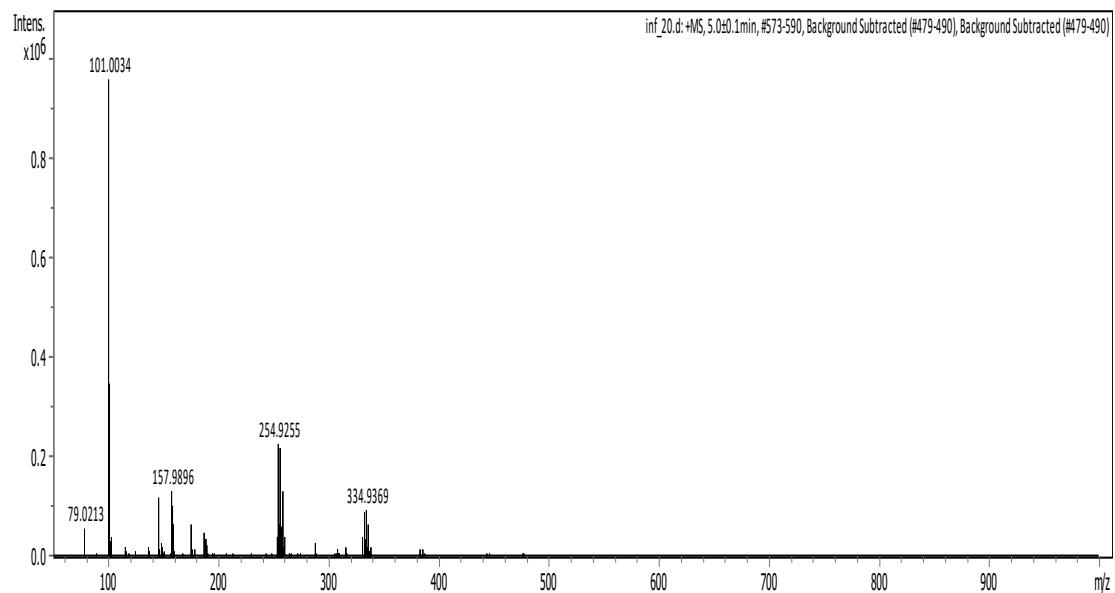
15) Figure S13. ESI-HRMS spectra of **2** in DMSO/methanol



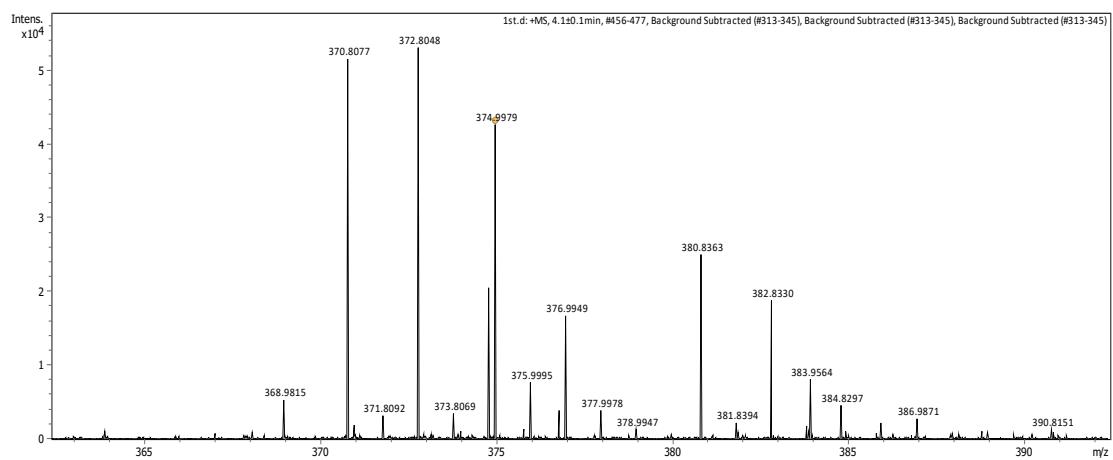
16) Figure S14. ESI-HRMS spectra of **3** in DMSO/methanol



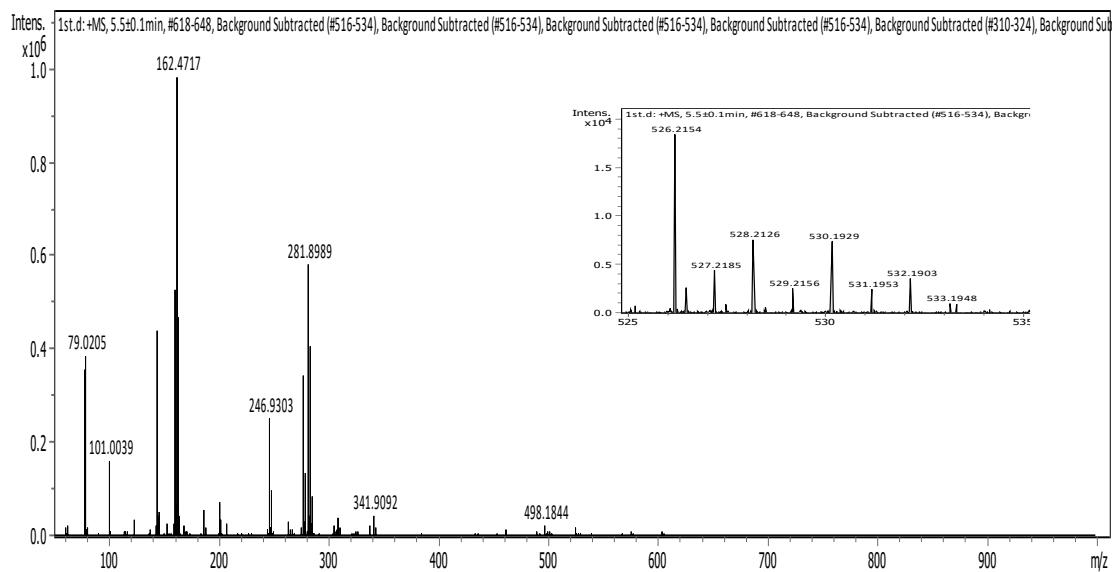
17) Figure S15. ESI-HRMS spectra of **4** in DMSO/methanol



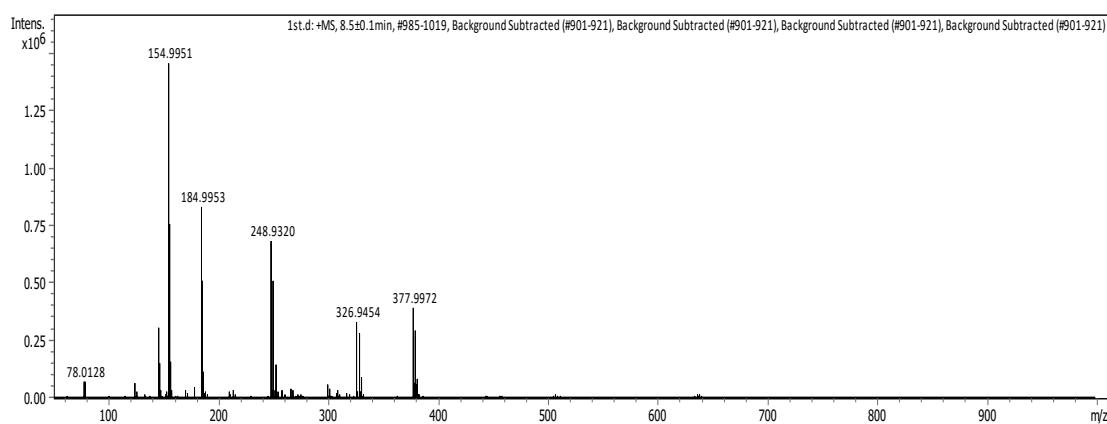
18) Figure S16. ESI-HRMS spectra of 5 in DMSO/methanol



19) Figure S17. ESI-HRMS spectra of **6** in DMSO/methanol



20) Figure S18. ESI-HRMS spectra of 7 in DMSO/methanol



21) General experimental part for ESI-HRMS

The QToF system was equipped with an electrospray ionization interface (ESI), operating in positive ionization mode, with the following operation parameters: capillary voltage 2500 V; end plate offset -500 V; nebulizer pressure 1.0 bar; drying gas 4.0 L min⁻¹; and gas temperature 180 °C. Full scan acquisition mode with a scan rate at 1Hz was employed, recording spectra over the m/z range 50-1000. A regular system maintenance protocol was followed to assure the good operation of the HRMS system, before analysis. After cleaning the source, transfer tube and cone, the sensitivity, mass accuracy and resolution were monitored and compared to installation values. Moreover, a QToF external calibration was performed using a sodium formate solution (10 mM), producing Na (NaCOOH)₁₋₁₄ clusters over the analysis m/z range. A blank solution (mixture of Methanol/DMSO, 50/50 v/v) was also infused for subtracting any background noise from the acquired spectra. The identification relied on the mass accuracy of the pseudomolecule ion and the distinctive isotopic pattern of the formed complexes, whereas further investigation of the high-resolution mass spectrometric spectra provided insights into the mechanisms that take place under dilution with DMSO.