

Supplementary Materials

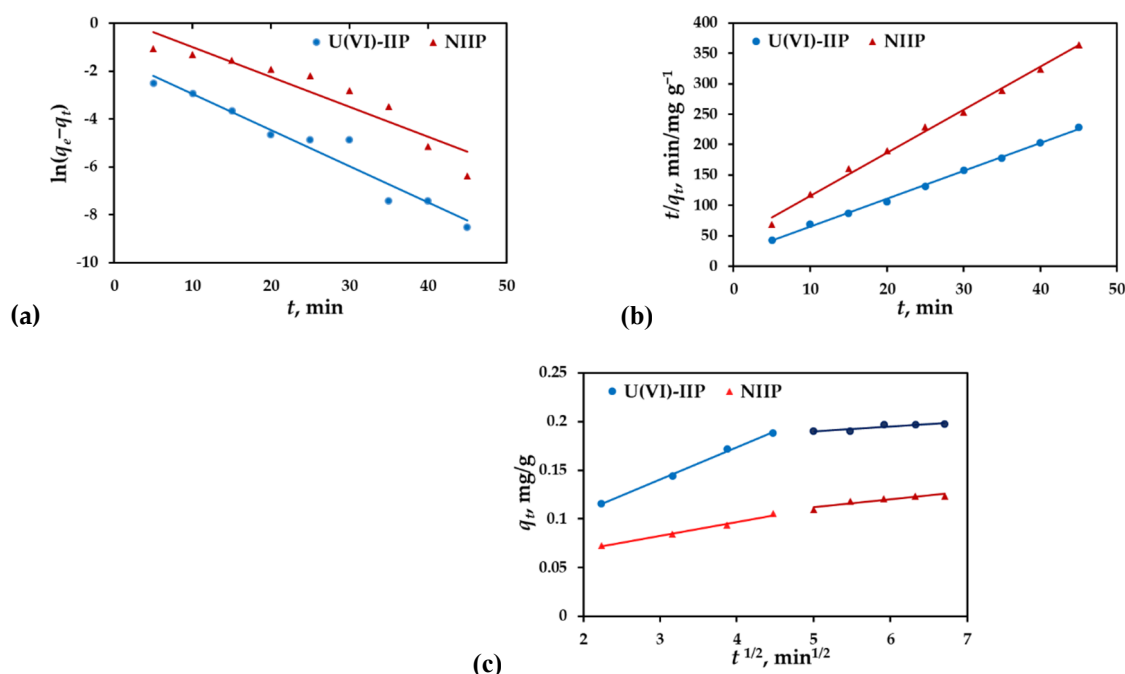
# Uranium Determination in Waters, Wine and Honey by Solid Phase Extraction with New Ion Imprinted Polymer

Valentin Georgiev, Ivanka Dakova \* and Irina Karadjova

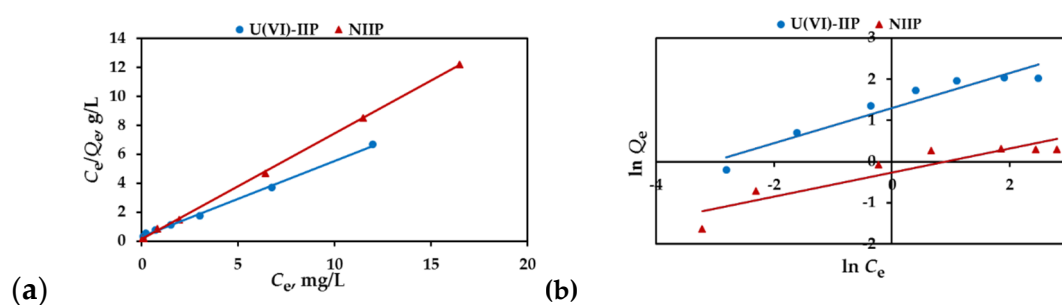
Faculty of Chemistry and Pharmacy, University of Sofia “St. Kliment Ohridski”, 1, J. Bourchier Blvd., 1164 Sofia, Bulgaria

\* Correspondence: i.dakova@chem.uni-sofia.bg

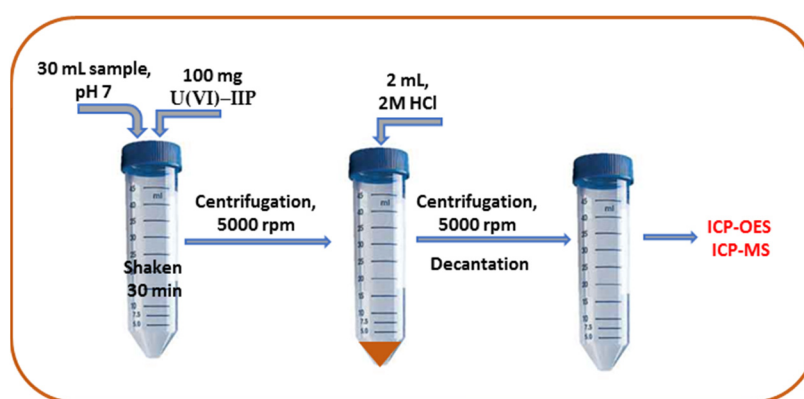
## Figures



**Figure S1.** Kinetic analysis of U(VI) adsorption on U(VI)-IIP and NIIP: the linear fitting curves of pseudo-first-order reaction (a) and pseudo-second-order reaction (b) and intra-particle diffusion model (c). (pH 7; sorbent dose = 100 mg/10 mL;  $C_0$  = 2 mg U(VI)/L, temperature 298 K).



**Figure S2.** Langmuir (a) and Freundlich (b) isotherms for adsorption of U(VI) on the U(VI)-IIP and NIIP.



**Figure S3.** Scheme of analytical procedure for U determination in surface waters using U(VI)-IIP.

## Tables

**Table S1.** Polymerization conditions for the preparation of copolymer gels (70 mg AIBN; 25 mL ACN; TMPTMA (0.96 mmol); U(VI)-PAR (0.12 mmol); T = 60 °C; 24 h) and nitrogen content, surface area ( $S_{\text{BET}}$ ), total pore volume ( $V_{\text{total}}$ ), average pore diameter ( $D_{\text{average}}$ ) and adsorption capacities of U(VI)-IIPs.

Copolymer gel	MAA (mmol)	N content (% wt.)	$S_{\text{BET}}$ (m <sup>2</sup> /g)	$V_{\text{total}}$ (cm <sup>3</sup> /g)	$D_{\text{average}}$ (nm)	Capacity (mg/g)
U(VI)-PAR-1	0.58	2.25	7.0	0.04	24	0.96
U(VI)-PAR-2	1.16	2.63	6.2	0.06	20	1.29
U(VI)-PAR-3	1.80	2.85	6.4	0.02	20	1.55
U(VI)-PAR-4	2.16	3.10	6.5	0.05	22	1.89
U(VI)-PAR-5	2.50	2.96	6.5	0.04	24	1.80

**Table S2.** Degree of elution ( $D_{\text{E}}$ , %) for U(VI) from U(VI)-IIP using different eluents (10 mL).

Desorption agent	Concentration	$D_{\text{E}}$ , %
HCl	0.5 mol/L	48±4
	1 mol/L	83±3
	2 mol/L	96±2
	3 mol/L	99±2

**Table S3.** Parallel analysis of tap water by the proposed analytical method and direct Alfa spectrometry.

U content, Bq/L	Tap water 1		Tap water 2	
	Proposed analytical method*	Alpha spectrometry	Proposed analytical method	Alpha spectrometry
<sup>234</sup> U	0.29±0.02	0.27±0.03	0.48±0.03	0.50±0.04
<sup>238</sup> U	0.28±0.03	0.27±0.02	0.47±0.04	0.46±0.03

\*—in this case ICP-MS was used as instrumental method.

**Table S4.** Comparative analysis by the proposed method and standard method based on microwave digestion of wine/honey sample and ICP-MS measurement.

Sample	U content	
	Proposed analytical method, µg/L	ICP-MS after digestion, µg/L
Red wine (Cabernet)	0.74±0.04	0.77±0.02
White wine (Muskat)	0.43±0.03	0.41±0.02
Honey (sunflower)	1.43±0.12	1.51±0.08
Honey (lime)	3.52±0.21	3.58±0.09

**Table S5.** Comparison of different U(VI) ion imprinted polymer sorbents.

Chelating agent/Matrix	Capacity, mg/g	Instrumental method	LOQ, µg/L	Ref.
2,4-dioxopentan-3-yl methacrylate – ethylene glycol dimethacrylate	15.3	ICP-OES		[29]
oleic acid coated magnetic particles	1.06	ICP-OES		[50]
1-hydroxy-2-(prop-2'-enyl)-9,10-anthraquinone – ethylene glycol dimethacrylate	12.4	UV-VIS		[27]
Salicylaldoxime / magnetic IIP	1.2	ICP-OES	-	[49]
Salicylaldoxime / γ-methacryloxypropyltrimethoxysilane modified magnetic nanoparticles – MAA – ethylene glycol dimethacrylate	5.4	ICP-OES		[38]
Salicylaldoxime / 4-vinylpyridine – MAA – ethylene glycol dimethacrylate	15.3	UV-VIS	5	[25]
5,7-dichloroquinoline-8-ol / 4-vinylpyridine – styrene – divinyl benzene	34	UV-VIS	2	[22]
N-hydroxyethylacrylamide and 1-vinylimidazole@ magnetic microspheres	146	ICP-OES	0.8 mg/L	[39]
IIP-grafted silica	-	ICP-OES	0.09 (LOD)	[34]
4-(2-Pyridylazo)resorcinol / MAA – TMPTMA	1.89	ICP-OES ICP-MS	0.15 0.003	This work