

Supplementary Materials

Flame-Made Calcium Phosphate Nanoparticles with High Drug Loading for Delivery of Biologics

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Table S1. Calculation of loading capacity of mesoporous SiO₂ nanoparticles in mg LL-37/g particle according to data presented in Braun *et al.* [1].

Particles	Diameter (nm)	Radius (nm)	Particle Volume (m ³)	Mass of a single particle (g)	Number of Particles in 5 mg
NSN	307.9	153.95	1.53·10 ⁻²⁰	3.36·10 ⁻¹⁴	1.49·10 ¹¹
MSNc	294.6	147.3	1.34·10 ⁻²⁰	2.94·10 ⁻¹⁴	1.69·10 ¹¹

Particles	Adsorption (μmol LL-37/particle) [1]	μmol LL-37 in 5 mg	mg LL-37 in 5 mg (MW _{LL-37} =4493.3 g/mol ^{**})	mg LL-37 / g particle
NSN	1.8·10 ⁻¹³	0.0268	0.120	24.06
MSNc	8.5·10 ⁻¹³	0.144	0.649	129.74

* Assuming density of SiO₂ 2.2 g/cm³ [2].

**According to the provider.

Table S2. LL-37 release at pH 7.4 after 2 h, 6 h and 24 h at 25°C and 37°C. After 24 h, there is 1.1 % and 1.2 % of LL-37 released at 25°C and 37°C, respectively.

Temperature (°C)	Time (h)	% LL-37 released
25	2	0.3
	6	0.6
	24	1.1
37	2	0.5
	6	0.8
	24	1.2

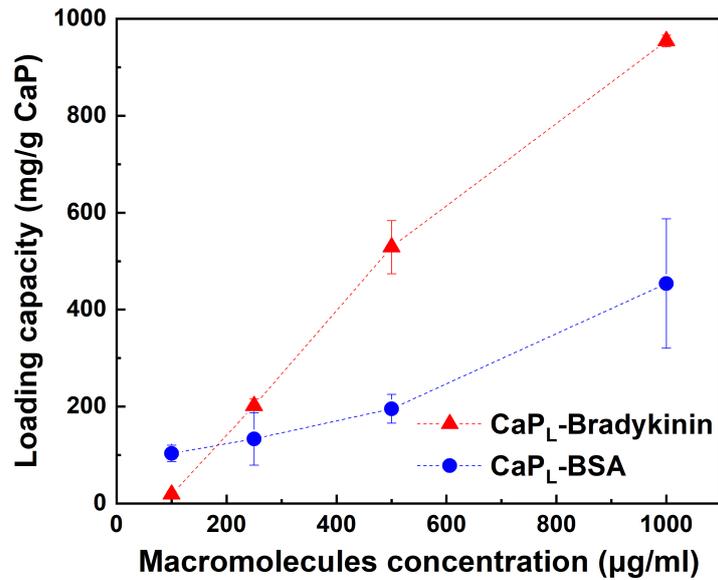


Figure S1. Effect of concentration of BSA and Bradykinin on the loading capacity of CaP_L nanoparticles after incubation for 6 h at room temperature (PBS pH 7.4, particle concentration 500 µg/ml). Data are reported as mean ± standard deviation, for at least 3 independent triplicates.

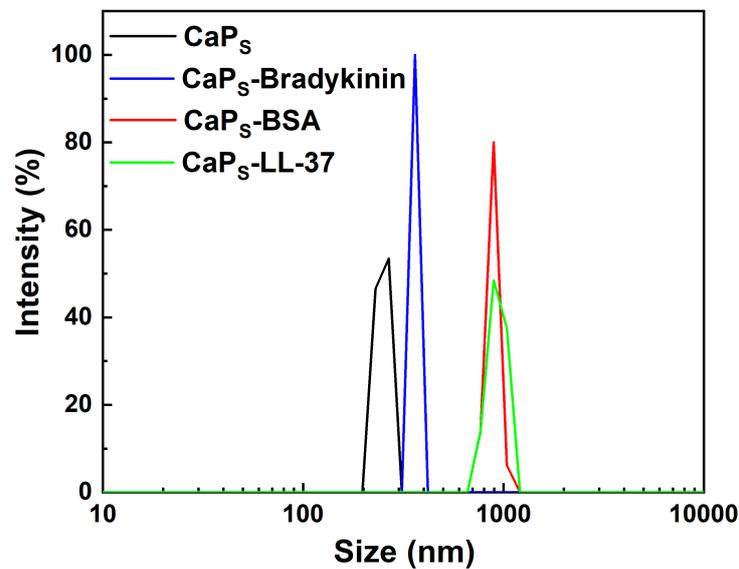


Figure S2. Size distribution of CaP_s nanoparticles (intensity % data) before and after loading with Bradykinin, BSA and LL-37 in PBS pH 7.4, as determined by DLS measurements (particle concentration 100 µg/ml).

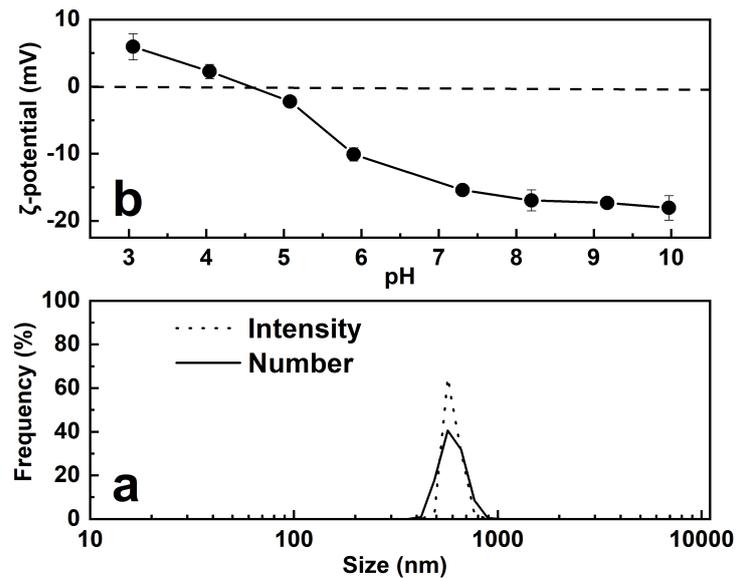


Figure S3. (a) Size distribution (both number and intensity data are presented) of CaPI nanoparticles in PBS pH 7.4 as determined by DLS measurements (particle concentration 100 $\mu\text{g/ml}$); and (b) ζ -potential profile of CaPI nanoparticles as determined by titration at different pH (particle concentration 100 $\mu\text{g/ml}$).

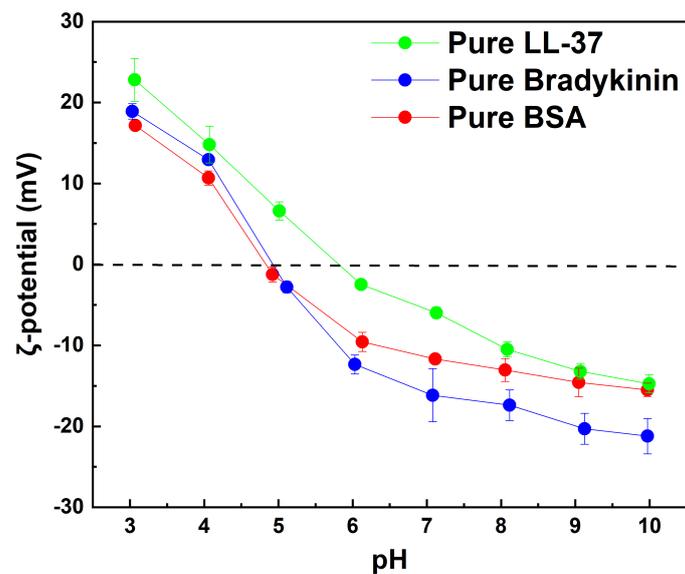


Figure S4. ζ -potential profile of pure BSA, bradykinin and LL-37 in PBS pH 7.4 as determined by titration at different pH (macromolecules concentration $\sim 100 \mu\text{g/ml}$). BSA and bradykinin have similar isoelectric points (~ 5), whereas the isoelectric point of LL-37 is ~ 6 .

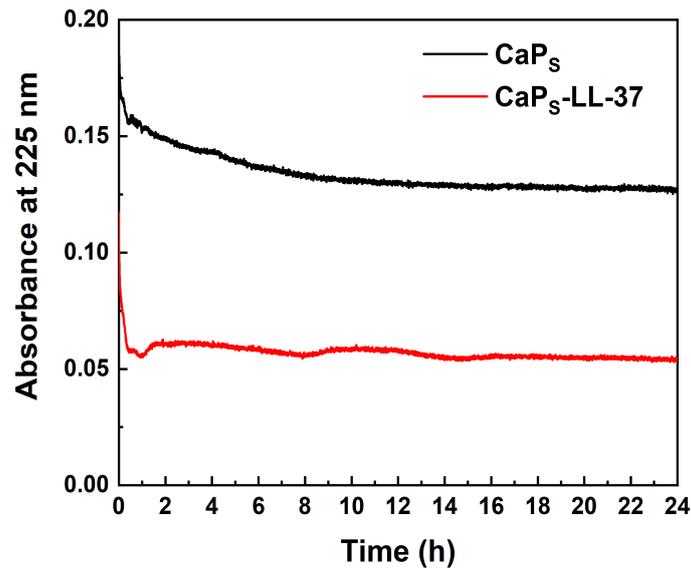


Figure S5. Absorbance at $\lambda = 225$ nm as a function of time for CaPs and CaPs-LL-37 nanocarriers of initial particle concentration $100 \mu\text{g/ml}$ in PBS. The light beam was aligned to monitor the absorbance of the top suspension layer. CaPs absorbance is stabilized after ~ 8 h whereas CaPs-LL-37 nanoparticles sedimented much rapidly (after ~ 2 h).

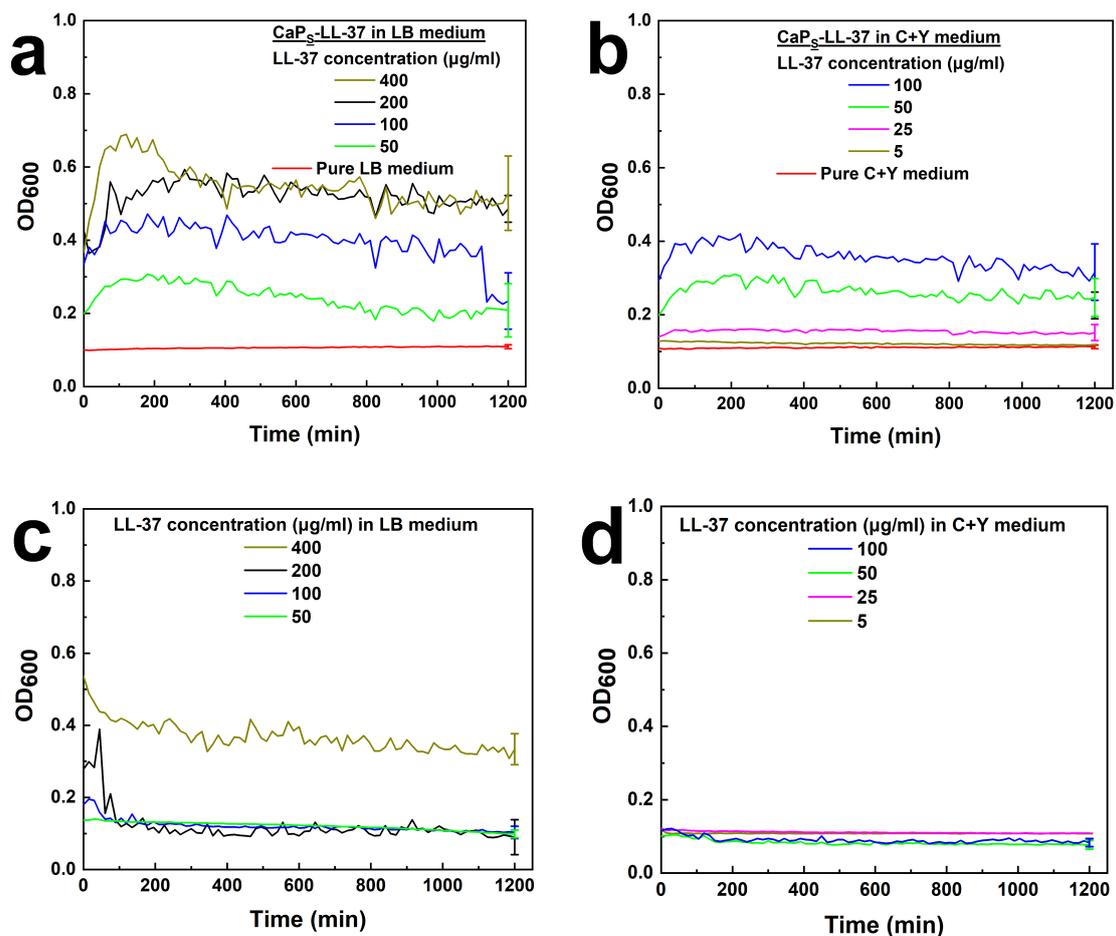


Figure S6. Absorbance values of LL-37-loaded CaPs nanoparticles in (a) LB medium and; (b) C+Y medium along with absorbance of pure media as measured in the bioscreen instrument at 600 nm; Absorbance values at 600 nm of pure LL-37 in LB (c) and C+Y (d) media. These values represent

background values that were used for the correction of the growth curves (Figure 7 of the main paper). Measurements had been performed in triplicate and mean values are presented with representative error bars.

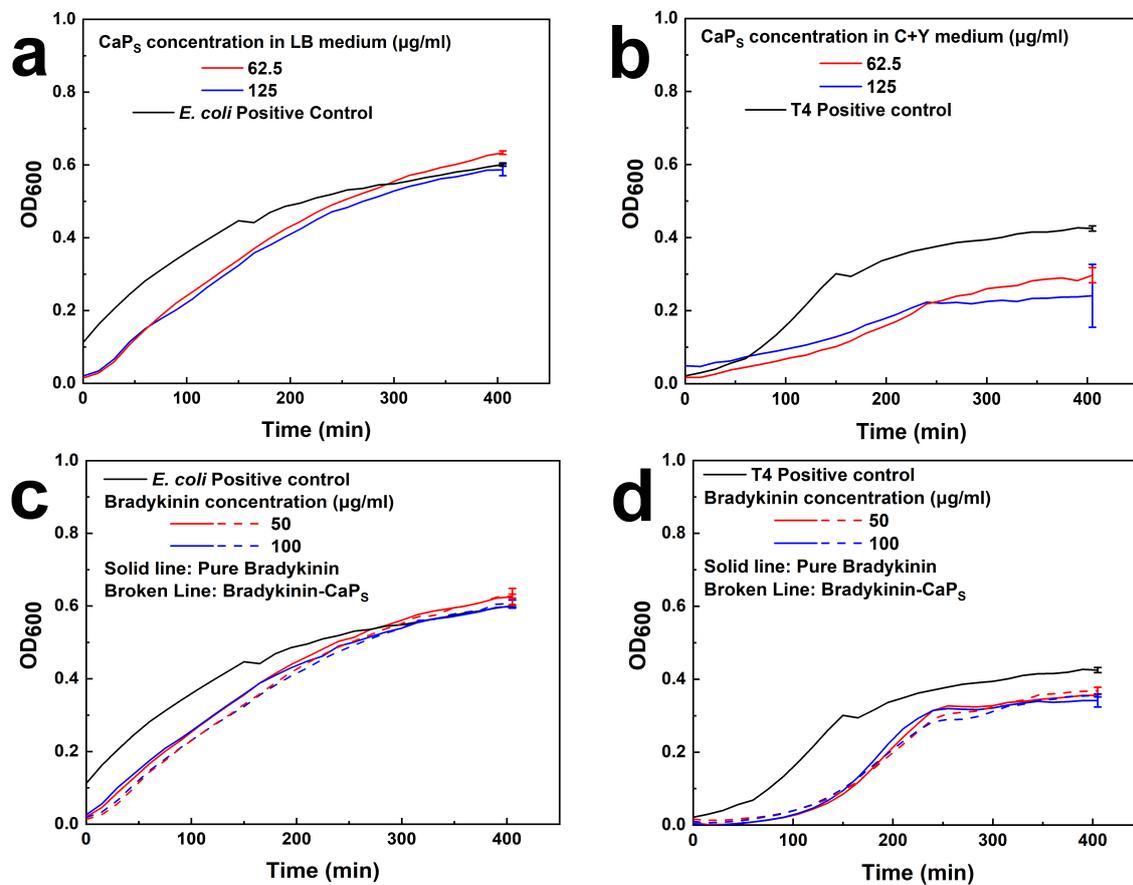


Figure S7. Effect of CaP_s nanoparticle presence on (a) *E. coli* and (b) *S. pneumoniae* growth after subtraction of media absorbance (Figure S6a and b). Nanoparticle dose is calculated taken into consideration that LL-37 loading is ~800 mg/g particle. Thus, for 100 and 50 µg/ml LL-37 concentration, particle concentration is 125 and 62.5 µg/ml, respectively; *E. coli* (c) and *S. pneumoniae* (d) growth in the presence of pure bradykinin and bradykinin-loaded CaP_s nanoparticles. Bradykinin was used as a control peptide in order to confirm that the observed antibacterial activity of the LL-37-loaded CaP_s nanoparticles would be attributed to the presence of LL-37 and not to the nanoparticles. The observed antibacterial activity (Figure 7) is attributed to the LL-37 peptide and not to the CaP_s nanoparticles. Measurements had been performed in triplicate and mean values are presented with representative error bars.

References

1. Braun, K.; Pochert, A.; Lindén, M.; Davoudi, M.; Schmidtchen, A.; Nordström, R.; Malmsten, M. Membrane interactions of mesoporous silica nanoparticles as carriers of antimicrobial peptides. *J. Colloid Interface Sci.* **2016**, *475*, 161–170.
2. Liu, J.; Zong, G.; He, L.; Zhang, Y.; Liu, C.; Wang, L. Effects of Fumed and Mesoporous Silica Nanoparticles on the Properties of Sylgard 184 Polydimethylsiloxane. *Micromachines* **2015**, *6*, 855–864.