Supplementary Information

Porous Silk Fibroin/Cellulose Hydrogels for Bone Tissue Engineering via a Novel Combined Process Based on Sequential Regeneration and Porogen Leaching

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Figure S1: Stress-strain curves of all samples and control. Number of samples measured: S5C1.0 n=5; S5C0.5 n=5; S5C0.3 n=6; S5C0 n=4.





Figure S2: Diagrams of deconvoluted peaks in amide I band region. Bands for peak allocation to secondary structure are: β -sheet: 1611 cm⁻¹, 1619 cm⁻¹, 1626 cm⁻¹, 1699 cm⁻¹. α -helix/random coil: 1631 cm⁻¹, 1641 cm⁻¹, 1649 cm⁻¹. Turns and bends: 1659 cm⁻¹, 1668 cm⁻¹, 1674 cm⁻¹, 1682 cm⁻¹.



SF matrix /cellulose scaffold composite structure

Figure S3: Cellulose framework structure embedded in a SF matrix. (A) Cellulose framework: By omitting methanol treatment of samples, a cellulose-rich, very stable hydrogel formed ("Scaffold" spectrum); that sample was compared with regenerated cellulose, degummed silk fibroin and lyophilized SF/cellulose hydrogel by ATR FTIR. (B) SF matrix: ATR and transmission spectra (KBr) normalized to the cellulose-specific band area 1100 cm⁻¹ to 960 cm⁻¹ of a lyophilized hydrogel. The larger amide band area in ATR mode, indicates a relatively increased SF content in the surface-near region compared to the bulk.



Figure S4: Different bulk and surface composition. Ratios of silk fibroin (SF) specific amide I and II band area and cellulose-specific band area at 1100 cm⁻¹ to 960 cm⁻¹ (respectively area 1 and 2 in A and B) of all samples decrease with increasing cellulose content in transmission mode (KBr) (C). In ATR mode ratios of samples do not differ, but show a lower ratio compared to the SF control. This indicates a richer SF structure in the surface regions of the hydrogel compared to bulk composition.